

# GOPEN ACCESS

**Citation:** Améndola L, Ratuski A, Weary DM (2021) Individual differences in rat sensitivity to CO<sub>2</sub>. PLoS ONE 16(1): e0245347. https://doi.org/10.1371/ journal.pone.0245347

Editor: Satoshi Ikemoto, National Institute on Drug Abuse, UNITED STATES

Received: June 14, 2020

Accepted: December 28, 2020

Published: January 22, 2021

**Copyright:** © 2021 Améndola et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its <u>Supporting</u> Information.

**Funding:** This study was funded by a Discovery grant (RGPIN-2016-04620) from Canada's Natural Science and Engineering Research Council to D.M. W. In addition, L.A. was supported by a CONACyT PhD scholarship (no. 381124) and the UBC Charles River Scholarship in Animal Welfare. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

# RESEARCH ARTICLE

# Individual differences in rat sensitivity to CO<sub>2</sub>

# Lucía Améndola, Anna Ratuski, Daniel M. Weary \*

Animal Welfare Program, University of British Columbia, Vancouver, British Columbia, Canada

\* dan.weary@ubc.ca

# Abstract

Feelings of fear, anxiety, dyspnea and panic when inhaling carbon dioxide  $(CO_2)$  are variable among humans, in part due to differences in CO<sub>2</sub> sensitivity. Rat aversion to CO<sub>2</sub> consistently varies between individuals; this variation in aversion may reflect CO<sub>2</sub> sensitivity, but other personality traits could also account for individual differences in aversion. The aims of this study were to 1) assess the stability of individual differences in rat aversion to CO<sub>2</sub>, 2) determine if individual differences in sweet reward motivation are associated with variation in aversion to CO<sub>2</sub>, and 3) assess whether variation in aversion to CO<sub>2</sub> is related to individual differences in motivation to approach gains (promotion focus) or maintain safety (prevention focus). Twelve female Sprague Dawley rats were exposed multiple times at three different ages (3, 9 and 16 months old) to  $CO_2$  in approach-avoidance testing to assess motivation to avoid CO<sub>2</sub> against motivation to gain sweet rewards. Rats were also tested for motivation to find hidden sweet rewards, and for their motivation to approach rewards or darkness. Tolerance to CO<sub>2</sub> increased with repeated exposures and was higher at older ages. Individual differences in aversion to CO2 were highly repeatable but unrelated to motivation for sweet rewards or the strength of promotion and prevention focus. These results indicate that individual differences in aversion to CO<sub>2</sub> reflect variation in CO<sub>2</sub> sensitivity.

# Introduction

People report feelings of fear, anxiety, dyspnea and panic during  $CO_2$  inhalation (for a review see [1]). This emotional response to  $CO_2$  inhalation varies among individuals. With a single inhalation of 35%  $CO_2$ , around 24% of healthy humans experience panic attacks [2, 3]. Between 43 to 94% of panic disorder patients experience PAs after a single inhalation of 35%  $CO_2$  [1]. When inhaling 35%  $CO_2$ , the anxiety experienced by healthy people and the panic attacks experienced by panic disorder patients are highly consistent between exposures [4, 5]. This between subject variability in the subjective emotional experience is often referred to as variation in  $CO_2$  sensitivity [1].

Rats respond to  $CO_2$  exposure with defence behaviours [6–9], and are motivated to avoid this agent [9–11]. Rats have also been used as translational models for understanding the underlying mechanisms of the emotional response to  $CO_2$  inhalation [12, 13]. Rat behavioural responses to  $CO_2$  are highly variable. For example, during forced (i.e. unavoidable) exposure to  $CO_2$  "escape attempts" have ranged between 0 to 34 among rats [7], with 50% of rats **Competing interests:** The authors have declared that no competing interests exist.

showing an increase in locomotion [9], and 20% of rats moving around the cage perimeter [14]. Aversion to  $CO_2$  is also variable among rats. For example, in one study the latency to avoid  $CO_2$  varied between individuals from 7 s to 48 s in an aversion-avoidance setting (in which the cost of avoiding the CO<sub>2</sub> delivered in a dark compartment was escaping to a CO<sub>2</sub>free compartment that was brightly lit) [10]. In an approach-avoidance setting (in which the cost of escaping to a  $CO_2$ -free compartment was the loss of sweet rewards), the threshold of aversion ranged between 11 to 19% CO<sub>2</sub> between rats [9, 11]. In more recent work we found that variation in rat behaviour was consistent between two exposures to  $CO_2$ , during forced exposure, aversion- and approach-avoidance testing and we found that rats that consistently showed higher responses to CO<sub>2</sub> forced exposure were consistently less tolerant of CO<sub>2</sub> when tested in aversion-avoidance [15]. These results suggest that variation in rat responses to CO<sub>2</sub> is linked to consistent individual differences in CO2 sensitivity; i.e. like humans, rats may vary in the emotional experience elicited by  $CO_2$  (for a review, see [16]). The first aim of the current study was to determine whether individual differences in rat aversion to  $CO_2$  in approachavoidance tests assessed at 3, 9 and 16 months of age, are stable and consistent through multiple exposures.

Individual differences in aversion to  $CO_2$  could be caused by behavioural differences elicited by testing contingencies specific to the approach-avoidance setting. In approach-avoidance tests, exposure to  $CO_2$  is paired with access to sweet rewards that rats are motivated to approach [11]. An underlying but untested assumption is that the strength of motivation to approach the sweet rewards is similar among rats. However, motivation for sucrose is known to consistently vary among rats [17–19], so it is possible that variation in rat aversion to  $CO_2$ in approach-avoidance tests is due to individual variability in motivation for sweet rewards. Thus, the second aim of our study was to assess if individual differences in rat responses to  $CO_2$  in an approach-avoidance test are associated with variation in sweet reward motivation.

Following regulatory focus theory [20], variation in rat behaviour in approach-avoidance could be related to individual differences in the strength of promotion and prevention motivations. Individuals focused on promotion are more motivated to approach gains and are more sensitive to their presence or absence; individuals focused on prevention are more motivated and sensitive to safety related incentives [21]. Promotion and prevention motivations are independent; hence individuals can be high or low in promotion or prevention motivations, or both [22]. These motivational foci are consistent over time in rats [22, 23]. Approach-avoidance tests involve both gain (sweet rewards) and safety ( $CO_2$ -free cage) incentives. Individual differences in promotion and prevention motivations could account for variation in  $CO_2$  thresholds of aversion. Promotion focused individuals may be maintaining safety by avoiding non-threatening  $CO_2$  concentrations. Thus, the final aim of this study was to determine if individual differences in regulatory focus are related to variation in  $CO_2$  aversion in approach-avoid-avo

# Methodology

All procedures were performed in accordance with the guidelines on care and use of rodents in research established by the Canadian Council on Animal Care and were approved by The University of British Columbia Animal Care Committee (protocol A15-0071).

#### Subjects and housing

In a previous study by our group, individual differences in rat responses to  $CO_2$  were detected with a sample size of 12 rats [15]; therefore, 12 Sprague-Dawley rats were used in this study.

Rats were obtained as surplus stock from the University of British Columbia as an effort to reduce the total number of animals used; however, only female rats were available at the beginning of the current study. Rats were 3 months old and weighed  $328 \pm 45$  g (mean  $\pm$  standard deviation) at the beginning of the study. All subjects were marked with an animal marker (Ketchum Manufacturing Inc., ON, Canada) and housed in groups of three on a 12 h light/ dark cycle at controlled temperature and humidity ( $21 \pm 0.4$ °C and  $52 \pm 11$ %, respectively). The housing system consisted of two cages (20 cm x 50 cm x 40 cm) connected by a red polycarbonate tube (7.6 cm diameter, 15.0 cm long). Both cages contained bedding (Biofresh, Absorption Corp, WA, USA) and environmental enrichment (e.g. cardboard boxes, hammocks, PVC pipes, and shredded paper towels). Rats were provided ad libitum food (Rat Diet PMI 5012, Lab Diets, Land O'Lakes, Inc., MN, USA) and tap water. Rats were also provided daily access to a playpen (i.e. a highly enriched large cage; following [24]) for 30 min/d.

**Handling and transport.** Rats were habituated to handling during the week before experiments started (following [24]). For all sessions, rats were individually transported in a cage covered with black plastic. All habituation, training and test trials were conducted during the light cycle between 900 h and 1700 h. Each rat was habituated, trained, or tested only once per day at similar hours each day across all tests, and only separated from cage-mates for a maximum of 40 min per day. Test cages and the apparatus were cleaned with a combination of water and isopropanol (70%), and bedding was replaced before the next session.

#### Experiment 1: Aversion to CO<sub>2</sub>

**Apparatus.** An approach-avoidance apparatus was used to assess repeatability in aversion to  $CO_2$  through multiple exposures. The apparatus consisted of a larger top cage (one of the rat home cages) positioned 20 cm above a smaller bottom cage (Fig 6A). Both cages contained bedding. The top cage was connected to the bottom cage via a transparent acrylic tube fitted with plastic cleats to improve traction. A plastic sliding door was attached to the entrance of the connecting tube at the top cage of the tube. The top cage was covered with a wire lid, and the bottom cage was covered with a clear acrylic lid. To allow for comparability between studies, gases were delivered from an inlet in the acrylic lid [9, 11, 15, 24]. The lid also contained two scavenging outlets inlet (Fig 1A); the scavenging outlets were connected to a wall exhaust system by a plastic scavenger hose.

Two flow meters ( $CO_2$ : Western Medica, OH, USA; air: Dwyer instruments, Inc., NI, USA) were used to regulate gas flow. Gases (air and  $CO_2$ ) were delivered from compressed gas cylinders (Praxair, BC, Canada), through a clear vinyl tube inserted in the gas inlet.

**Habituation, training and testing procedures.** Subjects were trained to eat 20 sweet rewards (Cheerio; Honey Nut Cheerios TM, General Mills Inc., MN, USA) in the bottom cage of the apparatus. At the beginning of each training session, the subject was placed in the top cage and allowed to explore the apparatus for 5 min, with air (4 L min<sup>-1</sup>) flowing into the bottom cage at all times. The experimenter then tapped her fingers on the side of the cage, gave the rat one sweet reward in the top cage, and closed the sliding door to block access to the bottom cage. The sliding door remained closed for 60 s while 20 sweet rewards were placed in a dish in the bottom cage. The sliding door was opened and the rat was able to descend into the bottom cage to consume the sweet rewards. The training session ended as soon as the rat left the bottom cage (i.e. shoulders crossed into the tube exiting the bottom cage). A rat was considered to have met the training criterion when, during three consecutive training trials, it stayed in the bottom cage for 5 min or ate all 20 sweet rewards, whichever occurred first.

Once trained, rats were repeatedly exposed to  $CO_2$  gradual-fill (3 months of age: 26%  $CO_2$  cage vol. min<sup>-1</sup>; 9 and 16 months of age: 20%  $CO_2$  cage vol. min<sup>-1</sup>) in the approach-avoidance

a. Approach-avoidance apparatus



Fig 1. Experimental apparatus. a) Approach-avoidance apparatus used to assess aversion to  $CO_2$ . Measurements were: The top cage 20 cm x 50 cm x 40 cm, bottom cage 20 cm x 45 cm x 24 cm, connecting tube 10 cm diameter x 45 cm long, and plastic sliding door 10 cm x 10 cm. b) Modified approach-avoidance apparatus used to evaluate motivation for sweet rewards, the test cage measured 20 cm x 45 cm x 24 cm and the ice cube trays 32 cm x 12 cm x 4 cm. c) Modified open field arena used to assess promotion and prevention motivation focus, the arena was made of white acrylic glass (100 cm x 100 cm x 61 cm) and contained two smaller acrylic glass boxes (10 cm3) placed against the center of two adjacent walls of the arena (treat and dark locations).

https://doi.org/10.1371/journal.pone.0245347.g001

apparatus. During  $CO_2$  trials, air flow was substituted with  $CO_2$  flow as soon as the rat started eating the sweet rewards. We ran one control trial (air flow of 4 L min<sup>-1</sup>) after every two  $CO_2$  exposures. The trial ended when the rat left the bottom cage. Latency (s) to exit the bottom cage was recorded by direct observations. If a rat failed to stay for 5 min or eat all 20 sweet rewards in a control trial, the previous  $CO_2$  trial was excluded and the rat was re-trained before continuing in  $CO_2$  trials. Rats were re-trained until the training criterion was met at 9 and 16 months of age (for order and length of the experiments, see Fig 2).

Assessment of CO<sub>2</sub> concentrations. With no animal present in the approach-avoidance apparatus, we conducted twelve CO<sub>2</sub> flow trials for each of the two flowrates used (26% CO<sub>2</sub> chamber vol. min<sup>-1</sup> and 20% CO<sub>2</sub> chamber vol. min<sup>-1</sup>) to assess changes in CO<sub>2</sub> concentration during gradual-fill. A clear plastic sampling tube, connected to an oxygen analyzer (Series 200, Alpha Omega Instrument Corporation, RI, USA), was inserted through the inlet in the middle of the acrylic glass lid. The oxygen analyzer readings were video recorded during filling (5 min). Every 0.2 s CO<sub>2</sub> concentrations were estimated from changes in oxygen concentrations using the formula CO<sub>2</sub> (t = x) = 100 –([O<sub>2</sub> (t = x) \* 100] / O<sub>2</sub> (t = 0).



Fig 2. Experiments timeline. Order of testing across the three experiments (i.e. aversion to CO2, sweet reward motivation and regulatory focus). In all three experiments rats were trained, retrained or tested every weekday but not on weekends.

#### **Experiment 2: Sweet reward motivation**

**Apparatus.** A modified approach-avoidance apparatus was used for this test. During baseline, the apparatus remained the same as described for the approach-avoidance test. During test sessions, the bottom cage was replaced with a new test cage measuring 20 cm x 45 cm x 24 cm. The test cage was covered with a wire lid and contained two ice cube trays with 12 holes each, and was covered with autoclaved sand (Fig 1B).

**Training and testing procedure.** Rats were habituated once and tested three times for sweet reward motivation. At the beginning of each session, the rat was placed into the top cage, and could freely move between the top and bottom cages for 5 min. The rat was then given a signal by the experimenter to receive one sweet reward in the top cage, and the sliding door was closed for 60 s. During this period the baseline bottom cage was replaced with the test cage. The test cage contained 20 sweet rewards placed in ice tray holes and hidden underneath a layer of sand. One sweet reward was left on top of the sand in the middle of the cage. The rat was then allowed to descend to the bottom test cage to search for and consume the rewards. The session ended if the rat left the test cage (i.e. shoulders crossed into the tube exiting the cage) without carrying a sweet reward, or if the subject had left the cage carrying a sweet reward.

For the training trial, the sweet rewards were distributed in 6 adjacent holes of the ice tray, with 3 to 4 sweet rewards per hole. For rats that consumed fewer than 15 sweet rewards during their training trial (n = 5 rats), training was repeated a second time. In the first test trial, the sweet rewards were distributed into 9 reward holes, separated by empty holes, with 2 to 3 sweet rewards per hole. In the second test trial, sweet rewards were evenly distributed throughout the tray with only one sweet reward per reward hole and at least one empty hole between each reward. In the third test trial, the sweet rewards were randomly distributed throughout the tray at coordinates obtained from a random number generator with a maximum of 2 rewards per hole.

Any rewards remaining were counted at the end of the trial. All trials were video recorded and scored using Boris software (Version 7.0.9) [25]. A trained observer, blind to rat identity and trial number, scored the videos for the number of sweet rewards consumed and searching time between each consecutive reward found (s). Inter-observer reliability was estimated from 10 videos scored by the trained observer and another independent observer (number of sweet rewards consumed: r = 0.99; searching time: r = 0.99).

#### **Experiment 3: Regulatory focus**

**Apparatus.** Following Franks and colleagues [23], a modified open field arena was used for regulatory focus profiling. The modified open field arena was made of white acrylic and contained two smaller acrylic boxes placed against the center of two adjacent walls of the arena (treat and dark locations; Fig 1C). The arena was illuminated with red light and white light that provided an average light intensity of  $82 \pm 1.6$  lux (mean  $\pm$  standard deviation) at the center of the arena floor.

**Habituation and testing procedure.** Rats were habituated twice and tested twice in the modified open field arena. Before each trial, a variety of food rewards (20 Cheerios: Honey Nut Cheerios TM, General Mills Inc., MN, USA; 10 sunflower seeds: Raw Sunflower Seeds, Western Family, Overwaitea Food Group LP, BC, Canada; 1 yogurt drop: Drops Yogurt Flavoured Treats, Living World, Hagen Inc., QC, Canada; 2 peanut M&Ms: Mars Canada Inc., ON, Canada; 20 peanuts: Peanuts Roasted in the Shell, Western Family, Overwaitea Food Group LP., BC, Canada; 1 he small boxes (treat location) inside the arena, and the other small box was left empty (dark location). At the beginning of each trial, the subject was introduced to the arena in the farthest corner from and equidistant to the treat and dark locations. Rats were left in the arena for 10 min; if the rat approached the dark location, whichever occurred first.

All trials were video recorded and scored by a trained observer, blind to rat identity and trial number. The size of rats was bigger than that of the treat and dark locations; therefore, and following Franks et al. [23], the observer scored the amount of time rats spent within 20 cm of the treat and dark locations (s) using Boris software. Another independent observer scored 4 videos to estimate inter-observer reliability (treat location time: r = 0.99; dark location time: r = 0.99).

#### Data analysis

Analyses were carried out with R (R Development Core Team, Version 3.4.1) and RStudio (RStudio, Inc., Version 1.0.136). The model residuals and data were visually assessed for normality. Results are reported as mean  $\pm$  standard error.

**Experiment 1: Aversion to CO<sub>2</sub>.** We estimated the CO<sub>2</sub>% concentrations at the time when rats exited the bottom cage (i.e.  $CO_2$ % avoided), using the average concentration of  $CO_2$ at each time point (measured every 0.2 s) during the  $12 \text{ CO}_2$  flow trials. Random effects models are a useful tool to handle unbalanced or incomplete data, and it has been shown that the power to detect individual differences in these models is improved by the inclusion of data from individuals with only a few observations and that removal of individuals with a low number of observations is unjustified [26]. Hence, we used all available observations from all subjects using a linear mixed model ("nlme" R-package; an alternative analysis considering each age separately is presented as Supporting Methods S1 in S1 File). The model presented here, included the response variable  $CO_2$ % avoided in approach-avoidance tests, age (3, 9 and 16 months of age) as a fixed factor, exposure number (within age) as a covariate, the interaction between age and exposure number, and series identity (i.e. unique combination of the individual rat by the age at which observations were taken) within rat identity as a random intercept. We found that weight as fixed effect did not significantly affect aversion to  $CO_2$ , hence was not included in the model (results not shown). Age and exposure number were both mean centered and standardized to 2 standard deviations [27]. We assessed the fit of this model as essentially equivalent to a model that included exposure number as a random slope and series identity within rat identity as random intercept (see Supporting Methods S2 in S1 File). We

assessed the power to detect significant random intercepts and slopes given the current study sampling structure [26] and found sufficient power (0.87) to detect differing random intercepts by rat but low power to detect differing slopes by rat (Supporting Methods S3 in S1 File; further model diagnostics are presented in Supporting Methods S4 in S1 File). The significance of the random intercept was evaluated using the likelihood ratio test (LRT). We estimated repeatability (R; "rptR" R-package) of  $CO_2$ % avoided adjusted for age and within age exposure number (adjusted repeatability for Gaussian data [28]). The point estimate of aversion to  $CO_2$ for each individual rat was calculated as the average best linear unbiased predictors (BLUPs) of the random effects obtained from 1000 simulations ("arm" R-package). This method has been used in behavioural ecology studies to reduce biases in the estimates, arising from for example habituation or increased number of repeated tests in older animals [29].

**Experiment 2: Sweet reward motivation.** To assess individual differences in sweet reward motivation, the total number of rewards consumed, and the total searching time were included as response variables in two linear mixed models. In the models, trial number was included as fixed factor and rat identity as random intercept. LRTs were used to assess the significance of the random intercept, and repeatability (R) across trials was assessed.

We then estimated the average number of sweet rewards consumed and total searching time per rat across trials. The relationship between the two measures of rat motivation for sweet rewards and the average BLUPs of  $CO_2$ % avoided in approach-avoidance tests was assessed using Pearson correlation tests.

**Experiment 3: Regulatory focus.** To assess consistency in promotion (and prevention) focus, we used Pearson correlation to examine the percentage of time spent in the treat (and dark) location in the two test trials. For each rat, we estimated the average percentage of test time spent in the treat (and dark) location across the two trials. Again, we used Pearson correlation to assess the relationship between promotion (and prevention) focus and the average BLUPs of  $CO_2\%$  avoided.

**Sample disposition.** For Experiment 1, some rats failed to meet training criterion after six training trials in approach-avoidance within each age; these rats were not tested. We tested nine, nine and six rats at 3, 9 and 16 months of age, respectively. At age 16 months, eight rats were re-trained in approach-avoidance; however, two rats had to be euthanized due to mammary tumor development. The remaining six rats were clinically healthy. Due to repeated failure to meet training criterion during control trials (four consecutive trials), not all rats were tested with  $CO_2$  for the same number of exposures at each age (see Table 1). For experiments 2 and 3 we tested 11 rats–one of the rats tested in approach-avoidance failed to follow handling procedures and was excluded from these experiments. We used these 11 rats to assess the relationship between aversion to  $CO_2$ , sweet reward motivation and regulatory focus (see Table 1).

#### Results

#### Experiment 1: Aversion to CO<sub>2</sub>

During control trials, rats left the cage after  $4.8 \pm 1.8$  min and ate  $18 \pm 3$  Cheerios. In all test trials rats avoided CO<sub>2</sub> before recumbency. Concentrations of CO<sub>2</sub> avoided by rats increased with repeated exposures (F<sub>1,148</sub> = 17.83, p < 0.0001; n = 12; Fig 3). Aversion to CO<sub>2</sub> was affected by age (F<sub>2,10</sub> = 6.09, p < 0.02; n = 12). Compared to 3 months of age ( $8.4 \pm 0.9\%$  CO<sub>2</sub>), rats tolerated higher CO<sub>2</sub> concentrations at 9 months of age (p = 0.02; 10.9 ± 0.9% CO<sub>2</sub>). We found no evidence for an interaction between age and exposure number (F<sub>2,148</sub> = 0.57, p = 0.57; n = 12). Rat identity and series identity within rat explained 45% (Likelihood ratio test: LR = 61.67, p < 0.001; n = 12) and 5% (LR = 1.45, p = 0.21; n = 12) of the variation in the CO<sub>2</sub> concentrations avoided, respectively. CO<sub>2</sub>% avoided was repeatable (rat identity adjusted

	Experiment 1			Experiment 2	Experiment 3
	Number of exposures to CO <sub>2</sub>			Number of trials	
Rat identity	3 months	9 months	16 months	9 months	9 months
1	0	4	3	0	0
2	9	9	0	3	2
3	10	9	0	3	2
4	7	0	0	3	2
5	9	9	3	3	2
6	10	0	0	3	2
7	10	0	0	3	2
8	0	9	0	3	2
9	7	7	2	3	2
10	0	10	3	3	2
11	10	10	3	3	2
12	10	9	3	3	2

Table 1. Sample disposition for experiments 1 (aversion to CO<sub>2</sub>), 2 (sweet reward motivation) and 3 (regulatory focus), specifying number of exposures or trials for each individual rat and the age at which rats were tested.

https://doi.org/10.1371/journal.pone.0245347.t001

repeatability: R = 0.46, p < 0.01; series identity: R = 0.07, p = 0.06; n = 12; Fig 4); with average CO<sub>2</sub> concentrations avoided ranging between 4 and 15% among rats.

#### **Experiment 2: Sweet reward motivation**

Trial number did not affect rewards consumed or total searching time (sweet rewards consumed:  $F_{2,20} = 0.33$ , p = 0.72; searching time:  $F_{2,20} = 0.19$ , p = 0.83; n = 11; Fig 5). Rats spent on average 174 ± 17.2 s searching for rewards, and consumed on average 14.2 ± 1.04 sweet rewards. The random intercept (rat identity) explained 56% (LRT = 9.9, p < 0.01, n = 11) and 60% (LRT = 11.61, p < 0.001, n = 11) of the variation in searching time and rewards consumed, respectively. Searching time and sweet rewards consumed were repeatable (R = 0.56, p < 0.0001, and R = 0.60, p < 0.001, respectively; n = 11). However, aversion to CO<sub>2</sub> was not related to searching time (Pearson correlation test: r = 0.13, p = 0.71; n = 11) or rewards consumed (r = 0.13, p = 0.71; n = 11) in sweet reward motivation trials (Fig 6).





https://doi.org/10.1371/journal.pone.0245347.g003



Fig 4. Individual differences in aversion to CO<sub>2</sub>, arranged from the least to the most tolerant rat. Bigger and solid shapes indicate the mean and error bars show the standard error for each individual rat within each age. Smaller and transparent shapes represent observation from each rat at each age. Each age is presented with a different shape (circles: 3 months, triangles: 9 months, squares: 16 months) and each color represents an individual rat.

#### **Experiment 3: Regulatory focus**

Rats spent on average 31% and 51% of the test time in the treat and dark locations, respectively. Across the two test trials, rats consistently varied in the percentage of time spent in these locations (treat: r = 0.80, p < 0.01; dark: r = 0.81, p < 0.01; n = 11 rats; Fig 7). Aversion to CO<sub>2</sub> was not related to the percentage of time spent in the treat (r = 0.34, p = 0.3; n = 11) or dark locations (r = -0.09, p = 0.79; n = 11).

#### Discussion

In agreement with previous studies [15, 24], we found that rat aversion to  $CO_2$  consistently varied among rats, ranging between 4 and 15%  $CO_2$ . As frequently occurs in longitudinal studies, despite the effort to obtain measurements of aversion to  $CO_2$  from all subjects at all time points, data from some rats were missing for some ages. Despite this unbalance data, our method of analysis had sufficient power test variation among individuals. To our knowledge, the current study is the first to show that rat individual thresholds of aversion to  $CO_2$  are stable and highly repeatable across repeated exposures and across different ages (R = 0.46; the average repeatability estimates across behaviours and among taxa has been shown to be 0.37 [30]).



**Fig 5. Individual differences in sweet reward motivation.** Individual rat (n = 11) mean ( $\pm$  SE) a) searching time, and b) rewards consumed, across three sweet reward motivation trials. Rat identity follows that shown in Fig 4.



**Fig 6. Relationship between aversion to CO<sub>2</sub> and sweet reward motivation.** Each dot represents an individual rat's point estimate (obtained from the BLUPs of the random effects of 1000 simulations) of aversion to CO<sub>2</sub> and their average a) searching time, and b) rewards consumed, from the sweet reward motivation trials (n = 11).

https://doi.org/10.1371/journal.pone.0245347.g006



Fig 7. Regulatory focus consistency for individual rats (n = 11) tested in two trials; results are shown separately for measures of a) treat location time (promotion motivation), and b) dark location time (prevention motivation). Rat identity is given by a number following the order presented in Fig 4.

Previous studies have reported high between-individual variation in rat responses to CO<sub>2</sub> during forced exposure [7, 9, 14], choice [31, 32], and aversion tests [10, 11, 33], but the source of this variation remained unexplained. Rat defence behaviours are plastic, varying with environmental familiarity (i.e. habituation) [34-36], situational contingencies (for example, threat proximity and possibility to escape) [37–39], and with specific conditions prior to or during testing [40-45]. Although we found that the thresholds of aversion to CO<sub>2</sub> were higher at older ages, at 3 months of age rats were exposed to a higher flowrate than that used at later ages, impeding our ability to draw age-related inferences. Contrary to the general trend, as it can be seen in Fig 4, rat number 12 experienced a decrease in aversion to  $CO_2$  measured at ages 9 and 16 months when compared to age 3 months. This could indicate the presence of individual differences in plasticity; however, the current study lacks power to make strong claims about rat differences in slopes. Consistent with previous results from our research group [9], we found that thresholds of aversion to CO<sub>2</sub> increased over repeated exposures at 3 and 9 months of age. This result is also consistent with human studies showing that chemoreceptor sensitivity [46] and feelings of anxiety [47, 48] decrease with habituation to CO<sub>2</sub> inhalation. Overall, these results indicate that aversion to CO<sub>2</sub> is plastic and sensitive to habituation.

Behaviours consistent across time and contexts are often referred to as personality traits. These individual differences are more or less permanent characteristics that distinguish individuals from one another [49, 50]. For example, within the same strain, the degree to which rats explore novel environments is consistent between four and eight months of age [51]. Individual differences in this behavioural trait are heritable; two lines, originating from rats that differed in active avoidance acquisition (i.e. Roman high avoidance and Roman low avoidance), consistently differ in their degree of exploration of novel environments [52]. Our results showed that rat thresholds of aversion to  $CO_2$  vary among individuals, are highly repeatable, and can be considered a lasting characteristic of the individual (i.e. personality trait).

In the current study we tested individual differences in motivation to access a sweet reward using a modified approach-avoidance apparatus. Traditionally, rat motivation is measured through a progressive ratio schedule, in which the cost of gaining a reward is increased. Animals continue to invest as the required effort increases, until the cost is higher than the value of the reward [53]. In the current study, rat motivation was assessed in an experimental setting similar to that used to assess differences in foraging wild rodents [54, 55]. The individual's motivation to engage in a goal directed behaviour, like searching for food, is affected by expectancy and value [56, 57]. Motivation is expected to be higher as the likelihood of success and the value of the resources increase. When foraging in a patch, animals experience a decrease in the rate at which resources are found as they consume the available items (diminishing returns); this decrease imposes a trade-off between investing time searching for resources and leaving the patch (marginal value theorem) [58]. In the current experiment, rats searched for access to sweet rewards that were hidden in a layer of sand, and rewards were progressively dispersed across trials. The assumption was that rats would invest time searching and digging to gain access to the rewards until the required effort surpassed the value of the reward.

We found that rat motivation for sweet rewards varied among individuals. For example, one rat invested on average 5 min searching and consumed all rewards, and another rat consumed on average just 6 rewards and spent less than 40 s searching. We also found that motivation for sweet rewards was highly repeatable and not affected by repeated testing. These results align with what has been reported in the literature; rat preference and motivation for sucrose is a stable personality trait [19, 59] that does not change with repeated testing [19]. High sucrose consumers ingest more than double the intake of low consumers [18, 60, 61], but among-individual variation in sucrose preference is not related to variation in food consumption [60]. Under a progressive ratio schedule, high consumers work harder than low consumers to earn sucrose [17].

We found no evidence that individual differences in motivation for sweet rewards were related to aversion to  $CO_2$ . However, it is important to note that rats that showed low motivation for sweet rewards frequently failed to meet the training criterion for the approach-avoidance test. This result suggests that a bias of approach-avoidance tests is that only reward-motivated rats are likely to be selected.

In the current study rats spent a similar amount of time in the treat location (31% of the test time) but less time in the dark location (51% of the test time), compared to results from Franks and colleagues [23]. It is likely that the lower time in the dark location was due to methodological differences. Franks and colleagues kept the light off for 30 s, or while the subject stayed in the dark location, whichever was longer. In our study the light was turned on after 30 s or when the rat left the dark location, whichever occurred first. We argue that our experimental methodology allows for the assessment of prevention and promotion foci since rats frequently brought food rewards to consume in the dark location, and rats consistently varied in their motivation to approach gains (promotion motivation) and pursue darkness (prevention motivation). These results correspond to those previously reported. For example, rats that consistently pursued darkness in the modified open field, also consistently spent more time burying a noxious object [23]. High prevention motivated rats avoided risk, but also approach potential threats to maintain safety. In the current study, we found no evidence for a relationship between individual differences in the strength of promotion or prevention motivation and rat aversion to  $CO_2$ , indicating that personality differences in regulatory focus are not related to aversion to CO<sub>2</sub> in approach-avoidance.

Variation among human subjects in the felt experience (i.e. conscious awareness of emotions) during  $CO_2$  inhalation is well documented. The increase in feelings of anxiety is eight times higher in individuals that are more responsive to  $CO_2$ , than that of less responsive individuals [62]. Feelings of anxiety and experiences of panic due to ~ 7%  $CO_2$  inhalation are consistent between repeated inhalations [62, 63]. Vulnerability to  $CO_2$ -induced anxiety and panic increases in people diagnosed with panic disorder [3, 64] and individuals with a firstdegree relative diagnosed with panic disorder [4, 65, 66]. Thus, human  $CO_2$  sensitivity involves stable individual differences in the emotional response to  $CO_2$ . We found that individual differences in rat thresholds of aversion to  $CO_2$  were stable and consistent, and not related to sweet reward motivation or the strength of promotion and prevention motivations. It is likely that individual differences in the affective states experienced are the underlying cause of among-rat variation in aversion to  $CO_2$  (i.e.  $CO_2$  sensitivity), indicating that some rats experience a more aversive emotional response when exposed to  $CO_2$ .

# Conclusion

Variation in rat aversion to  $CO_2$  was repeatable through multiple exposures and across three different ages but was not related to individual differences in motivation for sweet rewards, promotion or prevention foci. These results indicate that individual differences in aversion to  $CO_2$  reflects variation in  $CO_2$  sensitivity.

# **Supporting information**

**S1 File.** (DOCX)

S1 Data. (XLSX)

# Acknowledgments

We thank Becca Franks, Catherine Schuppli, Marina Von Keyserlingk and Joanna Makowska for their helpful comments and suggestions. We thank Nicola Adderley for data collection, Emma Piercey for behavioural coding and Delea Carrillo and Jackson Lai for assistance in caring for our rats. We especially want to thank the three anonymous reviewers who's critical assessment and detailed and thoughtful comments were crucial for the improvement of this manuscript.

# **Author Contributions**

Conceptualization: Lucía Améndola, Daniel M. Weary. Data curation: Lucía Améndola, Anna Ratuski. Formal analysis: Lucía Améndola, Daniel M. Weary. Funding acquisition: Daniel M. Weary. Investigation: Lucía Améndola. Methodology: Lucía Améndola, Daniel M. Weary. Project administration: Lucía Améndola, Anna Ratuski.

Resources: Daniel M. Weary.

Supervision: Daniel M. Weary.

Writing - original draft: Lucía Améndola.

Writing - review & editing: Lucía Améndola, Anna Ratuski, Daniel M. Weary.

#### References

- Colasanti A, Esquivel G, J Schruers K, J Griez E. On the psychotropic effects of carbon dioxide. Curr. Pharm. Des. 2012 Dec 1; 18(35):5627–37. <u>https://doi.org/10.2174/138161212803530745</u> PMID: 22632467
- van Beek N, Griez E. Reactivity to a 35% CO<sub>2</sub> challenge in healthy first-degree relatives of patients with panic disorder. Biol. Psychiatry 2000 May 1; 47(9):830–5. https://doi.org/10.1016/s0006-3223(99) 00265-6 PMID: 10812042
- Monkul ES, et al. History of suffocation, state-trait anxiety, and anxiety sensitivity in predicting 35% carbon dioxide-induced panic. Psychiatry Res. 2010 Sep 30; 179 (2): 194–197. <a href="https://doi.org/10.1016/j.psychres.2009.06.015">https://doi.org/10.1016/j.psychres.2009.06.015</a> PMID: 20478634
- Coryell W, Arndt S. The 35% CO<sub>2</sub> inhalation procedure: test–retest reliability. Biological Psychiatry 1999 Apr 1; 45(7): 923–7. https://doi.org/10.1016/s0006-3223(98)00241-8 PMID: 10202581
- Verburg K, Pols H, De Leeuw M, Griez E. Reliability of the 35% carbon dioxide panic provocation challenge. Psychiatry Res. 1998 May 8; 78(3): 207–14. <u>https://doi.org/10.1016/s0165-1781(98)00009-2</u> PMID: 9657425
- Chisholm J, De Rantere D, Fernandez NJ, Krajacic A, Pang DSJ. Carbon dioxide, but not isoflurane, elicits ultrasonic vocalizations in female rats. Lab. Anim. 2013 Oct; 47(4): 324–7. <u>https://doi.org/10.1177/0023677213493410 PMID: 23828852</u>
- Niel L, Weary DM. Behavioural responses of rats to gradual-fill carbon dioxide euthanasia and reduced oxygen concentrations. Appl. Anim. Behav. Sci. 2006 Nov 1; 100(3–4): 295–308. https://doi.org/10. 1016/j.applanim.2005.12.001
- Winter A, Ahlbrand R, Naik D, Sah R. Differential behavioral sensitivity to carbon dioxide (CO<sub>2</sub>) inhalation in rats. Neuroscience. 2017 Mar 27; 346:423–33. https://doi.org/10.1016/j.neuroscience.2017.01. 003 PMID: 28087339
- 9. Niel L, Kirkden RD, Weary DM. Effects of novelty on rats' responses to CO<sub>2</sub> exposure. Appl. Anim. Behav. Sci. 2008 May 1; 111(1–2):183–94. https://doi.org/10.1016/j.applanim.2007.06.004
- Wong D, Makowska IJ, Weary DM. Rat aversion to isoflurane versus carbon dioxide. Biol. Lett. 2013 Feb 23; 9(1): 20121000. https://doi.org/10.1098/rsbl.2012.1000 PMID: 23256183
- Kirkden RD, Niel L, Lee G, Makowska IJ, Pfaffinger MJ, Weary DM. The validity of using an approachavoidance test to measure the strength of aversion to carbon dioxide in rats. Appl. Anim. Behav. Sci. 2008 Nov 1; 114(1–2): 216–34. https://doi.org/10.1016/j.applanim.2008.03.001
- Johnson PL, et al. Orexin 1 and 2 Receptor Involvement in CO<sub>2</sub>-induced panic-associated behavior and autonomic responses. Depress. Anxiety 2015 Sep; 32(9): 671–83. <u>https://doi.org/10.1002/da.22403</u> PMID: 26332431
- **13.** Leibold NK, et al. CO<sub>2</sub> exposure as translational cross-species experimental model for panic. Transl. Psychiatry 2016 Sep; 6(9): 885. https://doi.org/10.1038/tp.2016.162 PMID: 27598969
- Smith W, Harrap SB. Behavioural and cardiovascular responses of rats to euthanasia using carbon dioxide gas. Lab. Anim. 1997 Oct 1; 31(4): 337–346. https://doi.org/10.1258/002367797780596130 PMID: 9350705
- Améndola L, Weary DM. Evidence for consistent individual differences in rat sensitivity to carbon dioxide. PLoS One. 2019; 14(4): e0215808. https://doi.org/10.1371/journal.pone.0215808 PMID: 31017958
- Améndola L, Weary DM. Understanding rat emotional responses to CO<sub>2</sub>. Transl. Psychiatry 2020 Jul; 10(1):1–2. https://doi.org/10.1038/s41398-019-0665-5 PMID: 32066695
- Brennan K, Roberts DCS, Anisman H, Merali Z. Individual differences in sucrose consumption in the rat: Motivational and neurochemical correlates of hedonia. Psychopharmacology 2001 Sep 1; 157(3): 269–276. https://doi.org/10.1007/s002130100805 PMID: 11605082
- Desousa NJ, Wunderlich GR, De Cabo C, Vaccarino FJ. Individual differences in sucrose intake predict behavioral reactivity in rodent models of anxiety. Pharmacol. Biochem. Behav. 1998 Aug 1; 60(4): 841– 846. https://doi.org/10.1016/s0091-3057(98)00069-0 PMID: 9700967
- Tõnissaar M, Herm L, Rinken A, Harro J. Individual differences in sucrose intake and preference in the rat: Circadian variation and association with dopamine D2 receptor function in striatum and nucleus accumbens. Neurosci. Lett. 2006 Jul 31; 403 (1–2): 119–124. https://doi.org/10.1016/j.neulet.2006.04. 023 PMID: 16682119
- Higgins ET. Beyond pleasure and pain. Am. Psychol. 1997 Dec; 52 (12): 1280. <u>https://doi.org/10.1037//0003-066x.52.12.1280 PMID: 9414606</u>

- Shah J, Higgins ET. Expectancy x value effects: Regulatory focus as determinant of magnitude and direction. J. Pers. Soc. Psychol. 1997 Sep; 73 (3): 447. https://doi.org/10.1037//0022-3514.73.3.447 PMID: 9294896
- Franks B, Higgins ET, Champagne FA. A theoretically based model of rat personality with implications for welfare. PLoS One. 2014; 9 (4): e95135. https://doi.org/10.1371/journal.pone.0095135 PMID: 24755737
- Franks B, Higgins ET, Champagne FA. Evidence for individual differences in regulatory focus in rats, Rattus norvegicus. J. Comp. Psychol. 2012 Nov; 126 (4): 347. <u>https://doi.org/10.1037/a0027244</u> PMID: 22390620
- Améndola L, Ratuski A, Weary DM. Variation in the onset of CO<sub>2</sub>-induced anxiety in female Sprague Dawley rats. Sci. Rep. 2019 Dec 12; 9 (1): 19007. https://doi.org/10.1038/s41598-019-55493-0 PMID: 31831816
- Friard O, Gamba M. BORIS: a free, versatile open-source event-logging software for video/audio coding and live observations. Methods Ecol. Evol. 2016; 7: 1325–1330. https://doi.org/10.1111/2041-210X.12584
- Martin JG, Nussey DH, Wilson AJ, Réale D. Measuring individual differences in reaction norms in field and experimental studies: a power analysis of random regression models. Methods Ecol Evol. 2011 Aug; 2(4): 362–374.
- Araya-Ajoy YG, Mathot KJ, Dingemanse NJ. An approach to estimate short-term, long-term and reaction norm repeatability. Methods Ecol Evol. 2016 Dec; 6(12): 1462–1473.
- Nakagawa S. & Schielzeth H. Repeatability for Gaussian and non-Gaussian data: A practical guide for biologists. Biol. Rev. 2010 Nov; 85(4): 935–956. <u>https://doi.org/10.1111/j.1469-185X.2010.00141.x</u> PMID: 20569253
- Dingemanse NJ, Moiron M, Araya-Ajoy YG, Mouchet A, Abbey-Lee RN. Individual variation in agedependent reproduction: Fast explorers live fast but senesce young?. J Anim Ecol. 2020 Feb; 89(2): 601–613. https://doi.org/10.1111/1365-2656.13122 PMID: 31618450
- Bell AM, Hankison SJ, Laskowski KL. The repeatability of behaviour: a meta-analysis. Anim. Behav. 2009 Apr 1; 77(4): 771–783. https://doi.org/10.1016/j.anbehav.2008.12.022 PMID: 24707058
- Leach MC, Bowell VA Allan TF, Morton DB. Aversion to gaseous euthanasia agents in rats and mice. Comp. Med. Jun 15; 52(3): 249–57. PMID: 12102571
- Leach MC, Bowell VA, Allan TF, Morton DB. Measurement of aversion to determine humane methods of anaesthesia and euthanasia. Anim. Welf. 2004 Feb 1; 13: S77–86.
- Niel L, Stewart SA, Weary DM. Effect of flow rate on aversion to gradual-fill carbon dioxide exposure in rats. Appl. Anim. Behav. Sci. 2008 Jan 1; 109(1): 77–84. <u>https://doi.org/10.1016/j.applanim.2007.02.</u> 004
- Fernandes C, File SE. The influence of open arm ledges and maze experience in the elevated plusmaze. Pharmacol. Biochem. Behav. 1996 May 1; 54(1): 31–40. https://doi.org/10.1016/0091-3057(95) 02171-x PMID: 8728536
- Romero RD, Chen WJA. Gender-related response in open-field activity following developmental nicotine exposure in rats. Pharmacol. Biochem. Behav. 2004 Aug 1; 78(4): 675–81. https://doi.org/10.1016/ j.pbb.2004.04.033 PMID: 15301921
- Treit D, Menard J, Royan C. Anxiogenic stimuli in the elevated plus-maze. Pharmacol. Biochem. Behav. 1993 Feb 1; 44(2):463–9. https://doi.org/10.1016/0091-3057(93)90492-c PMID: 8446680
- Eilam D. Die hard: A blend of freezing and fleeing as a dynamic defense—Implications for the control of defensive behavior. Neurosci. Biobehav. Rev. 2005 Jan 1; 29(8):1181–91. <u>https://doi.org/10.1016/j.</u> neubiorev.2005.03.027 PMID: 16085311
- Blanchard RJ, Fukunaga KK, Blanchard DC. Environmental control of defensive reactions to a cat. Bull. Psychon. Soc. 1976 Sep 1; 8(3):179–81. https://doi.org/10.3758/BF03335118
- McGregor IS, Schrama L, Ambermoon P, Dielenberg RA. Not all 'predator odours' are equal: Cat odour but not 2, 4, 5 trimethylthiazoline (TMT; fox odour) elicits specific defensive behaviours in rats. Behav. Brain Res. 2002 Jun 1; 129 (6): 1–16. https://doi.org/10.1016/S0166-4328(01)00324-2
- Archer J. Tests for emotionality in rats and mice: A review. Anim. Behav. 1973; 21: 205–235. <a href="https://doi.org/10.1016/s0003-3472(73)80065-x">https://doi.org/10.1016/s0003-3472(73)80065-x</a> PMID: 4578750
- Davis H, Pérusse R. Human-based social interaction can reward a rat's behavior. Anim. Learn. Behav. 1988 Mar 1; 16 (1): 89–92. https://doi.org/10.3758/BF03209048
- 42. Eilam D. Open-field behavior withstands drastic changes in arena size. Behav. Brain Res. 2003 Jun 16; 142(1–2), 53–62. https://doi.org/10.1016/s0166-4328(02)00382-0 PMID: 12798265

- **43.** Lapin IP. Only controls: Effect of handling, sham injection, and intraperitoneal injection of saline on behavior of mice in an elevated plus-maze. J. Pharmacol. Toxicol. Methods. 1995 Oct 1; 34 (2): 73–77. https://doi.org/10.1016/1056-8719(95)00025-d PMID: 8563035
- 44. Pritchard LM, Van Kempen TA. & Zimmerberg B. Behavioral effects of repeated handling differ in rats reared in social isolation and environmental enrichment. Neurosci. Lett. 2013 Mar 1; 536: 47–51. https://doi.org/10.1016/j.neulet.2012.12.048 PMID: 23313592
- Walsh RN, Cummins RA. The open-field test: A critical review. Psychol. Bull. 1976; 83: 482. <u>https://doi.org/10.1037/0033-2909.83.3.482</u> PMID: <u>17582919</u>
- 46. McMahon ME, Boutellier U, Smith RM, Spengler CM. Hyperpnea training attenuates peripheral chemosensitivity and improves cycling endurance. J Exp Biol. 2002 Dec 15; 205 (24): 3937–43. PMID: 12432015
- Van den Hout MA, der Molen V, Margo G, Griez E, Lousberg H, Nansen A. Reduction of CO2-induced anxiety in patients with panic attacks after repeated CO2 exposure. Am J Psychiatry. 1987 Aug 1; 144 (8): 788–791. https://doi.org/10.1176/ajp.144.6.788 PMID: 3109265
- Li W, et al. Air hunger and ventilation in response to hypercapnia: effects of repetition and anxiety. Physiol Beha. 2006 Jun 15; 88(1–2): 47–54. <u>https://doi.org/10.1016/j.physbeh.2006.03.001</u> PMID: 16626764
- Dingemanse NJ, Kazem AJN, Réale D, Wright J. Behavioural reaction norms: animal personality meets individual plasticity. Trends Ecol. Evol. 2010 Feb 1; 25(2): 81–89. <u>https://doi.org/10.1016/j.tree.2009</u>. 07.013 PMID: 19748700
- Réale D, Reader SM, Sol D, McDougall PT, Dingemanse NJ. Integrating animal temperament within ecology and evolution. Biol. Rev. 2007 May; 82 (2): 291–318. <u>https://doi.org/10.1111/j.1469-185X.</u> 2007.00010.x PMID: 17437562
- Cavigelli SA, McClintock MK. Fear of novelty in infant rats predicts adult corticosterone dynamics and an early death. Proc. Natl. Acad. Sci. U. S. A. 2003 Dec 23; 100 (26): 16131–16136. https://doi.org/10. 1073/pnas.2535721100 PMID: 14673078
- Steimer T, La Fleur S, Schulz PE. Neuroendocrine correlates of emotional reactivity and coping in male rats from the Roman high (RHA/Verh)-and low (RLA/Verh)-avoidance lines. Behav. Genet. 1997 Nov 1; 27(6): 503–512. https://doi.org/10.1023/a:1021448713665 PMID: 9476359
- Hodos W. Progressive ratio as a measure of reward strength. Science. 1961 Sep 29; 134(3483): 943– 944. https://doi.org/10.1126/science.134.3483.943 PMID: 13714876
- Kotler BP, Brown JS. Rates of Seed Harvest by Two Species of Gerbilline Rodents. J. Mammal. 1990 Dec 7; 71(4): 591–6. https://doi.org/10.2307/1381798
- Smith RJ. Harvest Rates and Escape Speeds in Two Coexisting Species of Montane Ground Squirrels. J. Mammal. 1995 Feb 23; 76(1): 189–95. https://doi.org/10.2307/1382327
- Wigfield A, Eccles JS. Expectancy-value theory of achievement motivation. Contemp. Educ. Psychol. 2000; 76: 49–78. https://doi.org/10.1006/ceps.1999.1015 PMID: 10620382
- 57. Tolman EC. Principles of performance. Psychol. Rev. 1955 Sep; 62(5): 315. <u>https://doi.org/10.1037/</u> h0049079 PMID: 13254969
- Charnov EL. Optimal foraging, the marginal value theorem. Theor. Popul. Biol. 1976; 129–136. <a href="https://doi.org/10.1016/0040-5809(76)90040-x">https://doi.org/10.1016/0040-5809(76)90040-x</a> PMID: 1273796
- 59. Sills TL, Vaccarino FJ. Individual differences in sugar consumption following systemic or intraaccumbens administration of low doses of amphetamine in nondeprived rats. Pharmacol. Biochem. Behav. 1996 Aug 1; 54(4): 665–70. https://doi.org/10.1016/0091-3057(96)00024-x PMID: 8853187
- DeSousa NJ, Bush DEA, Vaccarino FJ. Self-administration of intravenous amphetamine is predicted by individual differences in sucrose feeding in rats. Psychopharmacology. 2000 Jan 1; 148(1): 52–8. https://doi.org/10.1007/s002130050024 PMID: 10663417
- Gosnell BA. Sucrose intake predicts rate of acquisition of cocaine self- administration. Psychopharmacology. 2000 Apr 1; 149(3): 286–92. https://doi.org/10.1007/s002130000375 PMID: 10823410
- 62. Poma SZ, et al. Characterization of a 7% carbon dioxide (CO<sub>2</sub>) inhalation paradigm to evoke anxiety symptoms in healthy subjects. J. Psychopharmacol. 2005 Sep; 19(5): 494–503. <u>https://doi.org/10.1177/0269881105056533 PMID: 16166187</u>
- Roberson-Nay R, et al. Temporal stability of multiple response systems to 7.5% carbon dioxide challenge. Biol. Psychol. 2017 Mar 1; 124: 111–8. <u>https://doi.org/10.1016/j.biopsycho.2017.01.014</u> PMID: 28163046
- Gorman JM, et al. Physiological changes during carbon dioxide inhalation in patients with panic disorder, major depression, and premenstrual dysphoric disorder: Evidence for a central fear mechanism. Arch. Gen. Psychiatry. 2001 Dec 1; 52(12): 1048–60. https://doi.org/10.1001/archpsyc.58.2.125 PMID: 11177114

- Perna G, Cocchi S, Allevi L, Bussi R, Bellodi L. A long-term prospective evaluation of first-degree relatives of panic patients who underwent the 35% CO<sub>2</sub> challenge. Biol. Psychiatry. 1999 Feb 1; 45(3): 365–367. https://doi.org/10.1016/s0006-3223(98)00030-4 PMID: 10023515
- 66. Van Beek N, Griez E. Reactivity to a 35% CO<sub>2</sub> challenge in healthy first-degree relatives of patients with panic disorder. Biol. Psychiatry 2000 May 1; 47 (9), 830–835. https://doi.org/10.1016/s0006-3223(99) 00265-6 PMID: 10812042