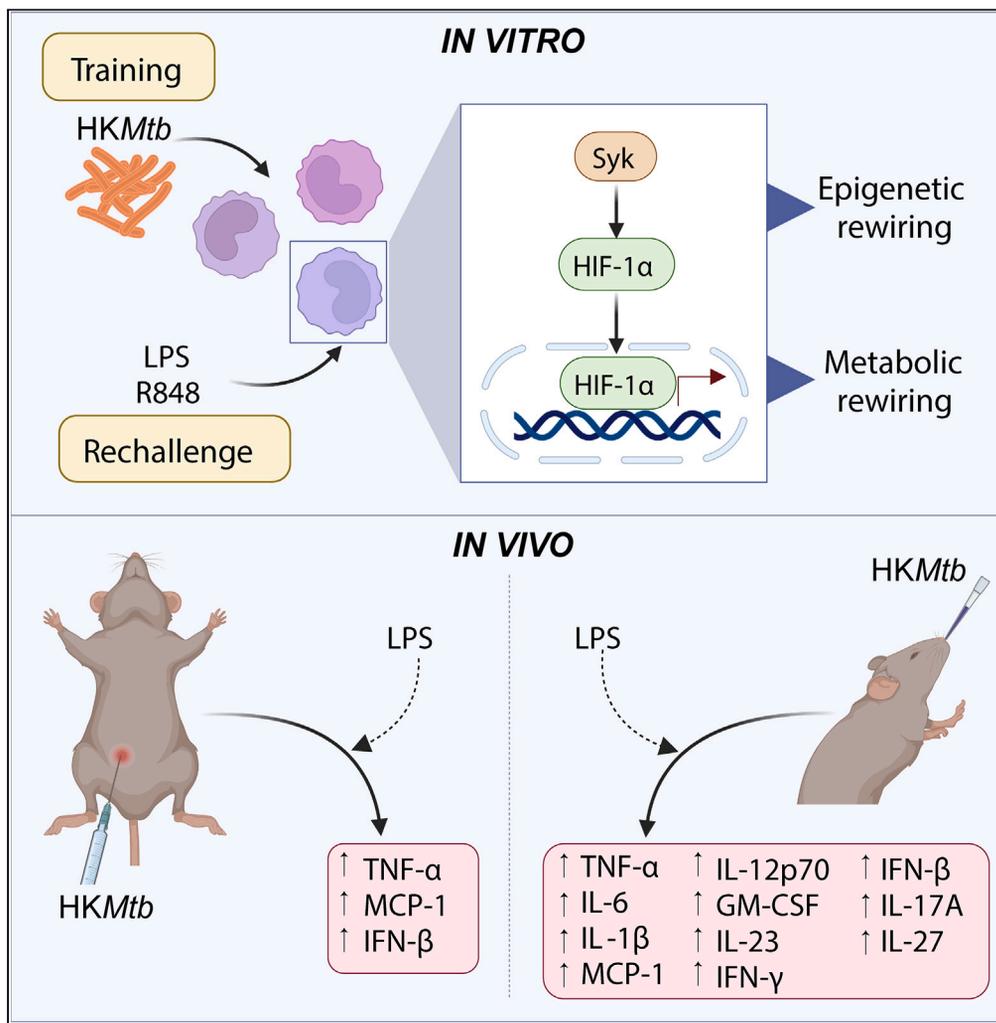


Article

Heat-killed *Mycobacterium tuberculosis* induces trained immunity *in vitro* and *in vivo* administered systemically or intranasally



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Highlights

Heat-killed *Mycobacterium tuberculosis* (HKMtb) induces trained immunity (TI)

HKMtb-induced TI relies on epigenetic and metabolic reprogramming

Syk and HIF-1α mediate the induction of TI by HKMtb

Intranasal administration of HKMtb generates robust trained systemic responses

Minute et al., iScience 27, 108869
February 16, 2024 © 2024 The Authors.
<https://doi.org/10.1016/j.isci.2024.108869>



Article

Heat-killed *Mycobacterium tuberculosis* induces trained immunity *in vitro* and *in vivo* administered systemically or intranasally

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SUMMARY

Trained immunity (TI) represents a memory-like process of innate immune cells. TI can be initiated with various compounds such as fungal β -glucan or the tuberculosis vaccine, Bacillus Calmette-Guérin. Nevertheless, considering the clinical applications of harnessing TI against infections and cancer, there is a growing need for new, simple, and easy-to-use TI inducers. Here, we demonstrate that heat-killed *Mycobacterium tuberculosis* (HKMtb) induces TI both *in vitro* and *in vivo*. In human monocytes, this effect represents a truly trained process, as HKMtb confers boosted inflammatory responses against various heterologous challenges, such as lipopolysaccharide (Toll-like receptor [TLR] 4 ligand) and R848 (TLR/7/8 ligand). Mechanistically, HKMtb-induced TI relies on epigenetic mechanisms in a Syk/HIF-1 α -dependent manner. *In vivo*, HKMtb induced TI when administered both systemically and intranasally, with the latter generating a more robust TI response. Summarizing, our research has demonstrated that HKMtb has the potential to act as a mucosal immunotherapy that can successfully induce trained responses.

INTRODUCTION

A complex network of cells, tissues, and molecules integrates our immune systems to protect our bodies against harmful insults. Traditionally, the immune system has been considered a dichotomy: the innate immune system, which provides a first line of defense, and the adaptive immune system, which confers long-term immunity through the generation of specific immune responses. In the last decade, however, this paradigm has been challenged due to the revelation of a fascinating phenomenon: trained immunity (TI).¹

TI refers to the ability of innate immune cells such as monocytes or macrophages to develop a memory-like effect that results in an improved proinflammatory response to a secondary challenge.¹ An essential attribute of this memory is its heterologous capacity for recall, given that a boosted inflammatory response can be generated by stimuli unrelated to those that induced the training process.² Mechanistically, the epigenetic reprogramming of inflammatory promoters drives the characteristic long-lasting proinflammatory trained response, in particular, the enrichment of specific histone acetylation and methylation.^{1,3} Of note, metabolic rewiring is also relevant for the training process by providing key cofactors required to establish the characteristic epigenetic TI markers.⁴ Altogether, TI is considered an innate immune memory process.²

The current insights regarding the training process are mostly attributed to β -glucan as a TI inducer because β -glucan was the first stimulus studied in this context and it uncovered the mechanisms involved in the TI process.¹ β -glucan is a type of polysaccharide found in fungal cell walls, and it is recognized by the Dectin-1 receptor. This recognition triggers a PI3K/mTOR/HIF-1 α -mediated pathway, activating the glycolytic Warburg effect and TI-related epigenetic reprogramming.^{3,5} Interestingly, β -glucan can be found either in a soluble or in a particulate form, which conditions its immunomodulatory properties, given that Dectin-1 signaling is only activated by particulate β -glucan.⁶ Depending on their physical properties, soluble forms of β -glucan, such as laminarin, are known to inhibit subsequent Dectin-1 signaling.⁷ Along these lines, TI triggering has been experimentally achieved by using particulate forms of β -glucan as an inducer, both *in vitro*¹ and *in vivo*.^{3,8} A commercially available β -glucan peptide consisting of a highly ramified, high-molecular-weight polysaccharide linked to a polypeptide portion has been reported to induce TI *in vivo*⁹; nevertheless, its particulate nature has not been fully elucidated.

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<https://doi.org/10.1016/j.isci.2024.108869>



Bacillus Calmette-Guérin (BCG), a live-attenuated strain of *Mycobacterium bovis* originally developed as a vaccine against tuberculosis, is a well-established TI inducer.¹⁰ Epidemiological studies have shown that BCG vaccination not only confers protection against tuberculosis but also enhances immune responses against non-tuberculosis-related heterologous infections.¹¹ Currently, BCG is one of the gold-standard stimuli to induce TI in humans, both *in vitro*¹² and *in vivo*.^{13,14} Mechanistically, the signaling axis NOD2 receptor/Rip2 kinase is responsible for the training induced by BCG,¹⁰ in which PI3K/mTOR¹² and autophagy¹⁵ also play a role. Thus, BCG represents a component with training capabilities that is used in clinical practice.

In our search for alternative TI inducer formulations, we studied MV130, a complex immunotherapy composed of various proportions of whole heat-inactivated gram-positive (90%) and gram-negative (10%) bacteria, originally indicated for treating recurrent respiratory tract infections.¹⁶ We had reported that MV130 generates TI, conferring heterologous protection against fungal and viral infections.^{17,18} MV130 modulates immune responses through Rip2 kinase and MyD88,¹⁶ although the role of these signaling pathways in the training process has not been addressed. Nevertheless, MV130 introduced the novelty of a heat-killed bacterial preparation with the capability of training innate immune cells by modulating their metabolism and epigenetic landscape.¹⁷

Interestingly, the *in vivo* induction of TI by these 3 components is achieved by various routes of administration. β -glucan is administered systemically in mouse models, primarily after intraperitoneal injection.^{3,8} BCG has been injected intradermally in humans,¹¹ and typically subcutaneously in mice,¹⁹ with some intravenous²⁰ or mucosal/intranasal^{21,22} attempts. Lastly, MV130 is administered as a sublingual spray in humans^{23,24} and mostly intranasally in mice.^{16,18} Comparative studies to address the relevance of the administration route employing a defined TI inducer are scarce; however, at least for BCG, mucosal delivery confers a stronger TI induction than intradermal vaccination in terms of enhanced innate cytokine production.²²

The nature of these stimuli and their administration route determine their potential clinical application for triggering TI. The particulate nature of β -glucan limits its transition from mouse models to clinical applications because it requires systemic administration. However, there is no doubt regarding the clinical use of BCG in clinical practice as an intradermal vaccine, or even by intravesical instillation as immunotherapy against bladder cancer.²⁵ However, due to the live nature of BCG, it bears the risk of localized or disseminated infections, depending on the patient's immunological status²⁶ and the route of administration. Although MV130 as a heat-killed bacteria-based mucosal immunotherapy has shown actual success in protecting against heterologous infections, with an excellent safety profile,^{27,28} MV130's polybacterial nature could hamper the dissection of relevant mechanistic clues in certain clinical scenarios, such as autoimmunity.^{29,30}

Linking the capacity of the mycobacterial vaccine BCG to induce TI, with the background provided by MV130 regarding heat-killed bacteria as a TI inducer, we here demonstrate the capacity of a commercial preparation of heat-killed *Mycobacterium tuberculosis* (HKMtb) as a TI inducer in human monocytes. This demonstration supports a recently published study using environmental *Mycobacterium manresensis* contained in a food supplement as a TI inducer.³¹ Furthermore, the HKMtb-induced trained heterologous response against both the Toll-like receptor (TLR) 4 ligand lipopolysaccharide (LPS) and the TLR7/8 ligand R848 relies on spleen tyrosine kinase (Syk), hypoxia-inducible factor (HIF)-1 α , and epigenetic reprogramming, generating a glycolytic rewiring. Of note, HKMtb induced TI *in vivo* when administered both intraperitoneally and intranasally. Altogether, our results indicate that a simple and commercially available HKMtb preparation constitutes a TI inducer. Furthermore, considering that our *in vivo* data reinforce the importance of the administration route of these compounds, more relevant from the clinical perspective is that intranasal/mucosal delivery of heat-killed mycobacterial preparations generates robust TI responses.

RESULTS

HKMtb induces TI against heterologous challenges

The BCG vaccine, which is a live-attenuated mycobacteria-based vaccine, has been found to induce TI. Another known TI inducer is MV130, a complex mix of heat-killed bacteria. Based on these examples, we investigated whether heat-killed mycobacteria could also trigger TI. To assess the validity of this hypothesis, we selected a commercially available preparation of HKMtb for a prototypical *in vitro* long-term TI experiment.¹ Peripheral blood mononuclear cells (PBMCs) were exposed to β -glucan (as a standard TI inducer) or HKMtb. After 1 day, stimuli and non-adherent cells were washed out, and adherent cells rested in a complete medium for 5 days. A flow cytometry analysis of these cultures confirmed that they were primarily CD14⁺ monocytes/macrophages ($\approx 90\%$) (Figure S1A). Next, the medium was removed and the cells were stimulated with a panel of pathogen-associated molecular patterns, including LPS as a TLR4 ligand, R848 as a TLR7/8 ligand, Pam3CSK4 as a TLR2 ligand, and CpG as a TLR9 ligand. The production of tumor necrosis factor alpha (TNF- α) in the supernatants was analyzed to evaluate the training process (Figure 1A). Note that the 2 latter stimuli produced negligible amounts of this proinflammatory cytokine, independent of pretreatment with either β -glucan or HKMtb (data not shown). As a control, β -glucan training generated increased TNF- α production upon restimulation (Figure S1B), confirming the performance of our model. Next, HKMtb-pretreated monocytes showed a robust enhanced TNF- α production in response to both LPS and R848 (Figure 1B), suggesting that the mycobacterial preparation induced TI.

At this stage, we evaluated whether the training effect induced by HKMtb depended on its concentration. A titration experiment was conducted, using HKMtb concentrations ranging from 1 μ g/mL to 100 μ g/mL. Note that the experiments shown in Figure 1B were performed with 10 μ g/mL concentrations. As shown in Figure 1C and 1A robust TI was only revealed against LPS at 10 μ g/mL, whereas boosted TNF- α production upon secondary R848 stimulation was detected at both 10 and 1 μ g/mL of HKMtb. Interestingly, β -glucan-induced training was comparable across a wide range of concentrations (Figure S1C). Therefore, considering these results, 10 μ g/mL of HKMtb was employed as the training stimulus for the subsequent experiments.

Following our previous study,⁸ our aim with regard to the training stimulations of total PBMCs was to perform these challenges under more native conditions, maintaining the blood's full mononuclear cellular environment. Afterward, the washing step removes non-adherent cells,

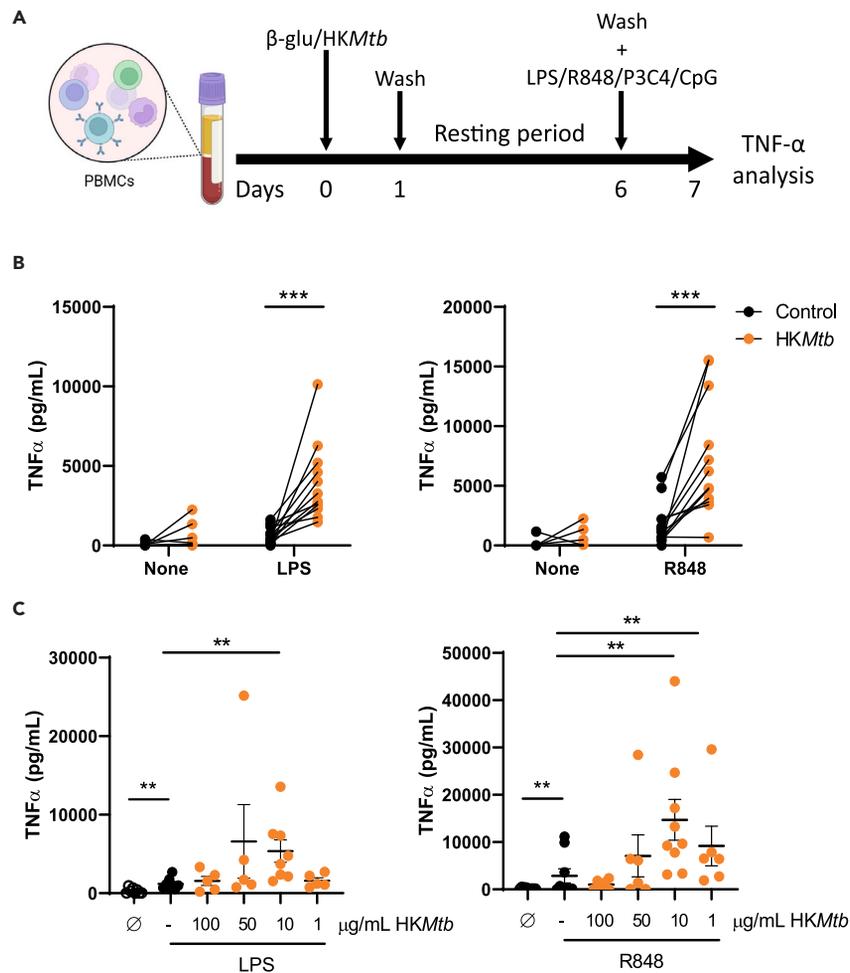


Figure 1. Heat-killed *Mycobacterium tuberculosis* induces trained immunity

(A) Trained immunity *in vitro* model in peripheral blood mononuclear cells (PBMCs). Created with [BioRender.com](https://www.biorender.com).

(B and C) PBMCs were trained with 10 $\mu\text{g}/\text{mL}$ (B) or the indicated concentrations (C) of heat-killed *Mycobacterium tuberculosis* (HKMTb) for 1 day, washed, and left to rest for 5 days. After washing, remaining adherent monocytes were rechallenged with lipopolysaccharide (LPS) (TLR4 ligand) (left panels) or R848 (TLR7/8) (right panels) for another day; supernatants were collected, and TNF- α was analyzed by enzyme-linked immunosorbent assay (ELISA). Each dot represents an independent donor. (B) $N = 4$, $n = 12$. Paired Student's *t* test comparing control (not trained) versus trained cells. (C) $N = 3$, $n = 6-9$. Data are represented as mean \pm SEM. Paired Student's *t* test comparing control (not trained) versus trained cells or non-treated (\emptyset) versus LPS or R848 stimulation, and (** $p < 0.01$; *** $p < 0.001$). See also [Figure S1](#).

primarily leaving adherent monocytes/macrophages ([Figure S1A](#)) to be secondarily stimulated. Nonetheless, to confirm that the observed training effects were intrinsic to monocytes, these cells were magnetically purified from PBMCs and subjected to the *in vitro* training scheme. As shown in [Figures 2A](#) and [2B](#), β -glucan as well as HKMTb induced increased TNF- α production in purified monocytes following both LPS and R848 secondary stimulation.

To further explore the TI process induced by HKMTb, a broad panel of cytokines was analyzed after training with the mycobacterial stimulus and secondary rechallenge with either LPS or R848. Cytokines such as inducible protein-10 (IP-10), interleukin (IL)-6, transforming growth factor (TGF)- β 1, and IL-12p70 were upregulated in response to LPS upon HKMTb-trained conditions, whereas IL-17A and interferon (IFN)- γ production was enhanced following both LPS and R848 ([Figure 2C](#)). Interestingly, the levels of the immunoregulatory cytokine IL-10 were downregulated in HKMTb-trained cells after restimulation ([Figure 2C](#)), reinforcing the proinflammatory trained phenotype generated after training with HKMTb.

Altogether, these data demonstrate that HKMTb induces TI in human monocytes, with a specific profile of response against secondary heterologous challenges.

HKMTb-induced TI relies on epigenetic reprogramming, hypoxia-inducible factor-1 α , and Syk kinase

The epigenetic reprogramming generated by the training stimulus constitutes the underlying mechanism responsible for the long-term trained process.² To assess the ability of epigenetic rewiring to sustain HKMTb-induced TI, we used the methyltransferase inhibitor

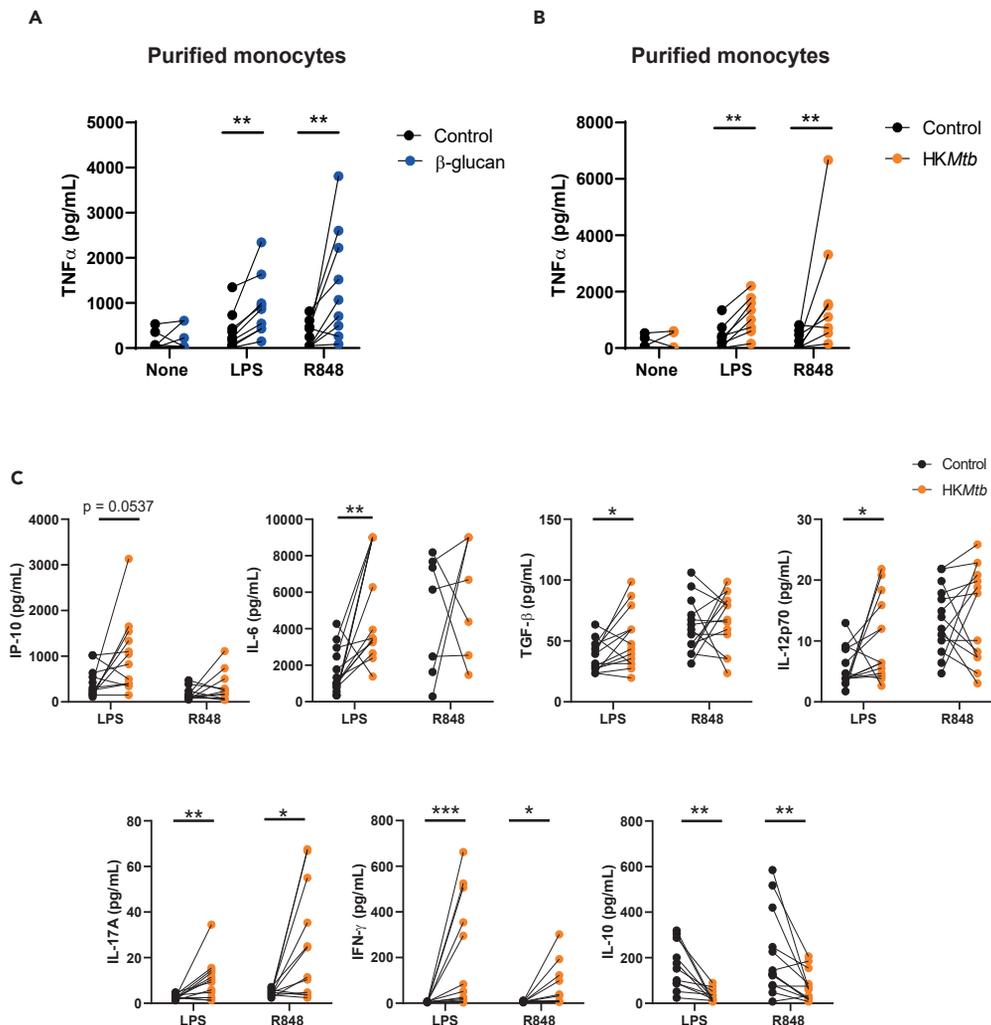


Figure 2. Heat-killed *Mycobacterium tuberculosis* induces a robust trained immunity response against lipopolysaccharide and R848

(A and B) Monocytes were magnetically purified from peripheral blood mononuclear cells (PBMCs), plated, and subjected to the protocol indicated in Figure 1A. Ten $\mu\text{g}/\text{mL}$ of both β -glucan and heat-killed *Mycobacterium tuberculosis* (HKMtb) were used as training stimuli, and lipopolysaccharide (LPS) or R848 as secondary challenge. TNF- α was analyzed by enzyme-linked immunosorbent assay (ELISA).

(C) PBMCs were trained with 10 $\mu\text{g}/\text{mL}$ of HKMtb following the protocol indicated in Figure 1A, rechallenging with LPS or R848. Supernatants were collected, and a panel of cytokines was analyzed by a multiparametric cytokine determination kit. Each dot represents an independent donor. (A, B) $N = 3$, $n = 9$. (C) $N = 6$, $n = 14$. Paired Student's *t* test comparing control (not trained) versus trained cells. (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$). Specific *p* value is shown for clarification.

5'-deoxy-5'-(methylthio) adenosine (MTA), a widely accepted TI regulator.^{1,8} Preincubation of PBMCs with MTA dampened HKMtb-induced TI in response to both LPS and R848 (Figure 3A). To further confirm the generation of an epigenetic rewiring by HKMtb stimulation, chromatin immunoprecipitation assays capturing trimethyl-Histone H3 activating residue on the *TNFA* promoter³ showed enrichment following HKMtb training (Figure 3B). These results confirmed that TI induced by HKMtb depends on epigenetic reprogramming.

HIF-1 α is critical for the innate training process induced by β -glucan.³ To address whether the activity of this transcription factor is relevant for the TI induced by HKMtb, we employed the HIF-1 α -specific inhibitor, PX-478.³² Based on this experimental approach, we observed a significant impairment of HKMtb-induced TI after secondary stimulation with both LPS and R848 when PBMCs were pretreated with PX-478 (Figure 3C), indicating the role of HIF-1 α in HKMtb-induced training. Importantly, quantitative polymerase chain reaction (qPCR) analyses showed a clear induction on the expression of this transcription factor following HKMtb training (Figure 3D), further illustrating that HKMtb-induced TI relies on HIF-1 α .

As previously mentioned, metabolic reprogramming toward glycolysis is also critical for the induction of TI in myeloid cells.⁴ Lactate production following HKMtb training was analyzed as a surrogate of glycolytic metabolism, showing a clear increase (Figure 3E). Thus, these data indicate that HKMtb induces TI, involving metabolic and epigenetic reprogramming and HIF-1 α . Moving upstream, HKMtb is known to be recognized by the C-type lectin receptor (CLR), macrophage inducible C-type lectin (Mincle),³³ triggering a primary signaling pathway involving the proximal adaptor Syk.³⁴ To explore whether HKMtb-induced TI relied on this signaling module, we used the Syk inhibitor,

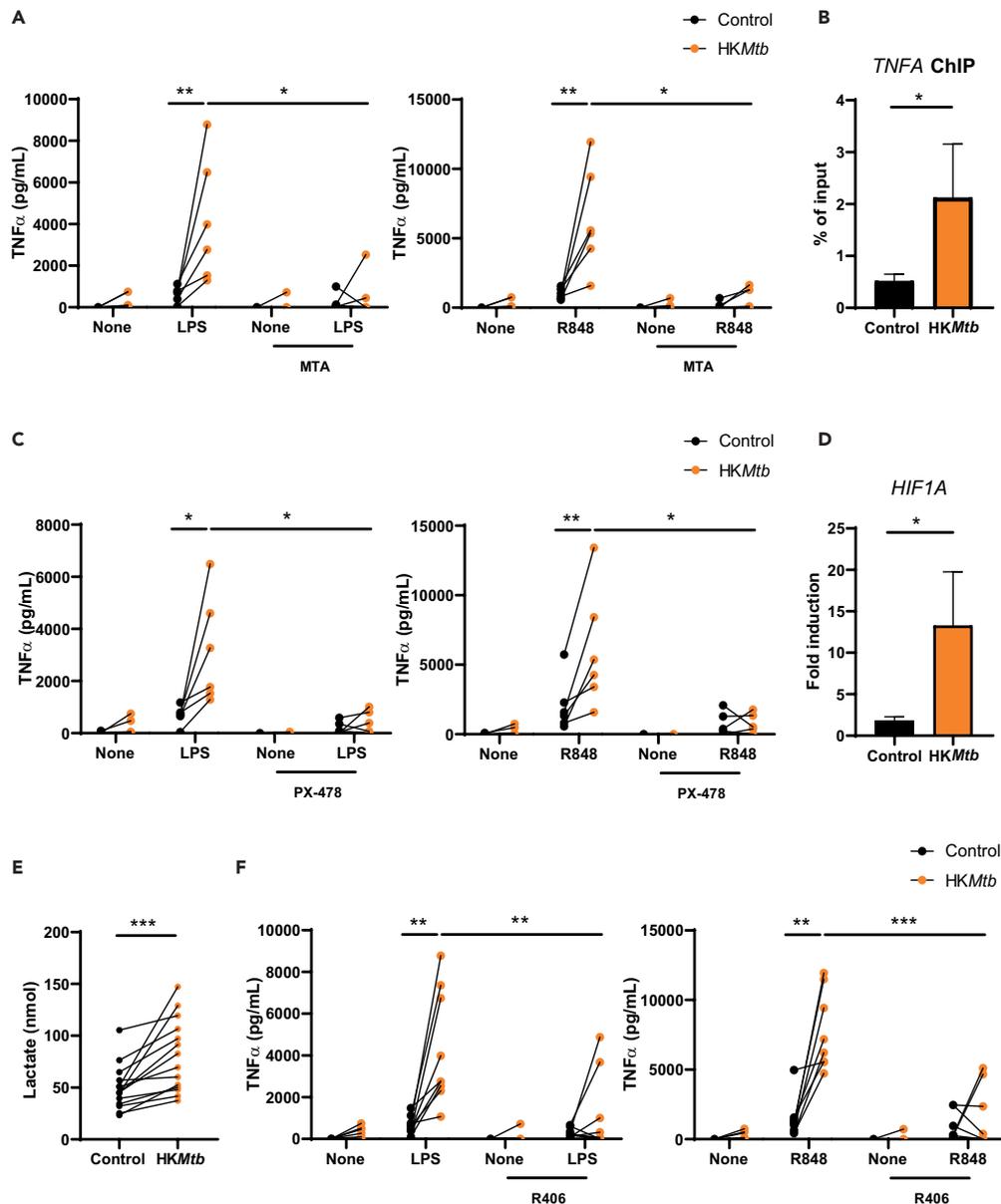


Figure 3. Heat-killed *Mycobacterium tuberculosis*-induced trained immunity relies on epigenetic reprogramming, hypoxia-inducible factor-1 α , and Syk kinase

(A, C, and F) Peripheral blood mononuclear cells (PBMCs) were incubated with the methyltransferase inhibitor 5'-deoxy-5'-(methylthio) adenosine (MTA) (A), the hypoxia-inducible factor (HIF)-1 α inhibitor PX-478 (C), and the Syk inhibitor R406 (F) before training with heat-killed *Mycobacterium tuberculosis* (HKMtb) for 1 day. Cells were then washed and left to rest for 5 days. Remaining adherent monocytes were washed and rechallenged with lipopolysaccharide (LPS) or R848 for another day, and tumor necrosis factor (TNF) α in the supernatants was analyzed by enzyme-linked immunosorbent assay (ELISA).

(B, D and E) PBMCs were trained with HKMtb for 1 day. Cells were then washed and left to rest for 5 days (B) Chromatin immunoprecipitation (ChIP) was performed against trimethyl-Histone H3 (Lys4), analyzing the enrichment of this residue at the *TNFA* promoter by quantitative polymerase chain reaction (qPCR). (D) Total RNA was extracted and *HIF1A* expression was analyzed by qPCR. (E) Supernatants were collected, and lactate was analyzed by a colorimetric assay. Each dot represents an independent donor. (A–D, F) N = 2–3, n = 6–9. (E) N = 6, n = 14. (B, D) Data are represented as mean \pm SEM. Paired Student's t test comparing control (not trained) versus trained cells or exposed versus not exposed to the various inhibitors (A, C, F). (*p < 0.05; **p < 0.01; ***p < 0.001).

R406.³⁵ HKMtb-induced TI was impaired by Syk inhibition upon both LPS and R848 rechallenge (Figure 3F), indicating that HKMtb-induced TI depends on Syk kinase, most probably through the Mincle receptor.

Altogether, these mechanistic results indicate that epigenetic reprogramming underlies HKMtb-induced TI in an HIF-1 α - and Syk-dependent manner, also involving a glycolytic switch, independent of the secondary heterologous stimuli.

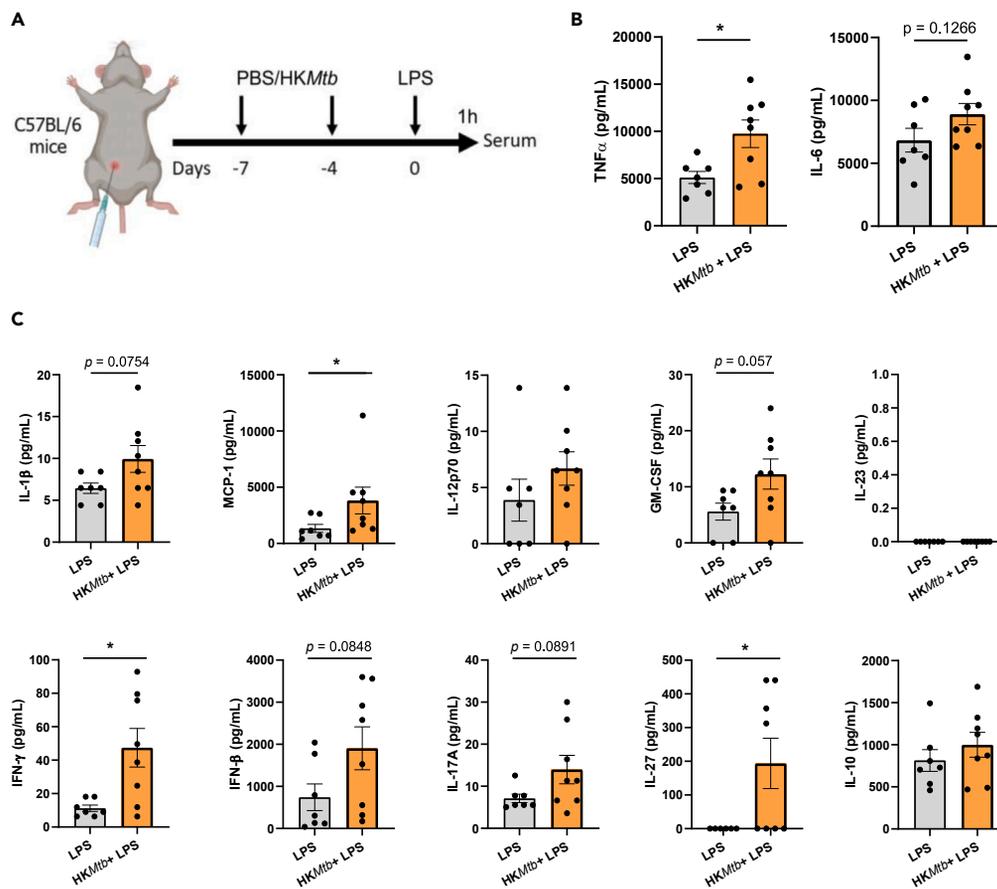


Figure 4. In vivo systemic administration of heat-killed *Mycobacterium tuberculosis* induces trained immunity

(A) *In vivo* model of training systemically by 2 intraperitoneal injections of heat-killed *Mycobacterium tuberculosis* (HKMTb) or control with phosphate-buffered saline (PBS), followed by a secondary intraperitoneal lipopolysaccharide (LPS) challenge for measuring serum cytokines. Created with [BioRender.com](https://www.biorender.com).

(B) Serum tumor necrosis factor (TNF) α and interleukin (IL)-6 were analyzed by enzyme-linked immunosorbent assay (ELISA).

(C) Serum IL-1 β , monocyte chemoattractant protein (MCP)-1, IL-12p70, granulocyte-macrophage colony-stimulating factor (GM-CSF), IL-23, interferon (IFN)- γ , IFN- β , IL-17A, IL-27, and IL-10 were analyzed by a multiparametric cytokine determination kit. (B, C) $n = 7$ –8 mice per experimental condition. Unpaired Student's t test or Mann-Whitney test comparing mice trained or not with HKMTb ($*p < 0.05$). For clarity, some p values are indicated.

In vivo systemic administration of HKMTb induces TI

Once we had established that HKMTb induces TI *in vitro*, we examined its training capabilities *in vivo*. Intraperitoneal injection of β -glucan is a well-established administration route resulting in TI induction.^{1,3} We adopted this schedule to test whether HKMTb ignites a boosted inflammatory reaction against systemic heterologous LPS administration (Figure 4A). Serum levels of TNF- α and IL-6, quantified by enzyme-linked immunosorbent assay (ELISA), were chosen as the main readouts. Intraperitoneal training with HKMTb generated boosted TNF- α production following LPS stimulation, with a trend in the same direction in the case of IL-6 (Figure 4B). To go deeper in this analysis, we performed a multiparametric assay to assess a panel of serum cytokines in these settings. Our findings show that HKMTb training induced higher levels of MCP-1, IFN- γ , and IL-27, with a similar trend for IL-1 β , granulocyte-macrophage colony-stimulating factor (GM-CSF), IFN- β , and IL-17A (Figure 4C). These data indicate that the systemic administration of HKMTb trains mice against a heterologous LPS challenge, generating a mildly enhanced proinflammatory profile.

Intranasal administration of HKMTb robustly induces TI *in vivo*

Previously, we had demonstrated that the heat-killed polybacterial immunotherapy, MV130, consistently induces TI when administered intranasally.^{17,18} Therefore, we examined here whether this route of administration could also be used to induce TI with HKMTb, comparatively evaluating the potential responses with those observed upon systemic administration (see Figure 4). Mice were administered HKMTb intranasally following the schedule previously tested for MV130 (3 times per week for 2 weeks) and challenged with LPS 1 week after the last HKMTb inoculation (Figure 5A). As previously performed, serum levels of TNF- α and IL-6, analyzed by ELISA, were chosen as the primary endpoints. Levels of both proinflammatory cytokines were enhanced in those mice trained with HKMTb (Figure 5B). Furthermore, the multiparametric serum analysis indicated a general and robust increase in the amount of proinflammatory cytokines detected in the HKMTb-trained mice, with higher levels of virtually all cytokines, except the immunomodulatory cytokine IL-10 (Figure 5C).

Considering the high proinflammatory profile observed with the intranasal HKMtb administration scheme, we sought signs of potential autoimmune reactions. To this end, we analyzed the impact of HKMtb-mediated training on some steady-state inflammatory features, in the absence of secondary LPS stimulation. No signs of splenomegaly were found in the HKMtb-trained mice at the end of the 3-week training period (Figure S2A), nor did we find basal levels of circulating TNF- α (Figure S2B). These data indicate that the trained status generated by intranasal HKMtb administration is not accompanied by a proinflammatory profile unless a secondary stimulation takes place.

Considering the potential clinical application of HKMtb-induced trained responses, we evaluated whether this training generated positive reactions when employing the gold-standard method to identify an exposure to *Mycobacterium*. Thus, we tested purified protein derivative (PPD)-specific responses by exposing splenocytes to tuberculin obtained from *Mycobacterium tuberculosis*.³⁶ The production of IFN- γ was evaluated in an IFN- γ release assay (IGRA).³⁶ As shown in Figure 5D, HKMtb intranasal training conferred positive responses against PPD.

In summary, the administration of HKMtb through the intranasal route confers a robust trained response *in vivo*, with no apparent autoimmune symptoms and trackable by PPD test.

DISCUSSION

The heterologous nature of TI is opening new opportunities in vaccinology, promoting the development of TI-based vaccines (TibVs), which are defined as vaccine formulations that seek the induction of TI in innate immune cells.²⁸ The goal of these vaccines, unlike conventional formulations based on antigens plus adjuvant, is the priming of proinflammatory responses against a broad spectrum of heterologous insults, mainly infections.^{37,38} The potential application of these formulations would be the generation of a broad prophylaxis against pathogens lacking specific vaccination, or even to improve the protection conferred by antigen-specific vaccines.²⁸ This concept was applied during the COVID-19 pandemic before severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-specific vaccines were developed.^{18,39}

The growing clinical relevance of these training processes has sparked an urgency to identify and apply new TI inducers in clinical settings. Herein, we describe that a commercially available preparation of HKMtb induces TI, both in human monocytes *in vitro* and in mouse *in vivo* models. As mentioned in the Introduction, other mycobacteria, such as BCG, are the gold-standard stimuli to induce TI, both *in vitro*¹² and *in vivo* in humans.^{13,14} Nevertheless, the use of HKMtb as a TI inducer provides some advantages over BCG. The key differentiating factor is the use of metabolically inactive (killed) mycobacteria instead of live-attenuated mycobacteria, such as BCG. This difference is relevant because the use of heat-killed bacteria to induce TI overcomes some of BCG's limitations. For instance, it avoids the risk of infection due to uncontrolled bacterial growth in immunodeficient patients.²⁶ In addition, the logistic and clinical handling of heat-killed bacteria would be much easier, facilitating its distribution and implementation. Furthermore, our data indicate that HKMtb induces TI through the airways, opening the possibility to be self-administered by patients as a nasal or sublingual spray, as occurs with MV130.²⁷

An alternative heat-killed mycobacterium included in a food supplement, *Mycobacterium manresensis*, has also been described as an *in vitro* TI inducer.³¹ In these experiments, Percoll-purified adherent mononuclear cells after CD3 lymphocyte depletion were employed, reaching almost 80% of CD14⁺ monocytes/macrophages.³¹ Note, IFN- γ from lymphocytes has been described as responsible for TI induction in monocytes following a *Plasmodium falciparum* encounter.⁴⁰ Along these lines, we initially trained total PBMCs, aiming to reproduce the encounter of blood circulating mononuclear immune cells with HKMtb as a training stimulus, revealing the trained phenotype in adherent monocytes upon secondary heterologous stimulations. Thus, our initial setting did not allow us to discriminate a potential cooperation between lymphocytes and monocytes for the generation of the HKMtb-mediated trained response. To clarify this concern, monocytes cells were magnetically purified from total PBMCs and trained with HKMtb, confirming that the mycobacterial preparation trains isolated monocytes in a cell-intrinsic manner.

To reach potential clinical applications, it is important to understand the molecular mechanisms implicated in HKMtb-induced TI. Based on our findings, we can determine a signaling pathway implicating Syk/HIF-1 α leading to epigenetic and metabolic reprogramming. Syk kinase is a proximal molecule implicated in the signaling triggered by most of the activating CLRs^{34,41}; among these receptors, Dectin-1 is key to inducing TI in response to the prototypical inducer β -glucan, also activating HIF-1 α .³ According to our data, Syk participates in the induction of TI by HKMtb, suggesting that this kinase is shared for the direct and the training-related signaling pathway, most probably downstream Mincle.³³ Of note, the scaffold protein FcR γ is necessary for the signaling triggered by downstream Mincle, leading to Syk activation.^{34,41} This feature suggests that other FcR γ -coupled CLRs activating Syk might also induce TI.

Resiquimod, the commercial name for R848, has been extensively tested as a vaccine adjuvant in various contexts, such as experimental protein-based vaccine candidates,⁴² or antiviral or antitumor vaccines.^{43,44} One of the aims of TibV is to improve the immunogenic responses of antigen-specific vaccines,²⁸ due to either low vaccine immunogenicity or poor target population response, for instance, in the elderly. In this context, the administration of TibV in a prophylactic manner (we postulate HKMtb in this case) would be necessary before administering vaccine formulations containing Resiquimod as an adjuvant. We have observed this regimen improving antigen-specific responses when combining MV130-induced TI and COVID-19 vaccination.¹⁸ Thus, our results raise the possibility of combining training with HKMtb with vaccines harboring Resiquimod as an adjuvant, thereby improving the immunogenicity of those vaccine formulations.

The epigenetic proinflammatory profile imprinted during the development of TI could also have detrimental effects due to uncontrolled inflammatory responses, potentially leading to autoimmunity.³⁰ In this sense, HKMtb training alone did not lead to substantial TNF- α production in the long term in human adherent monocytes in the absence of secondary stimulation. Additionally, neither splenomegaly nor circulating TNF- α was found in steady-state HKMtb-trained mice at the end of the 3-week scheme of intranasal administration. These results suggest a lack of uncontrolled proinflammatory reactions due to the exposure to HKMtb, although this aspect would need to be further addressed in clinical settings, as would the possibility that intranasal administration of HKMtb could generate PPD-specific responses.

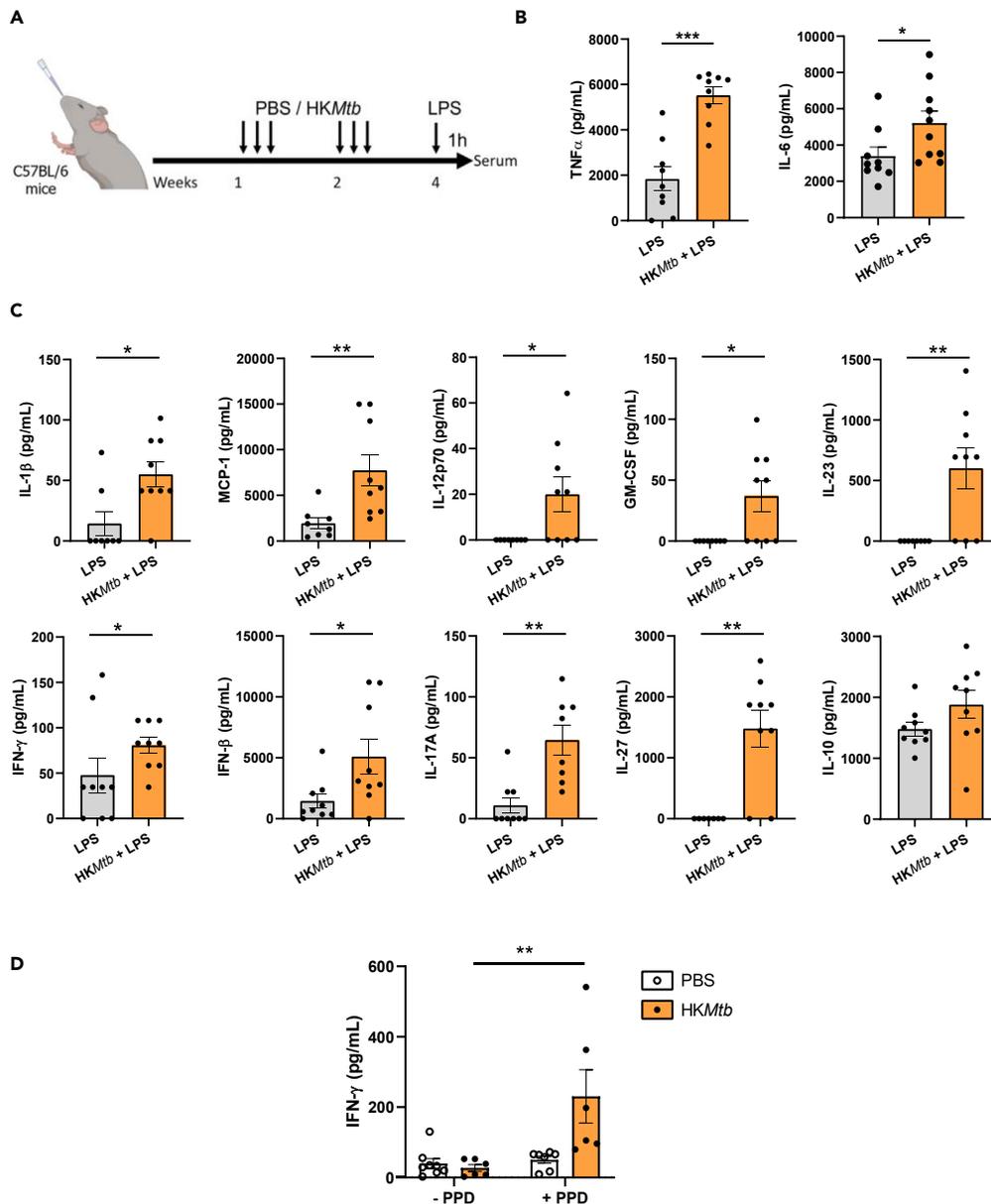


Figure 5. Intranasal administration of heat-killed *Mycobacterium tuberculosis* robustly induces trained immunity in vivo

(A) *In vivo* model of intranasal training. Mice were intranasally administered with phosphate-buffered saline (PBS) or heat-killed *Mycobacterium tuberculosis* (HKMtb) 3 times per week for 2 weeks. After a resting week, the mice were exposed to a secondary intraperitoneal lipopolysaccharide (LPS) challenge for measuring serum cytokines. Created with BioRender.com.

(B) Serum tumor necrosis factor (TNF) α and interleukin (IL)-6 were analyzed by enzyme-linked immunosorbent assay (ELISA).

(C) Serum IL-1 β , monocyte chemoattractant protein (MCP)-1, IL-12p70, granulocyte-macrophage colony-stimulating factor (GM-CSF), IL-23, interferon (IFN)- γ , IFN- β , IL-17A, IL-27 and IL-10 were analyzed by a multiparametric cytokine determination kit.

(D) Mice were intranasally administered HKMtb or PBS as indicated in panel A. Following the 1-week resting period and before lipopolysaccharide stimulation, the mice were euthanized, and spleens recovered. Single-cell suspensions were plated and re-stimulated or not with the purified protein derivative (PPD). Three days afterward, supernatants were collected and IFN- γ analyzed by ELISA. Data are represented as mean \pm SEM. (B, C) n = 6–9 mice per experimental condition. Unpaired Student's t test or Mann-Whitney test comparing mice trained or not with HKMtb (B, C) or restimulated or not with PPD (D) (*p < 0.05; **p < 0.01; ***p < 0.001). See also Figure S2.

Importantly, we have observed that HKMtb induces TI *in vivo* when administered both systemically through the intraperitoneal route and intranasally. Nevertheless, the trained response was more robust when the mucosal intranasal route of administration was chosen, given that we detected enhanced production of virtually all the analyzed cytokines, except IL-10, against the heterologous challenge. Notably, in one

clinical trial, the oral administration of alternative heat-killed mycobacteria included in a food supplement, *Mycobacterium manresensis*, did not induce TI, despite generating the trained process *in vitro*.³¹ This discrepancy points to the route of administration as a decisive factor to be considered for the induction of TI-based protective responses triggered by these mycobacterial preparations, as suggested by the effectiveness of BCG against SARS-CoV-2 infection when mice are vaccinated intravenously instead of subcutaneously.²⁰

Focusing on the strong induction of TI following intranasal administration of HKMtb, there are several examples of the power of this route for triggering TI. We have observed that the mucosal polybacterial immunotherapy MV130 confers protection against viral and fungal infection when provided through the airways.^{17,18} Indeed, successful results against pathologies of infectious etiology have been observed in clinical settings using MV130 when administered sublingually,^{27,45} indicating the effectiveness of the airway mucosae for the induction of TI. Along these lines, the induction of TI by BCG or the alternative *Mycobacterium tuberculosis* vaccine, MTBVAC, was stronger when non-human primates were vaccinated through the lung mucosae than intradermally.²² The exploitation of the intranasal route to administer immunomodulators of various types has been proposed as an effective approach to trigger antitumor immunity,⁴⁶ as well as to generate potent antigen-specific responses⁴⁷ that are comparatively stronger than, for example, subcutaneous vaccination.⁴⁸ In addition, despite preclinical data suggesting that intragastric administration of β -glucans could improve antitumor responses,^{49,50} suggestive of heterologous TI,^{51,52} no enhanced innate immune responses were observed in a controlled clinical trial in which participants ingested β -glucan once daily for 7 days.⁵³ Altogether, these data suggest the great potential of the intranasal route of administration to trigger potent TI responses.

Nonetheless, we can only speculate about the cellular mechanisms underlying this better TI induction through the airways because the current knowledge in this sense is scarce. A longer persistence in the airways of the training stimulus could play a role, perhaps due to a limited capacity of removal compared with the peritoneum in the case of intraperitoneal administration, or the liver when administering the training stimulus directly into the bloodstream. An alternative explanation would be the direct stimulation of TI-prone myeloid cells in the lungs, postulating a preponderant role for alveolar or interstitial macrophages in the lungs. Along these lines, MV130-induced intranasal TI is CCR2 independent,¹⁷ suggesting that the recruitment of monocytes from the bone marrow is not required to confer heterologous protection, pointing to local lung-resident populations.

In summary, we have herein described the capacity of HKMtb to induce TI *in vitro* through epigenetic rewiring in a Syk/HIF-1 α -dependent manner, leading to enhanced heterologous responses against LPS and R848. HKMtb induces TI *in vivo* as well, administered both systemically and intranasally, with the latter generating more robust proinflammatory trained responses. The presented data suggest an avenue whereby this immunotherapy could be deployed in pathophysiological contexts that require protective inflammatory responses.

Limitations of the study

The lack of a parallel comparison between HKMtb and *Mycobacterium manresensis* in the *in vivo* settings studied in this work (systemic and intranasal administration) represents a limitation of the study. Nevertheless, conclusions regarding this topic have been made strictly based on the provided experimental data and were only further extended in the [discussion](#) section based on solid references.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

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SUPPLEMENTAL INFORMATION

Supplemental information can be found online at <https://doi.org/10.1016/j.isci.2024.108869>.

ACKNOWLEDGMENTS

The laboratory of CdF is funded by the Instituto de Salud Carlos III through the projects CP20/00106 and PI21/01178 and co-funded by the European Union, Fundación para la Investigación Biomédica Hospital la Paz (FIBHULP - Luis Álvarez 2021), Scientific Foundation of the Spanish Association Against Cancer (IDEAS222745DELFI), Fundación Familia Alonso, Comunidad de Madrid (CAM) (IND2022-BMD-23669), and Immunotek. P.M.-M. and J.F.-P. are funded by Consejería de Educación y Universidades (CAM). L.M. is funded by ISCIII, Sara Borrel program (CD22/00042). L.B. and G.B. are funded by the European Union, NextGenerationEU.

AUTHOR CONTRIBUTIONS

C.d.F. designed the study and wrote the manuscript. L.M., M.B.-G., P.M.-M., J.F.-P., V.T., L.B.-R., G.B., and G.Z.-F. performed the experiments. N.A. performed critical analyses and provided essential reagents. N.A., E.L.-C., and C.d.F. discussed the results. All the authors have read, reviewed, and agreed to submit the manuscript for publication.

DECLARATION OF INTERESTS

The authors declare no competing interests.

Received: June 19, 2023

Revised: November 3, 2023

Accepted: January 8, 2024

Published: January 11, 2024

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
CD3 – BV510	Biolegend	Cat# 317332; RRID: AB_2561943
CD14 - SparkBlue550	Biolegend	Cat# 367148; RRID: AB_2820021
CD56 – BV750	Biolegend	Cat# 362556; RRID: AB_2801001
ChIPAb+ Trimethyl-Histone H3 (Lys4)	Millipore	Cat# 17-614; RRID: AB_11212770
Biological samples		
Blood samples of healthy health personnel	This paper	N/A
Chemicals, peptides, and recombinant proteins		
Ficoll-Plus	GE Healthcare	Cat# 17-1440-03
RPMI 1640 Medium	Thermo Fisher Scientific	Cat# 11594506
Phosphate-buffered saline (PBS)	Sigma	Cat# P4417-100TAB
Fetal bovine serum (FBS)	Thermo Fisher Scientific	Cat# 11560636
Pharm Lyse Lysing Buffer	BD	Cat# 555899
Penicillin Streptomycin	Gibco	Cat# 15140-148
Whole glucan particles	InvivoGen	Cat# tlrl-wgp
Heat-killed <i>Mycobacterium tuberculosis</i>	InvivoGen	Cat# tlrl-hkmt
Lipopolysaccharides (LPS) from <i>Escherichia coli</i> O111:B4	Sigma-Aldrich	Cat# L4391
R848 (Resiquimod)	InvivoGen	Cat# tlrl-r848
Pam3CSK4	InvivoGen	Cat# tlrl-pms
CpG ODN 2216	InvivoGen	Cat# tlrl-2216
5'-Deoxy-5'-(methylthio) adenosine (MTA)	Sigma-Aldrich	Cat# D5011
PX-478	MedChemExpress	Cat# HY-10231
R406 free base	MedChemExpress	Cat# HY-11108
Critical commercial assays		
Human TNF-alpha DuoSet ELISA	R&D Systems	Cat# DY210
Mouse TNF-alpha DuoSet ELISA	R&D Systems	Cat# DY410
Mouse IL-6 DuoSet ELISA	R&D Systems	Cat# DY406
Mouse IFN γ DuoSet ELISA	R&D Systems	Cat# DY485
KPL SureBlue TMB Microwell Substrate	Seracare	Cat# 5120-0076
LEGENDplex Mouse Inflammation Panel (13-plex) with Filter Plate	BioLegend	Cat# 740150
LEGENDplex human Essential Immune Response Panel (13-plex) with Filter Plate	BioLegend	Cat# 740930
Pan Monocyte Isolation kit	Miltenyi	Cat# 130-096-537
Magna ChIP A/G	Millipore	Cat# 17-10085
Monarch Total RNA Miniprep kit	New England Biolabs	Cat# T2010S
High-Capacity cDNA reverse transcription kit	Applied Biosystems	Cat# 10400745
PowerUp SYBR Green Master Mix	Applied Biosystems	Cat# A25742
Lactate Assay kit	Sigma-Aldrich	Cat# MAK064
Blue LIVE/DEAD Fixable staining	ThermoFisher	Cat# L23105

(Continued on next page)

Continued

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Experimental models: Organisms/Strains		
C57BL/6 mice	The Jackson Laboratory	RRID: MSR_JAX:000664
Oligonucleotides		
TNFA-ChIP Forward	CAGGCAGGTTCTCTTCTCTCT	N/A
TNFA-ChIP Reverse	GCTTTCAGTGCTCATGGTGT	N/A
HIF1A Forward	AGTGTACCCTAACTAGCCGAGGAA	N/A
HIF1A Reverse	CTGAGGTTGGTTACTGTTGGTACA	N/A
β -ACTIN Forward	CATGTACGTTGCTATCCAGGC	N/A
β -ACTIN Reverse	CTCCTTAATGTCACGCACGAT	N/A
Software and algorithms		
LEGENDplex software v.8	Biolegend	https://www.biolegend.com/en-us/legendplex
Prism version 8.3	GraphPad	https://www.graphpad.com/scientific-software/prism/
Microsoft Excel	Microsoft	https://products.office.com/
Biorender	Biorender.com	https://www.biorender.com/

RESOURCE AVAILABILITY**Lead contact**

Further information and requests for resources and reagents should be directed to and will be fulfilled by the lead contact, Carlos del Fresno (carlos.fresno@salud.madrid.org).

Materials availability

This study did not generate new unique reagents.

Data and code availability

- Data reported in this paper will be shared by the [lead contact](#) upon request.
- This paper does not report original code.
- Any additional information required to reanalyze the data reported in this paper is available from the [lead contact](#) upon request.

EXPERIMENTAL MODEL AND STUDY PARTICIPANT DETAILS**Blood samples**

Healthy volunteers of unspecified age and sex were recruited from the Blood Donor Service at La Paz University Hospital (Madrid, Spain). The experiments have been reviewed and approved by the La Paz University Hospital Ethics Committee (PI-5270) in the framework of the ISCIII grant PI21/01178 awarded to the PI of this project.

Mice

The mice we used were bred at the La Paz University Hospital Institute for Health Research (IdiPAZ). We employed mice from the Jackson Laboratory, mouse strain C57BL/6 (RRID: IMSR_JAX:000664), age-matched 6 to 8 weeks, regardless of sex. Otherwise, mice were included in the studies in a blinded fashion and randomly assigned to receive HKMtb or PBS/excipient (simple 1:1 randomization).

Experiments were approved by the "Dirección General de Agricultura, Ganadería y Alimentación" of Comunidad de Madrid as part of the project PROEX number 348.2/21.

METHOD DETAILS**Culture conditions of primary cells**

Fresh peripheral blood mononuclear cells (PBMCs) from ethylenediaminetetraacetic acid anticoagulant (EDTA) venous blood of healthy donors were isolated by gradient centrifugation with Ficoll-Paque (GE Healthcare Bio-Sciences, Cat# 17-1440-03). PBMCs were washed twice with phosphate buffered saline (PBS) (Sigma, Cat# P4417-100TAB) and counted by trypan blue staining. Thereafter, cells were resuspended in Roswell Park Memorial Institute (RPMI) medium (Thermo Fisher Scientific, Cat# 11594506) supplemented with antibiotics (0.1% penicillin

and 0.1% streptomycin, Gibco, Cat# 15140-148) and 10% fetal bovine serum (FBS) (Thermo Fisher Scientific, Cat# 11560636) (RPMIc). PBMCs were cultured at 37 °C at 5% CO₂ in a humidified incubator.

Trained immunity in PBMCs and purified monocytes

We plated 2×10^5 PBMCs in 100 μ L RPMIc per well in 96-well flat bottom plates. On day 0, cells were stimulated with 100 μ g/mL (or the indicated concentration) of β -glucan particles (InvivoGen, Cat# tlr-wgp) or 10 μ g/mL (or the indicated concentration) HKMtb (strain H37Ra, InvivoGen, Cat# tlr-hkmt-1) to a final volume of 200 μ L of RPMIc for 24 h at 37°C. Cells were then washed with PBS and allowed to rest for 5 days in RPMIc. On day 6, cells were washed again and further stimulated with 10 ng/mL lipopolysaccharide (LPS) from *Escherichia coli* O111:B4 (Sigma-Aldrich, Cat# L4391), 10 μ g/mL R848 (InvivoGen, Cat# tlr-r848), 10 ng/mL Pam3CSK4 (P3C4) (InvivoGen, Cat# tlr-pms), or 2,5 μ M CpG (ODN p2216) (InvivoGen, Cat# tlr-2216). After 24 h, supernatants were collected and stored at –20°C for subsequent processing. The composition of the cultures after the 5-day resting period was analyzed by flow cytometry with the Aurora 5L device (Cytex), using antibodies against CD3, (lymphocytes); CD14, (monocytes/macrophages); and CD56 (NKs).

Alternatively, monocytes were purified from PBMCs using the Pan monocyte isolation kit (Miltenyi, Cat# 130-096-537) following the manufacturer's instructions. We plated 2×10^4 purified monocytes in 100 μ L RPMIc per well in 96-well flat bottom plates and trained them according to the above-described workflow, using 10 μ g/mL of both β -glucan and HKMtb as training stimuli.

The bacterial concentration of HKMtb was determined by flow cytometry as follows.⁵⁴ Briefly, a calibration curve was prepared, using a log-phase growth culture of attenuated BCG, assuming that the whole culture was viable. Then, serial dilutions (1:2) were prepared and analyzed by flow cytometry. Bacterial viable units in the culture were calculated by plating in solid medium and counting colonies. Given that 10 μ g/mL of HKMtb contains 1.4×10^4 heat-killed bacteria, it corresponds to a 1:0.7 ratio (monocyte/HKMtb).

In vitro stimulation and peripheral blood mononuclear cell inhibition

When required, prior to β -glucan particles or HKMtb stimulation, cells were pre-incubated for 1 h with 1-mM epigenetic inhibitor 5'-Deoxy-5'-(methylthio) adenosine (MTA) (Sigma-Aldrich, Cat# D5011), or with 3- μ M Syk pathway inhibitor R406 (MedChemExpress, Cat# HY-11108), or for 16 h with 15- μ M HIF-1 α inhibitor PX-478 (MedChemExpress, Cat# HY-10231).

Chromatin immunoprecipitation assays (ChIP)

We plated 10^7 PBMCs in 5 ml RPMIc per well in p100 flat-bottom plates. On day 0, cells were stimulated with 10 μ g/mL HKMtb to a final volume of 10 ml RPMIc for 24 h at 37°C. Cells were then washed with PBS and allowed to rest for 5 days in RPMIc. ChIP was performed with the Magna ChIP A/G kit (Millipore) following the manufacturer's indications. Immunoprecipitated DNA and input DNA were amplified by means of quantitative PCR with specific primers for the promoter region of *TNFA*.

Lactate determination

PBMCs were stimulated with 10 μ g/mL of HKMtb in p96 wells as indicated before, collecting supernatants at the end of the 5-day resting period. Lactate was quantified in these supernatants (diluted 1/10) with the colorimetric assay Lactate Assay kit (Sigma-Aldrich).

HIF1A expression

We plated 10^7 PBMCs in 1 ml RPMIc per well in p6-well flat-bottom plates. On day 0, the cells were stimulated with 10 μ g/mL HKMtb to a final volume of 2 ml RPMIc, for 24 h at 37°C. Cells were then washed with PBS and allowed to rest for 5 days in RPMIc. At the end of the 5-day resting period, total RNA was extracted by means of the Monarch Total RNA Miniprep kit (New England Biolabs). The cDNA was prepared with the High-Capacity cDNA reverse transcription kit (Applied Biosystems). Quantitative PCR was performed with PowerUp SYBR Green Master Mix (Applied Biosystems) and primers for β -ACTIN and HIF1A. mRNA levels were normalized to β -ACTIN expression.

In vivo models of trained immunity

In one experiment, following a well-established *in vivo* trained immunity model,^{3,8} mice were intraperitoneally (i.p.) injected with 125 μ g HKMtb (8.75×10^5 bacteria) or PBS alone (as a control) on days -7 and -4. One week later, the mice were challenged i.p. with 5 μ g *E. coli* lipopolysaccharide serotype O111:B4 (Sigma-Aldrich, Cat# L4391) diluted in sterile PBS; 60 min later, blood was collected, clotted, and centrifuged, and serum was frozen at –20°C for subsequent analyses.

In another experiment, to evaluate an alternative administration route, mice were intranasally (i.n.) challenged with 12.5 μ g HKMtb (8.75×10^4 bacteria) or PBS 3 times a week for 2 weeks. One week after the last challenge, the mice were intraperitoneally administered 5 μ g *E. coli* lipopolysaccharide diluted in sterile PBS. Blood was collected 60 min later, and serum was frozen at –20°C for subsequent analyses.

Purified protein derivative assay

After intranasal administration of HKMtb according to the previous section, mice were euthanized before lipopolysaccharide rechallenge. Spleens were collected and single-cell suspensions prepared under sterile conditions. We plated 10^6 total splenocytes in U-bottom

96-well plates for 3 days, exposed or not to purified protein derivative (PPD), kindly provided by Dr. Aguiló. Supernatants were collected and IFN- γ analyzed by ELISA.

Cytokine measurement

A commercially available enzyme-linked immunosorbent assay (ELISA) kit (R&D Systems, Cat# DY210) was used to determine TNF- α concentration in cell culture supernatant samples from human donors. The manufacturer's instructions were followed to determine these concentrations.

The remaining cytokines (IL-4, IL-2, CXCL10 [IP-10], IL-1 β , CCL2 [MCP-1], IL-17A, IL-6, IL-10, IFN- γ , IL-12p70, CXCL8 [IL-8], TGF- β 1 [Free Active Form]) were measured in culture supernatants with the LEGENDplex human Essential Immune Response Panel (BioLegend, Cat# 740930), following the manufacturer's instructions. Briefly, plasma samples were incubated with premixed capture antibody-coated beads, washed, incubated with detection antibodies and streptavidin with phycoerythrin conjugate, acquired using a FACSCalibur flow cytometer (BD), and analyzed with Biolegend v8.0 software (Biolegend). Cytokines shown represent those whose detection was within the quantification range of the assay.

An ELISA kit was also used to determine TNF- α (R&D Systems, Cat# DY410) and IL-6 (R&D Systems, Cat# DY406) concentrations in mouse serum. The manufacturer's instructions were followed to determine these concentrations.

The remaining cytokines (IL-1 β , MCP-1, IL-12p70, GM-CSF, IL-23, IFN- γ , IFN- β , IL-17A, IL-27, and IL-10) were measured in mouse serum samples with the LEGENDplex Mouse Inflammation Panel, following the manufacturer's instructions (BioLegend, Cat# 740150).

QUANTIFICATION AND STATISTICAL ANALYSIS

Data are expressed as mean \pm SEM. Outliers were identified by means of a ROUT test. The D'Agostino–Pearson Normality test was performed for all the studied variables. We performed Student's t-test for 2 group comparisons of quantitative variables, either unpaired (t-test or Mann–Whitney) or paired (t-test or Wilcoxon). Only statistically significant differences were denoted, or *p*-values that might be visually confusing based on the charts. *P*-values are denoted as **p*<0.05, ***p*<0.01, ****p*<0.001 and explained in figure legends. In these figure legends, "N" represents the number of independent experiments performed and "n" the number of individuals included in the experiment shown, illustrated with single dots throughout. Prism Software was used to perform all the statistical analyses.