

Teaching Point
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An immunocompromised dialysis patient with skin and bone lesions

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Introduction

Patients with chronic kidney failure have disturbances of immune function involving both innate and adaptive systems. These result in both immunodepression which increases susceptibility to infection and immunoactivation leading to a chronic inflammatory state [1]. Dialysis treatment may further aggravate aspects of this, especially the induction of inflammation. In addition, there is a growing number of patients who have returned to dialysis programmes following transplant failure. Many of these will have been intensively immunosuppressed, often over years or decades. These patients are at particular risk.

Case report

A 60-year-old haemodialysis (HD) patient was admitted for investigation of a papulo-nodular rash predominantly affecting the extensor aspects of both arms (Figure 1). He had adult polycystic kidney disease and had developed end-stage renal failure 15 years previously, at which point he had commenced on HD. He had received a cadaveric renal transplant 12 years ago which had functioned for 10 years. For the past 2 years, he had been back on HD. Other past medical history included ischaemic heart disease, post-transplant diabetes mellitus and hypoadrenalism secondary to long-term steroid treatment. He had undergone splenectomy following abdominal trauma in the past. Two months prior to this admission, he had been hospitalized with abdominal pain and found to have a right paracolic abscess. This had been treated with drainage and antibiotics. At the time of this admission, he was receiving replacement doses of hydrocortisone, tacrolimus having been discontinued 18 months previously.

Shortly after admission, he sustained a fall. Subsequent radiographs showed pathological fractures of the right

ankle and of the middle phalanx of the left fifth finger (Figure 2). A little later, he developed severe rectal bleeding which failed to respond to medical management. Oesophagogastroduodenoscopy was unremarkable, but flexible sigmoidoscopy revealed fresh blood extending to the splenic flexure, suggesting a colonic cause. A mesenteric angiogram did not demonstrate active bleeding. He subsequently underwent sub-total colectomy with resolution of the bleeding.

Biopsies of the skin lesions showed non-caseating granulomata with central microabscesses and rod-like bacteria which were positive for Ziehl–Neelsen staining. A mycobacterial species was isolated after 5 days from a skin biopsy, suggestive of a rapidly growing atypical mycobacterium. The mycobacterium was subsequently identified as *Mycobacterium chelonae*. Histology of the appendix also demonstrated granulomatous inflammation, but staining for mycobacteria was negative. The radiographs of the right ankle and left hand were interpreted as compatible with mycobacterial infection of the bone causing pathological fracture of the ankle and dactylitis of the fifth middle finger. A diagnosis of disseminated atypical mycobacterial disease was made based on proven



Fig. 1. Skin involvement of disseminated disease with *M. chelonae* manifesting in papulo-nodular lesions on the forearm.



Fig. 2. Radiograph of the left hand demonstrating a pathological fracture through the middle phalanx of the little finger.

skin involvement and probable infection of bone and appendix. The colon exhibited mucosal ulceration but no granulomata. Considering the recent diagnosis of a disseminated opportunistic infection, haemorrhagic colitis secondary to co-existent cytomegalovirus (CMV) infection was felt to be highly likely. Indeed, CMV polymerase chain reaction (PCR) using blood was strongly positive.

He was treated with intravenous then oral clarithromycin and intravenous tobramycin [2]. In addition, he received intravenous ganciclovir then oral valganciclovir for the CMV infection. Subsequently, the skin lesions slowly resolved. There was also healing of the fractures. In addition, following 2 months of antiviral treatment, the CMV-PCR became negative. However, 4 months later, the patient died as a result of dialysis access failure.

Discussion

Infections with atypical mycobacteria, also known as non-tuberculous mycobacteria, are caused by a wide spectrum of ubiquitous acid-fast organisms, many of which have only recently been identified as pathogenic [3]. Infections with atypical mycobacteria commonly occur in the immunocompromised host, but can also manifest in immunocompetent individuals. The clinical picture varies depending on the species and includes soft tissue infections, chronic pulmonary and osteo-articular infections as well as disseminated disease. For many species, there is limited data on which to base treatment recommendations and the role of antimicrobial susceptibility tests is controversial [2]. Multi-drug combinations are often recommended to avoid emergence of resistance.

M. chelonae has been associated with disseminated soft tissue infections, localized cellulitis, osteomyelitis and catheter infections and appears to have a proclivity for renal transplant patients [4]. Skin biopsies from our patient clearly demonstrated granuloma with Ziehl–Neelsen positive rods, and *M. chelonae* was isolated from two specimens. However, the appendiceal biopsies only showed granuloma, with the stains being negative for acid-fast

bacilli. This may be explained by the prior exposure of the tissue to formalin. Indeed, atypical mycobacteria appear to be more sensitive to decontaminating agents than tuberculous mycobacteria [5]. Although microbiological or histological evidence for bone involvement cannot be provided, there is radiological evidence supporting this hypothesis. The lucencies in the right ankle and left little finger were described as atypical of brown tumours and in keeping with mycobacterial osteomyelitis. Despite the limited microbiological evidence for gut and bone involvement, we still feel this is a distinct possibility, though we think that the severe haemorrhagic colitis in this patient was probably related to co-existent CMV infection. To our knowledge, this would be the first report of infection with *M. chelonae* affecting these three organ systems concomitantly.

The risk factors for disseminated infection with atypical mycobacteria are treatment with immunosuppressant medication, steroid use and organ transplantation [4], all of which were present in the patient presented. Further contributing factors included post-transplant diabetes and previous splenectomy. The concurrent infection with CMV further reflects the immunocompromised state of our patient.

The mode of infection can only be speculated upon. There are several reports of infections with atypical mycobacteria occurring as a consequence of surgical and non-surgical procedures [6]. In immunocompetent individuals, infection may occur at the site of trauma or surgery, suggesting reduced local immunity. In our patient, the involvement of the appendix several weeks after radiologically guided drainage of a paracolic abscess may be attributable to inoculation with this ubiquitous pathogen which may then have spread from there.

Antimicrobial treatment of atypical mycobacterial disease is difficult, particularly in an HD patient. Clarithromycin is the only antibiotic which has been tested in a clinical trial and shown to be effective in treating disseminated cutaneous disease, albeit with a risk of developing resistance [7]. Isolates of *M. chelonae* are generally reported to be highly susceptible to clarithromycin [8] and tobramycin [9] and less susceptible to imipenem and ciprofloxacin [10]. The optimal duration of therapy for infections with atypical mycobacterial bacteria has not been established. However, the recommended minimal duration of combination antimicrobial therapy is 4 months for disseminated cutaneous disease and 6 months for osteomyelitis [11]. In patients on HD, the antimicrobial treatment is further complicated by pharmacokinetic alterations which can influence the efficacy and the duration of the treatment course and necessitate close monitoring of the clinical response to ensure effective treatment and monitoring of antibiotic levels to avoid toxicity.

Teaching points

- (1) *M. chelonae* is a rapidly growing mycobacterium, associated with disseminated soft tissue infections and osteomyelitis.

- (2) Risk factors for disseminated infection with atypical mycobacteria include long-term steroid use and organ transplantation. In the immunocompetent host, local infection may be found following trauma or surgery.
- (3) Adverse consequences of long-term immunosuppression may extend for months and even years following its cessation.
- (4) Co-existent opportunistic infections are common. The finding of one such infection in a patient should alert to the potential presence of others.

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Conflict of interest statement. None declared.

References

1. Kato S, Chmielewski M, Honda H *et al.* Aspects of immune dysfunction in end-stage renal disease. *Clin J Am Soc Nephrol* 2008; 3: 1526–1533
2. Griffith DE, Aksamit T, Brown-Elliott BA *et al.* An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am J Respir Crit Care Med* 2007; 175: 367–416
3. Jarzembowski JA, Young MB. Nontuberculous mycobacterial infections. *Arch Pathol Lab Med* 2008; 132: 1333–1341
4. Wallace RJ Jr., Brown BA, Onyi GO. Skin, soft tissue, and bone infections due to *Mycobacterium chelonae chelonae*: importance of prior corticosteroid therapy, frequency of disseminated infections, and resistance to oral antimicrobials other than clarithromycin. *J Infect Dis* 1992; 166: 405–412
5. Katoch VM. Infections due to non-tuberculous mycobacteria (NTM). *Indian J Med Res* 2004; 120: 290–304
6. Uslan DZ, Kowalski TJ, Wengenack NL *et al.* Skin and soft tissue infections due to rapidly growing mycobacteria: comparison of clinical features, treatment, and susceptibility. *Arch Dermatol* 2006; 142: 1287–1292
7. Wallace RJ Jr., Tanner D, Brennan PJ *et al.* Clinical trial of clarithromycin for cutaneous (disseminated) infection due to *Mycobacterium chelonae*. *Ann Intern Med* 1993; 119: 482–486
8. Brown BA, Wallace RJ Jr., Onyi GO *et al.* Activities of four macrolides, including clarithromycin, against *Mycobacterium fortuitum*, *Mycobacterium chelonae*, and *M. chelonae*-like organisms. *Antimicrob Agents Chemother* 1992; 36: 180–184
9. Swenson JM, Wallace RJ Jr., Silcox VA *et al.* Antimicrobial susceptibility of five subgroups of *Mycobacterium fortuitum* and *Mycobacterium chelonae*. *Antimicrob Agents Chemother* 1985; 28: 807–811
10. Wallace RJ Jr., Brown BA, Onyi GO. Susceptibilities of *Mycobacterium fortuitum* biovar. *fortuitum* and the two subgroups of *Mycobacterium chelonae* to imipenem, cefmetazole, cefoxitin, and amoxicillin-clavulanic acid. *Antimicrob Agents Chemother* 1991; 35: 773–775
11. Wallace RJ Jr., Swenson JM, Silcox VA *et al.* Treatment of nonpulmonary infections due to *Mycobacterium fortuitum* and *Mycobacterium chelonae* on the basis of in vitro susceptibilities. *J Infect Dis* 1985; 152: 500–514

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