**REVIEW ARTICLE** 



# **Applications of Gold and Silver Nanoparticles in Theranostics**

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# Abstract

Nanotechnology sculptures the current scenario of science and technology. The word nano refers 'small' which ranges from 10 to 100 nm in size. Silver and gold nanoparticles can be synthesized at nanoscale and have unique biological properties like antibacterial, anti-fungal, antiviral, antiparasitic, antiplatelet, anti-inflammatory, and anti-tumor activity. In this mini review, we shall discuss the various applications of silver and gold nanoparticles (AuNPs) in the field of therapy, imaging, biomedical devices and in cancer diagnosis. The usage of silver nanoparticles(AgNPs) in dentistry and dental implants, therapeutic abilities like wound dressings, silver impregnated catheters, ventricular drainage catheters, combating orthopedic infections, and osteointegration will be elaborated. Gold nanoparticles in recent years have garnered large importance in bio medical applications. They are being used in diagnosis and have recently seen a surge in therapeutics. In this mini review, we shall see about the various applications of AuNP and AgNP, and highlight their evolution in theranostics.

**Keywords** Silver nanoparticle · Gold nanoparticle · Nanotechology · Biomedical application · Evolution in theranostics

# Introduction

Nanotechnology is known as multifaceted field of science which deals with the particles at nanosized ranges from 10 to 100 nm. There are various types of nanomaterials used in fields of science and technology such as medicine, construction, agriculture, energy production, and food industry [1–5]. The characterizations of nanoparticles (NPs) are based on their shape (rod, triangular, polyhedral, octagonal, round) and among the various NPs, metal NPs has gained more attention in recent years due to their distinctive property. Their use is more primitive in comparison to other nanostructures and gold particles were used as medicine in India and China for ayurvedhic medicine preparation [6, 7]. Among metal

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NPs, gold and silver nanoparticles (AgNPs) have various potential application and uses due to their application in allied disciplines and biomedical properties. AgNP is used in cleansing up the environmental pollution, medical imaging techniques and also exhibit more in the biomedical field because of large surface area to volume ratio [8-10]. AgNP is a powerful disinfectant and antibacterial agent and is used as a topical silver sulfadiazine cream as an antibiotic for treating burn wounds. It also shows antibacterial activity with garlic, cinnamon extract and is reported to have anticancer, anti-inflamatory activity etc., as shown in Fig. 1 [11–15]. The recent advancements and developments of gold nanoparticle (AuNP) in medicine and in theranostics have also gained attention. They are being used in diagnosis and have recently seen a surge in therapeutics. AuNP has a deep connection with chemistry, and in the Roman era(735BC) they were used in art decorations [16]. It has been identified that gold nanoparticles are used in various field of sciences such as diagnosis, cancer treatment, antibacterial and anti viral agent [17]. Noble metal are those which are defiance to oxidation at higher temperatures and corrosion. Some of the metals used are palladium, osmium, ruthenium, rhodium, iridium, platinum, copper, silver, gold, etc. These metal nanoparticles play a prominent role in the development of nanobiosensors in point of care testing, gene deivery, gene detection, intracellular trafficking targeting, cancer treatment, preventing tuberculosis, HIV, keratitis, imaging and therapeutics [18]. Metal nanoparticles can produce excessive reactive oxygen species, protein damage, inflammation which leads to toxicity, and it can also produce hyperthermia locally. The fundamendal principle is that it exhibits photocatalytic and photothermal effects by which under the light the metal nanoparticles gets excited at different wavelength [19]. In this review, we will discuss about the various applications of AgNP and AuNP in the field of theranostics and biomedical science.

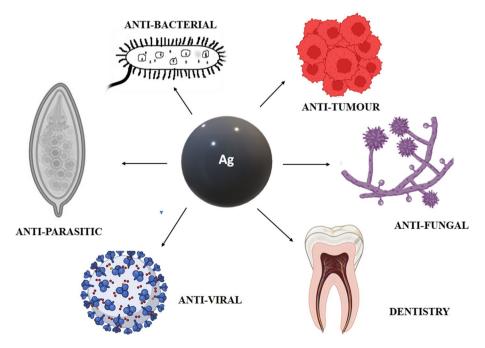


Fig. 1 Applications of silver nanoparticles

# **Biological Properties of Silver Nanoparticle**

## **Antibacterial Agent**

Multi-drug-resistant bacteria have become very prominent these days which has limited the resources for controlling infections using conventional therapies, such as, antibiotics, radiation etc. AgNPs is used as a superior antibacterial agent and has been found to be effective against a broad spectrum of vancomycin resistant strains in Gram-positive and Gram-negative bacteria [20–24]. Silver ions released by the AgNPs attaches to the thiol (SH) group of the sulfur and hydrogen present in the bacterial proteins and inhibits the bacterial growth [25–27]. AgNPs are potent against both anaerobic and aerobic bacteria and it functions by obstructing the bacterial electron transport chain system and precipitating the bacterial cellular proteins [28–30]. As AgNPs have larger surface area to volume ratio, it shows better efficiency [22, 31] and the mechanism of action are as follows,

- 1. The silver ions in AgNPs inhibits the function by reacting with phosphorous present in the DNA and sulfur containing proteins present in the bacterial membranes [32].
- 2. A nanometer scale silver provides firm attachment to the cell membrane and penetrates the bacterial cell wall [33, 34].
- 3. Ag<sup>+</sup> ions in AgNPs causes cell death by bombarding the electron transport chain in bacterial mitochondria [35].
- Continuous detachment of silver ions in the bacterial cell from AgNPs with an environment of lower pH produces free radicals, induces oxidative stress, and enhances the antibacterial activity [27, 36].
- AgNPs attaches to the bacteria and it penetrates inside the cell wall, further leading to death [31].

The shape, size, and concentration of the AgNPs determines the antibacterial efficacy. Studies state that enlarging the surface area of AgNPs can enhance the antibacterial activity [37]. Researchers have demonstrated that AgNPs possessed 50% inhibition activity against multidrug resistant bacteria (MDR)- Staphylococcus aureus and E. coli at a very low concentration of 20 µg/ml. At a further high concentration of about 40 µg /mL, it exhibited an efficient inhibition of both microorganisms. In another study, biosynthesized AgNP using marine macroalgae Padina species observed a good bacteriostatic activity against pathogenic Gram-positive organism like Bacillus subtilis, Staphylococcus aureus and Gram-negative bacteria like Pseudomonas aeruginosa, E. coli, Salmonella typhi. AgNPs with the concentration of 1mg/ml exhibited a higher sensitivity against *Pseudomonas aeruginosa* with diameter of zone of inhibition of  $13.33\pm0.76$  mm and *Staphylococcus aureus* of  $15.17\pm0.58$  mm respectively, where as the negative control possesed 0.00 mm. [38]. In another study, a comparative analysis was done with small AgNPs and PEGylated AgNPs of different molecular weight against a prime pathogen Staphylococcus aureus. In this study, 12 different sizes of AgNPs have been synthesized with a size range of 29.7 nm  $\pm$  0.02 to 35.5  $\pm$  0.02 nm in three different pH of 10,11 and 12. Excellent bactericidal activity was brought in by the PEGylated AgNPs with the zone of inhibition of about 29 mm at a pH 10. This was because PEG possessed a high hydrophilic property that eliminated more water and terminated the microbes [39]. Veerasamy et al have observed antimicrobial activity in the Green synthesized AgNP using Malia Azedarach at the concentrations of 25, 50, 75, 100 µg/ml in which highest zone of inhibition was observed at 100 µg/ml as 21 mm and 12 mm for *Pseudomonas aeruginosa* and *Bacillus subtilis* respectively [40].

# Antifungal Agent

Prolonged use of antifungal drugs leads to multidrug resistance especially for Candida species [41]. AgNPs coated reverse osmosis membrane exerted better antifungal activity against Candida albicans, Candida glabrata, Candida krusei, Candida tropicalis [24]. AgNP exerts antifungal property by interrupting the cell membrane and inhibits the normal asexual reproduction process by destroying the membrane integrity [42]. Mallmann et al., reported that AgNPs prepared by bio synthesis using SDS as stabilizer and reducing agent as ribose exhibited increased antifungal activity against C. albicans and C. tropicalis [43]. Additionally, AgNPs suppresses the growth of Aspergillus niger by 70% and Cladosporium *cladosporoides* by 90 %. Increasing the concentration of nanosilver minimizes the fungal growth in a dose dependent manner. AgNPs coated in cotton has shown antifungal activity against Aspergillus sp and the zone of inhibition was observed at  $14.33\pm3.51$  mm [44]. A study was done with biosynthesized AgNPs using *Malva parviflora* and leaf extract of Malva parviflora (LEMP) to monitor the antifungal activity against Alternaria alternata, Fusarium oxysporum Fusarium solani and Helminthosporium rostratum. With an average diameter of 50.6 nm, the AgNP mitigate the mycelia growth of *H. rostratum* at 88.6%, A.alternata at 83.0%, F.solani at 81.1% and F.oxyspoum at 80.7% where as the LEMP showed growth inhibition at 65.3% for F.solani, 54.7% F.oxyspoum, 53.6% H.rostratum and 45.6% for A.alternata [45]. Another study discussed about the pathogenic fungus which affects the quality of crops. To overcome this, mycosynthesized AgNPs using A. niger fungal isolate to invade against plant pathogenic fungi was utilized. At a concentration of about 10  $\mu$ g/ml, AgNPs showed inhibition of 91.0% for Fusarium oxysporum, 97.3% for Aspergillus flavvus, and 93.75% for Penicillin digitatum. This is because on treatment with Ag<sup>+,</sup> the DNA ruins its ability to replicate, thereby resulting in dormant expression of ribosomal subunit proteins and the enzymes needed for ATP production. [46]. A synergistic antifungal activity of Epoxiconazole and Lingustrum-lucidum leaf extract against Setosphaeria turcica showed the inhibitory percentage of 50 % against the organism at 170.20  $\mu$ g/ml concentration. At a ratio of 8:2 and 9:1 synergistic antifungal activity was seen in combination with epoxiconazole. This method gave a new perspective for the integrative control of plant pathogen [47].

# **Antiviral Agent**

Metal NPs like gold or silver exhibit antiviral activity against broad spectrum of virus by direct interaction with nanomaterial and viral surface proteins and reduce the infectivity of the viral cultured cells. Recent studies have shown that metal NPs are effective antiviral agents against HIV 1, Respiratory syncytial virus, HSV type1, monkey pox virus, Tacaribe virus and influenza virus [48–52]. AgNP coated with poly (N-vinyl-2-pyrrolidone) showed antiviral property against HIV1 by interacting with gp 120 [53]. AgNPs and polysaccharide coated AgNPs ranging from10-80 nm size was potent against monkey pox virus of Poxviridae family by blocking the virus – host cell binding and penetration [52]. AgNP at non-toxic concentration are used effectively to inhibit tacaribe virus (TCRV) of Arenaviridae family by inactivating the viral particles at their initial entry [54]. Saadh *et al.*, observed that AgNP integrated with epigallocatechingallate (EGCG) and doped with zinc

sulphate showed a potent antiviral activity against Avian influenza A virus H9N2. At a concentration of 50  $\mu$ M EGCG, it reduced the titre of AI H9N2 (logEID<sub>50</sub>/ml was 4.2). By increasing the concentration of EGCG >50  $\mu$ M, it did not show any enhanced antiviral effect. Effective result was obtained by combining EGCG with zinc sulphate showing logEID<sub>50</sub>/ml (1.5±0.6) [55]. Another study was on synthesizing AgNPs using *Lamprathus coccineus* and *Malephora lutea*. This study possessed that the hexane nanoextract of *L.coccineus* showed 50% inhibition against HAV-10 virus, HSV-1virus and CoxB4 virus with concentration 11.71 ng/ml, 36.36  $\mu$ g/ml and 12.74  $\mu$ g/ml respectively [56].

## Antiparasitic Agent

Saad and colleagues synthesized silver and copper NPs and studied the antiparasitic activity which showed remarkable decrease in the oocyte availability in Cryptosporidium par*vum.* They have also studied that AgNPs are effective against parasitic infection of *Enta*moeba histolytica and Cryptosporidium parvum. AgNPs expresses larvicidal activities against Aedes aegypti and Culex quinquefasciatus [57]. Allahverdiyev et al. demonstrated a study to assess the effects of AgNPs over the biological parameters of *Leishmania tropica*. AgNPs attack the parasites by impairing the lipophosphoglycan and glycoprotein 63 present in the surface of the parasite which is responsible for the infection. The study revealed that AgNPs have the potential to inhibit the promastigote's proliferation activity. And further, the AgNPs in the presence of increased UV light hinders the endurance of amastigotes in host cell [58]. A study on biosynthesis of AgNPs using Corn cob nanoxylan as a reducing and stabilizing agent observed a minimum inhibitory concentration (IC<sub>50</sub>) of 25 µg/mL against Leishmania amazonensis promastigotes [59]. In an in vitro study of AgNPs against Entamoeba histolytica trophozoite, the result showed a mortality percentage of 46.2 %, 42.4 % for 75 µg/ml and 46.2 %, 46.7 % for 100 µg/ml of concentration after 24 h and 48 h incubation respectively [60].

## Anti-inflammatory Agent

Silver-based NPs is powerful in preventing the bacterial infections and inflammation [61]. Intra-peritoneal injections of AgNPs in animal models showed decrease in the degree of post operative fibrous adhesions. Hebeish et al., have done the in vivo efficacy of albino rats' and grouped the 24 albino rats into 4 groups. Group 1 was the negative control and given 1 ml of saline. Group 2 & 3 rats were administered with two different AgNPs concentration of 250 ppm and 124 ppm respectively. Group 4 was the reference drug administered with indomethacin at concentration of 20 mg/kg rat. After one hour, all animals were given a sub planter injection of 1% carrageenan solution in 0.1ml of saline over the right hind paw and 0.1 ml saline in left hind paw. After four hours of administration, rat's both the paws were excised and weighed separately. The oedema percentage showed that the synthesized AgNP could significantly decrease in degree of rat oedema. The oedema percent of 250 ppm concentration of AgNPs gave the same effect as the standard drug indomethacin. Thus, it was indicated that AgNPs could reduce inflammation significantly [62]. Shensha et al., made a study on Nigella sativa oil mediated AgNPs to assess the antiinflammatory activity in male Wister rats which showed inhibitory concentration at 54.40% (1 h) and 60.30% (5 h) with the dose of 0.3 mg/kg bodyweight [63]. Researchers have biosynthesized AgNPs using Selaginella myosurus aqueous extract which showed significant inhibition of paw edema of Wistar rats of 44.30% (1h), 57.60% (5 h) and 60.50% (5 h) for the concentration 0.1, 0.2 and 0.4 mg/Kg (body weight) respectively [64].

## Antiplatelet Agent

Thrombotic disorders have become the remarkable problem in the medical field. Anticoagulant and thrombolytic therapy can lead to many bleeding complications. Recently Dakshayan et al. demonstrated the role of *Selaginella bryopetris* (Sanjeevini) plant extract supported AgNPs (SPE @AgNP) in platelets. Platelet aggregation assay was performed in platelet rich plasma with ADP and Epinephrin as agonists. SPE @AgNP inhibited ADP only and not the Epinephrin. Platelet exhibits its major role in arresting the bleeding in addition to the coagulation factors and they tend to aggregate at the injury site forming a platelet plug and it is vulnerable to the collagen present in endothelium [65]. Thrombin, ADP, epinephrine, thromboxane, thrombin, platelet activating factors etc. are the agonist that activates the platelets [66]. Formation of thrombus in arteries and vein occurs when platelets get hyperactivated. Thus, SPE @AgNP was used in the treatment of thrombotic disorders as a superior antiplatelet agent along with its anticoagulant activity [67]. Additionally, Shrivastava *et al.*, demonstrated that AgNPs inhibited the integrin mediated platelet functional responses like aggregation, adhesion, secretion to immobilized fibrinogen or collagen [26].

# Therapeutic Application of Silver Nanoparticle

#### Wound Dressing

AgNPs are used in wound dressing, in case of toxic epidermal and necrolysis, severe burns etc. AgNPs takes about 3.35 days on an average to heal the wound and the bacterial load decreases in wound area with no further effects [68]. AgNPs works more effectively when compared with 1 % silver sulfadiazine for superficial burns. In case of deep burns, AgNPs and 1 % silver sulfadiazine does the same work and AgNPs promotes the restoration of intact epidermal barrier but do not form any new tissue [69]. The application of biopolymer based biomaterial increases because of biocompatibility, biodegradability, non-immunogenicity and nontoxicity. The biopolymers used are gelatine, collagen, keratin, natural rubber proteins, polysaccharide. Collagen is a natural biopolymer and the usage of biopolymers in wound healing showed a better positive output in the clinical trials. Collagen sponges and glycosaminoglycans act as a double layered artificial skin and heals the wound. These act as a good carrier for AgNP to release the drug in a sustained manner [70]. Hasari et al., reported that silk based novel bilayer wound dressing material with gelatine is less toxic to skin cell than other wound dressing available and it also promotes the wound healing by increasing the collagen production [71]. Studies have shown that chitosan and keratin have wound healing effects and antimicrobial activity. Chitosan is effective for the treatment of chronic periodontitis [72]. Scientists have emerged with worlds first AgNP based commercially available wound dressing material (ACTICOAT: Smith and Nephew, UK) that covers the huge area of burns and increases the wound healing activity [73]. A study was done with konjac glucomann silver nanoparticle (KGM/AgNP) a composite sponge which possessed a better wound healing capacity and also exhibited good cytocompatibility. By animal experiments they have confirmed the activity of wound healing, and on the day 14, KGM/Ag3 wound healing capacity was increased to 99 % [74].

#### Silver Impregnated Catheters

The prevalence of central venous catheter (CVC)-related bloodstream infections were about 80,000 cases annually. However, the use of CVC in hospitals of the USA was around 5 million for 1 year [75]. Patients with tumour, intracerebral haemorrhage, subarachnoid haemorrhage were implanted with external ventricular drain (EVD) catheters usually for the therapy of acute hydrocephalus. The EVP catheters were significantly used for monitoring the draining CSF and intracranial pressure. Previously catheters were impregnated with antibiotics that decreased the colonization rates [76], which led to bacterial resistance. Therefore, a new trend of silver coated catheters was used in clinical field, in which the silver ions bind with an inert ceramic zeolite by the help of inorganic silver powder. Recent studies described that, there was a remarkable reduction in the colonization rate in silver impregnated CVC [77]. In a previous study, A.baumanni was made to form a biofilm to mimic the *in vivo* infection conditions and the CVC was coated with polydopamine and tested for the bactericidal activity. By surface characterization with field emission scanning electron microscopy, water contact angle (CA), Raman spectroscopy the results showed that dopamine coated AgNPs had a CA value of  $49.1 \pm 0.3$  [78]. Fichtner et al. conducted retrospective clinical analysis to see the contrasting efficacy between silver coated EVD catheters and standard non coated catheters. There was a significant reduction in the positive culture of CRI. Bacterial colonization was likely to be 4 times in EVD catheters in comparison to the standard non coated catheters. Another study showed that the growth of *Staphylococcus aureus* reduced in the silver coated EVD catheters [79].

#### Silver in Orthopaedics

Benchmark treatment for the arthritic disease is the artificial joint replacement. The use of bone cements like poly methyl methacrylate (PMMA) resulted in high rate of infection when integrated in the bones. Nanotechnology has evolved in the field of orthopaedics and trauma. Hence, nano silver coated bone cement resulted in outrageous antibacterial activity against broad range of bacteria including methicillin resistant Staphylococcus aureus (MRSA). Taken further, the nanoparticle did not show any cytotoxicity. Ultra high molecular weight polyethylene (UHMWPE) was used for the artificial joint replacement but the only disadvantage was with the wear and tear and associated debris generation, which led to inflammation and failure of joints in the body. This major problem was overcome by integration of AgNPs with bone cement. This protocol consequently reduced the polymer debris formation [20, 80]. The rate of infection was reduced by adding the AgNPs to the outer layer of the implants. A group of scientist demonstrated that there was an effective resistance to *Escherichia coli* by exposing it to modified titanium film. It also had a role in reducing the pin tract associated infections [69, 81]. Orthopaedic infections resulted in high morbidity when osteoblast like cells, and bone marrow mesenchymal stem cells were exposed to AgNPs and showed maximum inhibitory concentration of 25  $\mu$ g/ml [82].

#### Silver Surgical Meshes

Prosthetic mesh is not commonly used implanted devices for pelvic reconstructive surgery and hernia repair. The incidence of mesh associated infections ranges between 0.6% and 8.0% [83, 84]. In accordance to the one million herniorrhaphies there were around 30,000–50,000 prosthetic mesh infection in the USA [85]. Multiple antibacterial coatings were used on the medical devices such as urinary catheters, central venous catheters and surgical meshes to reduce the infection rate [86]. To reduce the occurrence of prosthetic mesh infections in the post pelvic and hernia surgery, nanocrystalline silver particles with polypropylene was used [87]. The antimicrobial activity of silver nanoparticle relies in the electrical state of the ions. Silver is biologically active in its soluble form as Ag<sup>+</sup>. Commonly used topicals like silver sulfadiazine, silver nitrate, has silver in the form of Ag<sup>+</sup>. Topical silver formulations were used two to twelve times a day in burn areas because of the rapid inactivation of silver ions with chloride or organic ions present in the wounds. In comparison to Ag<sup>+</sup>, Ag<sup>0</sup> does not form complex resulting in halt in rapid inactivation of microbes [88]. Nanocrystalline silver has decreased one million nosocomial infections in a year with the patients implanted with prosthetic material [89]. Cohen *et al.*, reported that polypropylene integrated with nanocrystalline silver particle (NCSP) showed that silver particles circulated inside the mesh and produced a zone of inhibition and higher inhibition efficacy against Staphylococcus aureus. The zone of inhibition increased in a dose dependent manner with increase in the concentration of silver [90]. NCSP also holds the property of anti-inflammatory agent. Secondary mechanism of NCSP is to repress tumour necrosis factor  $\alpha$  and interleukin (IL)-12 and initiate inflammatory cell apoptosis [91].

### Dentistry

AgNPs pops up as a promising agent used in dentistry. This property is because of their integration of antimicrobial property in dental biomaterials [92]. The major mechanism is by liberating cationic silver and its oxidative potential [93]. Using AgNPs in dentistry, is effective against multidrug resistant bacteria and for its prophylactic action. AgNPs are used in different streams like preventive dentistry, orthodontics, endodontics, periodontics and in oral dentistry. AgNPs inhibit the growth of Staphylococcus aureus, Streptococcus mitis, Streptococcus gordoinii biofilms. Additionally recent study states that AgNPs exert its antibacterial activity against Streptococcus sobrinus, Lactobacillus acidophilus, Lactobacillus casei, Streptococcus sanguinis, Enterococcus faecalis and Actinomyces actinomycetemcomitans [94]. Incorporation of AgNPs into polymers used as denture base and tissue conditioners in stomatitis have showed superior antimicrobial activity and capacity to fight against oral infections. Another study stated that modified denature base acrylic combined with AgNPs at 20.0 wt.% showed antifungal property [95]. AgNPs with smaller diameter size exerts good biofilm inhibition when compared with larger particles. Biological synthesis of AgNPs using neem, onion, and tomato with size measuring 26.2 to 33.3 nm showed antimicrobial activity against Staphylococcus aureus because of the high concentration of flavonoids and terpenoids in it [96]. Pérez-Díaz et al., reported that AgNPs inhibited the growth of planktonic Streptococcus mutans and killed the Streptococcus mutans biofilms. Thus, AgNPs play a significant role in dentistry and prevents dental caries [97]. Decrease in adhesion of biofilm and production of lactate by microorganism is seen in AgNPs treated with titanium disc -based composites [98]. A nano bacteriostatic agent silver Nano fluoride had inhibited the growth of *Streptococcus mutans* and it could be used once a year, easy to use, and had a better cost benefit ratio. This was a superior replacement for sodium fluorine [99]. Titanium micro implants with biopolymer coated AgNPs (Ti-BP-AgNp) showed largest zone of inhibition of  $50.58 \pm 4.88 \text{ mm}^2$  for *Streptococcus mutans*. The zone of inhibition for *S. sanguinis* was  $27 \pm 3.01 \text{ mm}^2$  and for *A. actinomycetemcometans* showed a smaller zone of inhibition of about  $25 \pm 3.06 \text{ mm}^2$  with the control showing no zone of inhibition [100].

## **Cancer Theranostics**

Cancer is a global menace and is caused by environmental effects and mutations in genes which activates the sequence of events in molecular level and leads to tumour formation. [101]. There are two major causes for cancer, they are external and internal factors. The external factors include radiation, viruses, chemical exposures, environmental conditions etc. [102]. Internal factors comprise the mutations, hormones and immune conditions which trigger the process of carcinogenesis [103]. Chronic low dose exposure to oxidative stress is also known to be a contributor for cancer onset by circumventing apotosis [104]. Many studies showed that AgNPs get localized inside perinuclear space of cytoplasm and endo lysosomal compartment cells by entering cells through endocytosis [105, 106]. AgNPs affect the respiration of cells and produce reactive oxygen species (ROS). As AgNPs are harmful to cells and proceeds to oxidative stress, damage of DNA, trigger the apoptosis and damage of mitochondria to cancer cells [107-112]. Studies showed that AgNPs can affect the activity of vascular endothelial growth factor which are involved in angiogenesis [113]. Theranostics is the mixture of diagnostics and therapy. Biosynthesized AgNP possess theranostic applications as it holds the anti-cancer property and used in targeted drug delivery and bioimaging vehicle [114]. It is a cost effective, safe, simple and eco-friendly approach [115]; the different applications of AgNPs in anti-cancer therspy is summarised in Table 1.

# Silver Nanoparticle in Imaging

#### Silver Nanoparticle as Photoacoustic Imaging

In the field of biomedical applications, research has increased in the area of designing and delivery of NPs to the specific organ. To know whether the nanoparticle has delivered to the diseased tissue and to also know about the intended function of NPs, a new combination of photoacoustic imaging modality and custom designed nano system has emerged [157, 158]. The main principle behind this is, first, the object will absorb light, then absorbed light energy will develop as heat and finally because of thermoelastic expansion, acoustic waves will be release out [159]. Photoacoustic imaging was first proposed by Oraevsky, for the use of biomedical applications [160]. AgNPs possess strong optical absorbance and scattering properties and are used as contrast agents for imaging therapies. With the light of wavelength 800 nm and radiation, the NP's injected 1cm deep in *ex vivo* pancreatic tissue could be detected using ultrasound imaging and photoacoustics. First mode is, AgNPs are engineered to target the tumour site specifically to the leaky blood vessels of the tumor and the reduced the rate of clearance due to lack of functional lymphatic vessels, and will retain the AgNPs [161]. The second one is, at the site of tumour, the AgNPs gets conjugated with

<ul> <li>No. Types of Silver nanoparticles Used</li> <li>Green synthesis of AgNPs using Ganoderma neo-japonicum Imazeki</li> <li>Green synthesis of AgNPs using <i>Escherichia fergusoni</i></li> <li>Green synthesis of AgNPs using sucrose</li> <li>Green synthesis of AgNPs using sucrose</li> <li>Green synthesis of AgNPs using <i>Saracum officinale</i></li> <li>Chemical biosynthesis of AgNPs using <i>Sargassum vulgare</i></li> <li>Green synthesis of AgNPs using <i>Sargassum vulgare</i></li> <li>Green synthesis of AgNPs using <i>Sargassum vulgare</i></li> <li>Green synthesis of AgNPs using <i>Sargassum vulgare</i></li> </ul>	Table	Table 1         Antitumor effects of AgNPs			
Green synthesis of AgNPs using Ganoderma neo-japonicum Imazeki Green synthesis of AgNPs using <i>Escheri-</i> <i>chia fergusoni</i> Green synthesis of AgNPs using sucrose Green synthesis of AgNPs using Taraxacum officinale Chemical biosynthesis of AgNPs using Saraasum phycocyanin extracted from Nostoc linckia as reducing agent Green synthesis of AgNPs using Sargassum vulgare Green synthesis of AgNPs using Piper longum leaf	S.No.	Types of Silver nanoparticles	Model used	Outcome	References
Green synthesis of AgNPs using Escheri- chia fergusoni Green synthesis of AgNPs using sucrose Green synthesis of AgNPs using Taraxacum officinale Chemical biosynthesis of AgNPs using phycocyanin extracted from Nostoc linckia as reducing agent as reducing agent Green synthesis of AgNPs using Sargassum wulgare Green synthesis of AgNPs using Piper longum leaf		Green synthesis of AgNPs using Ganoderma neo-japonicum Imazeki	Breast cancer cell lines	AgNP increased the production of hydroxyl radical and reactive oxygen species by inhibiting the cell viability. They play a prime role in apoptosis by activation of caspase 9 and DNA nuclear fragmenta- tion.	Gurunathan <i>et al.</i> , [116]
Green synthesis of AgNPs using sucrose Green synthesis of AgNPs using Taraxacum officinale Chemical biosynthesis of AgNPs using phycocyanin extracted from Nostoc linckia as reducing agent as reducing agent Green synthesis of AgNPs using Sargassum vulgare Green synthesis of AgNPs using Piper longum leaf	5	Green synthesis of AgNPs using Excheri- chia fergusoni	Breast cancer MCF 7 cell lines	Cytotoxicity effects of bio synthesized AgNP reduced the activation of LDH, increases the ROS production and results in apoptosis.	Gurunathan et al., [117]
Green synthesis of AgNPs using Taraxacum officinale Chemical biosynthesis of AgNPs using phycocyanin extracted from <i>Nostoc linckia</i> as reducing agent Green synthesis of AgNPs using <i>Sargassum</i> <i>vulgare</i> Green synthesis of AgNPs using <i>Piper</i> <i>longum</i> leaf	З.	Green synthesis of AgNPs using sucrose	Malignant skin melanoma (HT144 cell line) and squamous cell lung carcinoma (H157 cell line)	Observed a prominent antitumor activity against vincristine and methotrexate.	Nazir <i>et al.</i> ,[118]
Chemical biosynthesis of AgNPs using phycocyanin extracted from <i>Nostoc linckia</i> as reducing agent Green synthesis of AgNPs using <i>Sargassum</i> <i>vulgare</i> Green synthesis of AgNPs using <i>Piper</i> <i>longum</i> leaf	<del>4</del>		Liver hepatocellular carcinoma in HepG2 cell line	Possess an enhanced activity against the commercial AgNP and in increased cytotoxic effects against HepG2 cell line	Saratale <i>et al.</i> , [119]
Green synthesis of AgNPs using Sargassum vulgare Green synthesis of AgNPs using Piper longum leaf	5.	Chemical biosynthesis of AgNPs using phycocyanin extracted from <i>Nostoc linckia</i> as reducing agent	Human breast adenocarcinoma	Observed a significant cytotoxic activ- ity against MCF-7 cell line with the inhibitory concentration (IC <sub>50</sub> ) of about $27.79\pm2.3$ µg/ml	Naggar <i>et al.</i> ,[120]
Green synthesis of AgNPs using <i>Piper</i> longum leaf	6.		Human myeloblastic leukaemia in HL60 and HeLa cells	Prevents carcinogenesis related with irradiation by inhibiting lipid peroxida- tion – mediated reactive oxygen species generation which leads to apoptosis.	Govindaraju <i>et al.</i> , [121]
	7.	Green synthesis of AgNPs using <i>Piper</i> longum leaf	Hep 2 cell line	Observed effective cytotoxic effect of 94.02% at 500 μg/ml due to formation of ROS.	Jacob <i>et al.</i> , [122]
	∞.	Green synthesis of AgNPs using Inonotus obliquus extract	Human lung cancer in A549 cell line and breast cancer in MCF -7 cell line.	Cell lines shows significant cytotoxic effects.	Nagajyothi <i>et al.</i> , [123]

Table	Table 1 (continued)			
S.No.	Types of Silver nanoparticles Used	Model used	Outcome	References
.6	Green synthesis of AgNPs using Commelina nudiffora	ng Commelina HCT-116 Colon cancer cell line.	AgNP showed less toxicity when compared with AuNP and inhibitory concentration (IC <sub>50</sub> ) was 100 µg/ml.	Kuppusamy et al., [124]
10.	Green synthesis of AgNPs using Melia dubia leaf	Human breast cancer cell line (KB)	The inhibitory concentration (IC <sub>50</sub> ) was 31.2 µg/ml.	Kathiravan <i>et al.</i> , [125]
11.	Green synthesis of AgNPs using Dimocar- pus longan Lour	H1299 lung cancer cell line.	Possess inhibitory effect with (IC <sub>50</sub> ) value of 5.33±0.37 µg/ml and suppress the growth of H1299 tumours in SCID mice.	He <i>et al.</i> , [126]
12.	Cisplatin (cis-diamminedichloroplantium II) bound bio AgNPs using <i>Penicillium</i> , <i>Fusarium and Aspergillus</i> .	Prostate cancer cell line (PC -3).	Decreases the toxic effects and increases the efficacy against human prostate cancer	El-Sheikh <i>et al.</i> , [127]
13.	Green synthesis of AgNPs using root extract of <i>Erythrina indica</i>	<ul> <li>pg root extract Breast and lung cancer cell line (MCF-7 and Viability of the cells decreased with increase in concentration of AgNP 25µg/ml, the viability percentage i 23.89±0.39 for MCF-7 cell line an 13.86±0.95 for HEP G2 cell line. root mediated synthesis plays a rol cancer chemotherapy and chemopr tion.</li> </ul>	Viability of the cells decreased with increase in concentration of AgNP. At 25µg/ml, the viability percentage is 23.89±0.39 for MCF-7 cell line and 13.86±0.95 for HEP G2 cell line. This root mediated synthesis plays a role in cancer chemotherapy and chemopreven- tion.	Sre et al., [128]
14.	Green synthesis of AgNPs using Acalypha indica leaves extract.	Human breast cancer MDA-MB-231 cells	Four different concentrations were used, such as, 1, 10, 50, 100 µg/ml, in which 100 µg/ml AgNP exposed toxicity to some extent.	Krishnaraj.C <i>et al.</i> , [129]
15.	Green synthesis of AgNPs using leaf extract Human breast carcinoma (MCF-7 cells) of mistletoe <i>Dendrophthoe falcata</i> (L.f) Ettingsh	Human breast carcinoma (MCF-7 cells)	At merest dosage of 5 $\mu$ g/ml, of fabri- cated AgNP they observed the enhanced cytotoxic effect. This concentration is the IC <sub>50</sub> value.	Sathishkumar <i>et al.</i> , [130]
16.	Green synthesis of AgNPs using Datura inoxia leaves.	Human breast cancer (MCF-7 cell line)	The inhibitory concentration at 50% ( $IC_{50}$ ) was 20 µg/ml. It also seizes the cell cycle phase, suppresses the growth, finally induces apoptosis and exhibits the antiproliferative activity against MCF -7 cell line.	Gajendran <i>et al.</i> , [131]

Table	Table 1         (continued)			
S.No.	Types of Silver nanoparticles Used	Model used	Outcome	References
17.	Green Synthesis of AgNPs using Clinacan- thus Nutans leaves extract.	Oral squamous cell carcinoma cell line (HSC-4)	Observed a prominent cytotoxic effect at the concentration of $1.61 \pm 0.14 \ \mu g/ml$ by repressing the release of Bcl-2 protein.	Yakop <i>et al.</i> , [132]
18.	Green synthesis of AgNPs using Pimpinella anisum seeds	Human neonatal skin stromal cells (hSSCs) and colon cancer cells (HT115)	Observed lower cytotoxicity for bio- synthesized AgNP of about 51.39% in comparison with chemically synthesized one that showed 85.45%. And it is use- ful in pharmacological applications for producing nanodrugs.	AlSalhi <i>et al.</i> , [133]
19.	Biosynthesized AgNPs using aqueous fruit extract of Chaenomeles sinensis (CS)	Human breast cancer cell line (MCF-7)	At concentration of 0.01 µg/ml, the viability Keun Hyun Oh <i>et al.</i> , [134] percentage of cells was remarkably reduced.	Keun Hyun Oh et al., [134]
20.	Green synthesis of AgNPs using Saccharo- myces boulardii	Human breast cancer cell line (MCF-7)	$IC_{50}$ of bio-synthesized AgNP was about less than 10 μg/ml. This indicated that AgNP with low concentration exhibited almost 80% of inhibition of the cancer cells. They observed no significant changes in the higher concentration (10- 100 μg/ml)	Kaler <i>et al.</i> , [135]
21.	Green synthesis of AgNPs using Indigofera tinctoria leaf extract	Lung cancer cell line (A549)	IC <sub>50</sub> value of AgNP-tinctoria was71.92 ±0.76 μg/ml. This is because of the NPs induced ROS.	Vijayan <i>et al.</i> , [136]
22.	PVP coated AgNPs	Human lung cancer cell line (Alveolar cell line A549)	<ol> <li>Ag<sup>+</sup> with 0-10 μg/ml and AgNP with 0-20 μg/ml concentration exhibited similar toxic effects and a decrease in mitochondrial function.</li> <li>Study showed that oxidative stress was induced by both AgNP and Ag<sup>+</sup> by cor- relating with geno and cytotoxicity.</li> </ol>	Foldbjerg <i>et al.</i> , [137]

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Table 1	Table 1 (continued)			
S.No.	Types of Silver nanoparticles Used	Model used	Outcome	References
23.	Green synthesis of AgNPs using aqueous extract of <i>Phyllanthus emblica</i> ( <i>PE</i> ) fruit.	Laryngeal carcinoma cells (Hep2 cell line)	Observed a potent cytotoxic effect, the IC <sub>50</sub> value of PE alone was 30 μg/ml and PE-AgNP was 20 μg/ml.	Rosarin <i>et al.</i> , [138]
24.	Green synthesis of AgNPs using Padina tetrastromatica seaweed extract	Human breast cancer cell line (MCF-7)	With increased concentration, the percent- age of inhibition increased. The bio-AgNP showed a value of IC <sub>50</sub> as 86.7 µg/ml and AgNP value was 200 µg/ml. The observed cytotoxicity effect of AgNP was because of caspase 3 mediated apoptosis.	Selvi <i>et al.</i> , [139]
25.	Green synthesis of AgNPs using leaves of Vitex negundo	Human colon cancer (HCT15 cell line)	Proliferation of HCT-15 was inhibited with a concentration of 20 $\mu g/ml$ (IC <sub>50</sub> ) at 48h incubation. They exhibited a antiprolifera- tive effect by seizing the G <sub>0</sub> /G <sub>1</sub> - phase and induced programmed apoptosis.	Prabhu <i>et al.</i> , [140]
26.	Green synthesis of AgNPs using Artemisia turcomanica leaf extract	Gastric cancer (AGS cell line)	The IC <sub>50</sub> value of bio-synthesized AgNP was 4.88 µg/ml and commercial AgNP showed a value of 6.37 µg/ml. This study inferred that least concentration of bio-AgNP was sufficient to inhibit the cell growth when compared with commercial AgNP.	Mousavi <i>et al.</i> , [141]
27.	Green synthesis of AgNPs using Bacillus licheniformis in tumour bearing mice.	Dalton's lymphoma ascites (DLA cell line)	Observed a prominent decrease in tumour volume from 7.3ml to 2.6ml in the group of mice treated with AgNP with the concentration of 500 µg/ml in about 15 days.	Sriram <i>et al.</i> , [142]
28.	Chemical synthesis of AgNPs.	Glioma (U251 glioblastoma cells)	AgNP exhibited better inhibition over U251 glioma cells in comparison to AuNP. The $IC_{50}$ of AgNP was 75.9 µg/ml.	Liu et al., [143]
29.	Metal silver and PVP coated AgNPs on tumor bearing mice	Lymphoma	70% and 60% of mice survived at the day 35 with the metal silver and PVP AgNP administered at day 0.	Lara-Gonzalez et al., [144]

Table (	Table 1 (continued)			
S.No.	Types of Silver nanoparticles Used	Model used	Outcome	References
30.	Poly vinyl pyrrolidone -coated nano silver (PVP-AgNP) and bare nano silver (AgNP)	<ul> <li>Human hepatoma cell line (HepG2 cell line) 1. AgNP caused more DNA damage to and mice.</li> <li>HepG2 cells than PVP-AgNP, where: PVP-AgNP possessed more chromos aberration in comparison to AgNP.</li> <li>2. At the highest dose of 250 mg/mL, they observed no inhibitory effects in bone marrow cells of mice.</li> </ul>	<ol> <li>AgNP caused more DNA damage to HepG2 cells than PVP-AgNP, whereas PVP-AgNP possessed more chromosomal aberration in comparison to AgNP.</li> <li>At the highest dose of 250 mg/mL, they observed no inhibitory effects in the bone marrow cells of mice.</li> </ol>	Wang <i>et al.</i> , [145]
31.	Green synthesis of AgNPs using curcumin derivative (ST06)	Cervical cancer in HeLa cell line and EAC (Ehrlich Ascites carcinoma) tumour bear- ing mice.	<ol> <li>At a concentration of 1 μg/ml of ST06 and 1 μg/ml of ST06-AgNP, 50% of the cells were killed in a HeLa cell line.</li> <li>Inferred that at the concentration of 5 μg/ml (ST06-AgNP) intraperitoneally inhibited the tumour growth in a tumour bearing mice which did not affect the body weight.</li> </ol>	Murugesan <i>et al.</i> , [146]
3.	Green synthesis of AgNPs using Spinacia oleracea leaves.	Myoblast cancer (mouse C <sub>2</sub> C <sub>12</sub> cell and in zebra fish)	1. AgNP showed 100% inhibition of growth at low concentration of about 20 μg/ ml and AgNP, at a concentration of 100 µg/ml exhibited 20% viability of cells, whereas the plant extract did not possess any significant cytotoxic effects against $C_2 C_{12}$ cell. 2. In zebrafish embryo, AgNP was more toxic and exhibited 100% mortality at concentration of 3 µg/ml and AuNP showed the 100% mortality only at higher concentration of 300 mg/ml. Moreover, plant extract did not cause any mortality.	Ramachandran <i>et al.</i> , [147]

Table	Table 1 (continued)			
S.No.	Types of Silver nanoparticles Used	Model used	Outcome	References
33.	Green synthesis of AgNPs using poisonous plant <i>Cleistanthus collinus</i> extract.	Lung cancer cell line (A549 cell line) and mice.	<ol> <li>Poisonous plant at a finite dosage was used as an anticancer agent. The inhibi- tory concentration (IC<sub>50</sub>) was found to be 30 μg/ml.</li> <li><i>In vivo</i> histopathological findings in mice treated with bio-synthesized AgNP did not show any edema or inflammation in the organs. Thus, it could be used for diagnostic and therapeutic purposes.</li> </ol>	Kanipandian <i>et al.</i> , [148]
34.	Green synthesis of AgNPs using <i>Teucrium</i> polium leaf extract	Human gastric cancer (MNK45 cell line)	The IC <sub>50</sub> value of T. polium-AgNP is 68.2 µg/ml after 48h exposure.	Hashemi <i>et al.</i> , [149]
35.	Green synthesis of AgNPs using Albizia adianthifolia leaf extract.	Lung cancer (A549 cell line)	Observed significant cytotoxic effect and the cell viability percentage was 79% and 27% in the concentration of 10 μg/ml and 50 μg/ml, respectively.	Gengen et al., [150]
36.	Green synthesis of AgNPs using aqueous extract of <i>Punica granatum</i>	Lung cancer (A549 cell line)	Possessed cytotoxic effect to cancer cells but not to the normal cells. Potent cyto- toxicity i.e.,50% growth inhibition was observed after 48h at a concentration of 5 µg/ml.	Annu et al., [151]
37.	Green synthesis of AgNPs using walnut fruits (Juglans regia)	Breast cancer (MCF-7)	Observed a significant cytotoxic effect at the concentration of 60 µg/ml as 70% and 42% for AgNP and extract, respectively.	Khorrami et al.[152]
38.	Green synthesis of AgNPs using Aspergil- lus niger	Human colon cancer (HT29 cell line)	Highest cell viability percentage was at 10 µg/ml and lowest at 160 µg/ml with exposure time of 24-72h by exhibiting ROS mediated apoptosis.	Chengzheng <i>et al.</i> , [153]
39.	Green synthesis of AgNPs using alcoholic extract of Argemone Mexicana leaves.	Cervical cancer (SiHa human cervical cancer cell line)	Observed a decrease in percentage of cell viability (70-80%) with the concentration of 100µg/ml.	Jha <i>et al.</i> , [154]

Table	Table 1 (continued)			
S.No.	S.No. Types of Silver nanoparticles Used	Model used	Outcome	References
40.	Green synthesis of AgNPs using <i>Pseu-</i> domonas aeruginosa	Thyroid cancer (TCL1 cell line)	IC <sub>50</sub> value was observed as 48.5 µg/ml by Yang <i>et al.</i> , [155] increasing the lipid peroxidation, decreasing the mitochondrial membrane potential, reducing antioxidants and finally cell condensation took place.	Yang <i>et al.</i> , [155]
41.	Green synthesis of AgNPs using the extract Human hepatocellular carcinoma (HepG <sub>2</sub> of red sea weed <i>Pterocladiella capillacea</i> cell line)	Human hepatocellular carcinoma (HepG <sub>2</sub> cell line)	Infers that the level of cytotoxicity increased El Kassas <i>et al.</i> ,[156] with higher concentration. Untreated cells were used as negative control. The 50% of cell inhibition (IC <sub>30</sub> ) occurred at 3.7 μg/ml of concentration.	El Kassas <i>et al.</i> , [156]

antibodies and bind to the antigens present. By localizing the AgNPs at the tumour site, it provides photoacoustic contrast with normal tissues thereby useful in *in vivo* examination of tumour [16].

## Gold Nanoparticles and Their Applications

Over in the last half century, AuNPs have been developed in many ways. Synthesizing AuNPs is now more reliable and are high in yields. These AuNP have distinct traits, namely size and shape dependent electrical and optical features, surfaces that can be changed with ligands containing functional groups. With the help of these functional groups to hold the ligands, and other biomolecules such as antibodies, proteins , etc. the newly formed gold nanoconjugates possess a wider range of research in therapeutics [16]. AuNP have provided useful materials for various biomedical applications, such as material crystallizations, programmed assembly, conversion of NPs into dimers and trimers and then to DNA templates, detection methods and bioelectronics. In diagnosis of a disease, the process of binding between the analytes and AuNPs, the physicochemical properties of AuNPs can be altered. We shall see about the various applications of AuNP and highlight their evolution in theranostics [162] in the sections mentioned below.

### Medical Uses of Gold

Gold has been used in medicinal practices for many decades including Chinese and Indian medicines. They were mainly used for the treatment of arthritic diseases. But it was later found that nephrotoxicity was caused by gold due to the prolonged exposure. Hence, they were not used in medical practices until researchers developed a new compound called auranofins and aurothioglucose [163]. Yao et al, made a comparitive study of Gold clusters using Bovine serum albumin as template, Gold clusters using glutathione and Au nanoparticles with large particle diameter in both in vitro and in vivo. Gold nanoparticles acts by suppressing the proinflammatory mediators which is produced by lipopolyschharide. In comparing the above three, Au clusters with glutathione as template produced a better antiinflammatory effects thus it is one of potent nanodrug for treating Rheumatoid arthritis [164]. Osteoarthritis is kind of the arthritis which is attributed by swelling, stiffness and joint pain. Researchers studied the use of AuNPs for improvising the delivery of chondroitin sulfate. Chrondroitin sulfate is one of the drug for treating osteoarthritis. The combination of Chondroitin sulfate and AuNP augment the production of extracellular matrix and proliferation of chrondrocytes [165]. Since they are non-toxic, they are vividly used in biomedical applications as well as in gene and drug delivery. The optical intensity of AuNP has been subjugated for polynucleotide detections. Gold consists of a dielectric core and can be used suitably to make nanoshells, which are very useful in Surface enhanced Raman spectroscopy (SERS). AuNP has the ability to aggregate upon themselves when they interact with proteins. One of the basis for the quantitative method of colorimetric determination of proteins were the sudden colour change in the solution. This process helped in a better accurate result for many analytical methods including ELISA. AuNP is used in photothermal therapy. This is used in tumor therapy and infectious diseases. AuNP tend to become hot when they reach their maximum absorption in visible or near infrared region at the corresponding light wavelength. This kills the cells that are located around or inside the target cells. Today controlled and direct damaging of the tumor tissues is possible in cancer thermotherapy using laser radiation. But there are a lot of questions regarding its biodistribution, blood stream circulation, pharmacokinetics and toxicity. There are numerous research groups which use AuNP in their projects. There is a vast difference in the experiments, functionalizing methods, and dosages. Due to this, there has been major inconsistency with the data and the kinetics of biodistribution for toxicity estimation. Due to the development of efficient medical tools, AuNP has provided technologies for the functionalization of molecules providing stabilization *in vivo*. AuNP is now widely being used as problem solvers in bioimaging. [166]. AuNP have a great future in the medicine field, but one important concern is their safety level towards humans and environment. The AuNP toxicity depends on their physical dimensions and surface chemistry. Studies based on cytotoxicity of AuNP in human cells was done and the research showed that AuNP are nontoxic upto 250 mM, while ionic gold showed cytotoxicity at 25 mM. Lot of similar results were produced using AuNP in therapies [167].

### Gold Nanoparticles for Cancer Theranostics

Cancer therapy has grown rapidly in the past few years. But surgery with chemoradiotherapy still remains the go to procedure for fighting malignant cancer. Nanotechnology has recently been in the main topic in medical research, and many NPs have been studied for cancer therapy. The color of AuNP changes when there is increase in their size. Because of this unique property AuNP can be used in detecting various biomolecule and easily identify the tumor targets that are accumulated in vitro and in vivo. The most commonly used metal in cancer theranostics are gold and iron oxide because of their structure stability, variability of the size, controlled release and low toxicity during cancer theranostics [168]. Table 2 shows the different anti tumor effect of AuNPs.

AuNPs are used for both cell imaging and CT imaging in vivo. They are used mainly because they act as an alternative to X-ray based CT machines. They are used because of their better absorbent coefficient, easy attachment to the moieties and better body tolerance. AuNP has a great X-ray attenuation because of their high concentration and smaller size. Due to this it becomes easier to diagnose cancer with imaging [171]. The tissue absorption is very weak for light of wavelength >650 and <2,000 nm and that is the reason the NIR light is used for deep tumor imaging. AuNPs are the NIR-active probe for imaging the cancer cells in our body which is used for whole body scans. AuNPs are conjugated with anti-EGFR antibodies and used as a contrast agent in tumor imaging [172, 173].

#### Gold Nanoparticles for Cancer Therapy

#### Photothermal Therapy (PTT)

Gold has shown promising results in various cancer treatment such as photodynamic and photothermal therapies. This photothermal therapy is done through converting absorbed light into heat by nonradiative process. There are two main process:

- 1. The heat which is absorbed from energy conversion is given to the surrounding environment through the phonon relaxation within 100 ps.
- 2. A meticulous process of heating the electron and being cooled by the surrounding medium takes place, when the rate of heating is higher than the rate of cooling.

Table 2	Table 2 Antitumor effects of AuNPs			
S.No.	Type of Gold Nanoparticles used	Model used	Outcome	References
	Green synthesis of AuNPs using <i>Commelina</i> HCT-116 Colon cancer cell line. <i>nudiflora</i>	HCT-116 Colon cancer cell line.	AuNP at high concentration of 400 $\mu g'$ ml showed 90% cell death in HCT-116 cell line and the inhibitory concentration (IC <sub>50</sub> ) was 200 $\mu g'$ ml.	Kuppusamy <i>et al.</i> , [124]
6	Green synthesis of AuNPs using Acalypha indica leaves extract.	Human breast cancer MDA-MB-231 cells	Four different concentrations were used - 1, 10, 50, 100 µg/ml, higher toxic effects were showed by AuNP at 100 µg/ml posessing 40 % of cell toxicity. This exhibited AuNPs as a novel anticancer agent which could be used for the human breast cancer.	Krishnaraj.C <i>et al.</i> , [129]
З.	Biosynthesized AuNPs using aqueous fruit extract of <i>Chaenomeles sinensis</i> (CS)	Human breast cancer cell line (MCF-7)	Observed a significant cytotoxicity at >25 µg/ml.	Keun Hyun Oh et al., [134]
4	Green synthesis of AuNPs using Indigofera tinctoria leaf extract	Lung cancer cell line (A549)	$IC_{50}$ value of AuNP- tinctoria was 56.62 $\pm$ 0.86 µg/ml. This was because of the NPs induced ROS.	Vijayan <i>et al.</i> , [136]
5.	Chemical synthesis of AuNPs	Glioma (U251 glioblastoma cells)	The IC <sub>50</sub> of AuNP was 116.3 µg/ml which was less potent than AgNP.	Liu <i>et al.</i> , [143]
6.	Gold nanopartcles with hibiscus and cur- cumin extracts	Human colorectal cancer (HCT 116) and Michagen Cancer Foundation-7 (MCF-7)	The IC <sub>30</sub> of AuNP. Hibiscus against HCT-116 and MCF-7 was found to be $5.80\pm0.91$ µg/ml and $3.67\pm0.75$ µg/ml and for AuNP-Curcumin was $4.94\pm0.84$ µg/ml and $3.91\pm0.65$ µg/ml respectively.	Akhtar et al, 2022 [169]
	Gold nano bioconjugates with Elephantopus Scaber (linn,) hydro methalic extract.	Michagen Cancer Foundation-7 (MCF-7), A-549 cells, Squamous cell carcinoma (SCC-40) and Human colon cancer cell lines (COLO- 205)	The 50 % cell growth inhibition (GI $_{50}$ ) for AuNP was found to be < 10 µg/ml against MCF-7 cell line where as in A-549, SCC- 40, COL0-205 was 33.0, 28.3 and 24.7 µg/ml respectively. Thus AuNP was one of the good antiproliferative agent.	Shinde et al, 2022 [170]

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To use the photothermal therapy, a continuous wave laser is overlapped maximally with the AuNP resonance band. To achieve PTT using gold nanospheres the resonance of Continuous-Wave visible lasers should be in the visible region. This method can be used in treatment of shallow tumors. Recently, the development of antibody targeted gold nanospheres were done to primarily target EGFR on squalor carcinoma cells [172]. Researchers have developed new type of nanomatryoshkas which consists of PEG stabilizing ligands and concentric gold silica layers. In comparison to the existing gold-silica nanoshells which is approximately 150 nm, these nanomatryoshkas could facilitate higher concentration in tumors due to their ability to infiltrate the smaller AuNP in to the tissue. The survival rate of TNBC model was lesser than 1000 mm<sup>3</sup> which significantly decreased the size of the tumors by gold nanomatryoshkas and this was superior than the conventional silica-gold nanoshells. The results were taken under irradiation with a CW laser emitting 3W/ cm<sup>2</sup> at a wavelength of 808 nm [174].

## Photodynamic Therapy (PDT)

Photo synthesizers tend to convert the surrounding oxygen into a toxic reactive oxygen species when they are stimulated under a specific wavelength that might destroy the malignant cells in the surrounding proximity, which is now known as cancer PDT [175, 176]. To treat deeply buried tumors, AuNP exert PDT over NIR light activation. In addition, the incorporation of various photosensitizers with NIR active property into the AuNP can also be done with low dosages of organic photosensitizers and lasers with short exposure irradiation for PDT. When tested under an 808 nm laser, CS-AuNR-ICG NSs at the same time produced reactive oxygen species and hyperthermia, which attained complete inhibition of tumor growth in xenografted mice. In comaprison to PTT or PDT the combined therapy showed a drastically better therapeutic effectiveness [177]. The principle of photodynamic therapy is explained in Figure 2. Nanobiosensors has emerged as great tool in diagnosing cancer. Figure 3 describes the principle of nanobiosensor. Functionalized gold nanoparticles are immbolized on a template which can detect the analytes (proteins, toxins, antibodies, disease amrkers, cells etc.) with high specificity and sensitivity. The combination of analyte and recognition element which is attached to the gold nanoparticles gives a signal (heat, light, fluorescence, electrochemical change-current, potential and conduction, sound) which is converet into measurable electrical or optical signal. The amplifiers then amplify those signals to make them measurable using electronics [178].

## Use of Gold Nanoparticles in SARS-cov-2 Virus

#### COVID Test

AuNP can be used as a test kit to find out whether a person is infected by the virus or not. As we know the existing test kits require atleast 12 h to find whether the person is infected or not. In these critical 12 h the person might come in contact with another infected person and acquire the virus. With the help of AuNP we can reduce both the amount of time required for testing a person and the amount needed to spend for testing. These AuNP based assay test is done in an molecular level i.e. it checks the RNA of the person. This helps in accurate result for the identification of the virus. If this is the case, then it might prove positive for everyone because it'll be helpful in finding out who is infected and can

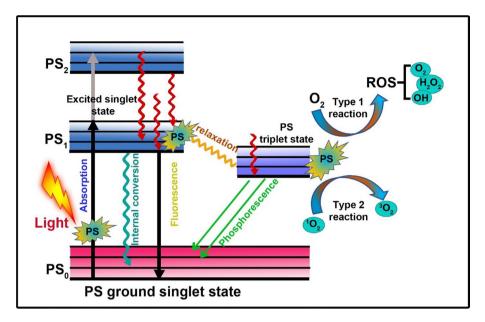


Fig. 2 Mechanism of Photodynamic therapy. PDT takes place through type I and type II reactions that can create ROS for killing the cancer cells

treat the person well before it gets worsened. Because the test kits available in the market doesn't give the results at the earliest time, this methododlogy can be helpful. [179].

## **Covid Vaccines**

The endurance of the human for the past 3 years become an dispute due to the outbreak of Covid 19. Severe Acute respiratory syndrome-Cov-2 (SARS-CoV-2) affects both humans and animals. Many researchers were in demand of developing Covid vaccines by collaborating with pharma companies. WHO reported on 9th June 2020 that among 136 vaccine candidates, 16 where nano based vaccines which are under clinical trials [180]. S proteins are the chief goal for the Corona virus vaccine production as it plays a role in its pathogenisis. Chen et al made a study using synthetic viral like particles as an effective vaccination tool in an avian model of Corona virus infection. Synthetic virus like particles were prepared by incubating the 100 nm gold nanoparticles in the optimal viral protein concentration solution. This enhanced the antibody titer, increased the lymphatic antigenic delivery, splenic T-cell response and decreased the infection symptoms. In comparison with the whole inactivated virus, synthetic viral like particles produced a better antiviral protection [181]. NPs in general are most preferred when it comes to destroy a deadly virus. This is mainly because of their nano size and the flexibility to alter a nanoparticle according to the virus. Covid 19 virus, an enveloped shape virus, which ranges from 50nm - 150nm in diameter has undergone many mutations. The exterior of this virus is covered by proteins in form of spikes which infects our cells. The genetic material of this virus is said to be ssRNA. This is one of the main reason for using a nanoparticle in killing this virus. Nanoparticle has the tendency to mimic a virus. This a good thing because it gives us the ability to directly attack the virus without affecting the nearby cells. The main

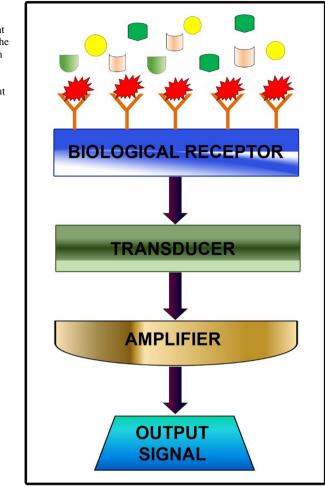


Fig. 3 Mechanism of Nanobiosensor. The biological receptor contains the recognition element that can specifically bind with the target molecule and covert them into measurable signal through the transducer. The signal gets amplified and recorded as output

path of these virus is to attack our respiratory system by binding with the cells in the respiratory system and affecting them with the help of protein spikes. There many ways to stop this from happening. As of now there are no confirmed therapeutic ways to destroy these viruses with the help of NPs. All of these are still in clinical trials. But it is well known that NPs is the way to destroy the virus efficiently [182].

# **Current Limitations**

As we have discussed above, AuNP gives us the possibility to be used in cancer diagnostics and therapeutics. But it is foolish to not consider the other side of the coin. These NPs might have side effects on health of the human beings. There are many studies in the cytotoxicity of AuNP, toxic effects of size, efficacy, response of NPs, biodistribution. But these researches give out contradicting results leaving us in a challenging situation. Absence of solid information on the effects of NPs could have serious effects and a negative impact on human health. Chaves et al., studied toxicity of gold nanoparticles by *in vitro* using HT- 29 and HepG2 cell lines and *in vivo* using Wister rats. For this study, reserchers used 10 ppm of 10, 30 or 60 nm gold nanoparticles for the experiments. AuNP increased the production of ROS in cells at 16 h and at the 32 h the overproduction was normalised. As a result of in vivo studies, AuNP produced an increase in protein carbonyl groups formation and lipid peroxidation, which was measured by Thiobarbituric acid reactive substance (TBARS). Gold content got accumulated in liver, intestine, spleen, faeces,urine and kidney of rats [183]. Li et al., proposed that AuNP obstructs the proliferation of cells by dysregulating the cell cycle genes and it also affects the genomic stability and DNA repair [184]. This findings shows that there is a side of bane for gold nanoparticles and it is very important to evaluate the toxicity of any metal and metaloxide nanoparticles [185].

# Conclusion

There are various metal NPs that as a predominant application in various fields of science. In which AuNP and AgNP has a vast range of application such as antimicrobial, antiinflamatory, diagnosis, anticancer agent and also has therapeutic activity such as in dentistry, orthopedics, cancer therapy, etc.,. which are mentioned above. Most of the researches are still in clinical trials. Some of them were already applied for treatment and diagnosis. Future research is necessary to conclude the safety aspects of nano silver and gold.

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**Data Availability** All the authors declare that all data and materials support their published claims and comply with field standards.

Code Availability Not Applicable

# Declarations

Ethics Approval • The manuscript is not be submitted to more than one journal for simultaneous consideration.

• The submitted work is original and should not have been published elsewhere in any form or language (partially or in full).

• A single study is not split up into several parts to increase the quantity of submissions and submitted to various journals or to one journal over time (i.e. 'salami-slicing/publishing').

• Results are presented clearly, honestly, and without fabrication, falsification or inappropriate data manipulation (including image based manipulation). Authors have adhered to discipline-specific rules for acquiring, selecting and processing data.

• No data, text, or theories by others are presented as if they were the author's own ('plagiarism'). Proper acknowledgements to other works have been given.

Informed Consent Not applicable

Consent to Participate Yes

#### Consent for Publication Yes

Conflict of Interest The authors declare no competing interests.

# References

- Singh, H., Du, J., Singh, P., & Yi, T. H. (2018). Ecofriendly synthesis of silver and gold nanoparticles by Euphrasia officinalis leaf extract and its biomedical applications. *Artificial Cells, Nanomedicine,* and Biotechnology, 46, 1163–1170. https://doi.org/10.1080/21691401.2017.1362417
- Singh, T., Shukla, S., Kumar, P., Wahla, V., Bajpai, V. K., & Rather, I. A. (2017). Application of nanotechnology in food science: perception and overview. *Frontiers in Microbiology*, 8, 1501. https:// doi.org/10.3389/fmicb.2017.01501
- Dasgupta, N., Ranjan, S., & Ramalingam, C. (2017). Applications of nanotechnology in agriculture and water quality management. *Environmental Chemistry Letters*, 15, 591–605. https://doi.org/10. 1007/s10311-017-0648-9
- Girigoswami, K., Viswanathan, M., Murugesan, R., & Girigoswami, A. (2015). Studies on Polymer-Coated Zinc Oxide Nanoparticles: UV-blocking Efficacy and in vivo Toxicity. *Materials Science and Engineering: C*, 56, 501–510. https://doi.org/10.1016/j.msec.2015.07.017
- Girigoswami, A., & Girigoswami, K. (2021). Nanotechnology in Detection of Food Toxins–Focus on the Dairy Products. *Biointerface Research in Applied Chemistry*, 11, 14155–14172. https://doi.org/ 10.33263/BRIAC116.1415514172
- Jamkhande, P. G., Ghule, N. W., Bamer, A. H., & Kalaskar, M. G. (2019). Metal nanoparticles synthesis: An overview on methods of preparation, advantages and disadvantages, and applications. *Journal of Drug Delivery Science and Technology*, 53, 101174. https://doi.org/10.1016/j.jddst.2019. 101174
- Ghosh, D., Girigoswami, A., & Chattopadhyay, N. (2012). Superquenching of coumarin 153 by gold nanoparticles. *Journal of Photochemistry and Photobiology A*, 242, 44–50. https://doi.org/10.1016/j. jphotochem.2012.05.027
- Ge, L., Li, Q., Wang, M., Ouyang, J., Li, X., & Xing, M. M. (2014). Nanosilver particles in medical applications: synthesis, performance, and toxicity. *International Journal of Nanomedicine*, 9, 2399. https://doi.org/10.2147/IJN.S55015
- Muthu, M. S., & Wilson, B. (2010). Multifunctional radionanomedicine: a novel nanoplatform for cancer imaging and therapy. *Nanomedicine*, 5(2), 169–171. https://doi.org/10.2217/nnm.09.107
- Haribabu, V., Girigoswami, K., & Girigoswami, A. (2021). Magneto-silver core-shell nanohybrids for theragnosis. *Nano-Struct.Nano-Objects*, 25, 100636. https://doi.org/10.1016/j.nanoso.2020. 100636
- Liu, X., Gao, P., Du, J., Zhao, X., & Wong, K. K. (2017). Long-term anti-inflammatory efficacy in intestinal anastomosis in mice using silver nanoparticle-coated suture. *Journal of Pediatric Surgery*, 52(12), 2083–2087. https://doi.org/10.1016/j.jpedsurg.2017.08.026
- Keshvadi, M., Karimi, F., Valizadeh, S., & Valizadeh, A. (2019). Comparative study of antibacterial inhibitory effect of silver nanoparticles and garlic oil nanoemulsion with their combination. *Biointerface Research in Applied Chemistry*, 9, 4560–4566. https://doi.org/10.33263/BRIAC96.560566
- Ali, G., Abd El-Moez, S., & Abdel-Fattah, W. (2019). Synthesis and characterization of nontoxic silver nano-particles with preferential bactericidal activity. *Biointerface Research in Applied Chemistry*, 9, 4617–4623.
- Kavya, J., Amsaveni, G., Nagalakshmi, M., Girigoswami, K., Murugesan, R., & Girigoswami, A. (2013). Silver Nanoparticles Induced Lowering of BCl2/Bax Causes Dalton's Lymphoma Tumour Cell Death in Mice. *Journal of Bionanoscience*, 7(3), 276–281. https://doi.org/10.1166/jbns.2013. 1135
- Girigoswami, A., Wafic, Y., Sharmiladevi, P., Haribabu, V., & Girigoswami, K. (2018). Camouflaged Nanosilver with Excitation Wavelength Dependent High Quantum Yield for Targeted Theranostic. *Scientific Reports*, 8, 16459. https://doi.org/10.1038/s41598-018-34843-4
- Giljohann, D. A., Seferos, D. S., Daniel, W. L., Massich, M. D., Patel, P. C., & Mirkin, C. A. (2010). Gold nanoparticles for biology and medicine. *Angewandte Chemie, International Edition*, 49(19), 3280–3294. https://doi.org/10.1002/anie.200904359
- 17. Maleki, M., Pourhassan-Moghaddam, M., Karimi, A., Akbarzadeh, A., Zarghami, N., & Mohammadi, S. (2019). Synthesis, characterisation, and application of chamomile gold nanoparticles in

molecular diagnostics: a new component for PCR kits. *Biointerface Research in Applied Chemistry*, 9(6), 4635–4641. https://doi.org/10.33263/BRIAC96.635641

- Azharuddin, M., Zhu, G. H., Das, D., Ozgur, E., Uzun, L., Turner, A. P., & Patra, H. K. (2019). A repertoire of biomedical applications of noble metal nanoparticles. *Chemical Communications*, 55(49), 6964–6996. https://doi.org/10.1039/c9cc01741k
- Ye, L., Cao, Z., Liu, X., Cui, Z., Li, Z., Liang, Y., Zhu, S., & Wu, S. (2022). Noble Metal-Based Nanomaterials as Antibacterial Agents. *Journal of Alloys and Compounds*, 2022, 164091. https://doi. org/10.1016/j.jallcom.2022.164091
- Alt, V., Bechert, T., Steinrücke, P., Wagener, M., Seidel, P., Dingeldein, E., et al. (2004). An in vitro assessment of the antibacterial properties and cytotoxicity of nanoparticulate silver bone cement. *Biomaterials.*, 25(18), 4383–4391. https://doi.org/10.1016/j.biomaterials.2003.10.078
- Panáček, A., Kvitek, L., Prucek, R., Kolář, M., Večeřová, R., Pizúrová, N., et al. (2006). Silver colloid nanoparticles: synthesis, characterization, and their antibacterial activity. *The Journal of Physical Chemistry*, 110(33), 16248–16253. https://doi.org/10.1021/jp063826h
- Lok, C. N., Ho, C. M., Chen, R., He, Q. Y., Yu, W. Y., Sun, H., et al. (2007). Silver nanoparticles: partial oxidation and antibacterial activities. *Journal of Biological Inorganic Chemistry*, 12(4), 527– 534. https://doi.org/10.1007/s00775-007-0208-z
- 23. Ovington, L. G. (2004). The truth about silver. Ostomy/Wound Management, 50(9A Suppl), 1S-10S.
- Manjumeena, R., Duraibabu, D., Sudha, J., & Kalaichelvan, P. (2014). Biogenic nanosilver incorporated reverse osmosis membrane for antibacterial and antifungal activities against selected pathogenic strains: an enhanced eco-friendly water disinfection approach. *Journal of Environmental Science and Health, Part A*, 49(10), 1125–1133. https://doi.org/10.1080/10934529.2014.897149
- Sanpui, P., Murugadoss, A., Prasad, P. D., Ghosh, S. S., & Chattopadhyay, A. (2008). The antibacterial properties of a novel chitosan–Ag-nanoparticle composite. *International Journal of Food Microbiology*, 124(2), 142–146. https://doi.org/10.1016/j.ijfoodmicro.2008.03.004
- Shrivastava, S., Bera, T., Singh, S. K., Singh, G., Ramachandrarao, P., & Dash, D. (2009). Characterization of antiplatelet properties of silver nanoparticles. ACS Nano, 3(6), 1357–1364. https://doi.org/ 10.1021/nn900277t
- Morones, J. R., Elechiguerra, J. L., Camacho, A., Holt, K., Kouri, J. B., Ramírez, J. T., et al. (2005). The bactericidal effect of silver nanoparticles. *Nanotechnology*, 16(10), 2346. https://doi.org/10.1088/ 0957-4484/16/10/059
- Leaper, D. J. (2006). Silver dressings: their role in wound management. *International Wound Journal*, 3(4), 282–294. https://doi.org/10.1111/j.1742-481X.2006.00265.x
- Barreiro, E., Casas, J. S., Couce, M. D., Sánchez, A., Seoane, R., Sord, O. J., et al. (2007). Synthesis and antimicrobial activities of silver (i) sulfanylcarboxylates. Structural isomers with identically or unequally coordinated Ag centers in an Ag 4 S 4 ring. *Dalton Transactions*, 28, 3074–3085. https:// doi.org/10.1039/b702936e
- Thomas, V., Yallapu, M. M., Sreedhar, B., & Bajpai, S. A. (2007). versatile strategy to fabricate hydrogel–silver nanocomposites and investigation of their antimicrobial activity. *Journal Colloid and Interface Science*, 315(1), 389–395. https://doi.org/10.1016/j.jcis.2007.06.068
- Lok, C. N., Ho, C. M., Chen, R., He, Q. Y., Yu, W. Y., Sun, H., et al. (2006). Proteomic analysis of the mode of antibacterial action of silver nanoparticles. *Journal of Proteome Research*, 5(4), 916–924. https://doi.org/10.1021/pr0504079
- Thomas, J. G., Chenoweth, C. E., & Sullivan, S. E. (2013). Iatrogenic Creutzfeldt-Jakob disease via surgical instruments. *Journal of Clinical Neuroscience*, 20(9), 1207–1212. https://doi.org/10.1016/j. jocn.2013.01.007
- Rai, M., Yadav, A., & Gade, A. (2009). Silver nanoparticles as a new generation of antimicrobials. Biotechnology Advances, 27(1), 76–83. https://doi.org/10.1016/j.biotechadv.2008.09.002
- Feng, Q. L., Wu, J., Chen, G. Q., Cui, F., Kim, T., & Kim, J. A. (2000). Mechanistic study of the antibacterial effect of silver ions on Escherichia coli and Staphylococcus aureus. *Journal of Biomedical Materials Research*, 52(4), 662–668. https://doi.org/10.1002/1097-4636(20001215)52:4%3C662:: AID-JBM10%3E3.0.CO;2-3
- Sondi, I., & Salopek-Sondi, B. (2004). Silver nanoparticles as antimicrobial agent: a case study on E. coli as a model for Gram-negative bacteria. *Journal Colloid and Interface Science*, 275(1), 177– 182. https://doi.org/10.1016/j.jcis.2004.02.012
- Song, H., Ko, K., Oh, L., & Lee, B. (2006). Fabrication of silver nanoparticles and their antimicrobial mechanisms. *European Cells & Materials*, 11(Suppl 1), 58.
- Ghodake, G., Kim, M., Sung, J. S., Shinde, S., Yang, J., Hwang, K., et al. (2020). Extracellular synthesis and characterization of silver nanoparticles—Antibacterial activity against multidrug-resistant bacterial strains. *Nanomaterials*, 10(2), 360. https://doi.org/10.3390/nano10020360

- Bhuyar, P., Rahim, M. H. A., Sundararaju, S., Ramaraj, R., Maniam, G. P., & Govindan, N. (2020). Synthesis of silver nanoparticles using marine macroalgae Padina sp. and its antibacterial activity towards pathogenic bacteria. *Beni-Suef University Journal of Basic and Applied Sciences*, 9(1), 1–15. https://doi.org/10.1186/s43088-019-0031-y
- Khan, B., Nawaz, M., Hussain, R., Price, G. J., Warsi, M. F., & Waseem, M. (2021). Enhanced antibacterial activity of size-controlled silver and polyethylene glycol functionalized silver nanoparticles. *Chemical Papers*, 75(2), 743–752. https://doi.org/10.1007/s11696-020-01335-7
- Thiruvengadam, V., & Bansod, A. V. (2020). Green Synthesis of Silver Nanoparticles Using Melia Azedarach and its Characterization, Corrosion and Antibacterial Properties. *Biointerface Research* in Applied Chemistry, 11(1), 8577–8586. https://doi.org/10.33263/BRIAC111.85778586
- Pulit, J., Banach, M., Szczygłowska, R., & Bryk, M. (2013). Nanosilver against fungi. Silver nanoparticles as an effective biocidal factor. *Acta Biochimica Polonica*, 60(4), 795–798. https://doi.org/ 10.18388/abp.2013\_2060
- Thirumalai Arasu, V., Prabhu, D., & Soniya, M. (2010). Stable silver nanoparticle synthesizing methods and its applications. *Research Journal of Biological Sciences*, 1, 259–270.
- Mallmann, E. J. J., Cunha, F. A., Castro, B. N., Maciel, A. M., Menezes, E. A., & Fechine, P. B. A. (2015). Antifungal activity of silver nanoparticles obtained by green synthesis. *Revista do Instituto de Medicina Tropical de São Paulo*, 57, 165–167. https://doi.org/10.1590/S0036-4665201500 0200011
- Fajar, M., Endarko, E., Rubiyanto, A., Malek, N., Hadibarata, T., & Syafiuddin, A. (2019). A green deposition method of silver nanoparticles on textiles and their antifungal activity. *Biointerface Research in Applied Chemistry*, 10, 4902–4907. https://doi.org/10.33263/BRIAC101.902907
- Al-Otibi, F., Perveen, K., Al-Saif, N. A., Alharbi, R. I., Bokhari, N. A., Albasher, G., et al. (2021). Biosynthesis of silver nanoparticles using Malva parviflora and their antifungal activity. *Saudi Journal of Biological Sciences*, 28(4), 2229–2235. https://doi.org/10.1016/j.sjbs.2021.01.012
- Al-Zubaidi, S., Al-Ayafi, A., & Abdelkader, H. (2019). Biosynthesis, characterization and antifungal activity of silver nanoparticles by Aspergillus niger isolate. *Journal Nanobiotechnology*, 1(1), 23–36. https://doi.org/10.26502/jnr.2688-8521002
- Huang, W., Yan, M., Duan, H., Bi, Y., Cheng, X., & Yu, H. (2020). Synergistic antifungal activity of green synthesized silver nanoparticles and epoxiconazole against Setosphaeria turcica. *Journal* of Nanomaterials, 2020, 9535432. https://doi.org/10.1155/2020/9535432
- Lara, H. H., Ixtepan-Turrent, L., Garza-Treviño, E. N., & Rodriguez-Padilla, C. (2010). PVPcoated silver nanoparticles block the transmission of cell-free and cell-associated HIV-1 in human cervical culture. *Journal Nanobiotechnology*, 8(1), 1–11. https://doi.org/10.1186/1477-3155-8-15
- Sun, L., Singh, A. K., Vig, K., Pillai, S. R., & Singh, S. R. (2008). Silver nanoparticles inhibit replication of respiratory syncytial virus. *Journal of Biomedical Nanotechnology*, 4(2), 149–158. https://doi.org/10.1166/jbn.2008.012
- Baram-Pinto, D., Shukla, S., Gedanken, A., & Sarid, R. (2010). Inhibition of HSV-1 attachment, entry, and cell-to-cell spread by functionalized multivalent gold nanoparticles. *Small*, 6(9), 1044– 1050. https://doi.org/10.1002/smll.200902384
- Papp, I., Sieben, C., Ludwig, K., Roskamp, M., Böttcher, C., Schlecht, S., et al. (2010). Inhibition of influenza virus infection by multivalent sialic-acid-functionalized gold nanoparticles. *Small*, 6(24), 2900–2906. https://doi.org/10.1002/smll.201001349
- Rogers, J. V., Parkinson, C. V., Choi, Y. W., Speshock, J. L., & Hussain, S. M. (2008). A preliminary assessment of silver nanoparticle inhibition of monkeypox virus plaque formation. *Nanoscale Research Letters*, 3(4), 129–133. https://doi.org/10.1007/s11671-008-9128-2
- Lara, H. H., Ayala-Nuñez, N. V., Ixtepan-Turrent, L., & Rodriguez-Padilla, C. (2010). Mode of antiviral action of silver nanoparticles against HIV-1. *Journal Nanobiotechnology*, 8(1), 1–10. https://doi.org/10.1186/1477-3155-8-1
- Speshock, J. L., Murdock, R. C., Braydich-Stolle, L. K., Schrand, A. M., & Hussain, S. M. (2010). Interaction of silver nanoparticles with Tacaribe virus. *Journal Nanobiotechnology.*, 8(1), 1–9. https://doi.org/10.1186/1477-3155-8-19
- Saadh, M. J., Aggag, M. M., Alboghdadly, A., Kharshid, A. M., Aldalaen, S. M., & Abdelrazek, M. A. (2021). Silver nanoparticles with epigallocatechingallate and zinc sulphate significantly inhibits avian influenza A virus H9N2. *Microbial Pathogenesis*, 158, 105071. https://doi.org/10. 1016/j.micpath.2021.105071
- Haggag, E. G., Elshamy, A. M., Rabeh, M. A., Gabr, N. M., Salem, M., Youssif, K. A., et al. (2019). Antiviral potential of green synthesized silver nanoparticles of Lampranthus coccineus and Malephora lutea. *Biomolecules*, 14, 6217. https://doi.org/10.2147/IJN.S214171

- Saad, A. H. A., Soliman, M. I., Azzam, A. M., & Mostafa, A. B. (2015). Antiparasitic activity of silver and copper oxide nanoparticles against Entamoeba histolytica and Cryptosporidium parvum cysts. *Journal of the Egyptian Society of Parasitology*, 45(3), 593–602. https://doi.org/10.12816/0017920
- Allahverdiyev, A. M., Abamor, E. S., Bagirova, M., Ustundag, C. B., Kaya, C., Kaya, F., et al. (2011). Antileishmanial effect of silver nanoparticles and their enhanced antiparasitic activity under ultraviolet light. *International Journal of Nanomedicine*, 6, 2705. https://doi.org/10.2147/IJN.S23883
- Silva Viana, R. L., Pereira Fidelis, G., Jane Campos Medeiros, M., Antonio Morgano, M., Chagas Faustino Alves, M. G., Domingues Passero, L. F., et al. (2020). Green synthesis of antileishmanial and antifungal silver nanoparticles using corn cob xylan as a reducing and stabilizing agent. *Biomolecules*, 10(9), 1235. https://doi.org/10.3390/biom10091235
- Zahra'a, A. A., Mustafa, T. A., Ardalan, N. M., & Idan, E. M. (2017). In vitro toxicity evaluation of silver nanoparticles on Entamoeba histolytica trophozoite. *Baghdad Science Journal*, 14(3), 509–515.
- Boateng, J., & Catanzano, O. (2020). Silver and silver nanoparticle-based antimicrobial dressings. *Therapeutic dressings and wound healing applications*, 2020, 157–184. https://doi.org/10.1002/ 9781119433316.ch8
- Winter, C. A., Risley, E. A., & Nuss, G. W. (1962). Carrageenin-induced edema in hind paw of the rat as an assay for antiinflammatory drugs. *Proceedings of the Society for Experimental Biology and Medicine*, 111(3), 544–547. https://doi.org/10.3181/00379727-111-27849
- Shehensha, S., & Jyothi, M. V. (2020). Anti-inflammatory Activity of Nigella sativa oil Mediated Silver Nanoparticles. *Pharmacognosy Magazine*, 12(5), 1086–1092. https://doi.org/10.5530/pj.2020.12. 153
- Kedi, P. B. E., Meva, F. E. A., Kotsedi, L., Nguemfo, E. L., Zangueu, C. B., Ntoumba, A. A., et al. (2018). Eco-friendly synthesis, characterization, in vitro and in vivo anti-inflammatory activity of silver nanoparticle-mediated Selaginella myosurus aqueous extract. *International Journal of Nanomedicine*, 13, 8537. https://doi.org/10.2147/IJN.S174530
- Devaraja, S., Nagaraju, S., Mahadeswaraswamy, Y., Girish, K., & Kemparaju, K. (2008). A low molecular weight serine protease: Purification and characterization from Hippasa agelenoides (funnel web) spider venom gland extract. *Toxicon*, 52(1), 130–138. https://doi.org/10.1016/j.toxicon.2008.04. 168
- Poon, M., & d'Oiron, R. (2000). Recombinant activated factor VII (NovoSeven®) treatment of platelet-related bleeding disorders. *Blood Coagulation & Fibrinolysis*, 11, S55–S68.
- Kenawy, H. I., Boral, I., & Bevington, A. (2015). Complement-coagulation cross-talk: a potential mediator of the physiological activation of complement by low pH. *Frontiers in Immunology*, *6*, 215. https://doi.org/10.3389/fimmu.2015.00215
- Huang, Y., Li, X., Liao, Z., Zhang, G., Liu, Q., Tang, J., et al. (2007). A randomized comparative trial between Acticoat and SD-Ag in the treatment of residual burn wounds, including safety analysis. *Burns*, 33(2), 161–166. https://doi.org/10.1016/j.burns.2006.06.020
- Chen, J., Han, C., Lin, X., Tang, Z., & Su, S. (2006). Effect of silver nanoparticle dressing on second degree burn wound. *Chinese Journal of Surgery*, 44(1), 50–52.
- Chandika, P., Ko, S. C., & Jung, W. K. (2015). Marine-derived biological macromolecule-based biomaterials for wound healing and skin tissue regeneration. *International Journal of Biological Macro*molecules, 77, 24–35. https://doi.org/10.1016/j.ijbiomac.2015.02.050
- Hasatsri, S., Yamdech, R., Chanvorachote, P., & Aramwit, P. (2015). Physical and biological assessments of the innovative bilayered wound dressing made of silk and gelatin for clinical applications. *Journal of Biomaterials Applications*, 29(9), 1304–1313. https://doi.org/10.1177/0885328214559138
- Dai, T., Tanaka, M., Huang, Y. Y., & Hamblin, M. R. (2011). Chitosan preparations for wounds and burns: antimicrobial and wound-healing effects. *Expert Review of Anti-Infective Therapy*, 9(7), 857– 879. https://doi.org/10.1586/eri.11.59
- 73. Burgess, R. (2012). Understanding nanomedicine: an introductory textbook. CRC Press.
- Chen, H., Lan, G., Ran, L., Xiao, Y., Yu, K., Lu, B., et al. (2018). A novel wound dressing based on a Konjac glucomannan/silver nanoparticle composite sponge effectively kills bacteria and accelerates wound healing. *Carbohydrate Polymers*, 183, 70–80. https://doi.org/10.1016/j.carbpol.2017.11.029
- Mermel, L. A. (2000). Prevention of intravascular catheter–related infections. Annals of Internal Medicine, 132(5), 391–402. https://doi.org/10.7326/0003-4819-132-5-200003070-00009
- Sheng, W. H., Ko, W. J., Wang, J. T., Chang, S. C., Hsueh, P. R., & Luh, K. T. (2000). Evaluation of antiseptic-impregnated central venous catheters for prevention of catheter-related infection in intensive care unit patients. *Diagnostic Microbiology and Infectious Disease*, 38(1), 1–5. https://doi.org/ 10.1016/s0732-8893(00)00166-8
- 77. Khare, M. D., Bukhari, S. S., Swann, A., Spiers, P., McLaren, I., & Myers, J. (2007). Reduction of catheter-related colonisation by the use of a silver zeolite-impregnated central vascular catheter in

adult critical care. The Journal of Infection, 54(2), 146–150. https://doi.org/10.1016/j.jinf.2006.03. 002

- Neethu, S., Midhun, S. J., Radhakrishnan, E., & Jyothis, M. (2020). Surface functionalization of central venous catheter with mycofabricated silver nanoparticles and its antibiofilm activity on multidrug resistant *Acinetobacter baumannii*. *Microbial Pathogenesis*, 138, 103832. https://doi.org/10.1016/j. micpath.2019.103832
- Fichtner, J., Güresir, E., Seifert, V., & Raabe, A. (2010). Efficacy of silver-bearing external ventricular drainage catheters: a retrospective analysis. *Journal of Neurosurgery*, 112(4), 840–846. https://doi. org/10.3171/2009.8.JNS091297
- Morley, K., Webb, P., Tokareva, N., Krasnov, A., Popov, V., Zhang, J., et al. (2007). Synthesis and characterisation of advanced UHMWPE/silver nanocomposites for biomedical applications. *European Polymer Journal*, 43(2), 307–314. https://doi.org/10.1016/j.eurpolymj.2006.10.011
- Zhao, Y., Xing, Q., Janjanam, J., He, K., Long, F., Low, K. B., et al. (2014). Facile electrochemical synthesis of antimicrobial TiO2 nanotube arrays. *International Journal of Nanomedicine*, 9, 5177. https://doi.org/10.2147/IJN.S65386
- Castiglioni, S., Cazzaniga, A., Locatelli, L., & Maier, J. A. (2017). Silver nanoparticles in orthopedic applications: New insights on their effects on osteogenic cells. *Nanomaterials*, 7(6), 124. https://doi. org/10.3390/nano7060124
- Cobb, W. S., Harris, J. B., Lokey, J. S., McGill, E. S., & Klove, K. L. (2003). Incisional herniorrhaphy with intraperitoneal composite mesh: a report of 95 cases. *The American Surgeon*, 69(9), 784–787.
- Read, R. C. (2004). Milestones in the history of hernia surgery: prosthetic repair. *Hernia*, 8(1), 8–14. https://doi.org/10.1007/s10029-003-0169-2
- Beard, J. H., Ohene-Yeboah, M., de Vries, C. R., & Schecter, W. (2015). Hernia and hydrocele. In H. T. Debas, P. Donkor, A. Gawande, D. T. Jamison, M. E. Kruk, & C. N. Mock (Eds.), *Essential Surgery: Disease Control Priorities* (Vol. 1, 3rd ed., pp. 151–171). The International Bank for Reconstruction and Development / The World Bank. https://doi.org/10.1596/978-1-4648-0346-8\_ch9
- Samuel, U., & Guggenbichler, J. (2004). Prevention of catheter-related infections: the potential of a new nano-silver impregnated catheter. *International Journal of Antimicrobial Agents*, 23, 75–78. https://doi.org/10.1016/j.ijantimicag.2003.12.004
- Klasen, H. (2000). Historical review of the use of silver in the treatment of burns. I. Early uses. *Burns*, 26(2), 117–130. https://doi.org/10.1016/s0305-4179(99)00108-4
- Dunn, K., & Edwards-Jones, V. (2004). The role of Acticoat<sup>™</sup> with nanocrystalline silver in the management of burns. *Burns*, 30, S1–S9. https://doi.org/10.1016/s0305-4179(04)90000-9
- Darouiche, R. O. (2004). Treatment of infections associated with surgical implants. *The New England Journal of Medicine*, 350(14), 1422–1429. https://doi.org/10.1056/NEJMra035415
- Cohen, M. S., Stern, J. M., Vanni, A. J., Kelley, R. S., Baumgart, E., Field, D., et al. (2007). In vitro analysis of a nanocrystalline silver-coated surgical mesh. *Surgical Infections*, 8(3), 397–404. https:// doi.org/10.1089/sur.2006.032
- Bhol, K., & Schechter, P. (2005). Topical nanocrystalline silver cream suppresses inflammatory cytokines and induces apoptosis of inflammatory cells in a murine model of allergic contact dermatitis. *The British Journal of Dermatology*, 152(6), 1235–1242. https://doi.org/10.1111/j.1365-2133. 2005.06575.x
- Zhang, Y., Zheng, Y., Li, Y., Wang, L., Bai, Y., Zhao, Q., et al. (2015). Tantalum nitride-decorated titanium with enhanced resistance to microbiologically induced corrosion and mechanical property for dental application. *PLoS One*, *10*(6), e0130774. https://doi.org/10.1371/journal.pone.0130774
- Porenczuk, A., Grzeczkowicz, A., Maciejewska, I., Gołaś, M., Piskorska, K., Kolenda, A., et al. (2019). An initial evaluation of cytotoxicity, genotoxicity and antibacterial effectiveness of a disinfection liquid containing silver nanoparticles alone and combined with a glass-ionomer cement and dentin bonding systems. *Advances in Clinical and Experimental Medicine*, 28(1), 75–83. https://doi. org/10.17219/acem/76160
- Fernandez, C. C., Sokolonski, A. R., Fonseca, M. S., Stanisic, D., Araújo, D. B., Azevedo, V., et al. (2021). Applications of Silver Nanoparticles in Dentistry: Advances and Technological Innovation. *International Journal of Molecular Sciences*, 22(5), 2485. https://doi.org/10.3390/ijms22052485
- Monteiro, D. R., Gorup, L. F., Takamiya, A. S., de Camargo, E. R., Filho, A. C. R., & Barbosa, D. B. (2012). Silver distribution and release from an antimicrobial denture base resin containing silver colloidal nanoparticles. *Journal of Prosthodontics*, 21(1), 7–15. https://doi.org/10.1111/j.1532-849X. 2011.00772.x
- Chand, K., Abro, M. I., Aftab, U., Shah, A. H., Lakhan, M. N., Cao, D., et al. (2019). Green synthesis characterization and antimicrobial activity against Staphylococcus aureus of silver nanoparticles

using extracts of neem, onion and tomato. RSC Advances, 9(30), 17002–17015. https://doi.org/10. 1039/C9RA01407A

- Pérez-Díaz, M. A., Boegli, L., James, G., Velasquillo, C., Sanchez-Sanchez, R., Martinez-Martinez, R. E., et al. (2015). Silver nanoparticles with antimicrobial activities against Streptococcus mutans and their cytotoxic effect. *Materials Science and Engineering: C*, 55, 360–366. https://doi.org/10. 1016/j.msec.2015.05.036
- Besinis, A., Hadi, S. D., Le, H., Tredwin, C., & Handy, R. (2017). Antibacterial activity and biofilm inhibition by surface modified titanium alloy medical implants following application of silver, titanium dioxide and hydroxyapatite nanocoatings. *Nanotoxicology*, 11(3), 327–338. https://doi.org/10. 1080/17435390.2017.1299890
- Nozari, A., Ajami, S., Rafiei, A., & Niazi, E. (2017). Impact of nano hydroxyapatite, nano silver fluoride and sodium fluoride varnish on primary teeth enamel remineralization: an in vitro study. *Journal* of Clinical and Diagnostic, 11(9), ZC97. https://doi.org/10.7860/JCDR/2017/30108.10694
- Venugopal, A., Muthuchamy, N., Tejani, H., Gopalan, A. I., Lee, K. P., Lee, H. J., et al. (2017). Incorporation of silver nanoparticles on the surface of orthodontic microimplants to achieve antimicrobial properties. *The Korean Journal of Orthodontics*, 47(1), 3–10. https://doi.org/10.4041/kjod.2017. 47.1.3
- Hollstein, M., Alexandrov, L., Wild, C., Ardin, M., & Zavadil, J. (2017). Base changes in tumour DNA have the power to reveal the causes and evolution of cancer. *Oncogene*, 36(2), 158–167. https:// doi.org/10.1038/onc.2016.192
- Manzoor, M., Khan, A. H. A., Ullah, R., Khan, M. Z., & Ahmad, I. (2016). Environmental epidemiology of cancer in South Asian population: risk assessment against exposure to polycyclic aromatic hydrocarbons and volatile organic compounds. *Arabian Journal for Science and Engineering*, 41(6), 2031–2043. https://doi.org/10.1007/s13369-016-2139-x
- Anand, P., Kunnumakara, A. B., Sundaram, C., Harikumar, K. B., Tharakan, S. T., Lai, O. S., et al. (2008). Cancer is a preventable disease that requires major lifestyle changes. *Pharm.Res.*, 25(9), 2097–2116. https://doi.org/10.1007/s11095-008-9661-9
- Ghosh, R., & Girigoswami, K. (2008). NADH dehydrogenase subunits are overexpressed in cells exposed repeatedly to H2O2. *Mutation Research, Fundamental and Molecular Mechanisms of Mutagenesis*, 638(1-2), 210–215. https://doi.org/10.1016/j.mrfmmm.2007.08.008
- Asharani, P., Hande, M. P., & Valiyaveettil, S. (2009). Anti-proliferative activity of silver nanoparticles. *BMC Cell Biology*, 10(1), 1–14. https://doi.org/10.1186/1471-2121-10-65
- Greulich, C., Diendorf, J., Simon, T., Eggeler, G., Epple, M., & Köller, M. (2011). Uptake and intracellular distribution of silver nanoparticles in human mesenchymal stem cells. *Acta Biomaterialia*, 7(1), 347–354. https://doi.org/10.1016/j.actbio.2010.08.003
- Kim, S., Choi, J. E., Choi, J., Chung, K. H., Park, K., Yi, J., et al. (2009). Oxidative stress-dependent toxicity of silver nanoparticles in human hepatoma cells. *Toxicology In Vitro*, 23(6), 1076–1084.
- Rosas-Hernández, H., Jiménez-Badillo, S., Martínez-Cuevas, P. P., Gracia-Espino, E., Terrones, H., Terrones, M., et al. (2009). Effects of 45-nm silver nanoparticles on coronary endothelial cells and isolated rat aortic rings. *Toxicology Letters*, 191(2-3), 305–313. https://doi.org/10.1016/j.toxlet.2009. 09.014
- 109. Hsin, Y. H., Chen, C. F., Huang, S., Shih, T. S., Lai, P. S., & Chueh, P. J. (2008). The apoptotic effect of nanosilver is mediated by a ROS-and JNK-dependent mechanism involving the mitochondrial pathway in NIH3T3 cells. *Toxicology Letters*, 179(3), 130–139. https://doi.org/10.1016/j.toxlet.2008.04. 015
- Sanpui, P., Chattopadhyay, A., & Ghosh, S. S. (2011). Induction of apoptosis in cancer cells at low silver nanoparticle concentrations using chitosan nanocarrier. ACS Applied Materials & Interfaces, 3(2), 218–228. https://doi.org/10.1021/am100840c
- 111. Ahamed, M., Karns, M., Goodson, M., Rowe, J., Hussain, S. M., Schlager, J. J., et al. (2008). DNA damage response to different surface chemistry of silver nanoparticles in mammalian cells. *Toxicology and Applied Pharmacology*, 33(3), 404–410. https://doi.org/10.1016/j.taap.2008.09.015
- 112. Sukirtha, R., Priyanka, K. M., Antony, J. J., Kamalakkannan, S., Thangam, R., Gunasekaran, P., et al. (2012). Cytotoxic effect of Green synthesized silver nanoparticles using Melia azedarach against in vitro HeLa cell lines and lymphoma mice model. *Process Biochemistry*, 47(2), 273–279. https:// doi.org/10.1016/j.procbio.2011.11.003
- 113. Kalishwaralal, K., Banumathi, E., Pandian, S. R. K., Deepak, V., Muniyandi, J., Eom, S. H., et al. (2009). Silver nanoparticles inhibit VEGF induced cell proliferation and migration in bovine retinal endothelial cells. *Colloids and Surfaces, B: Biointerfaces, 73(1)*, 51–57. https://doi.org/10.1016/j. colsurfb.2009.04.025

- Mukherjee, S., Chowdhury, D., Kotcherlakota, R., & Patra, S. (2014). Potential theranostics application of bio-synthesized silver nanoparticles (4-in-1 system). *Theranostics.*, 4(3), 316. https://doi. org/10.7150/thno.7819
- Yesilot, S., & Aydin, C. (2019). Silver nanoparticles; a new hope in cancer therapy? *Eastern Journal of Medicine*, 24(1), 111–116. https://doi.org/10.5505/ejm.2019.66487
- 116. Gurunathan, S., & Raman, J. Abd Malek, S.N.; John, P.A.; Vikineswary, S. (2013). Green synthesis of silver nanoparticles using Ganoderma neo-japonicum Imazeki: a potential cytotoxic agent against breast cancer cells *Int. Journal Nanomedicine*, 8, 4399. https://doi.org/10.2147/IJN. S51881
- 117. Gurunathan, S., Han, J. W., Dayem, A. A., Eppakayala, V., Park, J. H., Cho, S. G., et al. (2013). Green synthesis of anisotropic silver nanoparticles and its potential cytotoxicity in human breast cancer cells (MCF-7). *Journal of Industrial and Engineering Chemistry*, 19(5), 1600–1605. https://doi.org/10.3390/ijms17101603
- Nazir, S., Hussain, T., de Iqbal, M. M. K., Muazzam, A., & Ismail, J. M. (2011). Novel and costeffective green synthesis of silver nano particles and their in vivo antitumor properties against human cancer cell lines. *BioScience Technologies*, 2(6), 425–430.
- 119. Saratale, R. G., Benelli, G., Kumar, G., Kim, D. S., & Saratale, G. D. (2018). Bio-fabrication of silver nanoparticles using the leaf extract of an ancient herbal medicine, dandelion (Taraxacum officinale), evaluation of their antioxidant, anticancer potential, and antimicrobial activity against phytopathogens. *Environmental Science and Pollution Research*, 25(11), 10392–10406. https:// doi.org/10.1007/s11356-017-9581-5
- El-Naggar, N. E. A., Hussein, M. H., & El-Sawah, A. A. (2017). Bio-fabrication of silver nanoparticles by phycocyanin, characterization, in vitro anticancer activity against breast cancer cell line and in vivo cytotxicity. *Scientific Reports*, 7(1), 1–20. https://doi.org/10.1038/s41598-017-11121-3
- 121. Govindaraju, K., Krishnamoorthy, K., Alsagaby, S. A., Singaravelu, G., & Premanathan, M. (2015). Green synthesis of silver nanoparticles for selective toxicity towards cancer cells. *IET Nanobiotechnology*, 9(6), 325–330. https://doi.org/10.1049/iet-nbt.2015.0001
- 122. Jacob, S. J. P., Finub, J., & Narayanan, A. (2012). Synthesis of silver nanoparticles using Piper longum leaf extracts and its cytotoxic activity against Hep-2 cell line. *Colloids and Surfaces, B: Biointerfaces, 91,* 212–214. https://doi.org/10.1016/j.colsurfb.2011.11.001
- 123. Nagajyothi, P., Sreekanth, T., Lee, J. I., & Lee, K. D. (2014). Mycosynthesis: antibacterial, antioxidant and antiproliferative activities of silver nanoparticles synthesized from Inonotus obliquus (Chaga mushroom) extract. *Journal of Photochemistry and Photobiology A*, 130, 299–304. https:// doi.org/10.1016/j.jphotobiol.2013.11.022
- 124. Kuppusamy, P., Ichwan, S. J., Al-Zikri, P. N. H., Suriyah, W. H., Soundharrajan, I., Govindan, N., et al. (2016). In vitro anticancer activity of Au, Ag nanoparticles synthesized using Commelina nudiflora L. aqueous extract against HCT-116 colon cancer cells. *Biological Trace Element Research*, 173(2), 297–305. https://doi.org/10.1007/s12011-016-0666-7
- 125. Kathiravan, V., Ravi, S., & Ashokkumar, S. (2014). Synthesis of silver nanoparticles from Melia dubia leaf extract and their in vitro anticancer activity. *Spectrochimica Acta. Part A, Molecular* and Biomolecular Spectroscopy, 130, 116–121. https://doi.org/10.1016/j.saa.2014.03.10
- 126. He, Y., Du, Z., Ma, S., Liu, Y., Li, D., Huang, H., et al. (2016). Effects of green-synthesized silver nanoparticles on lung cancer cells in vitro and grown as xenograft tumors in vivo. *International Journal of Nanomedicine*, 11, 1879. https://doi.org/10.2147/IJN.S103695
- 127. El-Sheikh, S. M., Edrees, N., Hend, E. S., Khamis, T., Arisha, A. H., Metwally, M. M., et al. (2021, 1-9). Could Cisplatin Loading on Biosynthesized Silver Nanoparticles Improve Its Therapeutic Efficacy on Human Prostate Cancer Cell Line and Reduce Its In Vivo Nephrotoxic Effects? *Biological Trace Element Research*. https://doi.org/10.1007/s12011-021-02677-3
- Sre, P. R., Reka, M., Poovazhagi, R., Kumar, M. A., & Murugesan, K. (2015). Antibacterial and cytotoxic effect of biologically synthesized silver nanoparticles using aqueous root extract of *Erythrina indica* lam. Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy, 135, 1137–1144. https://doi.org/10.1016/j.saa.2014.08.019
- Krishnaraj, C., Muthukumaran, P., Ramachandran, R., Balakumaran, M., & Kalaichelvan, P. (2014). Acalypha indica Linn: biogenic synthesis of silver and gold nanoparticles and their cyto-toxic effects against MDA-MB-231, human breast cancer cells. *Biotechnology Reports*, *4*, 42–49. https://doi.org/10.1016/j.btre.2014.08.002
- Sathishkumar, G., Gobinath, C., Wilson, A., & Sivaramakrishnan, S. (2014). Dendrophthoe falcata (Lf) Ettingsh (Neem mistletoe): A potent bioresource to fabricate silver nanoparticles for anticancer effect against human breast cancer cells (MCF-7). Spectrochimica Acta Part A: Molecular and Biomolecular, 128, 285–290. https://doi.org/10.1016/j.saa.2014.02.096

- 131. Gajendran, B., Chinnasamy, A., Durai, P., Raman, J., & Ramar, M. (2014). Biosynthesis and characterization of silver nanoparticles from Datura inoxia and its apoptotic effect on human breast cancer cell line MCF7. *Materials Letters*, 122, 98–102. https://doi.org/10.1016/j.matlet.2014.02.003
- 132. Yakop, F., Abd Ghafar, S. A., Yong, Y. K., Saiful Yazan, L., Mohamad, H. R., Lim, V., et al. (2018). Silver nanoparticles Clinacanthus Nutans leaves extract induced apoptosis towards oral squamous cell carcinoma cell lines. *Artificial Cells, Nanomedicine, and Biotechnology*, 46(sup2), 131–139. https:// doi.org/10.1080/21691401.2018.1452750
- 133. AlSalhi, M. S., Devanesan, S., Alfuraydi, A. A., Vishnubalaji, R., Munusamy, M. A., Murugan, K., et al. (2016). Green synthesis of silver nanoparticles using Pimpinella anisum seeds: antimicrobial activity and cytotoxicity on human neonatal skin stromal cells and colon cancer cells. *International Journal of Nanomedicine*, 11, 4439. https://doi.org/10.2147/IJN.S113193
- Oh, K. H., Soshnikova, V., Markus, J., Kim, Y. J., Lee, S. C., Singh, P., et al. (2018). Biosynthesized gold and silver nanoparticles by aqueous fruit extract of Chaenomeles sinensis and screening of their biomedical activities. *Artificial Cells, Nanomedicine, and Biotechnology*, 46(3), 599–606. https://doi. org/10.1080/21691401.2017.1332636
- 135. Kaler, A., Jain, S., & Banerjee, U. C. (2013). Green and rapid synthesis of anticancerous silver nanoparticles by Saccharomyces boulardii and insight into mechanism of nanoparticle synthesis. *BioMed Research International*, 2013, 872940. https://doi.org/10.1155/2013/872940
- Vijayan, R., Joseph, S., & Mathew, B. (2018). Indigofera tinctoria leaf extract mediated green synthesis of silver and gold nanoparticles and assessment of their anticancer, antimicrobial, antioxidant and catalytic properties. *Artificial Cells, Nanomedicine, and Biotechnology*, 46(4), 861–871. https://doi.org/10.1080/21691401.2017.1345930
- Foldbjerg, R., Dang, D. A., & Autrup, H. (2011). Cytotoxicity and genotoxicity of silver nanoparticles in the human lung cancer cell line, A549. Archives of Toxicology, 85(7), 743–750. https://doi.org/ 10.1007/s00204-010-0545-5
- Rosarin, F. S., Arulmozhi, V., Nagarajan, S., & Mirunalini, S. (2013). Antiproliferative effect of silver nanoparticles synthesized using amla on Hep2 cell line. *Asian Pacific Journal of Tropical Medicine*, 6(1), 1–10. https://doi.org/10.1016/S1995-7645(12)60193-X
- Selvi, B. C. G., Madhavan, J., & Santhanam, A. (2016). Cytotoxic effect of silver nanoparticles synthesized from Padina tetrastromatica on breast cancer cell line. *Advances in Natural Sciences: Nano*science and Nanotechnology, 7(3), 035015. https://doi.org/10.1088/2043-6262/7/3/035015
- 140. Prabhu, D., Arulvasu, C., Babu, G., Manikandan, R., & Srinivasan, P. (2013). Biologically synthesized green silver nanoparticles from leaf extract of Vitex negundo L. induce growth-inhibitory effect on human colon cancer cell line HCT15. *Process Biochemistry*, 48(2), 317–324. https://doi.org/10. 1016/j.procbio.2012.12.013
- 141. Mousavi, B., Tafvizi, F., & Zaker, B. S. (2018). Green synthesis of silver nanoparticles using Artemisia turcomanica leaf extract and the study of anti-cancer effect and apoptosis induction on gastric cancer cell line (AGS). Artificial Cells, Nanomedicine and Biotechnology, 46(sup1), 499–510. https://doi.org/10.1080/21691401.2018.1430697
- Sriram, M. I., Kanth, S. B. M., Kalishwaralal, K., & Gurunathan, S. (2010). Antitumor activity of silver nanoparticles in Dalton's lymphoma ascites tumor model. *International Journal of Nanomedicine*, 5, 753. https://doi.org/10.2147/IJN.S11727
- 143. Liu, P., Jin, H., Guo, Z., Ma, J., Zhao, J., Li, D., et al. (2016). Silver nanoparticles outperform gold nanoparticles in radiosensitizing U251 cells in vitro and in an intracranial mouse model of glioma. *International Journal of Nanomedicine*, 11, 5003. https://doi.org/10.2147/IJN.S115473
- 144. Lara-González, J. H., Gomez-Flores, R., Tamez-Guerra, P., Monreal-Cuevas, E., Tamez-Guerra, R., & Rodríguez-Padilla, C. (2013). In vivo antitumor activity of metal silver and silver nanoparticles in the L5178Y-R murine lymphoma model. *Journal of Advances in Medicine*, 1308–1316. https://doi. org/10.9734/BJMMR/2013/3108
- 145. Wang, X., Li, T., Su, X., Li, J., Li, W., Gan, J., et al. (2019). Genotoxic effects of silver nanoparticles with/without coating in human liver HepG2 cells and in mice. *Journal of Applied Toxicology*, 39(6), 908–918. https://doi.org/10.1002/jat.3779
- 146. Murugesan, K., Koroth, J., Srinivasan, P. P., Singh, A., Mukundan, S., Karki, S. S., et al. (2019). Effects of green synthesised silver nanoparticles (ST06-AgNPs) using curcumin derivative (ST06) on human cervical cancer cells (HeLa) in vitro and EAC tumor bearing mice models. *International Journal of Nanomedicine*, 14, 5257. https://doi.org/10.2147/IJN.S202404
- 147. Ramachandran, R., Krishnaraj, C., Sivakumar, A. S., Prasannakumar, P., Kumar, V. A., Shim, K. S., et al. (2017). Anticancer activity of biologically synthesized silver and gold nanoparticles on mouse myoblast cancer cells and their toxicity against embryonic zebrafish. *Materials Science and Engineering: C*, 73, 674–683. https://doi.org/10.1016/j.msec.2016.12.110

- Kanipandian, N., Kannan, S., Ramesh, R., Subramanian, P., & Thirumurugan, R. (2014). Characterization, antioxidant and cytotoxicity evaluation of green synthesized silver nanoparticles using Cleistanthus collinus extract as surface modifier. *Materials Research Bulletin, 49*, 494–502. https://doi.org/10.1016/j. materresbull.2013.09.016
- Hashemi, S. F., Tasharrofi, N., & Saber, M. M. (2020). Green synthesis of silver nanoparticles using Teucrium polium leaf extract and assessment of their antitumor effects against MNK45 human gastric cancer cell line. *Journal of Molecular Structure*, 1208, 127889. https://doi.org/10.1016/j.molstruc.2020.127889
- Gengan, R., Anand, K., Phulukdaree, A., & Chuturgoon, A. (2013). A549 lung cell line activity of biosynthesized silver nanoparticles using Albizia adianthifolia leaf. *Colloids and Surfaces. B, Biointerfaces*, 105, 87–91. https://doi.org/10.1016/j.colsurfb.2012.12.044
- 151. Annu, M., Ahmed, S., Kaur, G., Sharma, P., Singh, S., & Ikram, S. (2018). Evaluation of the antioxidant, antibacterial and anticancer (lung cancer cell line A549) activity of Punica granatum mediated silver nanoparticles. *Toxicology Research*, 7(5), 923–930. https://doi.org/10.1039/c8tx00103k
- Khorrami, S., Zarrabi, A., Khaleghi, M., Danaei, M., & Mozafari, M. (2018). Selective cytotoxicity of green synthesized silver nanoparticles against the MCF-7 tumor cell line and their enhanced antioxidant and antimicrobial properties. *International Journal of Nanomedicine*, 13, 8013. https://doi.org/10.2147/ IJN.S189295
- 153. Chengzheng, W., Jiazhi, W., Shuangjiang, C., Swamy, M. K., Sinniah, U. R., Akhtar, M., et al. (2018). Biogenic synthesis, characterization and evaluation of silver nanoparticles from Aspergillus niger JX556221 against human colon cancer cell line HT-29. *Journal of Nanoscience and Nanotechnology*, 18(5), 3673–3681. https://doi.org/10.1166/jnn.2018.15364
- Jha, A. K., & Prasad, K. (2014). Green synthesis of silver nanoparticles and its activity on SiHa cervical cancer cell line. Advanced Materials Letters, 5(12), 501–505. https://doi.org/10.5185/amlett.2014.4563
- 155. Yang, J., Wang, Q., Wang, C., Yang, R., Ahmed, M., Kumaran, S., et al. (2020). Pseudomonas aeruginosa synthesized silver nanoparticles inhibit cell proliferation and induce ROS mediated apoptosis in thyroid cancer cell line (TPC1). Artificial Cells, Nanomedicine, and Biotechnology, 48(1), 800–809. https:// doi.org/10.1080/21691401.2019.1687495
- 156. El Kassas, H. Y., & Attia, A. A. (2014). Bactericidal application and cytotoxic activity of biosynthesized silver nanoparticles with an extract of the red seaweed Pterocladiella capillacea on the HepG 2 cell line. *Asian Pacific Journal of Cancer Prevention*, 15(3), 1299–13306. https://doi.org/10.7314/apjcp.2014. 15.3.1299
- Chen, H., Roco, M. C., Li, X., & Lin, Y. (2008). Trends in nanotechnology patents. *Nature Nanotechnology*, 3(3), 123–125. https://doi.org/10.1038/nnano.2008.51
- Roco, M. C. (2003). Nanotechnology: convergence with modern biology and medicine. *Current Opinion* in Biotechnology, 14(3), 337–346. https://doi.org/10.1016/s0958-1669(03)00068-5
- Xia, J., Yao, J., & Wang, L. V. (2014). Photoacoustic tomography: principles and advances. *Electromagn Waves (Camb)*, 147, 1–22. https://doi.org/10.2528/pier14032303
- Homan, K. A., Shah, J., Gomez, S., Gensler, H., Karpiouk, A. B., Brannon-Peppas, L., et al. (2010). Silver nanosystems for photoacoustic imaging and image-guided therapy. *Journal of Biomedical Optics*, 15(2), 021316. https://doi.org/10.1117/1.3365937
- Liu, Y., Miyoshi, H., & Nakamura, M. (2007). Nanomedicine for drug delivery and imaging: a promising avenue for cancer therapy and diagnosis using targeted functional nanoparticles. *International Journal of Cancer*, 120(12), 2527–2537. https://doi.org/10.1002/ijc.22709
- Yeh, Y. C., Creran, B., & Rotello, V. M. (2012). Gold nanoparticles: preparation, properties, and applications in bionanotechnology. *Nanoscale.*, 4(6), 1871–1880. https://doi.org/10.1039/c1nr11188d
- Yamashita, M. (2021). Auranofin: Past to Present and repurposing. *International Immunopharmacology*, 101(pt B), 108272. https://doi.org/10.1016/j.intimp.2021.108272
- 164. Zhao, Y., He, Z., Wang, R., Cai, P., Zhang, X., Yuan, Q., Zhang, J., Gao, F., & Gao, X. (2019). Comparison of the therapeutic effects of gold nanoclusters and gold nanoparticles on rheumatoid arthritis. *Journal of Biomedical Nanotechnology*, 15(11), 2281–2290. https://doi.org/10.1166/jbn.2019.2848
- Dwivedi, P., Nayak, V., & Kowshik, M. (2015). Role of gold nanoparticles as drug delivery vehicles for chondroitin sulfate in the treatment of osteoarthritis. *Biotechnology Progress*, 31(5), 1416–1422. https:// doi.org/10.1002/btpr.2147
- Dykman, L., & Khlebtsov, N. (2011). Gold nanoparticles in biology and medicine: recent advances and prospects. Acta Naturae, 3(2), 34–55.
- 167. Vecchio, G., Galeone, A., Brunetti, V., Maiorano, G., Rizzello, L., Sabella, S., et al. (2012). Mutagenic effects of gold nanoparticles induce aberrant phenotypes in Drosophila melanogaster. *Nanomedicine: Nanotechnology, Biology and Medicine*, 8(1), 1–7. https://doi.org/10.1016/j.nano.2011.11.001
- 168. Tan, E., Yin, P., Lang, X., Wang, X., You, T., & Guo, L. (2012). Functionalized gold nanoparticles as nanosensor for sensitive and selective detection of silver ions and silver nanoparticles by

surface-enhanced Raman scattering. The Analyst, 137(17), 3925–3928. https://doi.org/10.1039/c2an3 5670

- Akhtar, S., Asiri, S. M., Khan, F. A., Gunday, S. T., Iqbal, A., Alrushaid, N., Labib, O. A., Deen, G. R., & Henari, F. Z. (2022). Formulation of gold nanoparticles with hibiscus and curcumin extracts induced anti-cancer activity. *Arabian Journal of Chemistry*, 15(2), 103594. https://doi.org/10.1016/j.arabjc.2021. 103594
- Shinde, A. S., & Mendhulkar, V. D. (2022). Anticancer activity of gold nanobioconjugates synthesized from Elephantopus scaber (linn.) leaf extract. *Journal Canadian Research*. https://doi.org/10.4103/jcrt. JCRT\_1043\_20
- 171. Zhang, X. D., Wu, H. Y., Wu, D., Wang, Y. Y., Chang, J. H., Zhai, Z. B., Meng, A. M., et al. (2010). Toxicologic effects of gold nanoparticles in vivo by different administration routes *Int. Journal of Nanomedicine*, 5, 771. https://doi.org/10.2147/IJN.S8428
- Guo, J., Rahme, K., He, Y., Li, L. L., Holmes, J. D., & O'Driscoll, C. M. (2017). Gold nanoparticles enlighten the future of cancer theranostics. *International Journal of Nanomedicine*, 12, 6131. https://doi. org/10.2147/IJN.S8428
- 173. Sokolov, K., Follen, M., Aaron, J., Pavlova, I., Malpica, A., Lotan, R., et al. (2003). Real-time vital optical imaging of precancer using anti-epidermal growth factor receptor antibodies conjugated to gold nanoparticles. *Cancer Research*, 63(9), 1999–2004.
- Link, S., & El-Sayed, M. A. (2000). Shape and size dependence of radiative, non-radiative and photothermal properties of gold nanocrystals. *International Reviews in Physical Chemistry*, 19(3), 409–453. https://doi.org/10.1080/01442350050034180
- 175. Ayala-Orozco, C., Urban, C., Bishno, S., Urban, A., Charron, H., Mitchell, T., et al. (2014). Sub-100 nm gold nanomatryoshkas improve photo-thermal therapy efficacy in large and highly aggressive triple negative breast tumors. *Journal of Controlled Release*, 191, 90–97. https://doi.org/10.1016/j.jconrel.2014.07. 038
- Vimaladevi, M., Divya, K. C., & Girigoswami, A. (2016). Liposomal nanoformulations of rhodamine for targeted photodynamic inactivation of multidrug resistant gram negative bacteria in sewage treatment plant. *Journal of Photochemistry and Photobiology B: Biology, 162*, 146–152. https://doi.org/10.1016/j. jphotobiol.2016.06.034
- Allison, R. R., & Moghissi, K. (2013). Photodynamic therapy (PDT): PDT mechanisms. *Clinical Endos-copy*, 46(1), 24. https://doi.org/10.5946/ce.2013.46.1.24
- Metkar, S. K., & Girigoswami, K. (2019). Diagnostic biosensors in medicine–a review. *Biocatalysis and Agricultural Biotechnology*, 17, 271–283. https://doi.org/10.1016/j.bcab.2018.11.029
- Wang, J., Drelich, A. J., Hopkins, C. M., Mecozzi, S., Li, L., Kwon, G., et al. (2021). Gold nanoparticles in virus detection: Recent advances and potential considerations for SARS-CoV-2 testing development. *Wiley Interdisciplinary Reviews. Nanomedicine and Nanobiotechnology*, 2021, e1754. https://doi.org/10. 1002/wnan.1754
- WHO. (2020b). Draft landscape of COVID-19 candidate vaccines. World Health Organization. R & D Blue Prints. https://www.who.int/who-documents-detail-redirect/draft-landscape-of-covid-19-candidatevaccines. Accessed July 2020.
- 181. Chen, H. W., Huang, C. Y., Lin, S. Y., Fang, Z. S., Hsu, C. H., Lin, J. C., Chen, Y. I., Yao, B. Y., & Hu, C. M. (2016). Synthetic virus-like particles prepared via protein corona formation enable effective vaccination in an avian model of coronavirus infection. *Biomaterials, 106*, 111–118. https://doi.org/10.1016/j. biomaterials.2016.08.018
- Alphandéry, E. (2020). The potential of various nanotechnologies for coronavirus diagnosis/treatment highlighted through a literature analysis. *Bioconjugate Chemistry*, 31(8), 1873–1882. https://doi.org/10. 1021/acs.bioconjchem.0c00287
- Lopez-Chaves, C., Soto-Alvaredo, J., Montes-Bayon, M., Bettmer, J., Llopis, J., & Sanchez-Gonzalez, C. (2008). Gold nanoparticles: distribution, bioaccumulation and toxicity. In vitro and in vivo studies. *Nanomed.:Nanotechnol.Biol.Med*, 14(1), 1–2. https://doi.org/10.1016/j.nano.2017.08.011
- Li, J. J., Zou, L. I., Hartono, D., Ong, C. N., Bay, B. H., & Lanry Yung, L. Y. (2008). Gold nanoparticles induce oxidative damage in lung fibroblasts in vitro. *Advanced Materials*, 20(1), 138–142. https://doi.org/ 10.1002/adma.200701853
- Girigoswami, K. (2018). Toxicity of Metal Oxide Nanoparticles. Advances in Experimental Medicine and Biology, 1048, 99–122. https://doi.org/10.1007/978-3-319-72041-8\_7

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