



High-Quality Draft Genome Sequence of *Pseudomonas reidholzensis* Strain CCOS 865^T

Dominik Rutz,^{a,b} David Frasson,^b Martin Sievers,^b Jochen Blom,^c Fabio Rezzonico,^a  Joël F. Pothier,^a Theo H. M. Smits^a

^aEnvironmental Genomics and Systems Biology Research Group, Institute of Natural Resource Sciences, Zurich University of Applied Sciences (ZHAW), Wädenswil, Switzerland

^bMicrobiology and Molecular Biology Research Group, Institute of Chemistry and Biotechnology, Zurich University of Applied Sciences (ZHAW), Wädenswil, Switzerland

^cBioinformatics and Systems Biology, Justus-Liebig-Universität, Giessen, Germany

ABSTRACT We have sequenced and assembled the genome of *Pseudomonas reidholzensis* CCOS 865^T, which was isolated in 2014 from forest soil. Members of the genus *Pseudomonas* play important roles in environmental systems and are utilized in many biotechnological processes. The genome of this species may provide an important resource for the discovery of novel enzyme activities.

Bacteria in the genus *Pseudomonas* are some of the most ecologically important and genetically diverse organisms, and *Pseudomonas* strains can be isolated from a variety of environmental locations and contexts (1). They are involved in degradation, element cycling, and recycling of biogenic and xenobiotic compounds (2). Based on the enzymatic systems involved in the degradation of aromatic compounds originating from lignin biodegradation (3), pseudomonads have been used in catalytic processes for the biosynthesis of novel fine chemicals (4). *Pseudomonas reidholzensis* CCOS 865^T is a newly identified species within the *Pseudomonas putida* group and was isolated in 2014 from forest soil in Switzerland (5). Here, we report the draft genome sequence of the potential new biocatalyst *Pseudomonas reidholzensis* CCOS 865^T.

Genomic DNA of *P. reidholzensis* CCOS 865^T, grown overnight at 28°C in LB medium, was isolated using the NucleoSpin tissue kit (Macherey-Nagel, Düren, Germany) and fragmented using the Covaris E220 ultrasonicator (average target size, 550 bp). Library preparation was performed using the Illumina NeoPrep library system, according to the manufacturer's instructions. Genome sequencing was performed at the Zurich University of Applied Sciences (ZHAW) using 300-bp paired-end reads and a 550-bp insert library on an Illumina MiSeq instrument (5, 6). For the assembly process, a total of 1,731,754 reads were generated. The SeqMan NGen software 12.1.0 (DNASTar, Madison, WI) was used with standard settings for automatic assembly (5). After further assembly, the final draft genome sequence has a total of 45 contigs, with an N_{50} value of 261,911 bp, a length of 6,163,129 bp, and a G+C content of 64.09%. The genome was annotated in GenDB (7), while EDGAR version 2.3 (8) was used for comparative genomics against related pseudomonads.

We confirmed the species delineation against other species of the *P. putida* group by calculating the genome-to-genome distance (GGDC; version 2.1) values (9) and the average nucleotide identities based on BLAST (ANIb) with JSpeciesWS version 3.0.20 (10). An average ANIb of 82.92% ± 0.37% and GGDC of 28.9% ± 0.8% were obtained with *Pseudomonas guariconensis* LMG 27394^T (GenBank accession number [FMX000000000](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/entry.fcgi?accession=FMX000000000)), *Pseudomonas plecoglossicida* NBRC 103162^T (GenBank accession number [BBIV000000000](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/entry.fcgi?accession=BBIV000000000)), and *Pseudomonas* sp. strain GM84 (GenBank accession number [AKJC000000000](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/entry.fcgi?accession=AKJC000000000)). Based on these results, *P. reidholzensis* CCOS 865^T differs from other members of the *P. putida* group at the genome level. The first comparative

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Address correspondence to Joël F. Pothier, joel.pothier@zhaw.ch.

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genome analysis of *P. reidholzensis* CCOS 865^T revealed several gene clusters for the degradation of aromatic compounds, such as genes for mandelic acid, vanillin, or gallic acid (11–13), which are similar to those found in other *Pseudomonas* species.

Data availability. The draft genome sequence of *P. reidholzensis* CCOS 865^T was deposited at DDBJ/EMBL/GenBank under BioProject number [PRJEB28254](https://www.ncbi.nlm.nih.gov/bioproject/PRJEB28254) and the accession number [UNOZ00000000](https://www.ncbi.nlm.nih.gov/nuclseq/UNOZ00000000). The version described in this paper is version UNOZ01000000. Raw sequence reads (Illumina) have been deposited at EMBL under the accession number [ERR2814816](https://www.ncbi.nlm.nih.gov/nuclseq/ERR2814816).

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REFERENCES

- Kiil K, Binnewies TT, Willenbrock H, Hansen SK, Yang L, Jelsbak L, Ussery DW, Friis C. 2008. Comparative genomics of *Pseudomonas*, p 1–24. In Rehm BHA (ed), *Pseudomonas: model organism, pathogen, cell factory*. Wiley-VCH, Weinheim, Germany.
- Timmis KN. 2002. *Pseudomonas putida*: a cosmopolitan opportunist par excellence. *Environ Microbiol* 4:779–781. <https://doi.org/10.1046/j.1462-2920.2002.00365.x>.
- Bugg TD, Ahmad M, Hardiman EM, Singh R. 2011. The emerging role for bacteria in lignin degradation and bio-product formation. *Curr Opin Biotechnol* 22:394–400. <https://doi.org/10.1016/j.copbio.2010.10.009>.
- Poblete-Castro I, Becker J, Dohnt K, dos Santos VM, Wittmann C. 2012. Industrial biotechnology of *Pseudomonas putida* and related species. *Appl Microbiol Biotechnol* 93:2279–2290. <https://doi.org/10.1007/s00253-012-3928-0>.
- Frasson D, Opoku M, Picozzi T, Torossi T, Balada S, Smits THM, Hilber U. 2017. *Pseudomonas wadenswilerensis* sp. nov. and *Pseudomonas reidholzensis* sp. nov., two new species within the *Pseudomonas putida* group isolated from forest soil. *Int J Syst Evol Microbiol* 67:2853–2861. <https://doi.org/10.1099/ijsem.0.002035>.
- Rutz D, Frasson D, Sievers M, Blom J, Rezzonico F, Pothier JF, Smits THM. 2018. High-quality draft genome sequence of *Pseudomonas wadenswilerensis* CCOS 864^T. *Microbiol Resour Announc* 7:e01059-18. <https://doi.org/10.1128/MRA.01059-18>.
- Meyer F, Goesmann A, McHardy AC, Bartels D, Bekel T, Clausen J, Kalinowski J, Linke B, Rupp O, Giegerich R, Pühler A. 2003. GenDB—an open source genome annotation system for prokaryote genomes. *Nucleic Acids Res* 31:2187–2195. <https://doi.org/10.1093/nar/gkg312>.
- Blom J, Kreis J, Spänig S, Juhre T, Bertelli C, Ernst C, Goesmann A. 2016. EDGAR 2.0: an enhanced software platform for comparative gene content analyses. *Nucleic Acids Res* 44:W22–W28. <https://doi.org/10.1093/nar/gkw255>.
- Meier-Kolthoff JP, Auch AF, Klenk H-P, Göker M. 2013. Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinformatics* 14:60. <https://doi.org/10.1186/1471-2105-14-60>.
- Richter M, Rosselló-Móra R, Glöckner FO, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
- Tsou AY, Ransom SC, Gerlt JA, Buechter DD, Babbitt PC, Kenyon GL. 1990. Mandelate pathway of *Pseudomonas putida*: sequence relationships involving mandelate racemase, (S)-mandelate dehydrogenase, and benzoylformate decarboxylase and expression of benzoylformate decarboxylase in *Escherichia coli*. *Biochemistry* 29:9856–9862. <https://doi.org/10.1021/bi00494a015>.
- Priefert H, Rabenhorst J, Steinbüchel A. 1997. Molecular characterization of genes of *Pseudomonas* sp. strain HR199 involved in bioconversion of vanillin to protocatechuic acid. *J Bacteriol* 179:2595–2607. <https://doi.org/10.1128/jb.179.8.2595-2607.1997>.
- Nogales J, Canales Á, Jiménez-Barbero J, Serra B, Pingarrón JM, García JL, Díaz E. 2011. Unravelling the gallic acid degradation pathway in bacteria: the gal cluster from *Pseudomonas putida*: aerobic gallic acid degradation. *Mol Microbiol* 79:359–374. <https://doi.org/10.1111/j.1365-2958.2010.07448.x>.