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The Canine *POMC* Gene, Obesity in Labrador Retrievers and Susceptibility to Diabetes Mellitus

L.J. Davison (b), A. Holder, B. Catchpole, and C.A. O'Callaghan

Background: Diabetes mellitus (DM) in dogs is a common endocrinopathy with a complex genetic architecture. Disease susceptibility in several breeds is associated with polymorphisms in immune response genes, but in the Labrador retriever breed, no genetic associations with DM have been identified. A deletion in the pro-opiomelanocortin (*POMC*) gene in Labrador retrievers is associated with increased appetite and risk of obesity.

Hypothesis/Objectives: To characterize the *POMC* deletion in Labrador retrievers, to develop a simple genetic test for this mutation, and to test the hypothesis that the *POMC* gene deletion is associated with an increased risk of DM in this breed. **Animals:** Sixty-one non-diabetic Labrador retrievers aged >6 years and 57 Labrador retrievers with DM.

Ammais, Sixty-one non-diabetic Labrador retrievers aged 20 years and 57 Labrador retrievers with Divi.

Methods: Case–control genotyping study to compare the frequency of the *POMC* deletion in dogs with and without DM. After polymerase chain reaction (PCR) and sequencing to characterize the mutation, a PCR-based test was developed and validated using 2 different restriction fragment length polymorphism assays.

Results: A 14-base-pair deletion was confirmed and localized to exon 3 of the canine *POMC* gene. A PCR-based test for the deletion was successfully developed. There was no association between the presence of the *POMC* deletion mutation and DM in this population of Labrador retriever dogs (P = .31).

Conclusions and Clinical Importance: This study adds to the existing scientific literature indicating that there is little evidence for a direct link between obesity and DM in dogs.

Key words: Genetics; Endocrinology; Polymerase chain reaction; Pancreas.

Diabetes mellitus (DM) in dogs is a multifactorial disease, with genetic and environmental components thought to contribute to the development of the condition.^{1–3} The estimated prevalence of DM in dogs is 1 in 300^{4,5} and because most diabetic dogs are insulin-deficient, the disease has been compared to human type 1 diabetes and latent autoimmune diabetes of the adult.³ This comparison is supported by the presence of antibodies to pancreatic autoantigens in a number of cases^{6,7} and the lack of evidence of a direct link between obesity and development of diabetes in dogs, in contrast to human type 2 diabetes.⁸

The published genetic studies of DM in dogs are primarily based on a candidate gene approach.^{9–11} Within

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This paper has not been presented at any meetings.

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Abbreviations:

DM	diabetes mellitus
PCR	polymerase chain reaction
POMC	pro-opiomelanocortin
RFLP	restriction fragment length polymorphism
SNP	single nucleotide polymorphism

some breeds, certain haplotypes of the canine major histocompatibility complex (encoded by dog leukocyte antigen [DLA] genes) are associated with increased risk of DM,¹⁰ similar to the risk seen with some human leukocyte antigen (HLA) types and type 1 DM. More recently, using single nucleotide polymorphism (SNP) discovery and a candidate gene approach, polymorphisms in several other immune response and cytokine genes have been implicated as genetic risk factors in certain breeds.^{3,11–13} Polymorphisms in genes including those encoding IFNgamma, IL-10, IL-12 beta, IL-6, IL-4, PTPN22, and TNF-alpha have been implicated, with associations appearing to be most common in moderate risk breeds including terriers and spaniels. However, the genetic architecture of DM risk in dogs, like that of human type 1 and type 2 DM, is complex and it is also possible that specific genetic predispositions differ between breeds. Many SNPs are associated with protection or susceptibility in only a single breed, and interestingly, some SNPs are monomorphic (only 1 allele) within some breeds.^{11,12} Where DM risk variants are detected, they might only be important within a single breed (eg, IL10 polymorphisms in the Cavalier King Charles spaniel¹²).

A recent candidate gene study¹⁴ reported that a 14-base-pair deletion in the canine pro-opiomelanocortin (*POMC*) gene is associated with food motivation and obesity in Labrador retriever dogs. *POMC* mutations have also been associated with obesity in humans, with a subsequent increase in risk of obesity-related diseases

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Table 1. Descriptive statistics of the study population.

	Median Age	Male	Male	Female	Female	Sex	Total Number
	(Years) [Range]	Neutered (%)	Entire (%)	Neutered (%)	Entire (%)	Unknown (%)	of Dogs
Control population	9 [6–13]	19 (31)	13 (21)	25 (41)	2 (3)	2 (3)	61
Diabetic population	9 [6–13]	18 (32)	14 (25)	25 (43)	0 (0)	0 (0)	57

such as type 2 diabetes.¹⁵ Although there is very little evidence that obesity is a major risk factor for DM in dogs, obese dogs are hyperinsulinaemic.¹⁶ Additionally, in a recent epidemiological study of DM in first opinion practice in the UK, being recorded as being overweight was associated with a diagnosis of DM.⁴ Furthermore, research into the links between canine obesity and nondiestrus diabetes has not been undertaken from a breed-specific perspective, so this does not rule out an association between obesity and diabetes in certain breeds, similar to the association between type 2 DM and obesity in humans. Therefore, given the absence of any reported immune response gene associations with DM in Labrador retrievers, we hypothesized that the obesity-associated and appetite-associated POMC deletion might also be associated with increased risk of DM in Labrador retrievers, which may consequently be

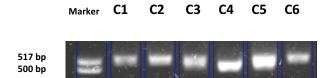


Fig 1. Agarose gel electrophoresis after the amplification of a 520-bp region of genomic DNA from 6 Labrador retriever dogs, using primers designed to flank exon 3 of *POMC* in the amplicon. The variability in the size of the amplicons indicates that some dogs were heterozygous for a 14-base-pair deletion (C3 and C5) and 1 dog was homozygous for the deletion (C4). This was confirmed by sequencing.

more similar to human type 2 than type 1 DM. The aims of this study were to develop a simple genetic test for the reported *POMC* deletion in Labrador retrievers and to use this test in a population of Labrador retrievers (DM-affected and unaffected) to determine whether the *POMC* deletion is present at a higher frequency in the diabetic population, supporting a potential role for obesity as a risk factor for DM in this breed.

Materials and Methods

Study Population

Diabetic Dogs. EDTA blood samples from diabetic Labrador retrievers were retrieved from the UK Canine Diabetes Register and Archive, at the Royal Veterinary College, University of London. The diagnosis was made by primary care veterinary practitioners, based on compatible clinical signs of polydipsia, polyuria, and weight loss and documented persistent hyperglycemia (>162 mg/dL), an elevation in serum fructosamine concentration, or both. Most dogs were receiving insulin treatment at the time of sampling. Dogs with early-onset diabetes (ie, <1 year of age), concurrent hyperadrenocorticism, and female entire dogs (likely to have diestrus diabetes), were excluded from the study.

Control Dogs. EDTA blood samples from non-diabetic Labrador retrievers over 6 years of age were selected from an archive of residual clinical samples at the Royal Veterinary College, University of London. Dogs over 6 years of age were selected because this provided appropriate age-matching for the diabetic samples and limited the likelihood of these dogs developing diabetes later in life.

All samples used for this study were initially collected for diagnostic purposes, and the residual blood volume remaining after completion of diagnostic testing was stored at -20° C until DNA extraction. Storage was undertaken with informed owner consent

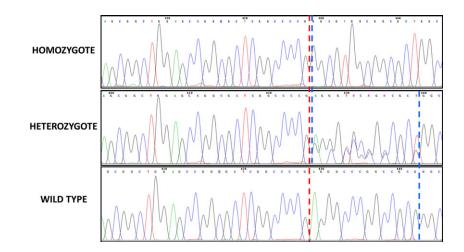


Fig 2. Sanger sequencing traces from dogs with homozygous, heterozygous, and wild-type genotypes for the 14-base-pair *POMC* deletion. The start point of the deletion is illustrated by a red-dashed line and the end point by a blue-dashed line.

and with institutional ethical review board approval from the Royal Veterinary College Ethics and Welfare Committee. DNA was extracted from EDTA blood using either a standard phenol: chloroform method or a Qiagen Blood and Tissue DNA extraction kit^a and sample quality was evaluated with a Nanodrop analyser^b before polymerase chain reaction (PCR). Where necessary, samples were re-extracted, diluted, or both to a suitable working concentration (5–20 ng/µL) before use.

Detection and Confirmation of POMC Mutation

Primers were designed to amplify a 560-base-pair (bp) fragment, containing canine *POMC* exon 3 (K9POMC-1 sense primer: 5'-GCGGCCCTAACCCTCTG-3' and K9POMC-2 antisense primer: 5'-GCCACCAGGCCGTACTC-3'), with an annealing temperature of 66°C found to be optimal. Primer 3 software¹⁷ was used to design the primers, and the *Canis lupus familiaris POMC* gene transcript (Transcript: POMC-201 ENSCAFT00000006656) was downloaded from Ensembl at http://www.ensembl.org/Canis_familiaris/Info/Index for this purpose. Primers were supplied by IDT

technologies^c, and all PCR reactions were performed on an MJ PTC-225 thermal cycler^d using Hi-fidelity Hotstart Q5 enzyme^c.

Amplicons were visualized under UV light with ethidium bromide, after 5% agarose gel electrophoresis at 100 V for 2 hours to allow optimal resolution of bands. Polymerase chain reaction products were purified with a Qiagen PCR cleanup kit^a and submitted for Sanger sequencing^f in a forward and reverse direction using the same primers as used for PCR. Based on these results, a further primer set was designed and tested, with the identified deletion located more centrally within the PCR product. This primer pair was used in all further PCR reactions (K9POMC-3 sense primer: 5'-AAGTACGTCATGGGCCACTT-3' and K9 POMC-4 antisense primer: 5'-TTTTGAACAGCGTCACCAGG-3') at an optimal annealing temperature of 68°C.

Where indicated, restriction fragment length polymorphism (RFLP) analysis was undertaken on PCR products in $1 \times$ NEB CutSmart buffer and either *ApaI* enzyme^e or *SacII* enzyme^e for 1 hour at 37°C. The online software NEB Cutter v2.0 at http:// nc2.neb.com/NEBcutter2/ was used to determine suitable enzymes for this purpose. After digestion, DNA was separated on 4%

A WT-LAB DEL-LAB ENSEMBL REFSEQ	GCTGGAG-CCGGCGCTCGGCCCCGAGGGCCCCGCCGCGGGCGTGGC GCTGGAG-CCGGCGCTCGGCCCCG-agGGCCCCCCCGCGGGCGTGGC GCTGGAG-CCGGCGCTCGGCCCC-agGGCCCCCCCGCGGGCGTGGC GCTGGAGNCCGGCGCTCGGCCCC-AGGGCCCCGCCGCGGGCGTGGC *******
В	
DEL-LAB UNIPROT WT-LAB	MPRSCCSRPGALLLALLLQASVEVSGWCLESSQCQDLTTESNLLACIRACKPDLSAETPV MPRSCCSRPGALLLALLLQASVEVSGWCLESSQCQDLTTESNLLACIRACKPDLSAETPV MPRSCCSRPGALLLALLLQASVEVSGWCLESSQCQDLTTESNLLACIRACKPDLSAETPV ************************************
DEL-LAB UNIPROT WT-LAB	LPGNGDEQPLAENPRKYVMGHFRWDRFGRRNGSAGQKREEEEVAAGGGRAPLPAGGPGPR LPGNGDEQPLAENPRKYVMGHFRWDRFGRRNGSAGQKREEEEVAAGGGRAPLPAGGPGPR LPGNGDEQPLAENPRKYVMGHFRWDRFGRRNGSAGQKREEEEVAAGGGRAPLPAGGPGPR **********************
DEL-LAB UNIPROT WT-LAB	GDGGELGLQEGKRSYSMEHFRWGKPVGKKRRPVKVYPNGAEDESAEAFPVEFKKELARQR GDGGELGLQEGKRSYSMEHFRWGKPVGKKRRPVKVYPNGAEDESAEAFPVEFKKELARQR GDGGELGLQEGKRSYSMEHFRWGKPVGKKRRPVKVYPNGAEDESAEAFPVEFKKELARQR *************************
DEL-LAB UNIPROT WT-LAB	LEPALGP-GR <i>GGAGRPGVRPGGGGRGGREGGREALQDGALPLGQPAQGQALRRLHELGEEP</i> LEPALGP-GPAAGVAALADLEYGLVAEAGAAEKKDDGPYKMEHFRWGSPPKDKRYGGFMSS LEPALGP <mark>B</mark> GPAAGVAALADLEYGLVAEAGAAEKKDDGPYKMEHFRWGSPPKDKRYGGFMSS ******* * * *
DEL-LAB UNIPROT WT-LAB	DAPGDAVQKRHHQERPQEGPVRAQRGRRLLFPGSPRKAPLLPALLPSRPPPAPDVLLAQA ERSQTPLVTLFKNAIIKNAHKKGQ* ERSQTPLVTLFKNAIIKNAHKKGQ*
DEL-LAB UNIPROT WT-LAB	VGGSPVVSLLKLPVVRK*

Fig 3. (A) illustrates a fragment of the canine *POMC* gene, demonstrating the delta-exon 3 canine POMC allele (DEL-LAB), missing 14 nucleotides compared with the wild-type allele (WT-LAB). Ensembl transcript POMC-201 ENSCAFT00000006656 (ENSEMBL) in this region is also shown, with a putative 2 base-pair intron (shown in lower case). The RefSeq (REFSEQ) transcript in this region is also shown, containing a single putative extra nucleotide "N" (in bold) (The NCBI website notes that the sequence of the model RefSeq transcript was modified relative to its source genomic sequence to represent the inferred CDS: inserted 1 base in 1 codon; Derived by automated computational analysis using gene prediction method: Gnomon). Neither the RefSeq nor the Ensembl canine POMC transcript shows the G nucleotide (in bold) which was present in both the wild-type and delta-exon 3 Labrador retriever POMC allele in this study. The deleted nucleotide sequence is **AGGGCCCGGCCGCG**, which differs by 1 nucleotide from the Ensembl canine reference sequence (Ensembl transcriptPOMC-201 ENSCAFT00000006656) at position 8 of 14 (C>G) of the 14-bp deleted fragment (shown in white). (**B**) illustrates potential impact of the 14-base-pair deletion on the amino acid sequence of wild-type Labrador retriever POMC (WT-LAB) and delta-exon 3 *POMC* (DEL-LAB) compared to the published UNIPROT sequence (http://www.uniprot.org/uniprot/E2RQ39.fasta). The WT sequence differs from the UNIPROT sequence by 1 amino acid, highlighted in black and white. The 14-base-pair delta-exon 3 allele leads to a frameshift mutation with the subsequent amino acid sequence being shown in italics. This frameshift disrupts the 90 amino acid terminal region of the protein fragment of *POMC* which is cleaved to form beta lipotropin and subsequently cleaved further to form beta endorphin and beta melanocyte-stimulating hormone.

agarose gels containing ethidium bromide for 60 minutes at 120 V and visualized under UV light. Sequence alignments were performed at http://www.bioinformatics.org/sms2/ and functional evaluation of the deletion at http://www.uniprot.org/uniprot/E2RQ39.

Statistical analysis was undertaken by R version 3.2.3.¹⁸ A chisquared analysis was performed to compare allele frequencies between diabetic and non-diabetic Labrador retrievers, using a 3×2 contingency table with a significant *P* value set at .05.

Results

A total of 118 Labrador retriever dogs were included in the study, of which 57 were diabetic and 61 were non-diabetic. The characteristics of these dogs are reported in Table 1. The mutation in the canine *POMC* gene was reported as a 14-base-pair deletion in exon 3,¹⁹ but more precise details of the deletion had not been published before this study being undertaken. To confirm this finding, a region of the *POMC* gene containing exon 3 was amplified (K9POMC-1 and K9POMC-2 primers) and sequenced in a subset of the dogs (Fig 1). The predicted amplicon size was 520 bp (wild-type allele) and 506 bp (POMC deletion allele).

Sanger sequencing was used to confirm the presence of a 14-base-pair deletion in the canine POMC gene, as previously reported¹⁹ (Fig 2), as this work was undertaken before the full publication of the mutation sequence.¹⁴ There is some ambiguity in the annotation of the dog genome sequence containing this deletion, comparing different databases. Based on the RefSeq published canine POMC gene (http:// www.ncbi.nlm.nih.gov/nuccore/XM_844370.4), the deletion lies in exon 3. However, based on the Ensembl transcript (POMC-201 ENSCAFT0000006656), rather than being in exon 3 as reported in an initial abstract,¹⁹ this deletion includes a small 2-base-pair "intron" (between exons 3 and 4), plus the first 12 base pairs of exon 4 (Fig 3A). Direct sequencing of genomic DNA from the Labrador retriever dogs indicated that the "N" at position 547 of the RefSeq coding sequence was absent and that there was an additional G nucleotide base at position 562, immediately preceding the AG, annotated as an intron in the Ensembl sequence (Fig 3A). Hereafter, the deletion is referred to as the delta-exon 3 allele.

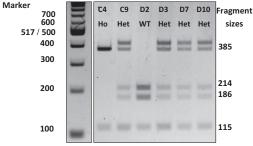
Once the mutation had been defined in detail, 2 new primer pairs were designed (K9POMC-3 and K9POMC-4) flanking the mutation in the centre of the amplicon, which facilitated RFLP analysis. An amplicon length difference as small as 14 base pairs can be difficult to resolve by agarose gel electrophoresis, particularly in heterozygotes; therefore, the wild-type and delta-exon 3 sequences were also examined for differential restriction enzyme cutting sites. Two restriction enzymes, ApaI and SacII were tested for this purpose. ApaI was predicted to cut the wild-type product into 4 fragments (211, 131, 107, and 85 bp), but would cut the delta-exon 3 allele into only 3 fragments (282, 131 and 107 bp). SacII was predicted to cut the wild-type product into 4 fragments (214, 185, 116, and 19 bp), but

would cut the delta-exon 3 allele into only 3 fragments (385, 116, and 19 bp). Agarose gel electrophoresis confirmed these RFLP profiles (Fig 4) with heterozygous dogs displaying a characteristic triple band, due to DNA heteroduplex formation. Using standard PCR, *ApaI/SacII* digestion, and agarose gel electrophoresis, 11 diabetic and 11 control dogs were screened for the delta-exon 3 mutation, demonstrating that this

A Undigested PCR products

Mark

(bp



B RFLP analysis following digestion with Apal enzyme

ker)	700 600 517 / 500 400 300		C4 Ho		D2 WT			Fragment sizes
	200]	-		-			282
	100	-	-	11		1	-	131 108

C RFLP analysis following digestion with SacII enzyme

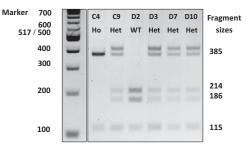


Fig 4. (A) Agarose gel electrophoresis of polymerase chain reaction (PCR) products after amplification of a 534-base-pair fragment of the canine *POMC* gene in 6 Labrador retriever dogs (C4, C9, D2, D3, D7, and D10). A combination of low voltage, high agarose gel content, and long running time allowed separation of the PCR products from dogs which had homozygous (Ho) or heterozygous (Het) or wild-type (WT) genotypes with respect to the 14-base-pair POMC deletion. The triple band in the heterozygotes represents the deleted allele, the wild-type allele, and a heteroduplex of both alleles. The same PCR products were digested with ApaI enzyme (**B**) or SacII enzyme (**C**) providing confirmation of the genotype, as the restriction enzyme digestion sites are not present in the POMC deletion allele. approach allowed the genotypes to be accurately identified.

Using this PCR/RFLP assay, the diabetic and nondiabetic Labrador retrievers were genotyped for the delta-exon 3 mutation, the results of which are summarized in Table 2. The minor allele frequency of the mutation in the total population of all 118 dogs was 0.207. No significant association was identified between the presence of the *POMC* delta-exon 3 allele and a diagnosis of DM in this population of Labrador retrievers.

Discussion

This study aimed to develop a simple genetic test for the POMC mutation in Labrador retrievers and to test the hypothesis that the deletion, previously associated with appetite and obesity in this breed,¹⁹ is also associated with risk of DM. Pro-opiomelanocortin is produced mainly by the pituitary gland and undergoes a wide range of post-translational cleavage and modification events in a tissue-specific manner²⁰ to generate melanocortins such as adrenocorticotropic hormone (ACTH), alpha, beta, and gamma melanocyte-stimulating hormones (MSH). POMC had been selected as a candidate gene in the canine obesity study^{15,19} because of the role of melanocortins in energy metabolism and appetite.^{14,15} In veterinary medicine, a polymorphism in the melanocortin 4 receptor (MC4R) gene has been associated with DM in overweight cats²¹ but to date, no associations between melanocortin/melanocortin receptor polymorphisms and DM in dogs have been established.¹⁹ Although obesity has not been directly linked to DM in dogs, it has been shown to cause insulin resistance²² and other metabolic derangements²³ which may have a detrimental impact on pancreatic function or glycemic control, and a recent epidemiological study reported that being overweight was associated with a diagnosis of DM in dogs.⁴

Using PCR and Sanger sequencing, the 14 bp *POMC* deletion was localized to a region of exon 3. This region of canine *POMC* exon 3 is not particularly well characterized in the dog genome assembly, with differences apparent when comparing genome browsers. Direct sequencing revealed that both RefSeq and Ensembl sequences (accessed April 25, 2016) were likely incorrect. The 2-base-pair "intron" in the Ensembl sequence is not valid as this cannot be spliced out of the primary transcript and might, instead, represent an acceptor site for

an alternative mRNA transcript with the donor site after exon 2. If this were the case, the resulting mRNA species would encode a protein lacking in γ -melanotropin, ACTH, and α -melanotropin, which seems unlikely. An additional G nucleotide was identified immediately upstream of the "AG" splice site, which, when combined with the AG and translated would result in an encoded protein more consistent with that found in other species. Taking this modified sequence as the reference, the 14base-pair deletion would lead to loss of 4 amino acids and a frameshift that would disrupt the coding sequence (Fig 3B), with a potential impact on expression of beta lipocortin, beta melanocyte-stimulating hormone, and beta endorphin.¹⁴

The RFLP analysis performed in this study confirmed the removal of a cleavage sites for both the *ApaI* enzyme and the *SacII* enzyme by the delta-exon 4 *POMC* deletion and allowed validation of a simple strategy for *POMC* genotyping based on PCR and agarose gel electrophoresis.

The results of genotyping 61 control and 57 diabetic Labrador retrievers did not demonstrate any association between the delta-exon 3 *POMC* deletion and susceptibility to DM. There are several possible reasons for this, including the presence of phenocopies of disease within this population, the potentially diverse genetic backgrounds of non-pedigree dogs identified by their owners as Labrador retrievers or the possibility that diabetes risk in Labrador retrievers is driven by a wide range of genes, each with small effects.

In addition to a genuine lack of association between POMC and DM in dogs, there are several other potential reasons for the findings of this study. It is possible that the study is underpowered, although this appears unlikely because the genetic architecture of dogs favors small sample sizes for within-breed genetic studies.²⁴ and the candidate gene study of POMC in which an association with obesity and appetite was initially discovered involved only 15 obese and 20 lean dogs.¹⁹ Unfortunately, data regarding weight and body condition score of the dogs included in the present analysis were not available, which is a considerable limitation, as DM is known to be a complex disease with both genetic and environmental triggers.¹ It is possible for example that maintenance of a lean bodyweight, even in an individual prone to obesity because of their deltaexon 3 *POMC* genotype, may override any increase in diabetes risk afforded by the genotype. This could be explored by a prospective study in which details about

 Table 2. Genotype data for 61 non-diabetic and 57 diabetic Labrador retrievers for the POMC delta-exon 4 mutation and chi-squared analysis of the distribution of alleles across the 2 populations.

	Control Dogs						
POMC Delta-Exon 3 Genotype	Observed	Expected	Chi-Squared	Observed	Expected	Chi-Squared	All Dogs
Wild-type	35	38.25	0.28	39	35.75	0.3	74
Heterozygous deletion	24	20.16	0.73	15	18.84	0.78	39
Homozygous deletion	2	2.58	0.13	3	2.42	0.14	5
Total	61			57			118

The chi-square statistic is 2.36. The *P*-value is .31. The result is not significant at P < .01.

appetite and obesity are collected alongside diabetes data. Additionally, although control dogs were selected to be over 6 years of age, to reduce the likelihood of dogs in this group developing DM in future, it is still possible some control dogs might still develop diabetes in later life, which is another limitation of this study.

Conclusions

This study finds no evidence that a common *POMC* 14-base-pair deletion in Labrador retrievers, reported to be associated with obesity and appetite, is associated with risk of DM in this breed. Splicing events in the canine *POMC* gene have not been fully elucidated, but the deletion is likely to cause a frameshift in the protein. Restriction fragment length polymorphism-based genotyping of the mutation can be performed rapidly and accurately using PCR, *ApaI*, or *SacII* enzyme digestion and agarose gel electrophoresis. This study adds to the body of evidence that supports a basis for DM in the Labrador retriever that is not obesity-driven.

Footnotes

- ^a Qiagen, Hilden, Germany
- ^b Nanodrop, Wilmington, DE
- ^c Coralville, IA
- ^d Bio-Rad, Hercules, CA
- ^e NEB, Ipswich, MA
- ^f BigDye 3.1 sequencing reagent, Thermo Fisher, Paisley, UK

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Conflict of Interest Declaration: Authors declare no conflict of interest.

Off-label Antimicrobial Declaration: Authors declare no off-label use of antimicrobials.

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