



Published in final edited form as:

*Oncogene*. 2015 May 28; 34(22): 2815–2822. doi:10.1038/onc.2014.238.

## DNA damage response and prostate cancer: defects, regulation and therapeutic implications

**Styliani Karanika, Theodoros Karantanos, Likun Li, Paul G. Corn, and Timothy C. Thompson**

Department of Genitourinary Medical Oncology – Research, The University of Texas MD Anderson Cancer Center, Houston, TX

### Abstract

DNA damage response (DDR) includes the activation of numerous cellular activities that prevent duplication of DNA lesions and maintain genomic integrity, which is critical for the survival of normal and cancer cells. Specific genes involved in the DDR such as *BRCA1/2* and *P53* are mutated during prostate cancer progression, while various oncogenic signaling such as Akt and c-Myc are activated, enhancing the replication stress and increasing the genomic instability of cancer cells. These events may render prostate cancer cells particularly sensitive to inhibition of specific DDR pathways, such as PARP in homologous recombination (HR) DNA repair and Chk1 in cell cycle checkpoint and DNA repair, creating opportunities for synthetic lethality or synergistic cytotoxicity. Recent reports highlight the critical role of androgen receptor (AR) as a regulator of DDR genes, providing a rationale for combining DNA-damaging agents or targeted DDR inhibitors with hormonal manipulation or AR inhibition as treatment for aggressive disease. The aims of this review are to discuss specific DDR defects in prostate cancer that occur during disease progression, to summarize recent advances in understanding the regulation of DDR in prostate cancer, and to present potential therapeutic opportunities through combinational targeting of the intact components of DDR signaling pathways.

### Keywords

DNA damage response; prostate cancer; DDR; defects; therapeutic implications

### INTRODUCTION

Prostate cancer remains the second most common cancer type in western societies. Despite recent therapeutic advances, metastatic castration-resistant prostate cancer (mCRPC) is incurable, and novel treatment approaches are needed. Abiraterone, a selective CYP17

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:[http://www.nature.com/authors/editorial\\_policies/license.html#terms](http://www.nature.com/authors/editorial_policies/license.html#terms)

**Correspondence to:** Dr. TC Thompson, Department of Genitourinary Medical Oncology – Research, Unit 18-3, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Boulevard, Houston, TX 77030, USA. Tel.: +1 713-792-9955; Fax: +1 713-792-9956; [timthomp@mdanderson.org](mailto:timthomp@mdanderson.org).

S. Karanika and T. Karantanos contributed equally to this work.

### CONFLICT OF INTEREST

The authors declare no conflict of interest.

inhibitor that decreases androgen biosynthesis within the tumor microenvironment, prolongs overall survival before and after chemotherapy in patients with mCRPC, but the duration of benefit is modest when compared to placebo (~5 and ~4 months, respectively).<sup>1, 2</sup> Similarly, enzalutamide, a potent androgen receptor (AR) inhibitor that impairs the nuclear translocation of AR, prolongs overall survival before and after chemotherapy in patients with mCRPC—but again only modestly, by ~2 and ~5 months, respectively, when compared to placebo.<sup>3, 4</sup> Thus, novel therapeutic strategies are needed.

Multiple lines of evidence link AR signaling to the DNA damage response (DDR) in prostate cancer cells. Clinically, radiation therapy (which promotes apoptosis via DNA damage) is more effective when combined with androgen-deprivation therapy (ADT) in treating patients with high-risk localized disease.<sup>5</sup> Apart from additive apoptotic effects of AR inhibition and radiation therapy, recent preclinical studies have elucidated a novel mechanism to explain this observation, whereby AR signaling regulates multiple genetic activities and pathways that influence the DDR.<sup>6, 7</sup> These results suggest that the exploration of these complex association may provide novel therapeutic opportunities for patients with mCRPC.

DNA is continually exposed to various insults causing a range of lesions such as single strand breaks (SSBs), double strand breaks (DSBs), bulky adducts, base mismatches, insertions and deletions and base alkylation.<sup>8</sup> Genomic instability refers to a high frequency of alterations within the genome of a cellular lineage. As genomic instability is deleterious to the organism, normal mammalian cells possess exquisite response mechanisms to avoid accumulation of DNA damage and maintain genomic integrity.<sup>9</sup> These mechanisms are known collectively as the DDR and include detection of DNA damage, accumulation of DNA repair factors and physical repair of the lesion.<sup>8</sup> This critical response program has two very well coordinated functions: (i) to prevent duplication and partitioning of the lesion into daughter cells and (ii) to repair the lesion. The cellular actions that manifest as cell cycle arrest following DNA damage are known as “checkpoint” functions and are considered as a critical part of the DDR.<sup>10</sup> Depending on the severity of the lesion and the capacity of the DDR system to repair it, cells will resume proliferation, become senescent (a state of irreversible cell cycle arrest), or undergo programmed cell death (apoptosis) to remove damaged DNA from the cellular population.<sup>10, 11</sup>

SSBs and DSBs are detected by specialized complexes recruiting ataxia-telangiectasia mutated (ATM) and ataxia-telangiectasia and Rad3 related (ATR) at the site of the lesion, leading to increased phosphorylation of H2AX. ATM recruitment, which is mainly related to DSBs leads to Chk2 activation and subsequent stabilization of p53, promoting G1/S cell cycle arrest through p21, and providing cells with time to repair the damage avoiding the replication of the damaged DNA.<sup>12</sup> ATR on the other hand which is mainly recruited to the site of SSBs phosphorylates and activates Chk1 which in turn phosphorylates Cdc25 and Wee1, leading to S and G2/M arrest replication recovery and initiation of DNA repair.<sup>12–15</sup>

Homologous recombination (HR) and non-homologous end joining (NHEJ) are the main mechanisms implicated in the repair of DSBs after the initial recognition.<sup>16,17</sup> HR takes place during the S phase of the cell cycle and promotes removal of a part of the DNA

(including the DSB) by using the homologous sister chromatid to mediate synthesis of new DNA. BRCA1, BRCA2, RAD51 and PALB2 are components of the enzymatic machinery involved in this process. NHEJ promotes DSB repair by joining the ends of the lesion together throughout the cell cycle through the function of DNA-PK complex, which consists of the Ku70 – Ku80 heterodimer and the DNA-PK catalytic subunit (DNA-PKcs).<sup>8, 18</sup> Interestingly, recent data suggest that AR regulates NHEJ and subsequently impacts DDR in prostate cancer cells.<sup>19</sup> It should be noted that both HR and NHEJ can be mutagenic but particularly NHEJ can cause deletion or mutation of DNA sequences at or around the DSB site.<sup>8</sup>

In contrast to normal cells, a universal characteristic of cancer cells is genomic instability due to defects in the mechanisms involved in repair of DNA damage. It is notable that genomic instability was found to be associated with worse prognosis in patients with prostate cancer.<sup>20</sup> Familial forms of breast and ovarian cancer are associated with mutations in homologous recombination (HR)–related genes, such as *BRCA1* and *BRCA2* (ref.21) while carriers of *BRCA1* and *BRCA2* mutations have an increased risk of prostate cancer.<sup>22,23</sup> Moreover, disruption of the ATM pathway, though loss of ATM itself or of downstream effector protein p53, has been observed in as many as 70% of tumors.<sup>24,25</sup> According to a recent study *P53* and *ATM* were found to be mutated in 40% and 8%, respectively, of CRPC cases examined by sequence analysis.<sup>26</sup> It is believed that dysfunctional DDR leads to accumulation of DNA lesions and promotes development of a precancerous phenotype<sup>27</sup>, while inactivation of critical DDR mediators such as ATM, and p53 leads to development of malignancy.<sup>27,28</sup>

Importantly, recent data suggest that defects in one component of DDR (e.g., *BRCA1* and *BRCA2* mutations affecting HR, implicated in the repair of double strand breaks [DSBs]) renders cancer cells specifically susceptible to inhibition of a second DDR defect (e.g., PARP1 inhibition affecting base excision repair [BER], implicated in the repair of single strand breaks [SSBs]), a concept referred to as “synthetic lethality”.<sup>29,30</sup> Synthetic lethality is defined as a type of genetic interaction whereby the co-occurrence of two genetic events results in cellular death. The presence of either event alone has no effect on cell viability, but the combination of the two leads to cell death. These events are detected only in cancer cells, so normal cells are spared, reducing the toxicity of therapy. In this context, DDR is critical for cancer cell survival but represents one event that can be exploited for therapy using agents that target a second event.

The pro-survival activities of DDR are realigned and misappropriated during cancer progression by activation of oncogenes such as Ras, Akt and Myc, which enhances replication stress.<sup>8,31–33</sup> Replication stress is defined as the harmful effect of DNA that is only partially replicated because of slow progression (or “stall”) of the replication forks, which can be caused by oncogene-induced hyper-replication that activates multiple origins of replication per S phase, by nucleotide pool imbalance or by DNA damage.<sup>34</sup> This leads to increased DNA damage, which eventually increases genomic instability to a level that is incompatible with cell survival. However, induction of replication stress by hyperactive growth factor and oncogene signaling in established cancer can lead to compensatory upregulation of DNA repair pathways, establishing a new paradigm in cancer therapy.<sup>34</sup>

Given the role of genomic instability in prostate cancer progression, recent evidence that agents targeting DDR may be effective in a subset of patients with prostate cancer and reports that AR signaling regulates multiple genetic activities and pathways that influence DDR, the aim of this review was defined to describe DDR involvement in development and progression of prostate cancer, crosstalk between AR and DDR signaling pathways, and therapeutic opportunities that result from targeting DDR especially at the lethal stage of this disease.

## DNA DAMAGE RESPONSE AND PROSTATE CANCER

### DNA damage response and prostate carcinogenesis

Inflammation is an important factor in prostate carcinogenesis. Regardless of etiology, inflammation produces cellular and genomic damage, induces secretion of cytokines and growth factors promoting cellular proliferation and angiogenesis and becomes more extensive over the lifetime of the individual.<sup>35–37</sup> Inflammatory lesions generate free radicals (e.g., nitric oxides and single oxygen species released from phagocytic inflammatory cells) that cause severe oxidative DNA damage within prostate epithelial cells. These molecular changes result in increased risk of permanent mutations, as the damaged cells may proliferate.<sup>38–41</sup> According to the results of a recent study which used an in vitro model of prostate cell inflammation exposure androgen sensitive prostate cancer cells to inflammatory cytokines led to loss of AR and downregulation of p53 signaling.<sup>42</sup> Interestingly, the administration of androgens restored p53/p21 function, inhibiting uncontrolled tumor growth related to DNA damage and genomic instability.<sup>42</sup>

Recent reports suggest a role for AR in the response to DNA damage during development of prostate cancer. In particular, Ide *et al.* showed that AR activation by testosterone promotes ATM activation and Chk2 phosphorylation in response to H<sub>2</sub>O<sub>2</sub>-induced DNA damage.<sup>43</sup> The authors suggested that testosterone suppresses prostate cancer initiation through activation of the DDR. These results are consistent with previous reports demonstrating that low testosterone levels are associated with advanced tumor stage, positive surgical margins and shorter overall survival.<sup>44,45</sup> Alternatively, other reports have associated high percentage of free testosterone with high grade prostate cancer<sup>46</sup>, highlighting the complexity of the relationship between AR and DDR with regard to prostate carcinogenesis.

A more recent report suggests that AR inactivation leads to telomere dysfunction, contributing to genomic instability and progression of prostate cancer.<sup>47</sup> Bowen *et al.* found that the prostate cancer suppressor NKX3.1, which is a target of AR, activates ATM, enhancing the DDR and thus contributing to DNA integrity in prostate epithelial cells.<sup>48</sup> Notably, ATM missense mutations and polymorphisms increase the risk of prostate cancer development.<sup>49,50</sup> According to these data, it is conceivable that impaired DDR may promote prostate carcinogenesis while AR may maintain genomic integrity in the earlier stages of the disease through DDR activation, mainly by activating the ATM/Chk2 pathway. Further, it is believed that reactive oxygen species (ROS)-induced unrepaired DNA damage may be one of the main mechanisms related to initiation of this disease.<sup>51</sup> Finally, it has been suggested that mutational or epigenetic inactivation of DDR components is selected for during neoplastic development, allowing malignant progression.<sup>8</sup>

## DDR defects related to prostate cancer

In established prostate cancers, multiple defects in various DDR components have been described. p53 was found to be mutated in 3–20% of prostate cancers at diagnosis<sup>52,53</sup> and its dysfunction has been associated with high-grade disease<sup>54</sup>, cancer recurrence, castration resistance and metastasis.<sup>55</sup> Wild-type p53 is rapidly activated and stabilized in response to a range of genotoxic insults, including ionizing irradiation<sup>56</sup>, UV light<sup>57</sup> and ribonucleotide deletion.<sup>58</sup> This results in cell cycle arrest in G<sub>1</sub> and G<sub>2</sub> phases through induction of p21, 14-3-3 $\sigma$  and GADD45 $\alpha$  and in apoptosis through upregulation of PUMA, BAX and BAK.<sup>59</sup> Inactivation of p53 renders cancer cells dependent on p53-independent mechanisms to promote cell cycle arrest for DNA damage repair, specifically ATR-Chk1– and ATM-Chk2–dependent checkpoints.<sup>60</sup> Moreover, Chk2 mutations are more frequent in patients with prostate cancer than in the general population, according to an earlier report.<sup>61</sup> Finally, Beltran *et al.* demonstrated that genomic alterations in HR DNA repair genes including *ATM* (8%) and *BRCA2* (12%) were detected in CRPC.<sup>62</sup> It should be mentioned that further studies are needed to establish the incidence of these mutations and explain their significance.

In general, the results of mechanism-based studies of DDR support the concept that aggressive cancers accumulating multiple defects in DDR due to deletions or mutations of DDR mediators and related defective cell cycle checkpoints may be particularly sensitive to DDR inhibition. In particular, loss of the ATM-Chk2-p53 component of the DDR creates a cancer with a DNA damage repair defect that can be exploited therapeutically with agents that lead to accumulation of DNA damage. For example, because defects in ATM-Chk2-p53 function render cancer cells more dependent on Chk1 to activate cell cycle checkpoints in the presence of DNA damage, inhibition of ATR and subsequent signaling mediated by Chk1 is a rational therapeutic strategy to push cancer cells to “mitotic catastrophe”. In support of this hypothesis, ATR and Chk1 inhibitors are effective in these cancer cells. Selective Chk1 inhibitors have been found to promote aberrant mitosis and increase cell death in cells harboring *P53* and *ATM-Chk2* mutations.<sup>63, 64</sup> Inhibition of Chk1 in p53-mutant prostate cancer is an example of synthetic lethality. What is more, Chk1 knockdown was found to increase the apoptotic effects of radiation in prostate cancer cells associated with decreased activation Cdc25C and Cdc2. Chk1 also knockdown confers radiosensitization in prostate cancer stem cells.<sup>65</sup> Similarly, AZD7762, a Chk1 and Chk2 inhibitor, abrogates G<sub>2</sub>/M arrest and leads to mitotic catastrophe associated with increased apoptosis, which enhances the cytotoxic effects of bendamustine, melphalan and doxorubicin in p53-deficient multiple myeloma cells.<sup>66</sup>

Another example of synthetic lethality based on DDR defects is the effect of PARP inhibitors in BRCA1-mutated cancer cells. Apart from the above described disrupted ATM-Chk2-p53 signaling another example of synthetic lethality in cancer cells related to DDR is defective HR. Particularly, it is known that defective HR secondary to *BRCA1* and *BRCA2* mutations may render cancer cells particularly sensitive to inhibition of SSB repair through PARP inhibitors.<sup>67</sup> Indeed, PARP inhibitors such as olaparib and niraparib present significant antitumor efficacy in ovarian, breast and based on more recent data in prostate cancer dysfunctional HR (e.g., *BRCA1* and *BRCA2* mutations).<sup>63,68</sup> Two recent studies have

demonstrated that male *BRCA1* mutation carriers younger than 65 years are more susceptible to prostate cancer compared to those who do not carry the mutation.<sup>69,70</sup> According to a recent report by Fong *et al.*, olaparib showed antitumor activity only in patients carrying *BRCA1* or *BRCA2* mutations.<sup>68</sup> Finally, Sandhu *et al.* demonstrated that three of four patients with a *BRCA2* mutation and an *ERG* rearrangement showed significant response to olaparib (10–34 months on treatment), while the fourth patient exhibited primary resistance to PARP inhibitor MK-4827 (ref.71). Together, these results suggest that PARP inhibitors are mainly effective in tumors with HR deficiency, but more studies are needed to identify novel predictive biomarkers (in addition to *BRCA1/2*) to stratify patients that will be good candidates for this therapy.

The known defects of DDR related to prostate cancer initiation and progression and the related opportunities for development of strategies to induce synthetic lethality are summarized in Figure 1.

### Oncogenic signaling and DDR in prostate cancer

During development of prostate cancer, dysfunction of tumor suppressors such as *PTEN*<sup>52</sup> and activation of oncogenic signaling<sup>72</sup> contribute to progression of the disease and resistance to hormonal therapy. Recent reports based on preclinical models demonstrated that multiple oncogenic events contribute to disease progression, tumor growth and metastatic potential.<sup>73–75</sup> Interestingly, it is believed that prostate cancer progression is strongly associated with genomic instability and more particularly telomere dysfunction.<sup>76–77</sup> Of note, during prostate cancer progression oncogenic signaling related to replication stress such as Akt and c-Myc are frequently induced.<sup>78</sup> These observations and results are consistent with the general concept that activation of oncogenic signaling promotes DNA replication stress, leading to increased incidence of DSBs and subsequent genomic instability.<sup>79</sup>

Activation of Akt signaling, which is very common during prostate cancer progression<sup>52</sup>, is known to modify the cellular response to DSBs. In particular, Akt activation has been related to phosphorylation of Chk1, Topbp1 and Brca1<sup>80–82</sup>, while the same signaling modulates focal accumulation of critical mediators of DSB repair such as RAD51, *BRCA1* and RPA.<sup>83–85</sup> Akt is known to physically associate with DNA-PKcs, while its inhibitors have been shown to reduce radiation-induced DNA-PKcs activation.<sup>86, 87</sup> These results suggest that Akt activation enhances the activity of DNA-PKcs and subsequent NHEJ. On the other hand, radiation-induced *BRCA1* and RAD51 foci formation and HR activation were strongly impaired in breast cancer cells with high Akt activity compared to cells with low Akt activity.<sup>88</sup> Moreover, Akt activation was found to be correlated with cytoplasmic retention of *BRCA1* and RAD51 foci instead of nuclear accumulation in sporadic breast cancer biopsies.<sup>84</sup> These data suggest that Akt activation, which has been shown to contribute to the progression of prostate cancer, activates NHEJ. However, in astrocytoma Akt was shown to suppress HR.<sup>89</sup> As mentioned above, NHEJ can be mutagenic; which is consistent with the association of *PTEN* deletion with gene rearrangements such as the *ERG* fusion gene during prostate cancer development<sup>90</sup> and synthetic lethality from targeting *PTEN*-deleted cells with PARP inhibitors.<sup>91</sup>



c-Myc has been implicated in the development and progression of prostate cancer<sup>52</sup>, and its activation increases the aggressiveness of prostate cancer with *PTEN* deletions in transgenic mice.<sup>92</sup> Interestingly, recent reports have demonstrated a connection between c-Myc and ATM signaling. In particular, Liyanage *et al.* showed that thymic lymphomas developed in ATM-knockdown mice are characterized by increased copies of chromosome 15, where the *c-MYC* gene maps.<sup>93</sup> Moreover, c-Myc activation has been related to ATM inactivation in numerous malignancies, including B-cell lymphomas.<sup>94</sup> It is believed that the DDR and subsequent apoptosis resulting from c-Myc upregulation and c-Myc-induced replication stress are both reduced upon ATM loss, while tumorigenesis is significantly accelerated.<sup>31</sup> One hypothesis is that, upon c-Myc upregulation, ATM mediates the DDR to repair DNA damage and releases c-Myc-induced replication stress. The results reported by Pusapati *et al.*, which were based on a mouse model of skin cancer, further supported the critical role of ATM in activating DDR upon c-Myc upregulation, which inhibits tumor growth.<sup>95</sup>

Mechanistically, c-Myc upregulation induces cell proliferation and DNA damage accumulation, leading to induced ATM activity and increased phosphorylation of numerous subsequent targets such as Chk2 and p53, which prolongs the G<sub>2</sub> phase of the cell cycle, contributing to DDR.<sup>31, 96</sup> Interestingly, induction of p53 in response to c-Myc overexpression requires ATM activity.<sup>95</sup> These results suggest that c-Myc activation induces replication stress and probably DNA damage as a result of ROS accumulation, leading to DDR mediated by ATM/ Chk2/ p53 signaling. Overall, mutations or deletions of this pathway will promote carcinogenesis induced by c-Myc upregulation. However, during disease progression the same defects may render these cells susceptible to inhibition of ATM-independent nodes such as Chk1 and ATR. Abundant c-Myc sensitizes a variety of cells to drug inhibition of either Chk1 or ATR.<sup>97, 98</sup> Similar results were observed for Ras-overexpressing tumors<sup>99</sup>. These results generally support the suggestion that aggressive cancers accumulating deletions of tumor suppressors, DDR defects and oncogenic activation are particularly sensitive to ATR/Chk1 inhibition.

The *TMPRSS2-ERG* chromosome fusion has been observed in 50–60% of prostate tumors<sup>100, 101</sup> and has been associated with increased proliferation and migration<sup>102</sup> and more aggressive forms of the disease.<sup>103</sup> This gene fusion results in androgen-regulated overexpression of oncogenes contributing to development of prostate cancer.<sup>104</sup> Brenner and Ateeq found that one of the top proteins interacting with *ERG* is DNA-PKcs<sup>105</sup>, a molecule that plays a critical role in NHEJ as discussed above. In particular, the authors showed that PARP1 and DNA-PKcs are both critical for activation of *ERG*-mediated transcription of a number of target genes, some of which are increased in metastatic disease.<sup>105</sup> In the same study, it was shown that prostate cancer cells are susceptible to olaparib when they express the *ERG* gene through potentiation of DSBs.<sup>105</sup> On the basis of these results, it was suggested that the *TMPRSS2-ERG* gene fusion could predict for sensitivity to PARP inhibition. Interestingly, Chatterjee *et al.* showed that PARP inhibition increased sensitivity to radiation therapy, especially in prostate cancer cells expressing the *TMPRSS2-ERG* fusion gene and deficient in *PTEN*, supporting this hypothesis.<sup>106</sup> However, a recent phase I clinical trial conducted in *BRCA1* mutation carriers and patients with sporadic prostate cancer showed no correlation between the activity of niraparib, another PARP inhibitor, and

*ETS* rearrangements.<sup>107</sup> Further studies are needed to clarify this question but this concept also highlights the importance of developing novel biomarkers of HR dysfunction to predict for sensitivity to PARP inhibitors such as olaparib and niraparib.

### The role of AR in DDR regulation

As described above, AR has been implicated in activation of DDR.<sup>43, 47</sup> The finding of Bowen *et al.* that AR target gene *NKX3.1* activates ATM suggests that AR may stimulate ATM-mediated DDR.<sup>48</sup> Recent reports support the role of AR in DDR regulation during prostate cancer progression, highlighting multiple therapeutic opportunities provided by combining hormonal manipulation with DNA-damaging approaches such as radiation therapy.

Schiewer *et al.* demonstrated that PARP1 mediates AR binding to chromatin, promoting transcription of AR target genes such as *PSA*, *TMPRSS2* and *ERG*.<sup>108</sup> The authors showed that PARP1 activity is essential to maintain AR function in genetically modified mouse embryonic fibroblasts and that PARP1 activity is upregulated in CRPC, enhancing the AR activity under ADT.<sup>108</sup> Moreover, according to that report, PARP1 inhibition sensitizes only AR-positive prostate cancer cells to genotoxic agents such as irradiation and docetaxel, while veliparib (ABT888), a PARP1 inhibitor, enhances the effects of castration *in vivo* in VCaP and LNCaP C4-2 xenografts and decreases prostate cancer cell proliferation in explants from human primary prostate tumors.<sup>108</sup> Given that PARP1 has a spectrum of activities, these data suggest that PARP1 promotes prostate cancer growth and survival, in part, by enhancing AR activity and regulating DDR, suggesting that PARP1 inhibition may improve the efficacy of hormonal therapy in patients with prostate cancer.

As previously described, the DNA-PK complex, consisting of the Ku70-Ku80 heterodimer and DNA-PKcs, is a key component of NHEJ. Ku proteins detect DSB ends and join them together, promoting DSB repair. Al-Ubaidi *et al.* in a recent report identified an interaction between AR and Ku-70 in prostate cancer tissues, while castration was found to be associated with reduction in Ku-70 protein levels.<sup>109</sup> Interestingly, low Ku-70 levels were related to low prostate-specific antigen levels and increased numbers of  $\gamma$ H2AX foci in prostate cancer samples.<sup>109</sup> These results support the role of AR regulation of DDR, particularly NHEJ activities; and that ADT can inhibit the repair of DSBs, leading to increased DNA damage in the absence of a genotoxic agent.

Goodwin *et al.* evaluated the hypothesis that AR regulates DSB repair and that AR inhibition may sensitize prostate cancer cells to DNA-damaging therapies such as irradiation. They showed that ADT and radiation synergize to decrease growth of androgen-sensitive and -insensitive AR-positive prostate cancer cells *in vitro*, while this combined therapeutic approach increases the incidence of DSBs as indicated by increased numbers of  $\gamma$ H2AX and *53BP1* foci in LNCaP C4-2 cells.<sup>19</sup> Interestingly, data presented in this report suggest that irradiation induced AR transcriptional activity, an effect that was reduced by addition of *N*-acetylcysteine, an ROS scavenger, and by administration of ADT.<sup>19</sup> The authors discovered that three genes were regulated by both androgens and radiation and upregulated in CRPC, namely, *XRCC2* and *XRCC3*, which encode the XRCC2 and XRCC3 proteins which promote strand transfer at the sites of DNA damage during HR<sup>110</sup>, and



*PRKDC*, which encodes the DNA-PKcs involved in the recruitment of repair factors in the sites of DNA damage during NHEJ.<sup>111</sup> Chromatin immunoprecipitation analysis showed that androgens promote expression of these genes under radiation. Moreover, the presence of androgens and radiation enhanced the activity of DNA-PK–dependent repair of DNA damage caused by radiation through NHEJ.<sup>19</sup> These results further support that a role for AR in the regulation of multiple genetic activities and pathways that influence DDR and DNA repair under genotoxic stress.

These results were further confirmed by Polkinghorn *et al.*, who showed that androgens induce expression of DDR genes, promoting repair of DNA damage caused by ionizing radiation. In contrast, novel anti-androgens such as ARN 509 inhibit the repair of DSBs.<sup>112</sup> Moreover, the combination of AR inhibition and radiation decreases the clonogenic survival of prostate cancer cells.<sup>112</sup> The authors also presented evidence that AR promotes expression of genes implicated in sensing DNA damage and other components of DDR such as BER and MMR.<sup>112</sup> A recent report by Li *et al.* showed that MYB is transcriptionally activated by androgen deprivation or genetic silencing of AR, and that AR and c-Myb share a subset of target genes that encode DDR proteins.<sup>113</sup> These results indicate that c-Myb may supplant AR as the dominant regulator of their common DDR target genes in prostate cancer cells that are resistant to AR inhibition.<sup>113</sup> Collectively, these data highlight the role of AR as a regulator of DDR and support the hypothesis that combining AR inhibition with genotoxic agents represents a rational approach for advanced prostate cancer.

## THERAPEUTIC IMPLICATIONS

While the concept of synthetic lethality is inherently plausible in rare familial prostate cancers carrying germline alterations in DDR genes (e.g., *BRCA1/2*), mutations of DDR genes are rare in sporadic cancers. However, AR-mediated regulation of DDR genes provides a rationale of synergistic combinations using novel androgen synthesis inhibitors and anti-androgens such as abiraterone and enzalutamide, respectively to inhibit or impair DDR. More specifically, inhibition of AR leads to downregulation of DDR function to create a DNA damage repair defect. If AR-inhibited tumors are then treated with an agent(s) that promotes DNA damage, synthetic lethality or synergistic cytotoxicity may result. In support of this hypothesis, combination of ADT with a PARP inhibitor (such as olaparib or niraparib) is one approach that has demonstrated encouraging results in preclinical models.<sup>108</sup> A clinical trial is currently underway (ClinicalTrials.gov ID NCT01576172) in which patients with mCRPC receive abiraterone with or without veliparib (ABT-888). Patients are stratified by their *ETS* gene fusion status (positive or negative) and then randomized to receive abiraterone alone or abiraterone + veliparib. We believe that this trial will inform about the potential for synergistic effects with combined ADT+PARP inhibition, although this is not a primary objective.

Since AR regulates ATM/Chk2 signaling, a second approach would be to combine ADT with ATR and/or Chk1 inhibitors, especially those bearing *p53* mutations and characterized by increased replication stress and genomic instability. A third approach would be to combine DDR inhibition with a cytotoxic DNA-damaging agent. Platinum-based chemotherapy regimens are active in aggressive prostate cancers with neuroendocrine and

anaplastic features.<sup>114</sup> Thus, combining an inhibitor of DDR with a platinum agent may represent a novel and promising approach to treating this lethal disease (Figure 2).

## CONCLUSION

DNA damage is considered one of the most frequent events contributing to development of both hematologic and epithelial neoplasms, including prostate cancer. Mutations and deletions of critical DDR “signaling nodes” regulated by ATM and Chk2 have been shown to increase the risk of prostate cancer development, while p53, an important mediator of DDR, is inactivated in most sporadic prostate cancers. Recent studies provide evidence supporting the role for AR in DDR activation and repair of DNA damage in prostate cancer cell survival through regulation of relevant genetic activities and pathways. Although there is controversy in the literature, the activation of oncogenic signaling such as Akt and c-Myc has been in general shown to increase DNA damage through replication stress enhancing genomic instability and disease aggressiveness. In cancer cells bearing defective DDR genes, the remaining active component of DDR becomes critical for tumorigenesis, since further inhibition of DDR may lead to genomic instability incompatible with cancer cell survival. Given a potential for AR to regulate genetic activities and pathways that influence DDR gene, particularly those related to ATM/Chk2 signaling, combination therapy with ADT plus a PARP inhibitor or ADT plus ATR and/or Chk1 inhibitors represents a rational therapeutic strategy. Combination of a DDR-targeted agent with a cytotoxic DNA-damaging agent (e.g. platinum agents) is another approach to be considered. Clinical trials based on these concepts are critical to determine which agents, conditions, and combinations of agents will benefit patients’ quality and longevity of life. It should also be noted that introducing DDR-targeted agents into clinical management of such a heterogeneous disease as mCRCP is not without potential consequences as evidenced by the emergence of de novo neoplasias following exposure to DNA damaging agents. However, characterizing specific DDR defects in each patient should provide a unique opportunity for personalized medicine utilizing rational therapy combinations that promote synthetic lethality or synergistic cytotoxicity.

## ACKNOWLEDGEMENTS

The authors thank Kathryn L Hale, MS, MLIS for her expert editorial assistance.

**Financial support:** This research is supported in part by the National Institutes of Health through MD Anderson’s Cancer Center Support Grant, 5 P30 CA016672

## REFERENCES

1. Ryan CJ, Smith MR, de Bono JS, Molina A, Logothetis CJ, de Souza P, et al. Abiraterone in metastatic prostate cancer without previous chemotherapy. *N Engl J Med*. 2013; 368:138–148. [PubMed: 23228172]
2. Fizazi K, Scher HI, Molina A, Logothetis CJ, Chi KN, Jones RJ, et al. Abiraterone acetate for treatment of metastatic castration-resistant prostate cancer: final overall survival analysis of the COU-AA-301 randomised, double-blind, placebo-controlled phase 3 study. *Lancet Oncol*. 2012 Oct; 13(10):983–992. [PubMed: 22995653]
3. Beer, TM.; Armstrong, AJ.; Sternberg, CN.; Higano, CS.; Iversen, P.; Lortol, Y., et al. Enzalutamide in men with chemotherapy-naïve metastatic prostate cancer (mCRPC): results of phase 3 PREVAIL

study. Presented at 2014 Genitourinary Cancers Symposium; January 30–February 1, 2014; San Francisco, California. Abstract LBA1.

4. Scher HI, Fizazi K, Saad F, Taplin ME, Sterberg CN, Miller K, et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. *N Engl J Med*. 2012; 367:1187–1197. [PubMed: 22894553]
5. Bolla M, Gonzalez D, Warde P, Dubois JB, Mirimanoff RO, Storme G, et al. Improved survival in patients with locally advanced prostate cancer treated with radiotherapy and goserelin. *N Engl J Med*. 1997 Jul 31; 337(5):295–300. [PubMed: 9233866]
6. Polkinghorn WR, Parker JS, Lee MX, Kass EM, Spratt DE, Iaquina PJ, et al. Androgen receptor signaling regulates DNA repair in prostate cancers. *Cancer Discov*. 2013 Nov; 3(11):1245–1253. [PubMed: 24027196]
7. Goodwin JF, Schiewer MJ, Dean JL, Schrecengost RS, de Leeuw R, Han S, et al. A hormone-DNA repair circuit governs the response to genotoxic insult. *Cancer Discov*. 2013 Nov; 3(11):1254–1271. [PubMed: 24027197]
8. Lord CJ, Ashworth A. The DNA damage response and cancer therapy. *Nature*. 2012; 481:287–294. Review. [PubMed: 22258607]
9. Ciccia A, Elledge SJ. The DNA damage response: making it safe to play with knives. *Mol Cell*. 2010; 40:179–204. [PubMed: 20965415]
10. D'Adda di Fagagna F. Living on a break: cellular senescence as a DNA-damage response. *Nat Rev Cancer*. 2008; 8:512–522. [PubMed: 18574463]
11. Taylor RC, Cullen SP, Martin SJ. Apoptosis: controlled demolition at the cellular level. *Nature Rev Mol Cell Biol*. 2008; 9:231–241. [PubMed: 18073771]
12. Goto H, Izawa I, Li P, Inagaki M. Novel regulation of checkpoint kinase 1: Is checkpoint kinase 1 a good candidate for anti-cancer therapy? *Cancer Sci*. 2012 Jul; 103(7):1195–1200. [PubMed: 22435685]
13. Martín Y, Domínguez-Kelly R, Freire R. Novel insights into maintaining genomic integrity: Wee1 regulating Mus81/Eme1. *Cell Div*. 2011 Dec 9; 6:21. [PubMed: 22152133]
14. Do K, Doroshow JH, Kummer S. Wee1 kinase as a target for cancer therapy. *Cell Cycle*. 2013 Oct 1; 12(19):3159–3164. [PubMed: 24013427]
15. Ma CX, Janetka JW, Piwnicka-Worms H. Death by releasing the breaks: CHK1 inhibitors as cancer therapeutics. *Trends Mol Med*. 2011 Feb; 17(2):88–96. [PubMed: 21087899]
16. Moynahan ME, Jasin M. Mitotic homologous recombination maintains genomic stability and suppresses tumorigenesis. *Nature Rev Mol Cell Biol*. 2010; 11:196–207. [PubMed: 20177395]
17. Lieber MR. NHEJ and its backup pathways in chromosomal translocations. *Nature Struct Mol Biol*. 2010; 17:393–395. [PubMed: 20368722]
18. Dvir A, Peterson SR, Knuth MW, Lu H, Dynan WS. Ku autoantigen is the regulatory component of a template-associated protein kinase that phosphorylates RNA polymerase II. *Proc Natl Acad Sci U S A*. 1992 Dec 15; 89(24):11920–11924.
19. Goodwin JF, Schiewer MJ, Dean JL, Schrecengost RS, de Leeuw R, Han S, Ma T, Den RB, Dicker AP, Feng FY, Knudsen KE. A hormone-DNA repair circuit governs the response to genotoxic insult. *Cancer Discov*. 2013 Nov; 3(11):1254–1271. [PubMed: 24027197]
20. Heaphy CM, Yoon GS, Peskoe SB, Joshu CE, Lee TK, Giovannucci E, Mucci LA, Kenfield SA, Stampfer MJ, Hicks JL, De Marzo AM, Platz EA, Meeker AK. Prostate cancer cell telomere length variability and stromal cell telomere length as prognostic markers for metastasis and death. *Cancer Discov*. 2013 Oct; 3(10):1130–1141. [PubMed: 23779129]
21. Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. *Nature*. 2011; 474:609–615. [PubMed: 21720365]
22. Leongamornlert D, Mahmud N, Tymrakiewicz M, et al. Germline BRCA1 mutations increase prostate cancer risk. *Br J Cancer*. 2012 May 8; 106(10):1697–1701. [PubMed: 22516946]
23. van Asperen CJ, Brohet RM, Meijers-Heijboer EJ, et al. Cancer risks in BRCA2 families: estimates for sites other than breast and ovary. *J Med Genet*. 2005 Sep; 42(9):711–719. [PubMed: 16141007]

24. Ding L, Getz G, Wheeler DA, Mardis ER, McLellan MD, Cibulskis K, et al. Somatic mutations affect key pathways in lung adenocarcinoma. *Nature*. 2008 Oct 23; 455(7216):1069–1067. [PubMed: 18948947]
25. Greenman C, Stephens P, Smith R, Dalgliesh GL, Hunter C, Bignell G, et al. Patterns of somatic mutation in human cancer genomes. *Nature*. 2007 Mar 8; 446(7132):153–158. [PubMed: 17344846]
26. Beltran H, Yelensky R, Frampton GM, Park K, Downing SR, et al. Targeted next-generation sequencing of advanced prostate cancer identifies potential therapeutic targets and disease heterogeneity. *Eur Urol*. 2013 May; 63(5):920–926. [PubMed: 22981675]
27. Bartkova J, Rezaei N, Liontos M, Karakaidos P, Kletsas D, Issaeva N, et al. Oncogene-induced senescence is part of the tumorigenesis barrier imposed by DNA damage checkpoints. *Nature*. 2006; 444:633–637. [PubMed: 17136093]
28. Halazonetis TD, Gorgoulis VG, Bartek J. An oncogene-induced DNA damage model for cancer development. *Science*. 2008; 319:1352–1355. [PubMed: 18323444]
29. Farmer H, McCabe N, Lord CJ, Tutt AN, Johnson DA, Richardson TB, et al. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature*. 2005; 434:917–921. [PubMed: 15829967]
30. Bryant HE, Schultz N, Thomas HD, Parker KM, Flower D, Lopez E, et al. Specific killing of BRCA2-deficient tumors with inhibitors of poly(ADP-ribose) polymerase. *Nature*. 2005; 434:913–917. [PubMed: 15829966]
31. Campaner S, Amati B. Two sides of the Myc-induced DNA damage response: from tumor suppression to tumor maintenance. *Cell Div*. 2012 Feb 28; 7(1):6. [PubMed: 22373487]
32. Ferrao PT, Bukczynska EP, Johnstone RW, McArthur GA. Efficacy of CHK inhibitors as single agents in MYC-driven lymphoma cells. *Oncogene*. 2012 Mar 29; 31(13):1661–1672. [PubMed: 21841818]
33. Abulaiti A, Fikaris AJ, Tsygankova OM, Meinkoth JL. Ras induces chromosome instability and abrogation of the DNA damage response. *Cancer Res*. 2006 Nov 1; 66(21):10505–10512. [PubMed: 17079472]
34. Curtin NJ. DNA repair dysregulation from cancer driver to therapeutic target. *Nat Rev Cancer*. 2012 Dec; 12(12):801–817. [PubMed: 23175119]
35. Lukas C, Melander F, Stucki M, Falck J, Bekker-Jensen S, Goldberg M, et al. Mdc1 couples DNA double-strand break recognition by Nbs1 with its H2AX-dependent chromatin retention. *EMBO J*. 2004; 23:2674–2683. [PubMed: 15201865]
36. Elkahwaji JE. The role of inflammatory mediators in the development of prostatic hyperplasia and prostate cancer. *Res Rep Urol*. 2012 Dec 31; 5:1–10. [PubMed: 24400229]
37. Balistreri CR, Candore G, Lio D, Carruba G. Prostate cancer: from the pathophysiologic implications of some genetic risk factors to translation in personalized cancer treatments. *Cancer Gene Ther*. 2014 Jan; 21(1):2–11. [PubMed: 24407349]
38. Malins DC, Johnson PM, Wheeler TM, Barker EA, Polissar NL, Vinson MA. Age-related radical-induced DNA damage is linked to prostate cancer. *Cancer Res*. 2001; 61(16):6025–6028. [PubMed: 11507046]
39. Malins DC, Johnson PM, Barker EA, Polissar NL, Wheeler TM, Anderson KM. Cancer-related changes in prostate DNA as men age and early identification of metastasis in primary prostate tumors. *Proc Natl Acad Sci USA*. 2003; 100(9):5401–5406.
40. De Marzo AM, Platz EA, Sutcliffe S, Xu J, Gronberg H, Drake CG, et al. Inflammation in prostate carcinogenesis. *Nat Rev Cancer*. 2007; 7 (4):256–269. [PubMed: 17384581]
41. Sciarra A, Di Silverio F, Salciccia S, Autran Gomez AM, Gentilucci A, et al. Inflammation and chronic prostatic diseases: evidence for a link? *Eur Urol*. 2007; 52 (4):964–972. [PubMed: 17618043]
42. Debelec-Butuner B, Alapinar C, Varisli L, Erbaykent-Tepedelen B, Hamid SM, Gonen-Korkmaz C, et al. Inflammation-mediated abrogation of androgen signaling: an in vitro model of prostate cell inflammation. *Mol Carcinog*. 2014 Feb; 53(2):85–97. [PubMed: 22911881]

43. Ide H, Lu Y, Yu J, China T, Kumamoto T, Koseki T, et al. Testosterone promotes DNA damage response under oxidative stress in prostate cancer cell lines. *Prostate*. 2012 Sep 15; 72(13):1407–1411. [PubMed: 22290195]
44. Imamoto T, Suzuki H, Yano M, Kawamura K, Kamiya N, Araki K, et al. The role of testosterone in the pathogenesis of prostate cancer. *Int J Urol*. 2008; 15(6):472–480. [PubMed: 18430151]
45. Schatzl G, Madersbacher S, Thurnidl T, Waldmüller J, Kramer G, Haitel A, et al. High-grade prostate cancer is associated with low serum testosterone levels. *Prostate*. 2001 Apr; 47(1):52–58. [PubMed: 11304729]
46. Albisinni S, De Nunzio C, Tubaro A, Barry WT, Banez LL, Freedland SJ. Greater percent-free testosterone is associated with high-grade prostate cancer in men undergoing prostate biopsy. *Urology*. 2012 Jul; 80(1):162–167. [PubMed: 22608797]
47. Zhou J, Richardson M, Reddy V, Menon M, Barrack ER, et al. Structural and functional association of androgen receptor with telomeres in prostate cancer cells. *Aging (Albany NY)*. 2013 Jan; 5(1):3–17. [PubMed: 23363843]
48. Bowen C, Ju JH, Lee JH, Paul TT, Gelmann EP. Functional activation of ATM by the prostate cancer suppressor NKX3.1. *Cell Rep*. 2013 Aug 15; 4(3):516–529. [PubMed: 23890999]
49. Dombernowsky SL, Weischer M, Allin KH, Bojesen SE, Tybjaerg-Hansen A, Nordestgaard BG. Risk of cancer by ATM missense mutations in the general population. *J Clin Oncol*. 2008 Jun 20; 26(18):3057–3062.
50. Angèle S, Falconer A, Edwards SM, Dörk T, Bremer M, Moullan N, et al. ATM polymorphisms as risk factors for prostate cancer development. *Br J Cancer*. 2004 Aug 16; 91(4):783–787. [PubMed: 15280931]
51. Khandrika L, Kumar B, Koul S, Maroni P, Koul HK. Oxidative stress in prostate cancer. *Cancer Lett*. 2009 Sep 18; 282(2):125–136. [PubMed: 19185987]
52. Taylor BS, Schultz N, Hieronymus H, Gopalan A, Xiao Y, Carver BS, et al. Integrative genomic profiling of human prostate cancer. *Cancer Cell*. 2010; 18:11–22. [PubMed: 20579941]
53. Muller, Patricia AJ.; Vousden, Karen H. p53 mutations in cancer. *Nature Cell Biology*. 2013; 15:2–8. [PubMed: 23263379]
54. Schlom T, Iwers L, Kirstein P, Jessen B, Köllermann J, Minner S, et al. Clinical significance of p53 alterations in surgically treated prostate cancers. *Mod Pathol*. 2008; 21:1371–1378. [PubMed: 18552821]
55. Eastham JA, Stapleton AM, Gousse AE, Timme TL. Association of p53 mutations with metastatic prostate cancer. *Clin Cancer Res*. 1995 Oct; 1(10):1111–1118. [PubMed: 9815901]
56. Kastan MB, Onyekwere O, Sidransky D, Vogelstein B, Craig RW. Participation of p53 protein in the cellular response to DNA damage. *Cancer Res*. 1991; 51:6304–6311. [PubMed: 1933891]
57. Maltzman W, Czyzyk L. UV irradiation stimulates levels of p53 cellular tumor antigen in nontransformed mouse cells. *Mol Cell Biol*. 1984; 4:1689–1694. [PubMed: 6092932]
58. Linke SP, Clarkin KC, Di Leonardo A, Tsou A, Wahl GM. A reversible, p53-dependent G0/G1 cell cycle arrest induced by ribonucleotide depletion in the absence of detectable DNA damage. *Genes Dev*. 1996; 10:934–947. [PubMed: 8608941]
59. Reinhardt HC, Schumacher B. The p53 network: cellular and systemic DNA damage responses in aging and cancer. *Trends Genet*. 2012 Mar; 28(3):128–136. [PubMed: 22265392]
60. Gabrielli B, Brooks K, Pavey S. Defective cell cycle checkpoints as targets for anti-cancer therapies. *Front Pharmacol*. 2012 Feb 2.3:9. [PubMed: 22347187]
61. Dong X, Wang L, Taniguchi K, Wang X, Cunningham JM, McDonnell K, et al. Mutations in CHEK2 associated with prostate cancer risk. *Am J Hum Genet*. 2003 Feb; 72(2):270–280. [PubMed: 12533788]
62. Beltran H, Yelensky R, Frampton GM, Park K, Downing SR, MacDonald TY, et al. Targeted next-generation sequencing of advanced prostate cancer identifies potential therapeutic targets and disease heterogeneity. *Eur Urol*. 2013 May; 63(5):920–926. [PubMed: 22981675]
63. Yap TA, Sandhu SK, Carden CP, de Bono JS. Poly(ADP-ribose) polymerase (PARP) inhibitors: Exploiting a synthetic lethal strategy in the clinic. *A Cancer J Clin*. 2011 Jan-Feb;61(1):31–49.

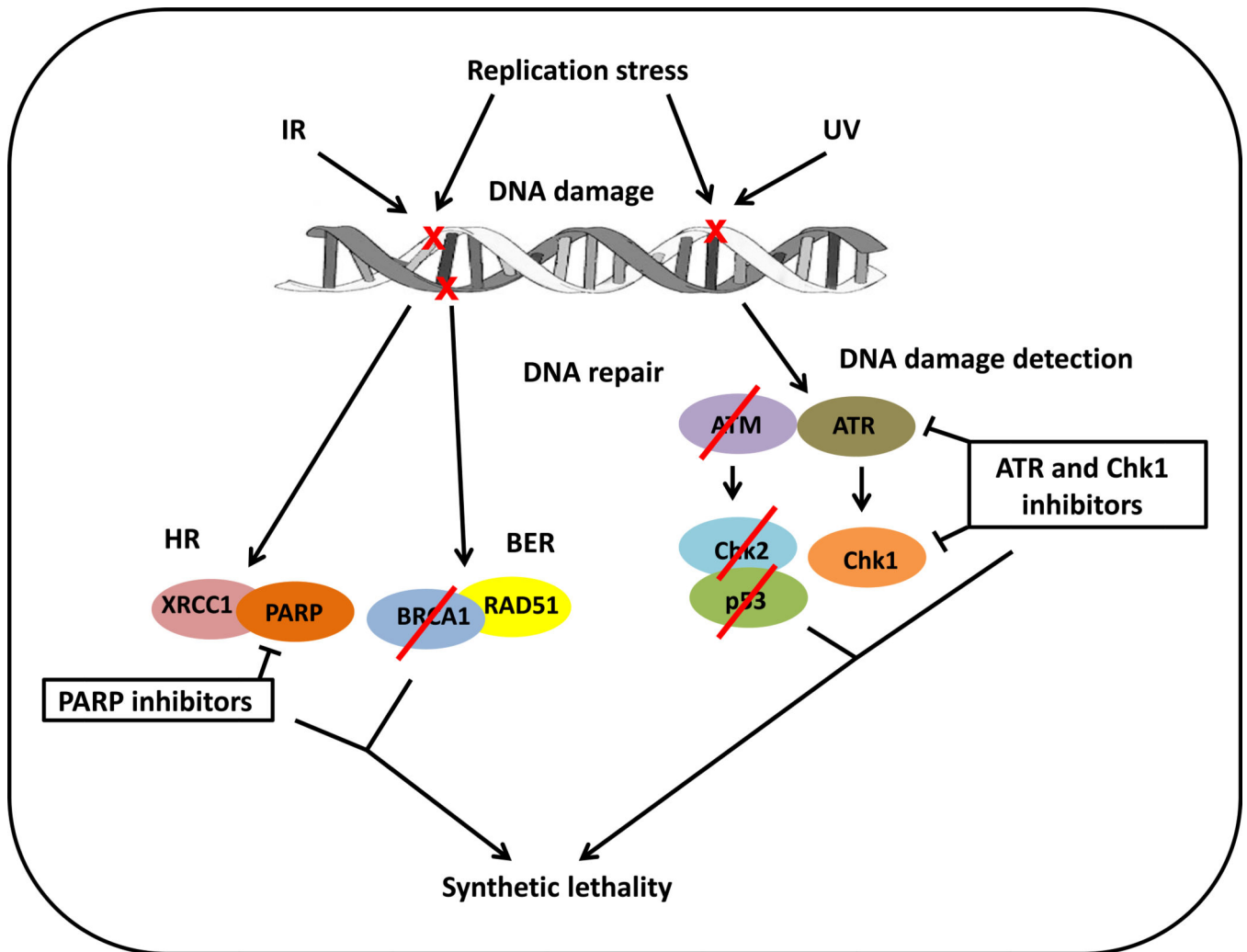


64. Edwards SM, Kote-Jarai Z, Meitz J, Hamoudi R, Hope Q, Osin P, et al. Two percent of men with early-onset prostate cancer harbor germline mutations in the BRCA2 gene. *Am J Hum Genet.* 2003; 72:1–12. [PubMed: 12474142]
65. Wang X, Ma Z, Xiao Z, Liu H, Dou Z, Feng X, Shi H. Chk1 knockdown confers radiosensitization in prostate cancer stem cells. *Oncol Rep.* 2012 Dec; 28(6):2247–2254. [PubMed: 23027394]
66. Landau HJ, McNeely SC, Nair JS, Comenzo RL, Asai T, Friedman H, et al. The checkpoint kinase inhibitor AZD7762 potentiates chemotherapy-induced apoptosis of p53-mutated multiple myeloma cells. *Mol Cancer Ther.* 2012 Aug; 11(8):1781–1788. [PubMed: 22653969]
67. Iglehart JD, Silver DP. Synthetic lethality--a new direction in cancer-drug development. *N Engl J Med.* 2009 Jul 9; 361(2):189–191. [PubMed: 19553640]
68. Fong PC, Boss DS, Yap TA, Tutt A, Wu P, Mergui-Roelvink M, et al. Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers. *N Engl J Med.* 2009 Jul 9; 361(2):123–134. [PubMed: 19553641]
69. Thompson D, Easton DF. Cancer incidence in BRCA1 mutation carriers. *J Natl Cancer Inst.* 2002; 94:1358–1365. [PubMed: 12237281]
70. Edwards SM, Kote-Jarai Z, Meitz J, Hamoudi R, Hope Q, Osin P, et al. Two percent of men with early-onset prostate cancer harbor germline mutations in the BRCA2 gene. *Am J Hum Genet.* 2003; 72:1–12. [PubMed: 12474142]
71. Sandhu SK, Omlin A, Hylands L, Miranda S, Barber LJ, Riisnaes R, et al. Poly (ADP-ribose) polymerase (PARP) inhibitors for the treatment of advanced germline BRCA2 mutant prostate cancer. *Ann Oncol.* 2013 May; 24(5):1416–1418. [PubMed: 23524863]
72. Thompson TC, Southgate J, Kitchener G, Land H. Multistage carcinogenesis induced by ras and myc oncogenes in a reconstituted organ. *Cell.* 1989; 56:917–930. [PubMed: 2538247]
73. Chen Z, Trotman LC, Shaffer D, Lin HK, Dotan ZA, Niki M, et al. Crucial role of p53-dependent cellular senescence in suppression of Pten-deficient tumorigenesis. *Nature.* 2005 Aug 4; 436(7051):725–730. [PubMed: 16079851]
74. Mulholland DJ, Kobayashi N, Ruscetti M, Zhi A, Tran LM, Huang J, et al. Pten loss and RAS/MAPK activation cooperate to promote EMT and metastasis initiated from prostate cancer stem/progenitor cells. *Cancer Res.* 2012 Apr 1; 72(7):1878–1889. [PubMed: 22350410]
75. Kim J, Roh M, Doubinskaia I, Algarroba GN, Eltoum IE, Abdulkadir SA. A mouse model of heterogeneous, c-MYC-initiated prostate cancer with loss of Pten and p53. *Oncogene.* 2012 Jan 19; 31(3):322–332. [PubMed: 21685943]
76. Ding Z, Wu CJ, Jaskelioff M, Ivanova E, Kost-Alimova M, Protopopov A, et al. Telomerase reactivation following telomere dysfunction yields murine prostate tumors with bone metastases. *Cell.* 2012 Mar 2; 148(5):896–907. [PubMed: 22341455]
77. Heaphy CM, Yoon GS, Peskoe SB, Joshu CE, Lee TK, Giovannucci E, et al. Prostate cancer cell telomere length variability and stromal cell telomere length as prognostic markers for metastasis and death. *Cancer Discov.* 2013 Oct; 3(10):1130–1141. [PubMed: 23779129]
78. Karantanos T, Corn PG, Thompson TC. Prostate cancer progression after androgen deprivation therapy: mechanisms of castrate resistance and novel therapeutic approaches. *Oncogene.* 2013 Dec 5; 32(49):5501–5511. [PubMed: 23752182]
79. Halazonetis TD, Gorgoulis VG, Bartek J. An oncogene-induced DNA damage model for cancer development. *Science.* 2008 Mar 7; 319(5868):1352–1355. [PubMed: 18323444]
80. Puc J, Keniry M, Li HS, Pandita TK, Choudhury AD, Memeo L, et al. Lack of PTEN sequesters CHK1 and initiates genetic instability. *Cancer Cell.* 2005; 7(2):193–204. [PubMed: 15710331]
81. Barré B, Perkins ND. A cell cycle regulatory network controlling NF- $\kappa$ B subunit activity and function. *EMBO Journal.* 2007; 26(23):4841–4855. [PubMed: 17962807]
82. Pedram A, Razandi M, Evinger AJ, Lee E, Levin ER. Estrogen inhibits ATR signaling to cell cycle checkpoints and DNA repair. *Molecular Biology of the Cell.* 2009; 20(14):3374–3389. [PubMed: 19477925]
83. Tonic I, Yu WN, Park Y, Chen CC, Hay N. Akt activation emulates Chk1 inhibition and Bcl2 overexpression and abrogates G2 cell cycle checkpoint by inhibiting BRCA1 foci. *Journal of Biological Chemistry.* 2010; 285(31):23790–23798. [PubMed: 20495005]



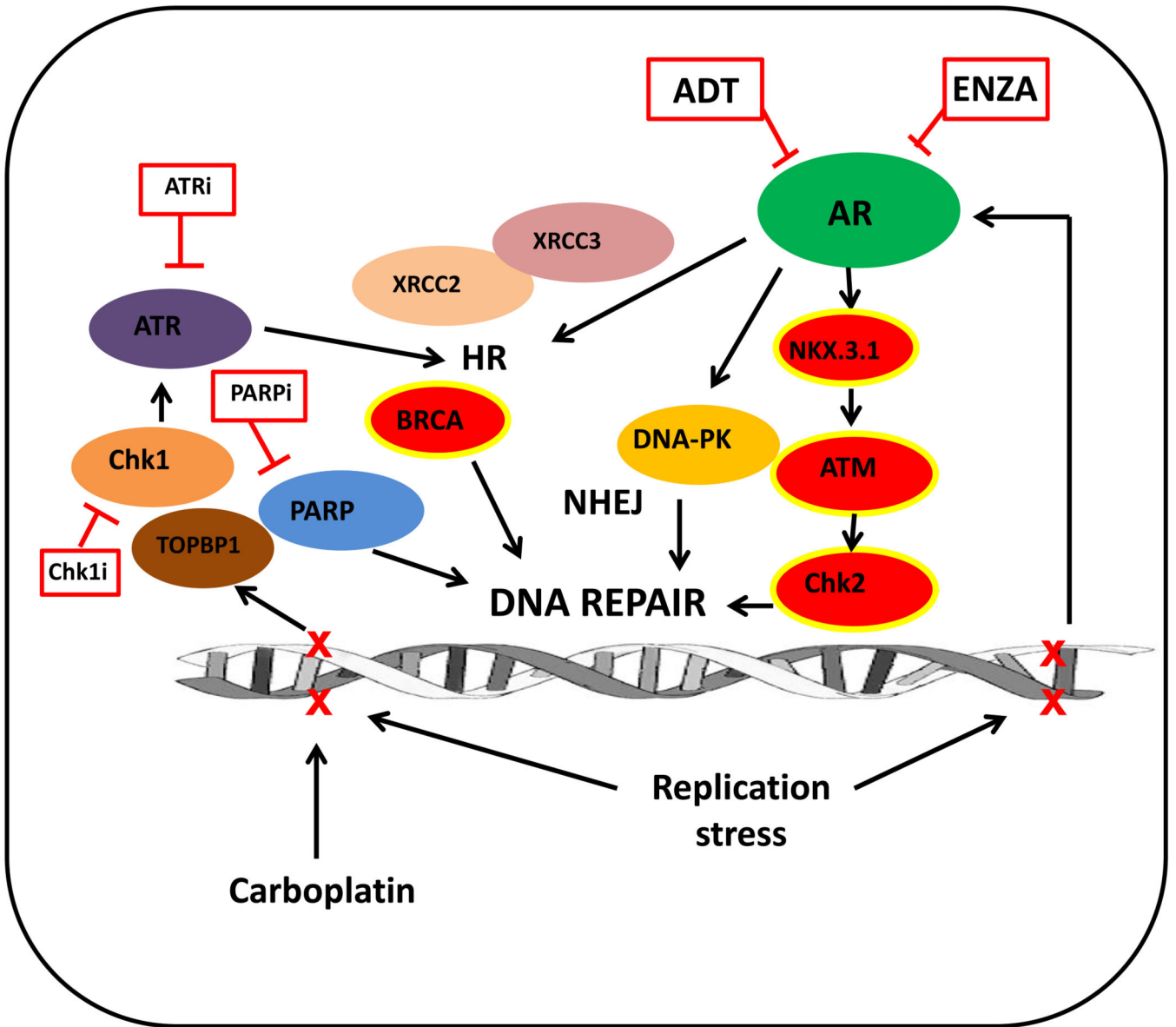
84. Plo I, Laulier C, Gauthier L, Lebrun F, Calvo F, Lopez BS. AKT1 inhibits homologous recombination by inducing cytoplasmic retention of BRCA1 and RAD5. *Cancer Research*. 2008; 68(22):9404–9412. [PubMed: 19010915]
85. Plo I, Lopez B. AKT1 represses gene conversion induced by different genotoxic stresses and induces supernumerary centrosomes and aneuploidy in hamster ovary cells. *Oncogene*. 2009; 28(22):2231–2237. [PubMed: 19398948]
86. Chen BP, Uematsu N, Kobayashi J, Lerenthal Y, Krempler A, Yajima H, Löbrich M, Shiloh Y, Chen DJ. Ataxia telangiectasia mutated (ATM) is essential for DNA-PKcs phosphorylations at the Thr-2609 cluster upon DNA double strand break. *Journal of Biological Chemistry*. 2007; 282(9): 6582–6587. [PubMed: 17189255]
87. Lavin MF, Kozlov S. DNA damage-induced signalling in ataxia-telangiectasia and related syndromes. *Radiotherapy and Oncology*. 2007; 83(3):231–237. [PubMed: 17512070]
88. Bishop AJ, Schiestl RH. Homologous recombination and its role in carcinogenesis. *Journal of Biomedicine and Biotechnology*. 2002; 2(2):75–85. [PubMed: 12488587]
89. McEllin B, Camacho CV, Mukherjee B, Hahm B, Tomimatsu N, Bachoo RM, Burma S. PTEN loss compromises homologous recombination repair in astrocytes: implications for glioblastoma therapy with temozolomide or poly(ADP-ribose) polymerase inhibitors. *Cancer Res*. 2010 Jul 1; 70(13):5457–5464. [PubMed: 20530668]
90. Han B, Mehra R, Lonigro RJ, Wang L, Suleman K. Fluorescence in situ hybridization study shows association of PTEN deletion with ERG rearrangement during prostate cancer progression. *Mod Pathol*. 2009 Aug; 22(8):1083–1093. [PubMed: 19407851]
91. Mendes-Pereira AM, Martin SA, Brough R, McCarthy A, Taylor JR, Kim JS, et al. Synthetic lethal targeting of PTEN mutant cells with PARP inhibitors. *EMBO Mol Med*. 2009 Sep; 1(6–7):315–322. [PubMed: 20049735]
92. Clegg NJ, Couto SS, Wongvipat J, Hieronymus H, Carver BS. MYC cooperates with AKT in prostate tumorigenesis and alters sensitivity to mTOR inhibitors. *PLoS One*. 2011 Mar 4.6(3):e17449. [PubMed: 21394210]
93. Liyanage M, Weaver Z, Barlow C, Coleman A, Pankratz DG, Anderson S, et al. Abnormal rearrangement within the alpha/delta T-cell receptor locus in lymphomas from *Atm*-deficient mice. *Blood*. 2000; 96:1940–1946. [PubMed: 10961898]
94. Korz C, Pscherer A, Benner A, Mertens D, Schaffner C, Leupolt E, et al. Evidence for distinct pathomechanisms in B-cell chronic lymphocytic leukemia and mantle cell lymphoma by quantitative expression analysis of cell cycle and apoptosis-associated genes. *Blood*. 2002; 99(12): 4554–4561. [PubMed: 12036888]
95. Pusapati RV, Rounbehler RJ, Hong S, Powers JT, Yan M, Kiguchi K, et al. ATM promotes apoptosis and suppresses tumorigenesis in response to *Myc*. *Proc Natl Acad Sci USA*. 2006; 103:1446–1451.
96. Guerra L, Albiñ A, Tronnersjö S, Yan Q, Guidi R, Stenerlöw B, Sterzenbach T, Josenhans C, Fox JG, Schauer DB, Thelestam M, Larsson LG, Henriksson M, Frisan T. *Myc* is required for activation of the ATM-dependent checkpoints in response to DNA damage. *PLoS One*. 2010; 5(1):e8924. [PubMed: 20111719]
97. Murga M, Campaner S, Lopez-Contreras AJ, Toledo LI, Soria R, Montana MF, et al. Exploiting oncogene induced replicative stress for the selective killing of *Myc*-driven tumors. *Nat Struct Mol Biol*. 2011; 18:1331–1335. [PubMed: 22120667]
98. Høglund A, Nilsson LM, Muralidharan SV, Hasvold LA, Merta P, Rudelius M, et al. Therapeutic implications for the induced levels of Chk1 in *Myc*-expressing cancer cells. *Clin Cancer Res*. 2011; 17:7067–7079. [PubMed: 21933891]
99. Gilad O, Nabet BY, Ragland RL, Schoppy DW, Smith KD, Durham AC, et al. Combining ATR suppression with oncogenic Ras synergistically increases genomic instability, causing synthetic lethality or tumorigenesis in a dosage-dependent manner. *Cancer Res*. 2010; 70:9693–9702. [PubMed: 21098704]
100. Clark J, Attard G, Jhavar S, Flohr P, Reid A, De-Bono J, et al. Complex patterns of ETS gene alteration arise during cancer development in the human prostate. *Oncogene*. 2008 Mar 27; 27(14):1993–2003. [PubMed: 17922029]

101. Mosquera JM, Perner S, Genega EM, Sanda M, Hofer MD, Mertz KD, et al. Characterization of TMPRSS2-ERG fusion high-grade prostatic intraepithelial neoplasia and potential clinical implications. *Clin Cancer Res.* 2008; 14:3380–3385. [PubMed: 18519767]
102. Carver BS, Tran J, Gopalan A, Chen Z, Shaikh S, Carracedo A, et al. Aberrant ERG expression cooperates with loss of PTEN to promote cancer progression in the prostate. *Nat Genet.* 2009; 41:619–624. [PubMed: 19396168]
103. Wang J, Cai Y, Ren C, Ittmann M. Expression of variant TMPRSS2/ERG fusion messenger RNAs is associated with aggressive prostate cancer. *Cancer Res.* 2006; 66:8347–8351. [PubMed: 16951141]
104. Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R, Sun XW, et al. Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. *Science.* 2005 Oct 28; 310(5748):644–648. [PubMed: 16254181]
105. Brenner JC, Ateeq B, Li Y, Yocum AK, Cao Q, Asangani IA, et al. Mechanistic rationale for inhibition of poly(ADP-ribose) polymerase in ETS gene fusion-positive prostate cancer. *Cancer Cell.* 2011 May 17; 19(5):664–678. [PubMed: 21575865]
106. Chatterjee P, Choudhary GS, Sharma A, Singh K, Heston WD, Ciezki J, et al. PARP inhibition sensitizes to low dose-rate radiation TMPRSS2-ERG fusion gene-expressing and PTEN-deficient prostate cancer cells. *PLoS One.* 2013; 8(4):e60408. [PubMed: 23565244]
107. Sandhu SK, Schelman WR, Wilding G, Moreno V, Baird RD, Miranda S, et al. The poly (ADP-ribose) polymerase inhibitor niraparib (MK4827) in BRCA mutation carriers and patients with sporadic cancer: a phase 1 dose-escalation trial. *Lancet Oncol.* 2013 Aug; 14(9):882–892. [PubMed: 23810788]
108. Schiewer MJ, Goodwin JF, Han S, Brenner JC, Augello MA, Dean JL, et al. Dual roles of PARP-1 promote cancer growth and progression. *Cancer Discov.* 2012 Dec; 2(12):1134–1149. [PubMed: 22993403]
109. Al-Ubaidi FL, Schultz N, Loseva O, Egevad L, Granfors T, Helleday T. Castration therapy results in decreased Ku70 levels in prostate cancer. *Clin Cancer Res.* 2013 Mar 15; 19(6):1547–1556. [PubMed: 23349316]
110. Thacker J. The RAD51 gene family, genetic instability and cancer. *Cancer Lett.* 2005; 219:125–135. [PubMed: 15723711]
111. Ciccio A, Elledge SJ. The DNA damage response: making it safe to play with knives. *Mol Cell.* 2010; 40:179–204. [PubMed: 20965415]
112. Polkinghorn WR, Parker JS, Lee MX, Kass EM, Spratt DE, Iaquinia PJ, et al. Androgen receptor signaling regulates DNA repair in prostate cancers. *Cancer Discov.* 2013 Nov; 3(11):1245–1253. [PubMed: 24027196]
113. Li, Likun; Chang, Wenjun; Yang, Guang; Ren, Chengzhen; Park, Sanghee; Karantanos, Theodoros; Karanika, Styliani, et al. Targeting Poly(ADP-Ribose) Polymerase and the c-Myb-Regulated DNA Damage Response Pathway in Castration-Resistant Prostate Cancer. *Sci Signal.* 2014 May 20.7(326) ra47.



**Figure 1. DDR defects implicated in initiation and progression of prostate cancer and opportunities for synthetic lethality**

Replication stress, irradiation (IR) and ultraviolet light (UV) promote DSBs and SSBs, respectively, leading to activation of multiple pathways regulating DNA repair, including ATM/Chk2/p53, ATR/Chk1 and PARP signaling. These events promote repair of the damage providing survival benefit to cancer cells under stimuli inducing genomic instability. During prostate cancer progression, genetic abnormalities such as polymorphisms and mutations or deletions of *p53*, *ATM*, *Chk2* and *BRCA1/2* (indicated by red bold slashes) have been reported, which make these pathways nonfunctional rendering other aspects of DDR critical for the cells' survival. Inhibition of alternative signaling (i.e., Chk1 and PARP) with DDR-targeted agents (Chk1, ATR and PARP inhibitors) may provide opportunities for synthetic lethality in tumors with defective DDR. Clinically, the identification of particular defects in the DDR system may create opportunities for personalized treatment in patients with aggressive, hormonal resistant prostate cancer. BER: base excision repair; HR: homologous recombination.



**Figure 2. Strategies for combination therapies in aggressive prostate cancers focusing on AR inhibition and androgen depletion**

Androgen receptor (AR) regulates multiple aspects of DDR during prostate cancer development. DNA damage induced by DNA-damaging agents such as carboplatin and replication stress activate DDR, many aspects of which, such as ATM/Chk2 and NHEJ, are mediated by AR. Mutations and deletions of BRCA and ATM/Chk2 signaling (molecules represented by red ovals with yellow outline) render Chk1/ATR and PARP critical for prostate cancer cell survival. Inhibition of AR by androgen-depletion therapy (ADT) or enzalutamide (ENZA) is expected to render prostate cancer cells particularly sensitive to inhibition of PARP (PARPi) and AR-independent DDR signaling such as Chk1/ATR (Chk1i and ATRi) with targeted agents. Finally, combination of DNA-damaging agents such as

carboplatin with inhibition of DDR may be a reasonable therapeutic approach for anaplastic (AR negative) disease. HR, homologous repair; NHEJ, nonhomologous end joining.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript