

Conversion of the CD8 lineage to CD4 T cells

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Mature T lymphocytes possess a CD4 or CD8 lineage-specific marker. These molecules also serve as a co-receptor which along with a specific T Cell Receptor (TCR) binds to MHC class II (MHCII) or class I (MHCI), respectively. MHCI is expressed by all nucleated cells, whereas MHCII by antigen-presenting cells (APC). CD8 T cells kill target cells through MHCI-based recognition. CD4 T cells recognize MHCII and act either as helper T (Th) cells potentiating immunity or as regulatory T (Treg) cells inducing tolerance.

The development of the adaptive immune repertoire is based on discrimination of self-antigens in the host against nonself-antigens from foreign invaders [1]. This concept has been serving as a pillar for immunology but faces a challenge to accommodate the interplay between the host immune system and the mutualistic microbiota. We tracked the clonal fate of CD4 and CD8 T cells at the interface with gut microbiota. We found that CD8 T cells cross-differentiated into MHCI-restricted CD4 T_h cells or Foxp3⁺ T_{reg} cells [2] (Figure 1). The CD8 lineage plasticity was found in two independent models of TCR-transgenic mice (OT1 and 8.3) and natural CD8 T cells from wildtype mice. The conversion from CD8 T cells to MHCI-restricted CD4 T_{reg} cells occurred in the gut-associated environment without regard to self-antigens, with a host-intrinsic plasticity amplified by microbiota. The MHCI-T_{reg} cell, in its physiological niche in the gut lamina propria or in a setting of adoptive transfer, potently suppressed inflammatory damage even in the apparent absence of cognate antigens [2].

Why is such an intrinsic plasticity built in the host immune system during evolution? The T cell repertoire consists of CD4 and CD8 T cells generated by thymic selection in newborns. With age, the thymus ceases to function but the thymic-derived T cell repertoire largely persists. In certain diseases, a portion or a subtype of T cells might be lost. Conceivably, evolution may have endowed mechanisms to restore the immune balance to cope with catastrophic damages, such as T cell depletion by viral infections or natural irradiation. The plasticity of the CD8 T cell lineage elucidated by our study has perhaps evolved as an alternative pathway to protect the integrity of the adaptive immune system. In the adverse event of CD4 T cell depletion, CD4 populations can be replenished via cross-differentiation from CD8 T cells. One of the fascinating aspects of this transition is the requirement of MHCII despite that the clonotype TCR recognizes MHCI-presentated antigens [2].

How is the CD8-to-CD4 lineage plasticity relevant to biomedicine? In the modern world, HIV infection perhaps represents the most known example of catastrophic loss of CD4 T cells and subsequent imbalance of the CD8 versus CD4 lineages. It remains to be seen whether the depletion of CD4 T cells in HIV infection triggers cross-differentiation from the CD8 lineage to CD4 T cells. If so, the conversion might not only replenish the CD4 T cell pool with new targets for infection by HIV but also exacerbate the depletion of CD8 populations, while creating a potent immunosuppression in the gut mucosa by the converted MHCI-T_{reg} cells.

An imbalance of CD8 versus CD4 T cells could also be medically introduced in a type of cancer immunotherapy, wherein a patient is subject to immunoablation conditioning and then receives adoptive transfer of CD8 T cells enriched for specificities against tumors. Tumors present an immunoprivileged microenvironment with some neoantigens but mostly self-antigens [3]. It remains to be examined whether

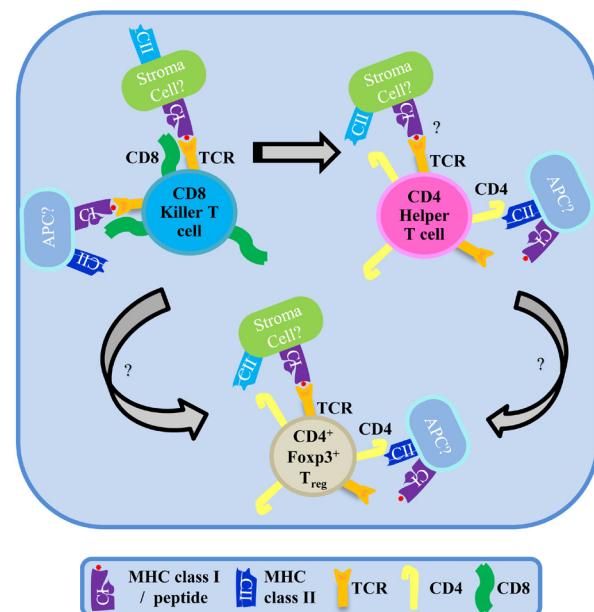


Figure 1: The mismatch to the rescue — a tale of the convert. The CD8 killer T cell lineage converts to MHCI-restricted CD4 T helper cells and CD4⁺Foxp3⁺ suppressor cells in the gut-associated environment. Despite the mismatch of the CD4 co-receptor on the converted CD4 T cells to MHCI-restricted TCR binding, the host-intrinsic plasticity of the CD8 T cell lineage may serve as an alternative pathway to induce “selfless” tolerance and restore immune balance, especially at the interface with microbiota.

such a microenvironment triggers CD8-to-CD4 lineage conversion and generation of MHCI-T_{reg} cells.

In autoimmune diseases, CD4 T cell depletion has been tested as a potential therapy. In our study, depletion of CD4 T cells in normal mice led to an increased population of CD4 T cells converted from the CD8 lineage. The converted CD4⁺Foxp3⁻ cells may help activation of CD8 T cells. Moreover, MHCI-T_{reg} cells might provide a potent antidote against autoimmune damage by CD8 T cells [2]. The conventional MHCII-restricted CD4⁺Foxp3⁺ T cells provide dominant tolerance through a variety of mechanisms [4], including cellular contact-based interaction between T_{reg} and pathogenic T cells [5]. One might envisage that MHCI-based recognition facilitates such a direct cellular contact. Of note, MHCI-restricted CD4 T cell clones exist in the natural repertoire of healthy humans [6]. A number of studies have also showed the possibilities of engineering human CD4 T cells recognizing MHCI-presented antigens by transducing MHCI-restricted TCR into CD4 T cells, including CD4 T_{reg} cells [7]. Along this direction, understanding the cellular and molecular mechanisms responsible for CD8-to-CD4 lineage conversion naturally at a clonal level will greatly benefit therapeutic translation of MHCI-restricted CD4 T cells, particularly MHCI-T_{reg} cells.

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REFERENCES

1. Burnet FM. Aust J Sci. 1957; 20: 67-69.
2. Lui JB, et al. Cell Rep. 2015; 10: 574-585.
3. Miska J, et al. Eur J Immunol. 2012; 42: 2584-2596.
4. Josefowicz SZ, et al. Annu Rev Immunol. 2012; 30: 531-564.
5. Miska J, et al. J Exp Med. 2014; 211: 441-456.
6. Strassman G, et al. J Immunol. 1984; 133: 1705-1709.
7. Plesa G, et al. Blood. 2012; 119: 3420-3430.