

RESEARCH ARTICLE

Effect of Voluntary Ethanol Consumption Combined with Testosterone Treatment on Cardiovascular Function in Rats: Influence of Exercise Training

Sheila A. Engi^{1,2}, Cleopatra S. Planeta^{1,2}, Carlos C. Crestani^{1,2*}

1 Laboratory of Pharmacology, School of Pharmaceutical Sciences, Univ. Estadual Paulista-UNESP, Araraquara, SP, Brazil, **2** Joint UFSCar-UNESP Graduate Program in Physiological Sciences, São Carlos, SP, Brazil

* cccrestani@yahoo.com.br



OPEN ACCESS

Citation: Engi SA, Planeta CS, Crestani CC (2016) Effect of Voluntary Ethanol Consumption Combined with Testosterone Treatment on Cardiovascular Function in Rats: Influence of Exercise Training. PLoS ONE 11(1): e0146974. doi:10.1371/journal.pone.0146974

Editor: Leonardo Barbosa Moraes Resstel, University of São Paulo, BRAZIL

Received: September 4, 2015

Accepted: December 23, 2015

Published: January 13, 2016

Copyright: © 2016 Engi et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are contained within the paper.

Funding: This work was supported by the São Paulo Research Foundation (FAPESP) grants # 2013/09715-2, 2012/14723-1, and 2012/14376-0; National Counsel of Technological and Scientific Development (CNPq) grant #456405/2014-3 CNPq, and PADCF UNESP. CSP is a CNPq research fellow. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

This study evaluated the effects of voluntary ethanol consumption combined with testosterone treatment on cardiovascular function in rats. Moreover, we investigated the influence of exercise training on these effects. To this end, male rats were submitted to low-intensity training on a treadmill or kept sedentary while concurrently being treated with ethanol for 6 weeks. For voluntary ethanol intake, rats were given access to two bottles, one containing ethanol and other containing water, three 24-hour sessions per week. In the last two weeks (weeks 5 and 6), animals underwent testosterone treatment concurrently with exercise training and exposure to ethanol. Ethanol consumption was not affected by either testosterone treatment or exercise training. Also, drug treatments did not influence the treadmill performance improvement evoked by training. However, testosterone alone, but not in combination with ethanol, reduced resting heart rate. Moreover, combined treatment with testosterone and ethanol reduced the pressor response to the selective α_1 -adrenoceptor agonist phenylephrine. Treatment with either testosterone or ethanol alone also affected baroreflex activity and enhanced depressor response to acetylcholine, but these effects were inhibited when drugs were coadministered. Exercise training restored most cardiovascular effects evoked by drug treatments. Furthermore, both drugs administered alone increased pressor response to phenylephrine in trained animals. Also, drug treatments inhibited the beneficial effects of training on baroreflex function. In conclusion, the present results suggest a potential interaction between toxic effects of testosterone and ethanol on cardiovascular function. Data also indicate that exercise training is an important factor influencing the effects of these substances.

Competing Interests: The authors have declared that no competing interests exist.

Introduction

Mental and substance use disorders are among major contributors to the burden of disease in the world [1]. Excessive ethanol consumption is the most prevalent condition among substance use disorders [1,2]. Cardiovascular dysfunctions constitute important complications associated with heavy ethanol use [3,4]. Indeed, several harmful cardiovascular effects have been reported following excessive ethanol consumption, including hypertension, cardiomyopathy, arrhythmia, coronary heart disease, and atherosclerosis [5,6]. Clinical and preclinical studies have demonstrated that alterations in contractile/relaxant properties of the vascular smooth muscle, changes in neuroendocrine function, impairment of baroreflex activity, and autonomic imbalance constitute important mechanisms underlying the negative cardiovascular effects of heavy ethanol consumption [3,4,6–8].

Abuse of androgenic—anabolic steroids (AASs) is also a serious public health problem [9,10]. For instance, clinical and preclinical studies have associated chronic AAS abuse with several cardiovascular dysfunctions, including hypertension, atherosclerosis, cardiac pathologies, impairment of baroreflex function, and changes in vascular function [11–13]. Most importantly, emerging data indicated that AAS abuse is associated with use of other substances. In fact, clinical evidence indicated that abuse of androgenic—anabolic steroids (AASs) was positively associated with ethanol use and dependence [14–16]. These findings are corroborated by preclinical studies showing that AAS can affect voluntary ethanol consumption and ethanol preference [17–19]. Despite the evidence that AAS and ethanol are co-abused, the potential toxic effects of the concomitant use of these substances are unknown.

Exercise is an important factor associated with ethanol consumption and AAS abuse. Indeed, a positive relationship between physical activity level and ethanol consumption have been demonstrated in humans across all ages [20]. To date, the factors related to this association in humans is unclear, but some authors have proposed that it would be an aware process of seeking of the exercise as a compensate mechanism for the excessive calories consumed from drinking [21,22]. However, evidence from preclinical studies has demonstrated that exercise can influence ethanol consumption and preference [23–27], possibly due to training-induced neuroplasticity in reward pathways [24]. This association is relevant to ethanol-evoked cardiovascular dysfunctions since previous studies have reported that exercise training attenuates the hypertension induced by ethanol [28,29]. However, the mechanisms underlying the beneficial cardiovascular effects of exercise in ethanol-treated animals are poorly understood.

The association between AAS abuse and exercise practice is well known [30]. Nevertheless, there is a lack in the literature of studies that investigated the influence of training in AAS-evoked cardiovascular changes [31]. Moreover, there is no evidence of the effect of exercise training on cardiovascular effects following combined use of ethanol and AAS. Therefore, our purpose in the present study was to evaluate the effects of voluntary ethanol consumption and testosterone treatment alone or in combination on basal values of arterial pressure and heart rate (HR), baroreflex activity, and blood pressure response to vasoactive agents in rats. Moreover, we investigated the possible protective effect of exercise training on these effects.

Materials and Methods

Animals

Sixty-seven male Wistar rats weighing approximately 200 g (50-days-old) in the beginning of the experiments were used. Animals were obtained from the animal breeding facility of the São Paulo State University-UNESP (Botucatu-SP, Brazil) and were housed in plastic cages in a temperature-controlled room at 24°C in the Animal Facility of the Laboratory of Pharmacology-

UNESP. They were kept under a 12:12 h light-dark cycle (lights on between 7:00h and 19:00h). Housing conditions and experimental procedures were carried out following protocols approved by the Ethical Committee for Use of Animal and Subjects of the School of Pharmaceutical Sciences/UNESP (approval# 18/2013), which complies with Brazilian and international guidelines for animal use and welfare.

Treatments

Voluntary ethanol consumption was performed using the *intermittent-access to 20% ethanol 2-bottle-choice drinking paradigm*, adapted from Simms et al. [32]. This is a free-choice method useful to estimate voluntary and spontaneous intake, as the animal is not forced to drink the ethanol solution and can choose whether to drink ethanol as well as the amount ingested over the time of exposure [32,33].

Rats were individually housed throughout the experiment and were given free access to two bottles during ethanol supply, one containing ethanol and other containing water. During the first 5 days (adaptation period), ethanol concentration was progressively increased daily (2%, 4%, 8%, 12%, 16%, or 20% v/v). On the 8th day, the intermittent access began, thus, rats were given 24h access to one bottle containing 20% ethanol and one bottle of water three times a week (Monday, Wednesday, and Friday) during 5 weeks. To determine the amount of ethanol consumed, the bottles were weighted before and after the 24h period of ethanol access. Values of ethanol consumed were normalized to body weight and consumption is presented as g/kg/24h. Rats had free access to standard laboratory food throughout the experiment.

Treatment with testosterone (10 mg/kg, subcutaneously) was realized daily for 14 consecutive days. The doses and treatment regimen of testosterone were based on our previous studies [13,34,35].

Exercise training

All animals were familiarized with exercise on a rodent treadmill (AVS Projetos, São Carlos, SP, Brazil) for one week. During the familiarization period, animals ran daily on the treadmill at a speed of 0.3 km/h and 0% grade for 10 min. No electrical stimulation was used to induce them to run [36]. Then, animals underwent a progressive maximal exercise test, which consisted on treadmill running with 0.3 km/h of increment each 3 min until exhaustion [37]. After the first maximal exercise test, animals were randomly allocated in sedentary and trained (both groups possessed the same physical capacity before training onset). Trained groups underwent a low-intensity training (50–60% of maximal exercise capacity, 0% grade) on the treadmill 1 h/day, 5 days/week for 6 weeks [37]. The sedentary groups were submitted once per week to a short period of mild exercise (10 min, 0.5 km/h, 0% grade) to keep them familiarized with treadmill environment and experimental procedures. Progressive maximal running test was repeated at weeks 4 and 6 in order to adjust training intensity and evaluate the efficacy of training protocol by comparing maximal capacity of sedentary and trained groups.

Surgical Preparation

Animals were anesthetized with tribromoethanol (250 mg/kg, i.p.) and a catheter was inserted into the abdominal aorta through the femoral artery for cardiovascular recording. A second catheter was implanted into the femoral vein for the infusion of drugs. Both catheters were tunneled under the skin and exteriorized on the animal's dorsum. The catheters were filled with a solution of heparin (50 UI/ml, Hepamax-S[®], Blausiegel, Cotia, SP, Brazil) diluted in saline (0.9% NaCl). After the surgery, rats were treated with a poly-antibiotic formulation with

streptomycins and penicillins (560 mg/ml/kg, i.m.) to prevent infection and the non-steroidal anti-inflammatory drug flunixin meglumine (0.5 mg/ml/kg, s.c.) for postoperative analgesia.

Measurement of Cardiovascular Parameters

The arterial cannula was connected to a pressure transducer (DPT100, Utah Medical Products Inc., Midvale, UT, USA). Pulsatile arterial pressure was recorded using an amplifier (Quad Bridge Amp, ML224, ADInstruments, NSW, Australia) and an acquisition board (PowerLab 4/30, ML866/P, ADInstruments, NSW, Australia). Mean (MAP), systolic (SAP), and diastolic (DAP) arterial pressure and HR values were derived from pulsatile arterial pressure recordings.

Infusion of vasoactive agents

Intravenous infusion of the α_1 -adrenoceptor agonist phenylephrine (70 μ g/ml at 0.4 ml/min/kg), the nitric oxide donor sodium nitroprusside (SNP) (100 μ g/ml at 0.8 ml/min/kg), and acetylcholine (10 μ g/ml at 1.2 ml/min/kg) was performed using an infusion pump (K.D. Scientific, Holliston, MA, USA) [8,13]. Phenylephrine caused incremental pressor effect while SNP and acetylcholine evoked incremental depressor responses.

Assessment of baroreflex activity

Paired values of MAP and HR changes evoked by phenylephrine and SNP infusion were plotted to generate sigmoid logistic functions. The logistic equation was as follows:

$$HR = P_1 + (P_2 - P_1) / (1 + \exp[BP_{50} - MAP / \text{slope}])$$

Where P_1 = lower HR plateau (bpm) (i.e., maximum reflex bradycardia), P_2 = upper HR plateau (bpm) (i.e., maximum reflex tachycardia), $P_2 - P_1$ = HR range (bpm), slope = the steepness of the curve, BP_{50} = the MAP at 50% of the HR range [38]. The average gain (G, bpm/mmHg) is the average slope of the curves between +1 and -1 standard derivations from BP_{50} [38].

Dose-response arterial pressure curves

The graded changes in MAP evoked by intravenous infusion of phenylephrine, SNP, and acetylcholine were plotted to generate dose—response curves [8,13]. Dose—effect curves were generated for each vasoactive agent by calculating the amount of drug infused and the MAP change each 2 s after starting the infusion. The maximal effect (E_{\max}) and the dose at 50% of the MAP range (ED_{50}) for each vasoactive agent were compared in all experimental groups.

Drugs

Phenylephrine hydrochloride (Sigma-Aldrich, St. Louis, MO, USA), sodium nitroprusside (Sigma-Aldrich), acetylcholine (Sigma-Aldrich) and tribromoethanol (Sigma-Aldrich) were dissolved in saline (0.9% NaCl). Ethanol (Labsynth, Diadema, SP, Brazil) was diluted in the drinking water. Testosterone propionate (PharmaNostra, Rio de Janeiro, RJ, Brazil) was dissolved in almond oil. Flunixin meglumine (Banamine[®], Schering-Plough, Cotia, SP, Brazil) and the poly-antibiotic preparation (Pentabiotico[®], Fort-Dodge, Brazil) were used as provided.

Experimental procedures

Different set of sedentary and trained animals were randomly allocated in four experimental groups: (i) control group (veh+veh), which animals were treated with almond oil (vehicle of

testosterone, 1 ml/kg, s.c.) and the vehicle of ethanol (water, v.o.) (sedentary: $n = 8$, trained: $n = 9$); (ii) testosterone group (T+veh), which animals were treated with testosterone (10 mg/kg, s.c.) and the vehicle of ethanol (sedentary: $n = 9$, trained: $n = 8$); (iii) ethanol group (veh+EtOH), which consumed ethanol (20% v/v, drinking water) and were treated with almond oil (sedentary: $n = 9$, trained: $n = 9$); and (iv) testosterone + ethanol group (T+EtOH), which consumed ethanol and were treated with testosterone (sedentary: $n = 7$, trained: $n = 8$). Exercise training on the treadmill and ethanol treatment started on the same day and were realized for 6 weeks. For voluntary ethanol consumption, during all period of ethanol supply animals were given free access to two bottles, one containing ethanol and other containing water. During the first week, ethanol concentration was progressively increased daily until reach 20%. After this period, rats were given 24 h access to one bottle containing 20% ethanol and one bottle of water three times a week (Monday, Wednesday, and Friday). In the last two weeks (weeks 5 and 6), animals underwent testosterone treatment concurrently with ethanol treatment and exercise training. Protocols of treatment were based on our previous studies demonstrating cardiovascular changes following 10 days of daily administration of testosterone, whereas alterations in autonomic activity and cardiovascular function evoked by ethanol are mainly observed after 4 weeks of treatment [8,13,34,35]. Twenty-four hours after drug treatments and exercise training completion, animals in all experimental groups were subjected to surgical preparation, and the cardiovascular tests were performed 24 hours later. A schematic representation of the complete experimental protocol is presented in Fig 1.

On cardiovascular test day, animals were transferred to the experimental room in their home box and allowed 60 min to adapt to experimental room conditions, such as sound and illumination, before starting experiments. In the sequence, animals were subjected to a 30-min period of basal cardiovascular recording. After that, they received intravenous infusion of phenylephrine, SNP, and acetylcholine in a random order.

Data Analysis

Data were expressed as mean \pm SEM. All analysis of cardiovascular function were realized using two-way ANOVA, with treatment (testosterone and/or ethanol) and exercise (sedentary vs trained) as independent factors. Ethanol consumption and treadmill performance were analyzed using three-way ANOVA, with treatment and exercise as main independent factors and time as repeated measurement. When interactions between the factors were observed in two- and three-way ANOVA, groups were compared using Bonferroni's *post hoc* test. Results of statistical tests with $P < 0.05$ were considered significant.

Results

Effects of ethanol and/or testosterone treatment and training on treadmill performance

Analysis of maximal running speed (km/h) in maximal exercise tests before the onset of testosterone treatment indicated a main effect of training ($F_{(1,65)} = 17$, $P < 0.0001$), but without influence of ethanol consumption ($F_{(1,65)} = 0.04$, $P > 0.05$) and time ($F_{(1,65)} = 0.1$, $P > 0.05$) (Fig 2A). Analysis also indicated a training x time interaction ($F_{(1,65)} = 8$, $P < 0.006$), but not training x treatment ($F_{(1,65)} = 0.3$, $P > 0.05$) or treatment x time ($F_{(1,65)} = 0.4$, $P > 0.05$) interactions. Analysis of treadmill performance after completion of drug treatments (ethanol and testosterone treatments) and exercise training indicated effect of training ($F_{(1,63)} = 55$, $P < 0.0001$), but without influence of treatments ($F_{(3,63)} = 0.1$, $P > 0.05$) and treatment x training interaction ($F_{(3,63)} = 1$, $P > 0.05$) (Fig 2B).

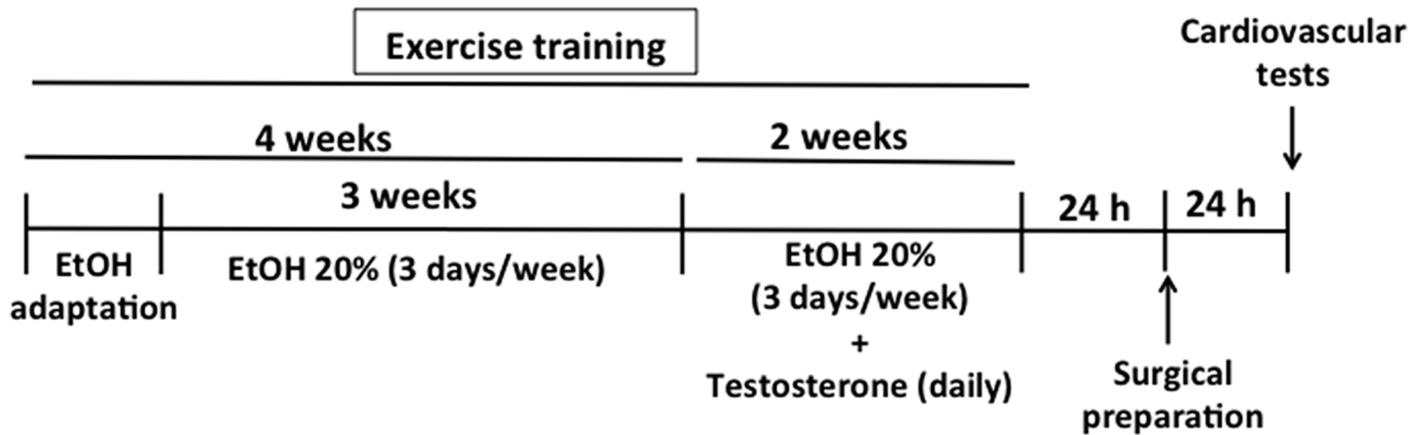


Fig 1. Schematic representation of the experimental protocol. Exercise training on the treadmill and ethanol treatment started on the same day and were realized for 6 weeks. During the first week (adaptation period), animals had continuous free access to two bottles, one containing ethanol and other containing water, and ethanol concentration was progressively increased daily until reach 20%. After this period, rats were given 24h concurrent access to one bottle containing 20% ethanol and other containing water three times a week (Monday, Wednesday, and Friday). In the last two weeks, animals underwent testosterone treatment concurrently with ethanol treatment and exercise training. Twenty-four hours after treatments and exercise training completion, animals in all experimental groups were subjected to surgical preparation, and the cardiovascular tests were performed 24 hours later. Rats had *ad libitum* food and water access throughout experimentation. EtOH—ethanol.

doi:10.1371/journal.pone.0146974.g001

Effects of exercise training and/or testosterone treatment on ethanol consumption

Analysis of ethanol intake before the onset of testosterone treatment indicated an effect over time ($F_{(5,202)} = 3, P < 0.01$), but without a significant effect of exercise training ($F_{(1,202)} = 0.03$,

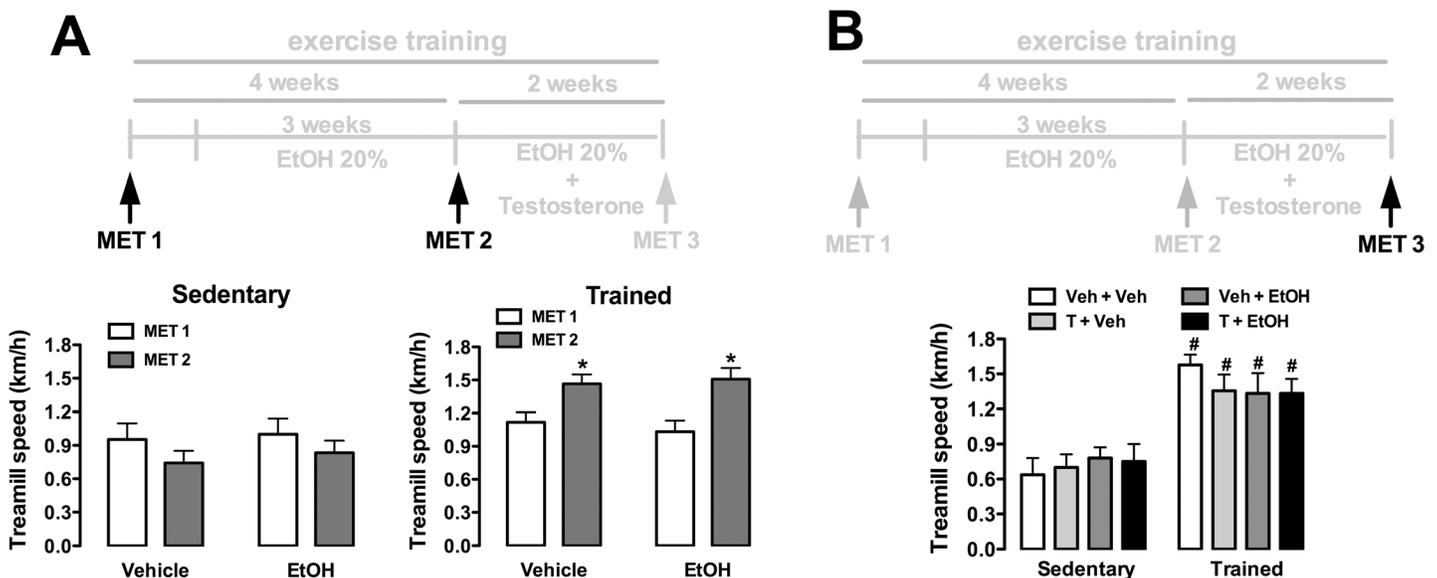


Fig 2. Maximal running speed (km/h) in maximal exercise tests (MET) in animals sedentary and subjected to exercise training on the treadmill (trained) treated with ethanol (EtOH) and/or testosterone (T). (A) Treadmill performance before the onset of T treatment in sedentary and trained animals treated with vehicle (water) or EtOH. The bars represent the mean \pm SEM. * $P < 0.05$ vs MET1. Three-way ANOVA followed by Bonferroni's *post hoc* test. ($n = 16-18$ /group). (B) Treadmill performance after completion of treatments and exercise training protocol in animals sedentary and trained treated with EtOH and/or T. The bars represent the mean \pm SEM. # $P < 0.05$ vs respective group sedentary. Two-way ANOVA followed by Bonferroni's *post hoc* test. ($n = 8-10$ /group).

doi:10.1371/journal.pone.0146974.g002

$P > 0.05$) and training x time interaction ($F_{(5,202)} = 1$, $P > 0.05$) (Fig 3A). Comparisons of ethanol consumption during testosterone treatment indicated an effect over time ($F_{(4,128)} = 8$, $P < 0.0001$), but without a significant effect of either exercise training ($F_{(1,32)} = 1$, $P > 0.05$) or testosterone treatment ($F_{(1,32)} = 1$, $P > 0.05$) (Fig 3B).

Effects of ethanol and/or testosterone treatment and exercise training in arterial pressure and hear rate

Analysis of both MAP, SAP, and DAP indicated no effect of either drug treatments (MAP: $F_{(3,59)} = 1$, $P > 0.05$; SAP: $F_{(3,59)} = 2$, $P > 0.05$; DAP: $F_{(3,59)} = 1$, $P > 0.05$) or exercise training (MAP: $F_{(1,59)} = 2$, $P > 0.05$; SAP: $F_{(1,59)} = 3$, $P > 0.05$; DAP: $F_{(1,59)} = 0.8$, $P > 0.05$) (Fig 4). However, analysis of HR indicated a main effect of drug treatments ($F_{(3,59)} = 8$, $P < 0.0003$), but without any influence of exercise training ($F_{(1,59)} = 0.01$, $P > 0.05$) and treatment x training interaction ($F_{(3,59)} = 0.6$, $P > 0.05$) (Fig 4). *Post-hoc* analysis revealed that testosterone treatment alone, but not in combination with ethanol ($P > 0.05$), reduced HR in sedentary animals ($P < 0.05$). This effect was not identified in trained rats ($P > 0.05$) (Fig 4).

Effects of ethanol and/or testosterone treatment and exercise training on baroreflex activity

Results of the analysis of baroreflex activity are presented in Fig 5. The analysis indicated significant influence of drug treatments (HR range: $F_{(3,59)} = 10$, $P < 0.0001$; BP_{50} : $F_{(3,59)} = 11$, $P < 0.0001$) and exercise training (HR range: $F_{(1,59)} = 17$, $P < 0.001$; BP_{50} : $F_{(1,59)} = 17$, $P < 0.0001$) as well as a treatment x training interaction (HR range: $F_{(3,59)} = 14$, $P < 0.0001$; BP_{50} : $F_{(3,59)} = 8$, $P < 0.0002$) for HR range and BP_{50} parameters. Analysis of P_1 and P_2 indicated a main effect of drug treatments (P_1 : $F_{(3,59)} = 5$, $P < 0.002$; P_2 : $F_{(3,59)} = 10$, $P < 0.0001$), but without influence of exercise (P_1 : $F_{(1,59)} = 0.1$, $P > 0.05$; P_2 : $F_{(1,59)} = 0.8$, $P > 0.05$) and treatment x training interaction (P_1 : $F_{(3,59)} = 2$, $P > 0.05$; P_2 : $F_{(3,59)} = 1$, $P > 0.05$). Analysis of the G indicated a main effect of exercise training ($F_{(1,59)} = 5$, $P < 0.03$) and a treatment x training interaction ($F_{(3,59)} = 3$, $P < 0.03$), but without influence of drug treatments ($F_{(3,59)} = 2$, $P > 0.05$). *Post-hoc* analysis revealed that exercise training reduced P_1 ($P < 0.05$) and increased G ($P < 0.05$) and HR range ($P < 0.05$), and these effects were not observed in animals treated with testosterone and/or ethanol ($P > 0.05$). Treatment with either testosterone or ethanol reduced both P_1 ($P < 0.05$) and P_2 ($P < 0.05$) in sedentary animals, but these effects were not identified in animals subjected to combined treatment with these substances ($P > 0.05$). Furthermore, exercise training restored all changes on baroreflex function evoked by either testosterone treatment or voluntary ethanol consumption.

Effects of ethanol and/or testosterone treatments and exercise training in arterial pressure changes evoked by vasoactive agents

Results of vascular reactivity to vasoactive agents are presented in Fig 6 and Table 1.

Phenylephrine. Intravenous infusion of the selective α_1 -adrenoceptor agonist phenylephrine dose-dependently increased arterial pressure in all experimental groups. Analysis of the E_{max} of the dose-response curves indicated a main effect of drug treatments ($F_{(3,59)} = 6$, $P < 0.002$), but without influence of exercise training ($F_{(1,59)} = 0.5$, $P > 0.05$) and treatment x training interaction ($F_{(3,59)} = 2$, $P > 0.05$). Comparison of ED_{50} values indicated a main effect of drug treatments ($F_{(3,59)} = 45$, $P < 0.0001$) and a treatment x training interaction ($F_{(3,59)} = 5$, $P < 0.006$), but without influence of the training ($F_{(1,59)} = 2$, $P > 0.05$). *Post-hoc* analysis revealed that combined treatment with testosterone and ethanol reduced E_{max} ($P < 0.05$) and increased

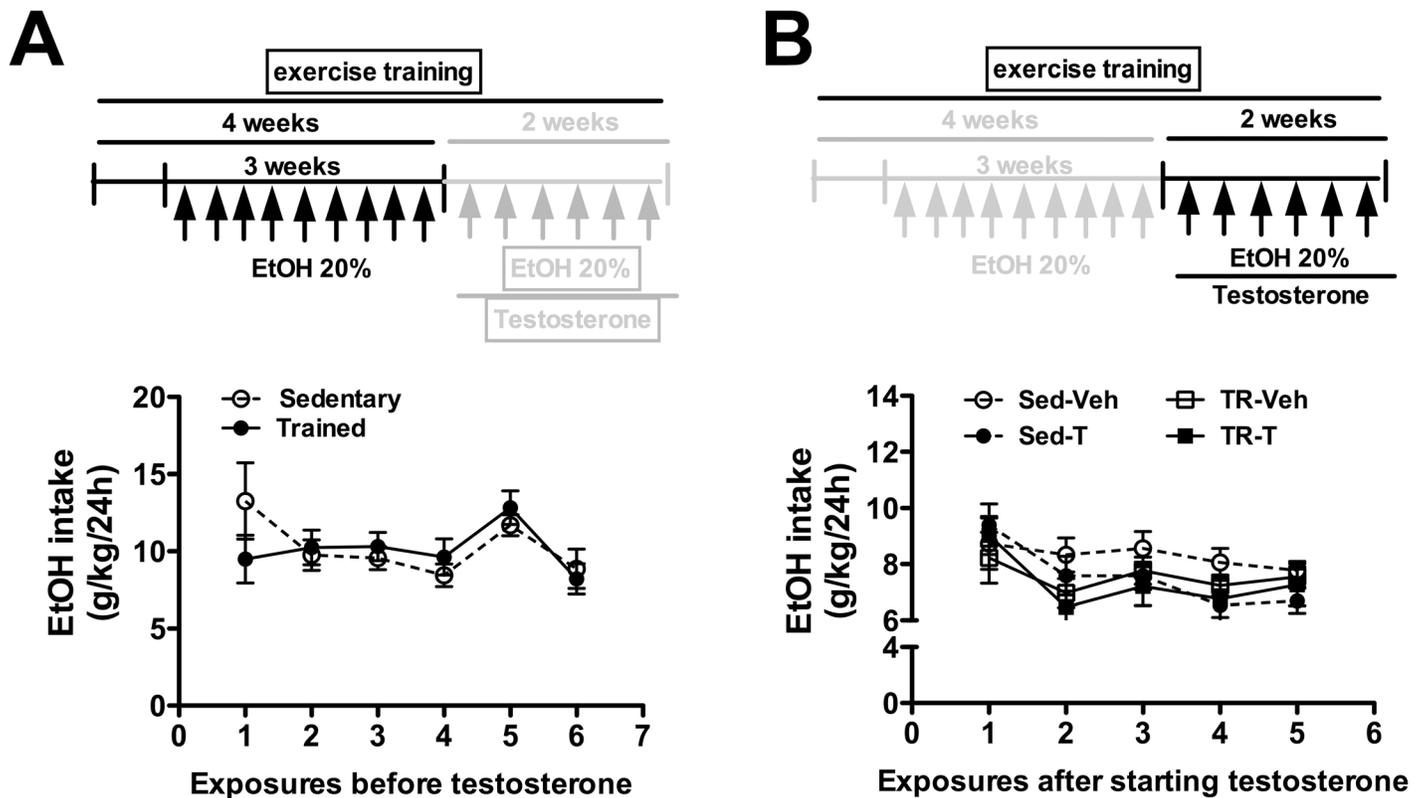


Fig 3. Voluntary ethanol intake (EtOH intake, g/kg/24h) in animals sedentary and subjected to exercise training on the treadmill (trained) treated with vehicle or testosterone (T). (A) Ethanol consumption before the onset of T treatment in sedentary and trained animals. The circles represent the mean \pm SEM. Two-way ANOVA followed by Bonferroni's *post hoc* test ($n = 16-18/\text{group}$). (B) EtOH intake during treatment with vehicle or T in animals sedentary and trained. The circles represent the mean \pm SEM. Three-way ANOVA followed by Bonferroni's *post hoc* test. ($n = 8-10/\text{group}$).

doi:10.1371/journal.pone.0146974.g003

ED₅₀ ($P < 0.05$) in sedentary animals. The effect in E_{max} ($P > 0.05$), but not ED₅₀ ($P < 0.05$), was restored by exercise training. Moreover, treatment with either testosterone or ethanol reduced ED₅₀ in trained rats ($P < 0.05$).

Acetylcholine. Intravenous infusion of acetylcholine dose-dependently reduced arterial pressure in all groups. Comparison of the E_{max} indicated a main effect of drug treatments ($F_{(3,59)} = 10, P < 0.0001$), but without influence of exercise ($F_{(1,59)} = 0.01, P > 0.05$) and treatment \times training interaction ($F_{(3,59)} = 1, P > 0.05$). Analysis of the ED₅₀ indicated a significant effect of drug treatments ($F_{(3,59)} = 5, P < 0.007$) and training ($F_{(1,59)} = 6, P < 0.02$), but without a treatment \times training interaction ($F_{(3,59)} = 0.2, P > 0.05$). *Post-hoc* analysis revealed that treatment with either testosterone ($P < 0.05$) or ethanol ($P < 0.05$) enhanced E_{max} in sedentary animals, whereas in trained animals only ethanol increased this parameter ($P < 0.05$).

Sodium nitroprusside. Systemic administration of the nitric oxide donor SNP dose-dependently reduced arterial pressure in all groups. Analysis of the E_{max} of the dose-response curves indicated a treatment \times training interaction ($F_{(3,59)} = 3, P < 0.04$), but without influence of either drug treatments ($F_{(3,59)} = 2, P > 0.05$) or training ($F_{(1,59)} = 0.8, P > 0.05$). Comparison of the ED₅₀ did not indicate a significant effect of either treatment ($F_{(3,59)} = 0.6, P > 0.05$) or training ($F_{(3,59)} = 0.01, P > 0.05$). *Post-hoc* analysis revealed that ethanol consumption increased E_{max} in trained animals ($P < 0.05$).

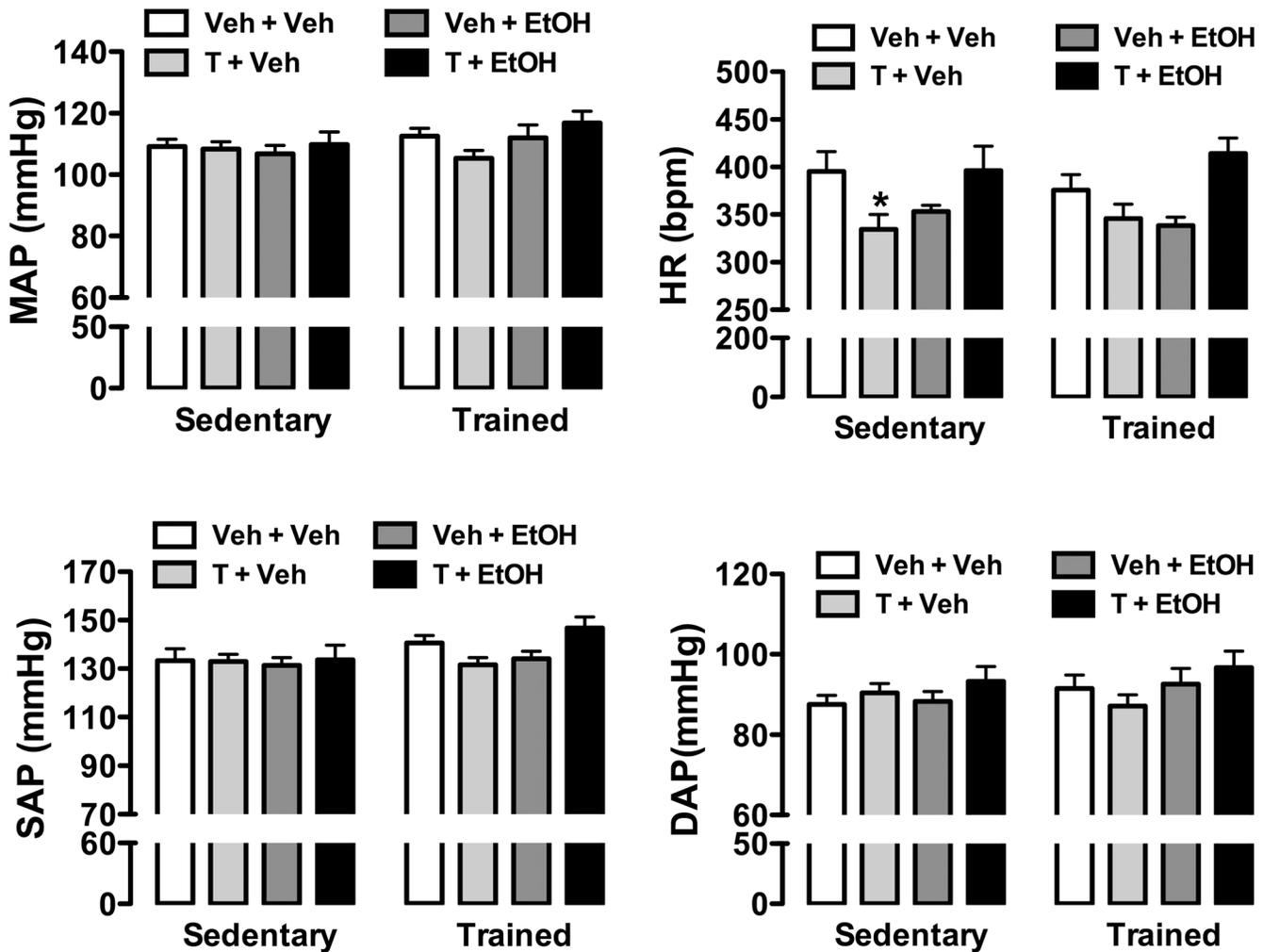


Fig 4. Mean (MAP), systolic (SAP), and diastolic (DAP) arterial pressure; and heart rate (HR) in animals sedentary and subjected to exercise training on the treadmill (trained) treated with ethanol (EtOH) and/or testosterone (T). The bars represent the mean±SEM. *P<0.05 vs respective group Veh+Veh within same condition. Two-way ANOVA followed by Bonferroni's *post hoc* test. (n = 8-9/group).

doi:10.1371/journal.pone.0146974.g004

Discussion

Present findings provide the first evidence of the effect of voluntary ethanol consumption combined with testosterone treatment on cardiovascular function of treadmill-trained rats. The main findings in the present study are: (i) exercise training improved the treadmill performance as evaluated in maximal exercise test, but neither ethanol consumption nor testosterone treatment affected this effect; (ii) voluntary ethanol consumption was not affected by either exercise training or repeated testosterone administration; (iii) voluntary ethanol consumption did not affect basal parameters of arterial pressure and HR, while testosterone treatment evoked resting bradycardia, which was not observed in animals submitted to combined treatment with ethanol or subjected to training on the treadmill; (iv) both testosterone treatment and voluntary ethanol consumption increased baroreflex-mediated bradycardia while tachycardia to blood pressure decrease was reduced. However, these baroreflex changes were not identified when substances were coadministered. Exercise training restored all alterations on

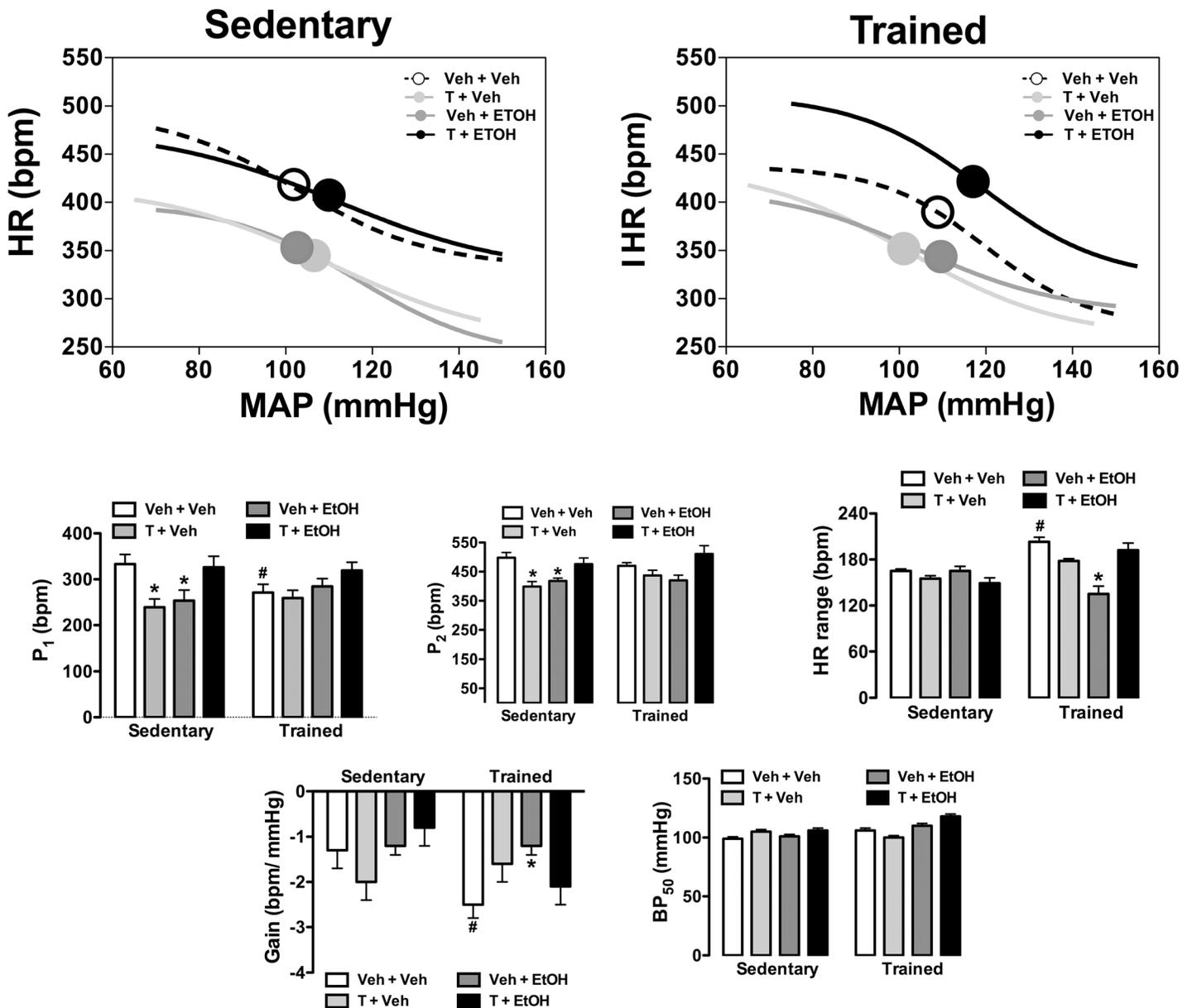


Fig 5. Analysis of baroreflex activity in animals sedentary and subjected to exercise training on the treadmill (trained) treated with ethanol (EtOH) and/or testosterone (T). (Top) Non-linear regression analysis of the baroreflex correlating mean arterial pressure change (MAP) evoked by intravenous infusion of phenylephrine and SNP and the reflex HR response (HR) in sedentary and trained animals treated with ethanol (EtOH) and/or testosterone (T). Symbols on sigmoid curves indicate the BP₅₀. (Bottom) Parameters derived from nonlinear regression analysis of the baroreflex in sedentary and trained animals treated with ethanol (EtOH) and/or testosterone (T). The bars represent the mean±SEM. *P<0.05 vs respective group Veh+Veh within same condition, #P<0.05 vs respective group sedentary. Two-way ANOVA followed by Bonferroni's *post hoc* test. (n = 8-9/group).

doi:10.1371/journal.pone.0146974.g005

baroreflex function; and (v) drug treatments affected vascular reactivity to vasoactive agents (see details below), which was influenced by exercise training.

Our findings are in line with previous data demonstrating an improvement in physical capacity following low-intensity training on treadmill [37,39]. Previous studies demonstrated that testosterone treatment increased the running wheel activity in hamsters and rats [40,41]. However, to the best of our knowledge, present study is the first evaluating the effect of

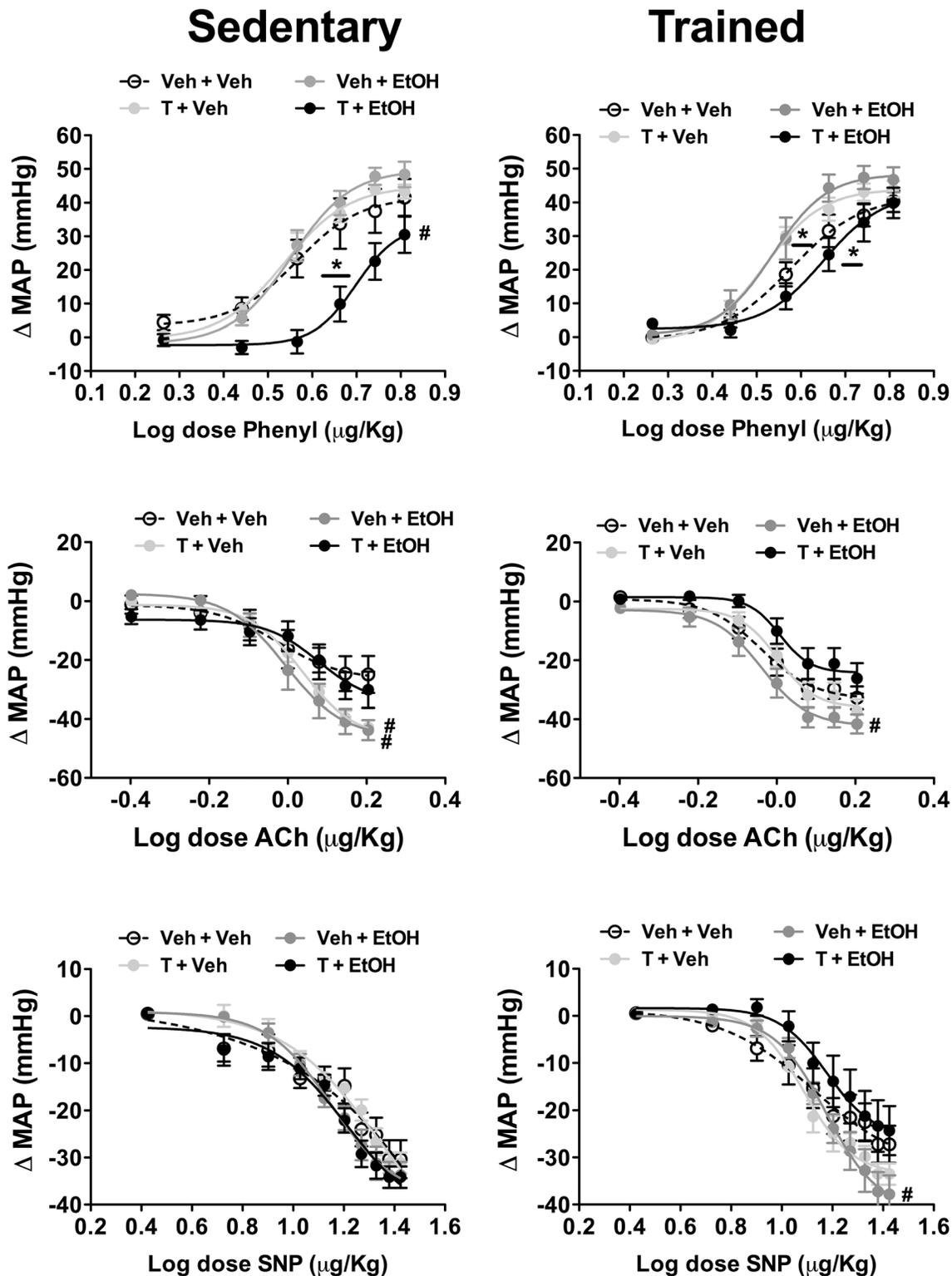


Fig 6. Mean arterial pressure change (Δ MAP) evoked by increasing concentrations of phenylephrine (Phenyl, top), acetylcholine (ACh, middle), and sodium nitroprusside (SNP, bottom) in in animals sedentary and subjected to exercise training on the treadmill (trained) treated with ethanol (EtOH) and/or testosterone (T). The circles represent the mean \pm SEM. * $P < 0.05$ vs respective Veh + Veh group within same condition for ED_{50} , # $P < 0.05$ vs respective Veh + Veh group within same condition for E_{max} . Nonlinear regression analysis. ($n = 8-9$ /group).

doi:10.1371/journal.pone.0146974.g006

Table 1. Maximal effect (E_{max}) and dose at 50% of the MAP range (ED_{50}) for phenylephrine (Phenyl), acetylcholine (Ach) and sodium nitroprusside (SNP) dose-response curves in animals sedentary and subjected to exercise training on the treadmill (trained) treated with ethanol (EtOH) and/or testosterone (T).

Group	Phenyl ED_{50}	E_{max}	Ach ED_{50}	E_{max}	SNP ED_{50}	E_{max}
Sedentary						
Veh + Veh	0.54±0.02	42±5	-0.03±0.04	-24±5	1.10±0.05	-31±4
T + Veh	0.53±0.007	43±2	0.02±0.005	-42±2*	1.17±0.02	-31±2
Veh + EtOH	0.54±0.008	49±3	-0.01±0.01	-43±3*	1.12±0.02	-33±2
T + EtOH	0.70±0.01*	29±4*	0.03±0.02	-30±5	1.11±0.01	-35±2
Trained						
Veh + Veh	0.57±0.01	40±3	-0.04±0.01	-32±3	1.10±0.04	-28±3
T + Veh	0.52±0.007*	44±2	-0.01±0.01	-38±3	1.09±0.01	-33±2
Veh + EtOH	0.53±0.01*	46±3	-0.05±0.01	-43±3*	1.15±0.02	-38±3*
T + EtOH	0.62±0.01* [#]	40±4	0.005±0.01	-27±3	1.15±0.02	-24±4

Values are mean ± SEM

* $P < 0.05$ vs respective Veh+Veh group within same condition

[#] $P < 0.05$ vs respective sedentary group. Two-way ANOVA followed by Bonferroni *post hoc* test.

doi:10.1371/journal.pone.0146974.t001

testosterone on treadmill performance in rodents. Results in humans demonstrated that hemodynamic and metabolic responses during an acute session of exercise on treadmill were impaired in AAS users [42]. Moreover, preclinical studies demonstrated that the cardiovascular beneficial effects of the exercise training were impaired by AAS administration in mice [43,44]. Thus, cardiovascular and metabolic negative effects may buffer a possible positive influence in performance related to anabolic actions of testosterone, thus explaining present findings. Regarding the ethanol, our findings are in line with earlier studies reporting that ethanol intake did not affect running wheel activity in rodents [24,25]. Present findings also corroborate clinical evidence that history of ethanol consumption does not affect exercise performance on treadmill [45].

The free-choice oral ethanol self-administration methods present face and construct validity as a model of human alcohol consumption once animals can choose whether to drink alcohol as well as the amount ingested over the time of exposure [33]. Therefore, in the present study we utilized the *intermittent access to 20% ethanol 2-bottle-choice drinking paradigm* for ethanol treatment [32]. The intermittent access to ethanol induces robust and reproducible levels of high voluntarily ethanol consumption over a long period of time without the use of any initiation procedures (e.g., sucrose fading or food and water deprivation) [32]. In fact, we detected ethanol intake in the range of 8–10 g/kg/24h throughout the experimental protocol, which is similar to those observed in alcohol-preferring rat strains [46] and in studies using protocols in which solution containing ethanol was the only source of liquid [47–50]. Previous results indicated that blood ethanol concentration following 30 minutes of voluntary ethanol consumption using the *intermittent-access to 20% ethanol drinking paradigm* ranged from 4 to 93 mg/dl in Wistar rats, and values significantly correlated with the amount of ethanol consumed [32]. Neither exercise nor testosterone affected ethanol consumption in the present study. Earlier studies reported that wheel running exercise reduced voluntary ethanol intake [23–27]. However, wheel running exercise has significant rewarding properties, so it has been proposed that a reduction in ethanol intake would be related to a substitution of rewarding effects of ethanol by hedonic properties of wheel running [24–26]. Regarding the influence of testosterone, previous studies have reported effect of AAS treatment in ethanol intake [17,18]. However, these studies

investigated ethanol consumption following either a single acute administration of AAS [17] or 1–3 weeks after completion of chronic AAS treatment [18]. Thus, differences in experimental protocol may explain the discrepancy.

Clinical and preclinical studies have demonstrated the development of hypertension following long-term ethanol consumption [3–6,51]. Most of the studies in animals evaluating the ethanol-evoked hypertension used models in which solutions containing high concentrations of ethanol were the only liquid source (e.g., [49,52–55]). In fact, to the best of our knowledge, present findings are the first investigating the impact of voluntary ethanol consumption on cardiovascular function in rodents. Therefore, differences in experimental procedures may explain the discrepancy between our findings and earlier studies. For instance, the ingestion of water through free access to a water bottle throughout the experimental procedure may buffer some ethanol effects, thus minimizing the cardiovascular consequences. Indeed, an increase in circulating vasopressin, possibly related to dehydration [56], has been implicated in ethanol-evoked hypertension in rodents [53]. A possible impact of voluntary ethanol consumption in circulating vasopressin and hematocrit deserve further investigations, but differences in impact of voluntary vs forced ethanol intake in these parameters may constitute possible mechanisms explaining the discrepancies. Moreover, contrary to continuous consumption in models of forced ethanol administration, in the present study ethanol treatment was intermittent, which may maximize the impact of behavioral and physiological compensatory mechanisms in ethanol-evoked hypertension.

The resting bradycardia following treatment with testosterone is in line with previous studies [11,13,57]. The mechanism underlying this effect remains unclear. However, steroids can cross the blood–brain barrier and expression of androgen receptors has been documented in brain areas regulating cardiovascular function [58]. Treatment with AAS has been related to an increase in sympathetic activity [59]. Thus, AAS-evoked bradycardia is likely mediated by an increase in cardiac parasympathetic tone rather than a reduction in sympathetic activity. Interestingly, ethanol consumption inhibited the resting bradycardia to testosterone, which may be related to a cardiac sympathoexcitation evoked by ethanol intake [60,61]. Exercise training abolished the testosterone-evoked bradycardia, which is in line with a recent report showing that the impairment of cardiac function evoked by AAS was completely restored by treadmill exercise training [62].

Exercise increased reflex bradycardia during blood pressure increase, which is in line with earlier evidence [63]. Previous studies demonstrated that arterial pressure and HR changes evoked by either testosterone or ethanol were followed by changes on baroreflex activity [3,13]. Accordingly, we observed that treatment with either ethanol or testosterone increased baroreflex-mediated bradycardia while tachycardia to blood pressure decrease was reduced. The baroreflex changes were not identified when substances were coadministered, thus further supporting an interaction between testosterone and ethanol on cardiovascular function. The baroreflex changes in testosterone-treated animals seems not to be related to the resting bradycardia since, for example, testosterone reduced the lower HR plateau (P_1) of baroreflex function by ~30% while basal HR was reduced by ~15%. Impairment of baroreflex activity has been implicated in the etiology and development of hypertension [64]. It was previously reported that forced ethanol intake impaired reflex bradycardia [8,54,65], and this effect has been implicated in ethanol-induced hypertension [8,50]. Therefore, discrepancy in effects on baroreflex function support the evidence of a different impact of voluntary vs forced ethanol intake on arterial pressure. Indeed, facilitation of baroreflex-mediated bradycardia may constitute a compensatory mechanism counteracting the emergence of hypertension induced by both voluntary ethanol consumption and testosterone treatment. Impairment of baroreflex activity is also

associated with overactivity of the sympathetic tone [66]. Thus, baroreflex changes do not seem to explain the resting bradycardia in testosterone-treated animals.

Training on the treadmill inhibited all changes on baroreflex function evoked by testosterone and ethanol treatments. As stated above, increased reflex bradycardia seems to be an important response counteracting arterial pressure changes, so that inhibition of these effects by exercise may be interpreted as a negative effect. Testosterone and ethanol also inhibited the effects of exercise training on baroreflex function as evidenced by inhibition of exercise-evoked facilitation of reflex bradycardia and increase in gain of baroreflex function, thus indicating that these substances may affect the beneficial cardiovascular effects of exercise training. These results are in line with previous data demonstrating that AAS suppressed cardiovascular adaptation to training [43,44]. Exercise training attenuated the hypertension induced by forced ethanol consumption [28,29]. However, to the best of our knowledge, present findings provide the first evidence of the impact of exercise on baroreflex function of ethanol-treated animals.

Testosterone treatment and ethanol consumption in combination, but not alone, reduced the pressor response to phenylephrine. A vascular hyperreactivity to vasoconstrictor agents has been associated with hypertension [67], so that reduced responsiveness to vasoconstrictor agents may be an important mechanism counteracting an increase on arterial pressure. The facilitation of depressor response to vasodilator agents following treatment with either testosterone or ethanol is in line with previous studies [8,13,54]. Our findings are also supported by evidence that testosterone acting directly in vascular wall induces relaxation of vascular smooth muscle [68]. Also, long-term ethanol consumption has been related to an inhibition of AChE activity [69,70], which can underlie the increased depressor response to acetylcholine in ethanol-treated animals. However, the absence of changes in pressor response to phenylephrine in ethanol-treated animals contrast with *in vitro* and *in vivo* studies reporting that forced ethanol consumption increased vascular reactivity to phenylephrine [4,8,50,54]. Nevertheless, these data are in line with arterial pressure results in which forced, but not voluntary, ethanol intake evoked hypertension.

Exercise training inhibited the reduction of pressor response to phenylephrine following combined treatment with testosterone and ethanol as well as the facilitation of depressor response to acetylcholine in testosterone-treated animals. These results contrast with the well-documented effects of exercise in increasing vascular nitric oxide availability and reducing vascular reactivity to α -adrenoceptor agonists [71]. A possible mechanism underlying the protector effect of training in these alterations on vascular reactivity can be an influence of exercise in pharmacokinetic of testosterone and ethanol, which in turn may affect the circulating levels of these substances. For instance, exercise training on treadmill can increase rates of ethanol clearance [72], while a decrease in metabolic clearance of testosterone was reported following an acute session of exercise [73]. The impact of training in testosterone clearance is less understood, but a study reported lower testosterone levels in runners vs sedentary subjects [74]. In addition, impairment in response of vascular relaxation to β -adrenoceptor agonist was reported following exercise training on treadmill [75]. Vasoconstrictor response to phenylephrine is counteracted by β_2 -adrenoceptor [76]. Thus, reduction in vasorelaxation response of β -adrenoceptors may also account to exercise effect in inhibiting the influence of the combined treatment with testosterone and ethanol in phenylephrine response. This mechanism may also underlie the facilitation in vascular responsiveness to phenylephrine observed in trained animals treated with either testosterone or ethanol. Facilitation of depressor response to SNP in trained rats treated with ethanol is line with evidence that exercise training increases vascular smooth muscle sensitivity to nitric oxide [71].

In summary, although combined treatment with testosterone and ethanol did not affect baseline arterial pressure and HR parameters, important changes on cardiovascular function

were identified, including reduction in pressor responsiveness to phenylephrine. Reduced vascular reactivity to vasoconstrictor agents may counteract other effects on cardiovascular function so inhibiting the emergence of changes in arterial pressure. Effects of treatment with testosterone and ethanol alone on baroreflex activity and depressor response to acetylcholine were inhibited when substances were coadministered. These results provide evidence that these substances are capable of mutually inhibiting the cardiovascular effects of each other, thus further supporting an interaction between their toxic effects on cardiovascular function. Regarding the influence of exercise training, we observed that training on the treadmill inhibited cardiovascular effects of drug treatments, but some effects were identified only in trained animals. Taken together, these results indicate the exercise as an important factor affecting the effects of ethanol and testosterone on cardiovascular function. Indeed, present data suggest that cardiovascular effects of these substances may be related, at least in part, to an inhibition of protector effects of exercise training on cardiovascular function.

Acknowledgments

The authors wish to thank Elisabete Z. P. Lepera and Rosana F. P. Silva for technical assistance. This work was supported by São Paulo Research Foundation (FAPESP) grants # 2013/09715-2, 2012/14723-1, and 2012/14376-0; National Counsel of Technological and Scientific Development (CNPq) grant #456405/2014-3, and PADC-FCF UNESP. CSP is a CNPq research fellow.

Author Contributions

Conceived and designed the experiments: SAE CSP CCC. Performed the experiments: SAE. Analyzed the data: SAE CSP CCC. Contributed reagents/materials/analysis tools: CSP CCC. Wrote the paper: SAE CSP CCC.

References

1. Whiteford HA, Degenhardt L, Rehm J, Baxter AJ, Ferrari AJ, Erskine HE, et al. (2013) Global burden of disease attributable to mental and substance use disorders: findings from the Global Burden of Disease Study 2010. *Lancet*.
2. Murray CJ, Vos T, Lozano R, Naghavi M, Flaxman AD, Michaud C, et al. (2012) Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 380: 2197–2223. doi: [10.1016/S0140-6736\(12\)61689-4](https://doi.org/10.1016/S0140-6736(12)61689-4) PMID: [23245608](https://pubmed.ncbi.nlm.nih.gov/23245608/)
3. Husain K, Ansari RA, Ferder L (2014) Alcohol-induced hypertension: Mechanism and prevention. *World J Cardiol* 6: 245–252. PMID: [24891935](https://pubmed.ncbi.nlm.nih.gov/24891935/)
4. Marchi KC, Muniz JJ, Tirapelli CR (2014) Hypertension and chronic ethanol consumption: What do we know after a century of study? *World J Cardiol* 6: 283–294. PMID: [24944758](https://pubmed.ncbi.nlm.nih.gov/24944758/)
5. Fernandez-Sola J (2015) Cardiovascular risks and benefits of moderate and heavy alcohol consumption. *Nat Rev Cardiol*.
6. Kawano Y (2010) Physio-pathological effects of alcohol on the cardiovascular system: its role in hypertension and cardiovascular disease. *Hypertens Res* 33: 181–191. doi: [10.1038/hr.2009.226](https://doi.org/10.1038/hr.2009.226) PMID: [20075936](https://pubmed.ncbi.nlm.nih.gov/20075936/)
7. Puddey IB, Zilkens RR, Croft KD, Beilin LJ (2001) Alcohol and endothelial function: a brief review. *Clin Exp Pharmacol Physiol* 28: 1020–1024. PMID: [11903307](https://pubmed.ncbi.nlm.nih.gov/11903307/)
8. Crestani CC, Lopes da Silva A, Scopinho AA, Ruginsk SG, Uchoa ET, Correa FM, et al. (2014) Cardiovascular alterations at different stages of hypertension development during ethanol consumption: time-course of vascular and autonomic changes. *Toxicol Appl Pharmacol* 280: 245–255. doi: [10.1016/j.taap.2014.08.012](https://doi.org/10.1016/j.taap.2014.08.012) PMID: [25151222](https://pubmed.ncbi.nlm.nih.gov/25151222/)
9. Kanayama G, Brower KJ, Wood RI, Hudson JI, Pope HG Jr. (2009) Anabolic-androgenic steroid dependence: an emerging disorder. *Addiction* 104: 1966–1978. doi: [10.1111/j.1360-0443.2009.02734.x](https://doi.org/10.1111/j.1360-0443.2009.02734.x) PMID: [19922565](https://pubmed.ncbi.nlm.nih.gov/19922565/)
10. UNODC (2010) World Drug Report 2010. United Nations Publication Sales No. E.10.XI.13.

11. Beutel A, Bergamaschi CT, Campos RR (2005) Effects of chronic anabolic steroid treatment on tonic and reflex cardiovascular control in male rats. *J Steroid Biochem Mol Biol* 93: 43–48. PMID: [15748831](#)
12. van Amsterdam J, Opperhuizen A, Hartgens F (2010) Adverse health effects of anabolic-androgenic steroids. *Regul Toxicol Pharmacol* 57: 117–123. doi: [10.1016/j.yrtph.2010.02.001](#) PMID: [20153798](#)
13. Engi SA, Cruz FC, Leao RM, Correa FM, Planeta CS, Crestani CC (2012) Effect of the single or combined administration of cocaine and testosterone on cardiovascular function and baroreflex activity in unanesthetized rats. *J Cardiovasc Pharmacol* 59: 231–240. doi: [10.1097/FJC.0b013e31823cc58b](#) PMID: [22030898](#)
14. Meilman PW, Crace RK, Presley CA, Lyerla R (1995) Beyond performance enhancement: polypharmacy among collegiate users of steroids. *J Am Coll Health* 44: 98–104. PMID: [8543731](#)
15. Yesalis CE, Kennedy NJ, Kopstein AN, Bahrke MS (1993) Anabolic-androgenic steroid use in the United States. *JAMA* 270: 1217–1221. PMID: [8355384](#)
16. McCabe SE, Brower KJ, West BT, Nelson TF, Wechsler H (2007) Trends in non-medical use of anabolic steroids by U.S. college students: results from four national surveys. *Drug Alcohol Depend* 90: 243–251. PMID: [17512138](#)
17. Gurkovskaya OV, Leonard ST, Lewis PB, Winsauer PJ (2009) Effects of pregnanolone and dehydroepiandrosterone on ethanol intake in rats administered ethanol or saline during adolescence. *Alcohol Clin Exp Res* 33: 1252–1264. doi: [10.1111/j.1530-0277.2009.00951.x](#) PMID: [19389187](#)
18. Johansson P, Lindqvist A, Nyberg F, Fahlke C (2000) Anabolic androgenic steroids affects alcohol intake, defensive behaviors and brain opioid peptides in the rat. *Pharmacol Biochem Behav* 67: 271–279. PMID: [11124391](#)
19. Lakoza GN, Barkov NK (1980) The role of testosterone in the development of experimental alcoholism. *Bull Narc* 32: 41–48. PMID: [6907025](#)
20. Piazza-Gardner AK, Barry AE (2012) Examining physical activity levels and alcohol consumption: are people who drink more active? *Am J Health Promot* 26: e95–104. doi: [10.4278/ajhp.100929-LIT-328](#) PMID: [22208422](#)
21. Barry AE, Piazza-Gardner AK (2012) Drunkorexia: understanding the co-occurrence of alcohol consumption and eating/exercise weight management behaviors. *J Am Coll Health* 60: 236–243. doi: [10.1080/07448481.2011.587487](#) PMID: [22420701](#)
22. French MT, Popovici I, Maclean JC (2009) Do alcohol consumers exercise more? Findings from a national survey. *Am J Health Promot* 24: 2–10. doi: [10.4278/ajhp.0801104](#) PMID: [19750956](#)
23. Brager AJ, Hammer SB (2012) Impact of wheel running on chronic ethanol intake in aged Syrian hamsters. *Physiol Behav* 107: 418–423. doi: [10.1016/j.physbeh.2012.09.011](#) PMID: [23022151](#)
24. Darlington TM, McCarthy RD, Cox RJ, Ehringer MA (2014) Mesolimbic transcriptional response to hedonic substitution of voluntary exercise and voluntary ethanol consumption. *Behav Brain Res* 259: 313–320. doi: [10.1016/j.bbr.2013.11.011](#) PMID: [24239693](#)
25. Ehringer MA, Hoft NR, Zunhammer M (2009) Reduced alcohol consumption in mice with access to a running wheel. *Alcohol* 43: 443–452. doi: [10.1016/j.alcohol.2009.06.003](#) PMID: [19801274](#)
26. Hammer SB, Ruby CL, Brager AJ, Prosser RA, Glass JD (2010) Environmental modulation of alcohol intake in hamsters: effects of wheel running and constant light exposure. *Alcohol Clin Exp Res* 34: 1651–1658. doi: [10.1111/j.1530-0277.2010.01251.x](#) PMID: [20569242](#)
27. McMillan DE, McClure GY, Hardwick WC (1995) Effects of access to a running wheel on food, water and ethanol intake in rats bred to accept ethanol. *Drug Alcohol Depend* 40: 1–7. PMID: [8746918](#)
28. Husain K, Mejia J, Lalla J (2006) Physiological basis for effect of physical conditioning on chronic ethanol-induced hypertension in a rat model. *Mol Cell Biochem* 289: 175–183. PMID: [16718371](#)
29. Husain K, Vazquez Ortiz M, Lalla J (2006) Physical training ameliorates chronic alcohol-induced hypertension and aortic reactivity in rats. *Alcohol Alcohol* 41: 247–253. PMID: [16467407](#)
30. Bahrke MS, Yesalis CE (2004) Abuse of anabolic androgenic steroids and related substances in sport and exercise. *Curr Opin Pharmacol* 4: 614–620. PMID: [15525553](#)
31. Rocha FL, Carmo EC, Roque FR, Hashimoto NY, Rossoni LV, Frimm C, et al. (2007) Anabolic steroids induce cardiac renin-angiotensin system and impair the beneficial effects of aerobic training in rats. *Am J Physiol Heart Circ Physiol* 293: H3575–3583. PMID: [17906098](#)
32. Simms JA, Steensland P, Medina B, Abernathy KE, Chandler LJ, Wise R, et al. (2008) Intermittent access to 20% ethanol induces high ethanol consumption in Long-Evans and Wistar rats. *Alcohol Clin Exp Res* 32: 1816–1823. doi: [10.1111/j.1530-0277.2008.00753.x](#) PMID: [18671810](#)
33. Sanchis-Segura C, Spanagel R (2006) Behavioural assessment of drug reinforcement and addictive features in rodents: an overview. *Addict Biol* 11: 2–38. PMID: [16759333](#)

34. Cruz FC, Alves FH, Leao RM, Planeta CS, Crestani CC (2013) Role of the bed nucleus of the stria terminalis in cardiovascular changes following chronic treatment with cocaine and testosterone: a role beyond drug seeking in addiction? *Neuroscience* 253: 29–39. doi: [10.1016/j.neuroscience.2013.08.034](https://doi.org/10.1016/j.neuroscience.2013.08.034) PMID: [23994153](https://pubmed.ncbi.nlm.nih.gov/23994153/)
35. Engi SA, Cruz FC, Leao RM, Spolidorio LC, Planeta CS, Crestani CC (2014) Cardiovascular complications following chronic treatment with cocaine and testosterone in adolescent rats. *PLoS One* 9: e105172. PMID: [25121974](https://pubmed.ncbi.nlm.nih.gov/25121974/)
36. Camargo LH, Alves FH, Biojone C, Correa FM, Resstel LB, Crestani CC (2013) Involvement of N-methyl-D-aspartate glutamate receptor and nitric oxide in cardiovascular responses to dynamic exercise in rats. *Eur J Pharmacol* 713: 16–24. doi: [10.1016/j.ejphar.2013.04.046](https://doi.org/10.1016/j.ejphar.2013.04.046) PMID: [23680118](https://pubmed.ncbi.nlm.nih.gov/23680118/)
37. Masson GS, Costa TS, Yshii L, Fernandes DC, Soares PP, Laurindo FR, et al. (2014) Time-dependent effects of training on cardiovascular control in spontaneously hypertensive rats: role for brain oxidative stress and inflammation and baroreflex sensitivity. *PLoS One* 9: e94927. PMID: [24788542](https://pubmed.ncbi.nlm.nih.gov/24788542/)
38. Crestani CC, Alves FH, Busnardo C, Resstel LB, Correa FM (2010) N-methyl-D-aspartate glutamate receptors in the hypothalamic paraventricular nucleus modulate cardiac component of the baroreflex in unanesthetized rats. *Neurosci Res* 67: 317–326. doi: [10.1016/j.neures.2010.05.001](https://doi.org/10.1016/j.neures.2010.05.001) PMID: [20472007](https://pubmed.ncbi.nlm.nih.gov/20472007/)
39. Sanches IC, Conti FF, Bernardes N, Brito JO, Caldini EG, Cavaglieri CR, et al. (2015) Impact of Combined Exercise Training on Cardiovascular Autonomic Control and Mortality in Diabetic Ovariectomized Rats. *J Appl Physiol* (1985): jap 00883 02014.
40. McGinnis MY, Lumia AR, Tetel MJ, Molenda-Figueira HA, Possidente B (2007) Effects of anabolic androgenic steroids on the development and expression of running wheel activity and circadian rhythms in male rats. *Physiol Behav* 92: 1010–1018. PMID: [17716697](https://pubmed.ncbi.nlm.nih.gov/17716697/)
41. Wood RI (2002) Oral testosterone self-administration in male hamsters: dose-response, voluntary exercise, and individual differences. *Horm Behav* 41: 247–258. PMID: [11971658](https://pubmed.ncbi.nlm.nih.gov/11971658/)
42. Maior AS, Simao R, de Salles BF, Alexander JL, Rhea M, Nascimento JH (2010) Acute cardiovascular response in anabolic androgenic steroid users performing maximal treadmill exercise testing. *J Strength Cond Res* 24: 1688–1695. doi: [10.1519/JSC.0b013e3181dc46c9](https://doi.org/10.1519/JSC.0b013e3181dc46c9) PMID: [20508475](https://pubmed.ncbi.nlm.nih.gov/20508475/)
43. Tagarakis CV, Bloch W, Hartmann G, Hollmann W, Addicks K (2000) Testosterone-propionate impairs the response of the cardiac capillary bed to exercise. *Med Sci Sports Exerc* 32: 946–953. PMID: [10795785](https://pubmed.ncbi.nlm.nih.gov/10795785/)
44. Sun M, Shen W, Zhong M, Wu P, Chen H, Lu A (2013) Nandrolone attenuates aortic adaptation to exercise in rats. *Cardiovasc Res* 97: 686–695. doi: [10.1093/cvr/cvs423](https://doi.org/10.1093/cvr/cvs423) PMID: [23338851](https://pubmed.ncbi.nlm.nih.gov/23338851/)
45. Hartung GH, Kohl HW, Blair SN, Lawrence SJ, Harrist RB (1990) Exercise tolerance and alcohol intake. Blood pressure relation. *Hypertension* 16: 501–507. PMID: [2228150](https://pubmed.ncbi.nlm.nih.gov/2228150/)
46. Bell RL, Rodd ZA, Lumeng L, Murphy JM, McBride WJ (2006) The alcohol-preferring P rat and animal models of excessive alcohol drinking. *Addict Biol* 11: 270–288. PMID: [16961759](https://pubmed.ncbi.nlm.nih.gov/16961759/)
47. Abdel-Rahman AA, Wooles WR (1987) Ethanol-induced hypertension involves impairment of baroreceptors. *Hypertension* 10: 67–73. PMID: [3596770](https://pubmed.ncbi.nlm.nih.gov/3596770/)
48. Kahonen M, Karjala K, Hutri-Kahonen N, Wu X, Jaatinen P, Riihioja P, et al. (1999) Influence of chronic ethanol consumption on arterial tone in young and aged rats. *Am J Physiol* 276: H464–471. PMID: [9950846](https://pubmed.ncbi.nlm.nih.gov/9950846/)
49. Ladipo CO, Adigun SA, Nwaigwe CI, Adegunloye BJ (2002) Chronic ethanol consumption alters vascular smooth muscle responses in rats. *Clin Exp Pharmacol Physiol* 29: 707–709. PMID: [12100004](https://pubmed.ncbi.nlm.nih.gov/12100004/)
50. Russ R, Abdel-Rahman AR, Wooles WR (1991) Role of the sympathetic nervous system in ethanol-induced hypertension in rats. *Alcohol* 8: 301–307. PMID: [1872991](https://pubmed.ncbi.nlm.nih.gov/1872991/)
51. Miller PM, Anton RF, Egan BM, Basile J, Nguyen SA (2005) Excessive alcohol consumption and hypertension: clinical implications of current research. *J Clin Hypertens (Greenwich)* 7: 346–351.
52. Chan TC, Sutter MC (1983) Ethanol consumption and blood pressure. *Life Sci* 33: 1965–1973. PMID: [6685805](https://pubmed.ncbi.nlm.nih.gov/6685805/)
53. Resstel LB, Scopinho AA, Lopes da Silva A, Antunes-Rodrigues J, Correa FM (2008) Increased circulating vasopressin may account for ethanol-induced hypertension in rats. *Am J Hypertens* 21: 930–935. doi: [10.1038/ajh.2008.189](https://doi.org/10.1038/ajh.2008.189) PMID: [18464746](https://pubmed.ncbi.nlm.nih.gov/18464746/)
54. Resstel LB, Tirapelli CR, Lanchote VL, Uyemura SA, de Oliveira AM, Correa FM (2006) Chronic ethanol consumption alters cardiovascular functions in conscious rats. *Life Sci* 78: 2179–2187. PMID: [16288925](https://pubmed.ncbi.nlm.nih.gov/16288925/)
55. Tirapelli CR, Legros E, Brochu I, Honore JC, Lanchote VL, Uyemura SA, et al. (2008) Chronic ethanol intake modulates vascular levels of endothelin-1 receptor and enhances the pressor response to endothelin-1 in anaesthetized rats. *Br J Pharmacol* 154: 971–981. doi: [10.1038/bjp.2008.157](https://doi.org/10.1038/bjp.2008.157) PMID: [18469849](https://pubmed.ncbi.nlm.nih.gov/18469849/)

56. Da Silva AL, Ruginsk SG, Uchoa ET, Crestani CC, Scopinho AA, Correa FM, et al. (2013) Time-course of neuroendocrine changes and its correlation with hypertension induced by ethanol consumption. *Alcohol Alcohol* 48: 495–504. doi: [10.1093/alcalc/agt040](https://doi.org/10.1093/alcalc/agt040) PMID: [23733506](https://pubmed.ncbi.nlm.nih.gov/23733506/)
57. Cruz FC, Engi SA, Leao RM, Planeta CS, Crestani CC (2012) Influence of the single or combined administration of cocaine and testosterone in autonomic and neuroendocrine responses to acute restraint stress. *J Psychopharmacol* 26: 1366–1374. doi: [10.1177/0269881112453210](https://doi.org/10.1177/0269881112453210) PMID: [22767371](https://pubmed.ncbi.nlm.nih.gov/22767371/)
58. Gorlick DL, Kelley DB (1986) The ontogeny of androgen receptors in the CNS of *Xenopus laevis* frogs. *Brain Res* 391: 193–200. PMID: [3697774](https://pubmed.ncbi.nlm.nih.gov/3697774/)
59. Kumai T, Tanaka M, Watanabe M, Matsumoto C, Kobayashi S (1994) Possible involvement of androgen in increased norepinephrine synthesis in blood vessels of spontaneously hypertensive rats. *Jpn J Pharmacol* 66: 439–444. PMID: [7723220](https://pubmed.ncbi.nlm.nih.gov/7723220/)
60. Zhang X, Abdel-Rahman AR, Wooles WR (1988) A differential action for ethanol on baroreceptor reflex control of heart rate and sympathetic efferent discharge in rats. *Proc Soc Exp Biol Med* 187: 14–21. PMID: [3340614](https://pubmed.ncbi.nlm.nih.gov/3340614/)
61. Sparrow MG, Roggendorf H, Vogel WH (1987) Effect of ethanol on heart rate and blood pressure in nonstressed and stressed rats. *Life Sci* 40: 2551–2559. PMID: [3600169](https://pubmed.ncbi.nlm.nih.gov/3600169/)
62. Bocalini DS, Beutel A, Bergamaschi CT, Tucci PJ, Campos RR (2014) Treadmill exercise training prevents myocardial mechanical dysfunction induced by androgenic-anabolic steroid treatment in rats. *PLoS One* 9: e87106. PMID: [24533053](https://pubmed.ncbi.nlm.nih.gov/24533053/)
63. Chen CY, DiCarlo SE (1996) Daily exercise and gender influence arterial baroreflex regulation of heart rate and nerve activity. *Am J Physiol* 271: H1840–1848. PMID: [8945899](https://pubmed.ncbi.nlm.nih.gov/8945899/)
64. Grassi G, Trevano FQ, Seravalle G, Scopelliti F, Mancia G (2006) Baroreflex function in hypertension: consequences for antihypertensive therapy. *Prog Cardiovasc Dis* 48: 407–415. PMID: [16714160](https://pubmed.ncbi.nlm.nih.gov/16714160/)
65. Abdel-Rahman AR, Dar MS, Wooles WR (1985) Effect of chronic ethanol administration on arterial baroreceptor function and pressor and depressor responsiveness in rats. *J Pharmacol Exp Ther* 232: 194–201. PMID: [4038417](https://pubmed.ncbi.nlm.nih.gov/4038417/)
66. Grassi G, Seravalle G, Dell'Oro R, Facchini A, Ilardo V, Mancia G (2004) Sympathetic and baroreflex function in hypertensive or heart failure patients with ventricular arrhythmias. *J Hypertens* 22: 1747–1753. PMID: [15311103](https://pubmed.ncbi.nlm.nih.gov/15311103/)
67. Chang HR, Lee RP, Wu CY, Chen HI (2002) Nitric oxide in mesenteric vascular reactivity: a comparison between rats with normotension and hypertension. *Clin Exp Pharmacol Physiol* 29: 275–280. PMID: [11985535](https://pubmed.ncbi.nlm.nih.gov/11985535/)
68. Perusquia M, Stallone JN (2010) Do androgens play a beneficial role in the regulation of vascular tone? Nongenomic vascular effects of testosterone metabolites. *Am J Physiol Heart Circ Physiol* 298: H1301–1307. doi: [10.1152/ajpheart.00753.2009](https://doi.org/10.1152/ajpheart.00753.2009) PMID: [20228257](https://pubmed.ncbi.nlm.nih.gov/20228257/)
69. Haboubi NA, Thurnham DI (1986) Effect of ethanol on erythrocyte acetylcholinesterase activity. *Ann Clin Biochem* 23 (Pt 4): 458–462. PMID: [3767274](https://pubmed.ncbi.nlm.nih.gov/3767274/)
70. Husain K, Somani SM (1998) Effect of exercise training and chronic ethanol ingestion on cholinesterase activity and lipid peroxidation in blood and brain regions of rat. *Prog Neuropsychopharmacol Biol Psychiatry* 22: 411–423. PMID: [9608610](https://pubmed.ncbi.nlm.nih.gov/9608610/)
71. McAllister RM, Jasperse JL, Laughlin MH (2005) Nonuniform effects of endurance exercise training on vasodilation in rat skeletal muscle. *J Appl Physiol* (1985) 98: 753–761.
72. Ardies CM, Morris GS, Erickson CK, Farrar RP (1989) Both acute and chronic exercise enhance in vivo ethanol clearance in rats. *J Appl Physiol* (1985) 66: 555–560.
73. Cadoux-Hudson TA, Few JD, Imms FJ (1985) The effect of exercise on the production and clearance of testosterone in well trained young men. *Eur J Appl Physiol Occup Physiol* 54: 321–325. PMID: [4065118](https://pubmed.ncbi.nlm.nih.gov/4065118/)
74. Wheeler GD, Wall SR, Belcastro AN, Cumming DC (1984) Reduced serum testosterone and prolactin levels in male distance runners. *JAMA* 252: 514–516. PMID: [6429357](https://pubmed.ncbi.nlm.nih.gov/6429357/)
75. Rogers PJ, Miller TD, Bauer BA, Brum JM, Bove AA, Vanhoutte PM (1991) Exercise training and responsiveness of isolated coronary arteries. *J Appl Physiol* (1985) 71: 2346–2351.
76. Davel AP, Ceravolo GS, Wenceslau CF, Carvalho MH, Brum PC, Rossoni LV (2012) Increased vascular contractility and oxidative stress in beta(2)-adrenoceptor knockout mice: the role of NADPH oxidase. *J Vasc Res* 49: 342–352. doi: [10.1159/000337486](https://doi.org/10.1159/000337486) PMID: [22627472](https://pubmed.ncbi.nlm.nih.gov/22627472/)