



## Complete Genome Sequence of a *Klebsiella pneumoniae* Strain Carrying *bla*<sub>NDM-1</sub> on a Multidrug Resistance Plasmid

💿 Sean Conlan,ª Anna F. Lau,ʰ NISC Comparative Sequencing Program,º Tara N. Palmore,ʰ Karen M. Frank,ʰ Julia A. Segreª

National Human Genome Research Institute, Bethesda, Maryland, USA<sup>a</sup>; National Institutes of Health Clinical Center, Bethesda, Maryland, USA<sup>b</sup>; NIH Intramural Sequencing Center, Rockville, Maryland, USA<sup>c</sup>

Here, we report the genome sequence of a  $bla_{NDM-1}$ -positive *Klebsiella pneumoniae* AATZP isolate cultured from a perirectal surveillance swab collected upon admission of a patient to the NIH Clinical Center in 2014. Genome sequencing of this isolate revealed three plasmids, including one carrying the  $bla_{NDM-1}$  gene encoding resistance to carbapenems.

Received 19 May 2016 Accepted 23 May 2016 Published 14 July 2016

**Citation** Conlan S, Lau AF, NISC Comparative Sequencing Program, Palmore TN, Frank KM, Segre JA. 2016. Complete genome sequence of a *Klebsiella pneumoniae* strain carrying *bla*<sub>NDM-1</sub> on a multidrug resistance plasmid. Genome Announc 4(4):e00664-16. doi:10.1128/genomeA.00664-16.

Copyright © 2016 Conlan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Sean Conlan, conlans@mail.nih.gov

Over the last two decades, there has been a steady and alarming global rise in multidrug-resistant bacteria (1). In particular, plasmid-borne carbapenemase-producing organisms (CPOs) have been cited as an immediate and even "catastrophic" threat to patient health (2, 3). The *Klebsiella pneumoniae* carbapenemase (KPC) enzyme has been the subject of considerable investigation because of its global prevalence (4, 5).

The New Delhi metallo-beta-lactamase (NDM-1), first identified in 2008, has become an important mechanism of carbapenem resistance with worldwide distribution (6). We previously sequenced a  $bla_{\rm NDM-1}$ -positive strain isolated in northeastern Ohio (7) but had never detected  $bla_{\rm NDM-1}$  in our hospital. In August of 2014, a *Klebsiella pneumoniae* isolate carrying the  $bla_{\rm NDM-1}$  gene was cultured from a perirectal surveillance swab collected upon admission. The patient came from India and had received extensive treatment in Indian hospitals for a malignancy. We performed whole-genome sequencing to identify whether the  $bla_{\rm NDM-1}$  gene was plasmid-borne and to enable the development of diagnostic assays for future epidemiological investigations.

Genomic DNA was prepared from an overnight culture, grown on blood agar using the Promega Maxwell 16 nucleic acid purification system (AS1030-tissue kit). Libraries for single-molecule real-time (SMRT) sequencing were constructed using the SMRTbell template kit, version 1.0. The DNA was size-selected for the range of 10 to 50 kb using a BluePippin with a 0.75% gel cassette. Sequencing was performed on the PacBio RSII using P5 polymerase binding and C3 sequencing kits with magnetic bead loading and 180-min acquisition. Genome assemblies were performed using HGAP3 and Quiver as part of SMRT Analysis version 2.3.

*Klebsiella pneumoniae* AATZP belongs to sequence type 147 and has a 5.35-Mb genome and three plasmids. pKPN-04f is 121,030 nucleotides (nt) and is largely syntenic with pKPHS1, a plasmid from *K. pneumoniae* HS11286 (accession no. NC\_ 016838) (8). Two regions of the pKPHS1 sequence are replaced in pKPN-04f, resulting in the loss of a CTX-M-14 extendedspectrum beta-lactamase and replacement of a restrictionmodification system. Interestingly, both pKPN-04f and pKPHS1 are predicted to be intact phages by the PHAST tool (9), suggesting a possible phage ancestor. The second plasmid, pKPN-041, is 38,384 nt and carries an aminoglycoside adenylyltransferase and two beta-lactamase genes. Finally, pNDM-1fa shares two large  $(\sim 20 \text{ kb})$  regions of identity (>99.8%) with plasmid 2 from the northeastern Ohio strain (7). The first region contains three antibiotic resistance genes: *bla*<sub>NDM-1</sub>, a downstream bleomycin resistance gene, and an upstream fluoroquinolone resistance gene. The second region contains plasmid partitioning genes and a restriction-modification system. In plasmid 2, those two regions are separated by a large region containing a tra conjugal transfer locus. In pNDM-1fa, that region is replaced with a class 1 integron that is 99.9% identical to one found in plasmids, like pHKU1 (10), and encodes resistance genes for a number of antibiotics, including aminoglycosides, beta-lactams, chloramphenicol, and rifampin. With few published complete bla<sub>NDM-1</sub>-positive Enterobacteriaceae genomes, these references serve to scaffold short-read assemblies for hospital transmission investigations and studies of plasmid evolution and diversity.

**Nucleotide sequence accession numbers.** This complete genome project has been deposited at DDBJ/EMBL/GenBank under the accession numbers CP014755 to CP014758.

## ACKNOWLEDGMENTS

This work was supported by the National Human Genome Research Institute and NIH Clinical Center Intramural Research Programs.

Sequencing was performed at the NIH Intramural Sequencing Center.

## FUNDING INFORMATION

This work, including the efforts of Sean Conlan, NFN NISC Comparative Sequencing Program, and Julia A Segre, was funded by HHS | NIH | National Human Genome Research Institute (NHGRI). This work, including the efforts of Anna F Lau, Tara N. Palmore, and Karen M Frank, was funded by HHS | NIH | NIH Clinical Center (Clinical Center).

## REFERENCES

1. Lockhart SR, Abramson MA, Beekmann SE, Gallagher G, Riedel S, Diekema DJ, Quinn JP, Doern GV. 2007. Antimicrobial resistance

among Gram-negative bacilli causing infections in intensive care unit patients in the United States between 1993 and 2004. J Clin Microbiol **45**: 3352–3359. http://dx.doi.org/10.1128/JCM.01284-07.

- Frieden T. 2013. Vital signs telebriefing on carbapenem-resistant *Enterobacteriaceae*. Centers for Disease Control and Prevention, Atlanta, GA. http://www.cdc.gov/media/releases/2013/t0305\_lethal\_cre.html.
- Department of Health. 2013. Antimicrobial resistance poses "catastrophic threat", says Chief Medical Officer. Press release. Department of Health, London, United Kingdom. https://www.gov.uk/government/new s/antimicrobial-resistance-poses-catastrophic-threat-says-chief-medicalofficer--2.
- Pitout JDD, Nordmann P, Poirel L. 2015. Carbapenemase-producing *Klebsiella pneumoniae*, a key pathogen set for global nosocomial dominance. Antimicrob Agents Chemother 59:5873–5884. http://dx.doi.org/ 10.1128/AAC.01019-15.
- Chen L, Mathema B, Chavda KD, DeLeo FR, Bonomo RA, Kreiswirth BN. 2014. Carbapenemase-producing *Klebsiella pneumoniae*: molecular and genetic decoding. Trends Microbiol 22:686–696. http://dx.doi.org/ 10.1016/j.tim.2014.09.003.
- 6. Dortet L, Poirel L, Nordmann P. 2014. Worldwide dissemination of the

NDM-type carbapenemases in Gram-negative bacteria. Biomed Res Int **2014**:249856. http://dx.doi.org/10.1155/2014/249856.

- 7. Van Duin D, Perez F, Rudin SD, Cober E, Hanrahan J, Ziegler J, Webber R, Fox J, Mason P, Richter SS, Cline M, Hall GS, Kaye KS, Jacobs MR, Kalayjian RC, Salata RA, Segre JA, Conlan S, Evans S, Fowler VG, Bonomo RA. 2014. Surveillance of carbapenem-resistant *Klebsiella pneumoniae*: tracking molecular epidemiology and outcomes through a regional network. Antimicrob Agents Chemother 58: 4035–4041. http://dx.doi.org/10.1128/AAC.02636-14.
- Liu P, Li P, Jiang X, Bi D, Xie Y, Tai C, Deng Z, Rajakumar K, Ou H-Y. 2012. Complete genome sequence of *Klebsiella pneumoniae* subsp. *pneumoniae* HS11286, a multidrug-resistant strain isolated from human sputum. J Bacteriol 194:1841–1842. http://dx.doi.org/10.1128/JB.00043-12.
- Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. Nucleic Acids Res 39:W347–W352. http://dx.doi.org/ 10.1093/nar/gkr485.
- Ho P-L, Chan J, Lo W-U, Lai EL, Cheung Y-Y, Lau TCK, Chow K-H. 2013. Prevalence and molecular epidemiology of plasmid-mediated fosfomycin resistance genes among blood and urinary *Escherichia coli* isolates. J Med Microbiol 62:1707–1713. http://dx.doi.org/10.1099/ jmm.0.062653-0.