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Regression of Hepatic Fibrosis and Evolution of Cirrhosis: A Concise Review

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Abstract: Fibrosis is not a unidirectional, linear process, but a dynamic one resulting from an interplay of fibrogenesis and fibrolysis depending on the extent and severity of a biologic insult, or lack thereof. Regression of fibrosis has been documented best in patients treated with phlebotomies for hemochromatosis, and after successful suppression and eradication of chronic hepatitis B and C infections. This evidence mandates a reconsideration of the term "cirrhosis," which implies an inevitable progression towards liver failure. Furthermore, it also necessitates a staging system that acknowledges the bidirectional nature of evolution of fibrosis, and has the ability to predict if the disease process is progressing or regressing. The Beijing classification attempts to fill this gap in contemporary practice. It is based on microscopic features termed "the hepatic repair complex," defined originally by Wanless and colleagues. The elements of the hepatic repair complex represent the 3 processes of fragmentation and regression of scar, vascular remodeling (resolution), and parenchymal regeneration. However, regression of fibrosis does not imply resolution of cirrhosis, which is more than just a stage of fibrosis. So far, there is little to no evidence to suggest that large regions of parenchymal extinction can be repopulated by regenerating hepatocytes. Similarly, the vascular lesions of cirrhosis persist, and there is no evidence of complete return to normal microcirculation in cirrhotic livers. In addition, the risk of hepatocellular carcinoma is higher compared with the general population and these patients need continued screening and surveillance.

Key Words: cirrhosis, hepatocellular carcinoma, regression of hepatic fibrosis, vascular lesions, vascular remodeling, hepatic repair complex, the Beijing classification, chronic hepatitis, nonalcoholic fatty liver disease

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L ike many radically disruptive ideas, the debate about regression of hepatic fibrosis and cirrhosis started in early 1960s. An early study on reversal of fibrosis was published by Hans Popper, one of the founders of hepatopathology.¹⁻⁴ The debate continued with much skepticism and little optimism over the next 4 decades,⁵ culminating with the pioneering publication of Wanless et al.⁶ The past 2 decades have however, provided substantial evidence for regression of hepatic fibrosis led to development of the Beijing classification for systematic and reproducible documentation of regression

versus progression of fibrosis.¹¹ Successful treatment of chronic liver diseases, accompanied by of regression of fibrosis, has additionally changed the diagnostic and therapeutic implications of the term "cirrhosis." Considered inevitably "end-stage" for almost 2 centuries, current evidence for regression of fibrosis begs us to reconsider the term "cirrhosis" in the face of timely diagnosis, accurate identification of the underlying etiologies, and effective therapies.¹²

In this review, we summarize the histopathologic parameters that have been demonstrated to support regression of fibrosis in chronic liver disease, elaborate the Beijing classification's relevance and clinical utility, and highlight the underexplored areas of inquiry such as the fate of vascular abnormalities associated with cirrhosis and the risk of hepatocellular carcinoma (HCC).

HEPATIC FIBROSIS: IS UNIDIRECTIONAL PROGRESSION INEVITABLE?

Hepatic fibrosis is defined as excessive deposition of collagen (type I and 1V) in the hepatic parenchyma. It is distinct from parenchymal collapse which results in approximation (collapse) of pre-existing reticulin network (collagen type III) when the intervening hepatocytes are lost in a disease process. Conventional systems for staging of fibrosis in chronic viral hepatitis, developed before effective therapies became available, considered fibrosis to be a linear, inevitably progressive process that proceeded from an early stage (portal fibrosis-Ishak stage 0 to 2, Metavir F0 to F1, Batts-Ludwig stage 0 to 2), through intermediate stages (fibrous septa, focal or frequent —Ishak stage 3 to 4, Metavir F2 to F3, Batts-Ludwig stage 3) to advanced stages of fibrosis (fibrous septa with diffuse nodularity/"established cirrhosis"-Ishak stage 5 to 6, Metavir F3 to F4, Batts-Ludwig stage 3 to 4).^{13–15} However, fibrosis is not a unidirectional, linear process, but rather a dynamic one resulting from an interplay of fibrogenesis and fibrolysis depending on the extent and severity of a biologic insult, or lack thereof. In short, fibrosis of the liver parenchyma at any given time represents the balance between injury and repair. Thus, when an underlying injurious agent is removed, the balance shifts towards repair, with fibrolysis leading to regression of fibrosis.

Initially limited to experimental animal studies, credible evidence now extends to human patients treated with phlebotomies for hemochromatosis, and after successful suppression and eradication of chronic hepatitis B and C infections.^{2,4,16–20} Partial regression of cirrhosis has also been documented in patients with biliary obstruction, Wilson disease, intestinal bypass-related cirrhosis, Indian childhood cirrhosis, autoimmune hepatitis, primary biliary cholangitis, and alcoholic liver disease.^{21–28} The earliest case reports and small case series documenting regression of fibrosis after therapeutic phlebotomies were published in 1950 to 1970s.^{29–33} One of the largest series on regression of

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fibrosis in hemochromatosis documented regression of fibrosis using METAVIR staging system.³⁴ The study documented 36 patients, between the ages of 18 to 75 with homozygous C282Y mutation, METAVIR stage F3 or F4 on the initial liver biopsy, and a minimal interval of 2 years between cessation of phlebotomies and a second liver biopsy. The pretherapeutic and posttherapeutic biopsies were compared independently by 2 liver pathologists, who were blinded to the clinical and biochemical data. The histologic parameters studied included portal fibrosis (MET-AVIR grading system), quantity of iron (semiquantitative grading system of Deugnier et al¹³),³¹ and amount of steatosis (a 4-grade scale). Regression of fibrosis, defined as a decrease of at least 2 METAVIR stages, was seen in 47% (17/36) patients. Among 23 patients with cirrhosis (F4) on their initial biopsy, fibrosis decreased in 10 (43.5%); from F4 to stage 0 in 1 patient, to stage 1 in 4 patients, to stage 2 in 3 patients, and to stage 3 in 2 patients. Among 13 patients with F3 fibrosis on initial biopsy, fibrosis decreased in 11 (84.6%); from F3 to stage 0 in 3 patients, to stage 1 in 6 patients, and to stage 2 in 2 patients.

DOES REGRESSION OF FIBROSIS IMPLY REVERSAL OF CIRRHOSIS?

Cirrhosis is not merely a "stage of fibrosis," but rather a diffuse process characterized by advanced fibrosis, regenerative nodule formation, and vascular remodeling. Vascular remodeling accompanies parenchymal damage and is fundamental to the progression of chronic liver diseases.

A hallmark of chronic liver diseases are regions of parenchymal extinction, originally defined as a parenchymal area with contiguous loss of hepatocytes.³⁵ The drop-out of hepatocytes is accompanied by loss of the local microvasculature, which causes vascular compromise and ischemia, leading to further parenchymal damage. A self-perpetuating cycle is set up with increasing vascular compromise, involving progressively larger branches of the hepatic and portal vein thus escalating the congestion and ischemia,³⁶ and leading to ever larger areas of parenchymal collapse and extinction. Therefore, Wanless et al³⁶ proposed a revised definition of parenchymal extinction as "a region with focal loss of contiguous hepatocytes and adjacent microvascular structures." The fate of vascular abnormalities in cirrhosis is an underexplored area and there is lack of substantial evidence to support the idea of complete regression of these lesions. Thus, regression of fibrosis does not equate to reversal of cirrhosis as vascular abnormalities persist in livers with regressed fibrosis.

HISTOLOGIC FEATURES OF FIBROSIS REGRESSION

The Hepatic Repair Complex

The "hepatic repair complex" defined by Wanless et al⁶ describes the elemental microscopic features that signal regression of fibrosis: delicate perforated septa, isolated thick collagen fibers, delicate periportal fibrous spikes, portal tract remnants, hepatic vein remnants with prolapsed hepatocytes, hepatocytes within portal tracts or splitting septae, minute regenerative nodules, and aberrant parenchymal veins. These features form the basis of the Beijing classification (discussed below), which broadly classifies them into 3 categories; fragmentation and regression of scar, vascular remodeling (resolution), and parenchymal regeneration (Fig. 1).¹¹

Fragmentation and Regression of Scar

An established cirrhotic nodule is a "regenerative" nodule surrounded completely by thick fibrous septae containing inflammatory infiltrate, ductular proliferation, and edema. The central hepatic veins are compressed, obliterated, thrombosed, and often extinct, with concomitant development of vascular collaterals shunts. The vascular shunts in the cirrhotic nodule act as independent microcirculatory units. With regression, the fibrous septa start to lose inflammatory infiltrates and edema thus becoming more compact, thin, delicate, and less cellular or completely acellular. Thinning of septa leads to perforation and fragmentation and eventual interruption by regenerating hepatocytes. The hepatocytes can often be seen splitting the septa. Other features suggestive of regressing fibrosis are the presence of periportal delicate fibrous spikes or "adhesions," and reduction in the amount of collagen in the portal tracts.

Vascular Remodeling

With resorption of collagen, the portal tracts at the periphery of cirrhotic nodules start losing collagen resulting in appearance of "portal tracts remnants"-paired artery/arteriole and bile duct with a thin, delicate rim of collagen. The small veins are often obliterated. As repair/regeneration persists, hepatocytes migrate into the portal tract stroma and can be seen in the proximity of ducts/arteries and even in the lumen of portal vein remnants. Partially or completely recanalized large hepatic veins can be seen with regression, however most veno-occlusive disease lesions persist as fibroelastotic cords of collagen. Migration of regenerating hepatocytes into these lesions is also seen. Other features include presence of telangiectatic sinusoids and unpaired arteries in the regions of parenchymal extinction. These "arterialized sinusoids" and 'capillarized channels" representing vascular shunts persist even when much of the fibrosis has regressed (Fig. 2).^{37–39}

Parenchymal Regeneration

Suppression of the underlying injurious agent (viral infection, autoimmune hepatitis, alcohol etc.) tilts the balance in favor of hepatocellular regeneration. Hepatocytes proliferate and repopulate foci of parenchymal extinction, hitherto rich in proliferating ductules and inflammatory cells. Resorption of sinusoidal collagen results in remodeling of these areas and appearance of "near-normal" hepatic plates. However, large regions of parenchymal extinction persist, thus a complete regression of cirrhosis is almost never seen (Fig. 2).³⁵

Incomplete Septal Cirrhosis

Incomplete septal cirrhosis, an enigmatic entity first described by Popper,⁴⁰ is a type of cirrhosis characterized by vaguely macronodular appearance delineated by delicate, incomplete septa. It is often associated with portal hypertension.⁴¹ Although incomplete septal cirrhosis has been a perplexing entity, there are documented reports of partially resolved cirrhosis following suppression of liver injury that appear as incomplete septal fibrosis.^{42,43} Wanless et al,⁶ based on a study of 52 cases, proposed that incomplete septal cirrhosis, in fact represents a stage of regression of cirrhosis. With regression of fibrosis, the septa around the cirrhotic nodules become thin and delicate with time, and with migration of hepatocyte buds are split apart and eventually fragmented, imparting the appearance of an "incomplete" nodule. This observation was supported by

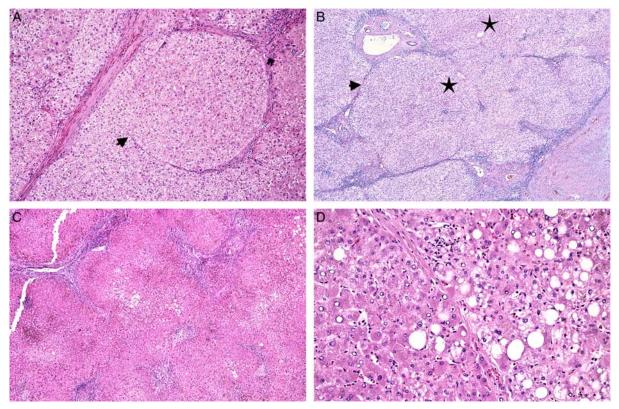


FIGURE 1. A, Regressing fibrosis seen as an incomplete nodule with progressively thinning septum (arrowhead) and regeneration of hepatocytes in a patient with cirrhosis due to nonalcoholic steatohepatitis. Hematoxylin and eosin ×100. B, Nodular liver parenchyma with incomplete septa. Compared with the portal tracts, fibrosis has regressed significantly with one thin septum (arrowhead) and aberrant vein remnants (stars) in a patient with cirrhosis due to nonalcoholic steatohepatitis. Hematoxylin and eosin ×100. C, Vaguely nodular liver parenchyma characterized by absence of complete septa and presence of portal tract remnants. Mild steatosis persists without ballooned hepatocytes or Mallory-Denk bodies in a patient with cirrhosis due to nonalcoholic steatohepatitis. Hematoxylin and eosin ×40. D, Hepatocyte buds are seen splitting the thin septum; the collagen band is significantly thinned out in the center compared with the periphery in a patient with cirrhosis due to nonalcoholic steatohepatitis. Hematoxylin and eosin ×100.

Theise et al¹¹ after assessment of 71 cases of treated chronic viral hepatitis B cirrhosis.

THE BEIJING CLASSIFICATION¹¹

The advent and wide-spread availability of successful therapies for suppression of hepatitis B viral replication and eradication of hepatitis C virus has compelled a reexamination of existing staging systems. Contemporary practice requires a staging system that take into consideration the bidirectional nature of evolution of fibrosis with the potential ability to predict if the disease process is progressing or regressing. The Beijing classification proposed by Theise and colleagues attempts to fill this gap by devising a method to assess whether fibrosis is progressing or regressing at a given period in time, as captured in a liver biopsy.

The Beijing classification was developed on a study of paired pre and post treatment biopsies from 71 patients successfully treated for chronic hepatitis B virus (HBV) infection. In addition to grading of necroinflammatory activity and staging of fibrosis, it provides a P-I-R (Progressive-Indeterminate-Regressive) score to evaluate the predominant pattern and quality of fibrosis: predominantly progressive, predominantly regressive or indeterminate for cases in which the pattern is not straightforward one way or the other. Evaluation is performed on a routine hematoxylineosin stain, combined with a trichrome or reticulin stain for assessment of fibrosis. Grading and staging are simplified into 3 grades of necroinflammatory activity and 3 stages of fibrosis, respectively. The 3 grades of necroinflammatory activity are "inactive" (portal inflammation only or rare foci of interface or lobular hepatitis; no confluent necrosis), "active, nonsevere" (varying degrees of interface and lobular hepatitis easily identified at low power; no confluent necrosis), and "active, severe" (confluent necrosis, perivenular drop out, bridging necrosis or parenchymal collapse). The 3 stages of fibrosis are "early" (no fibrosis or portal fibrosis), "intermediate" (fibrous septa, focal or frequent), and "advanced" (fibrous septa with focal or diffuse nodularity).

The defining histologic features of the progressive pattern are broad fibrous septa with loosely aggregated collagen fibers, edema, congestion, and inflammatory cells. In progressive disease, as parenchymal extinction evolves, the inflammatory activity is initially marked but gradually decreases and is masked by activation of macrophages and hepatic stellate cells, increasing deposition of collagen with broad fibrous septa formation, and appearance of ductular reactions. The predominantly regressive pattern is characterized by presence of thin, compact relatively acellular septa without edema, inflammation and other cellular infiltrates.

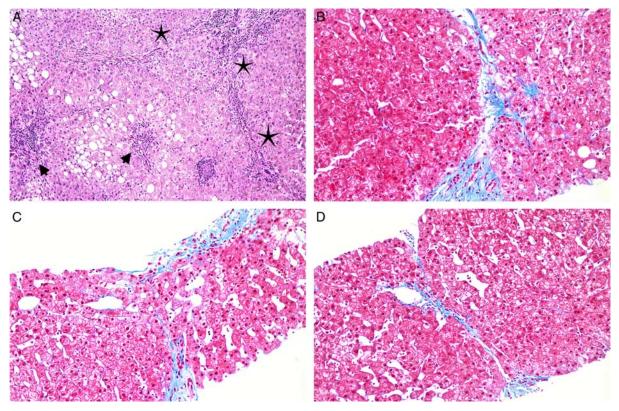


FIGURE 2. A, Regressed fibrous septa seen as periportal fibrosis (arrowheads) and regenerating hepatocytes splitting the septa (stars) in a patient with cirrhosis due to nonalcoholic steatohepatitis. Hematoxylin and eosin ×100. B, Fragmentation of septa and regression of scar seen as thinning of septae leading to perforation and fragmentation and eventual interruption by regenerating hepatocytes in a patient with cirrhosis due to chronic hepatitis C infection. Trichrome stain ×100. C, As repair/regeneration continues, hepatocytes migrate into the portal tract stroma and can be seen in the proximity of bile ducts and hepatic arteries in a patient with cirrhosis due to chronic hepatitis C. D, Resorption of a bridging septum between 2 portal tracts with negligible inflammatory infiltrate and resorption of sinusoidal collagen resulting into remodeling and appearance of "near-normal" hepatic plates. Trichrome stain ×40.

Septal fragmentation, recanalization of vascular thrombosis, and foci of disintegrating scar tissue become evident as regenerating hepatocytes begin to grown into and replace the scar tissue.

The prognostic value of the P-I-R score lies in its strong clinical correlations. In the pilot study, the P-I-R score reflected the prognosis of the disease, independent of necroinflammatory activity, fibrosis severity, or treatment experience. The score showed strong correlation with the treatment; *P/I* before treatment versus R after treatment. It also correlated with the 4 stage Laennec system for staging of cirrhosis, and thus may potentially serve as a surrogate marker for hepatic venous wedge pressures for risk stratification of portal hypertension. The Laennec system implies a sequential progression of cirrhosis from 4A to 4B to 4C; the Beijing classification's P-I-R score reflects the dynamic evolution between Laennec stages 4A, 4B, and 4C, thus incorporating the probability of regression from 4B and 4C to 4A.

DOES REGRESSION OF FIBROSIS DOWNGRADE THE RISK FOR HCC?

Pathogenesis of HCC is the result of accumulative mutational burden resulting from factors that affect the tumor and its microenvironment, the tumor

microenvironment (TME). Different etiological factors [HBV, hepatitis C viral (HCV), non alcoholic steatohepatitis, etc.] may elicit different pathways leading to hepatocarcinogenesis. The fibrous stroma, cellular elements, and vascular abnormalities in cirrhosis constitute the TME. When the injurious trigger is removed by effective therapies, the TME landscape changes due to decrease in inflammation and downregulation of the fibrogenic pathways. Despite this resolution, however, the driver mutations already imparted to hepatocytes in early disease, for instance, in HBV infection, do not vanish. The mutational burden rather becomes more complex and biodiverse with repeated cycles of injury and repair, resulting in genomic instability.44-47 Furthermore, hypoxia-induced vascular endothelial growth factor (VEGF) expression is linked to tumor diversity and TME polarization.48-51 With persistence of vascular lesions in cirrhosis, the hypoxic injury persists despite elimination of inflammation and regression of fibrosis.

HBV predisposes to hepatocarcinogenesis through direct and indirect mechanisms. One of the earliest direct mechanisms is HBV DNA integration into the host genome resulting in direct insertional mutations, leading to high levels of HBV replication, genomic instability and subsequent clonal expansion of tumor cells. In chronic HBV infection, HBV-encoded oncoproteins like hepatitis B virus X protein and truncated preS2/S proteins impair control of proliferation and transcription, thus contributing to malignant hepatocyte transformation. Hepatitis B virus X protein is also responsible for epigenetic changes of tumor suppressor genes, thus HBV associated HCCs have a higher rate of chromosomal alteration including TP53 mutations and relatively low rate of activating β -catenin mutations. HBV-related HCCs can arise in noncirrhotic livers, further supporting the direct role of the virus in hepatocarcinogenesis. Chronic high viral load and specific genotypes initiate a cascade of host immune response leading to recurrent necroinflammation, fibrosis, accelerated hepatocyte turnover and accumulation of mutations.^{52,53}

The principal oncologic drivers in chronic HCV infection are core protein and NS3 and NS5A proteins. HCV core is an RNA-binding protein, that is singularly capable of inducing hepatic steatosis, insulin resistance and HCC.^{54–56} It also promotes hepatocyte proliferation and its distinct mutations in genotype 1 patients confer a higher HCC risk, even after elimination of HCV.^{57,58} The NS3 protein of HCV promotes hepatic fibrosis in chronic liver disease while NS5A has a central role in viral replication and assembly, is capable of inducing hepatic steatosis, is protective against apoptosis and confers resistance against interferon α .^{59–64} The key cellular and molecular pathways underlying progression of chronic HCV infection to HCC are: the epidermal growth factor,^{65–70} signal transducer and activator of transcription 3^{71,72} transforming growth factor beta^{73–76} and VEGF.^{77,78} In its evolution, HCV infection is also associated with higher microvessel density compared with chronic hepatitis B patients.⁷⁹

In the face of ever-growing obesity epidemic in the developed and developing world, nonalcoholic fatty liver disease (NAFLD) has become one of the leading causes of end-stage liver disease.⁸⁰ Furthermore, it is the fastest growing cause of HCC.⁸¹ Inflammation in non alcoholic steatohepatitis is "metabolic" in nature⁸²; several pathways including insulin resistance, lipotoxicity, lipid peroxidation, and necroinflammation precipitate steatohepatitis.⁸³ With regression of fibrosis, most of the risk factors for development of HCC persist in NAFLD; lipid metabolic reprogramming involving carnitine palmitoyl transferase and fatty acid β -oxidation, being key mechanisms.⁸⁴ Similarly, the risk of HCC persists in cirrhosis secondary to hemochromatosis even after "clinically successful" therapeutic phlebotomies.^{85–90}

FUTURE PERSPECTIVE

With better understanding of the underlying cellular and molecular mechanisms of fibrosis, novel therapeutic strategies are under investigation to target these pathways.^{91–93} Although the significance of vascular lesions in the progression of liver disease is quite underappreciated, there is evidence to support and explore the potential role of anticoagulants, statins, and VEGF modulators as protective agents that can either slow down the progression or even reverse the vascular abnormalities.^{94–99}

CONCLUSIONS

The evidence in favor of regression of fibrosis in chronic liver disease is substantial and irrefutable. However, complete resolution of cirrhosis is yet to be demonstrated. So far, there is little to no evidence to suggest that large regions of parenchymal extinction can be repopulated by regenerating hepatocytes. Similarly, the vascular lesions of cirrhosis persist, and there is no evidence of complete return to normal microcirculation in cirrhotic livers. Despite regression of fibrosis, adequate suppression of viral load in chronic HBV infection, and successful treatment of chronic HCV infection, the risk of HCC is higher compared with the general population and these patients need continued screening and surveillance.^{100–103}

There is need for reassessing the timing of protocol biopsies for chronic viral hepatitis (HBV, HCV) and more studies are needed to explore the clinical utility and reproducibility of the Beijing classification. The vascular lesions of cirrhosis need to be studied and explored for their natural history and evolution. Cirrhosis, as a term either needs to be revised or discontinued as its historical diagnostic connotations and clinical implications would be increasingly less relevant in the face of modern therapeutic strategies.¹² And last but not the least, given the burden of NAFLD and a multitude of ongoing clinical trials, there is a need for documenting the regression of fibrosis and cirrhosis in a systematic, reproducible, and clinically relevant manner.

REFERENCES

- Hutterer F, Rubin E, Popper H. Mechanism of collagen resorption in reversible hepatic fibrosis. *Exp Mol Pathol.* 1964;3: 215–223.
- Brody JI, McKenzie D, Kimbal SG. Therapeutic phlebotomies in idiopathic hemochromatosis. *Am J Med Sci.* 1962;244: 575–586.
- 3. Bunton GL, Cameron SR. Regeneration of liver after biliary cirrhosis. Ann N Y Acad Sci. 1963;111:412–421.
- Quinn PS, Higginson J. Reversible and irreversible changes in experimental cirrhosis. *Am J Pathol.* 1965;47:353–370.
- 5. Bulletin of the World Health Organization. 1977;55:521-540.
- Wanless IR, Nakashima E, Sherman M. Regression of human cirrhosis. Morphologic features and the genesis of incomplete septal cirrhosis. *Arch Pathol Lab Med.* 2000;124:1599–1607.
- Ray MB. Regression of cirrhosis. A timely topic. Arch Pathol Lab Med. 2000;124:1589–1590.
- Chedid A. Regression of human cirrhosis. Arch Pathol Lab Med. 2000;124:1591.
- Chejfec G. Controversies in pathology. Is cirrhosis of the liver a reversible disease? Arch Pathol Lab Med. 2000;124:1585–1586.
- Geller SA. Coming or going? What is cirrhosis? Arch Pathol Lab Med. 2000;124:1587–1588.
- Theise ND, Jia J, Sun Y, et al. Progression and regression of fibrosis in viral hepatitis in the treatment era: the Beijing classification. *Mod Pathol.* 2018;31:1191–1200.
- Hytiroglou P, Snover DC, Alves V, et al. Beyond "cirrhosis": a proposal from the International Liver Pathology Study Group. Am J Clin Pathol. 2012;137:5–9.
- Deugnier YM, Loreal O, Turlin B, et al. Liver pathology in genetic hemochromatosis: a review of 135 homozygous cases and their bioclinical correlations. *Gastroenterology*. 1992;102:2050–2059.
- Knodell RG, Ishak KG, Black WC, et al. Formulation and application of a numerical scoring system for assessing histological activity in asymptomatic chronic active hepatitis. *Hepatology*. 1981;1:431–435.
- Intra-observer and interobserver variations in liver biopsy interpretation in patients with chronic hepatitis C. The French METAVIR Cooperative Study Group. *Hepatology*. 1994;20 (1 Pt 1):15–20.
- Marcellin P, Gane E, Buti M, et al. Regression of cirrhosis during treatment with tenofovir disoproxil fumarate for chronic hepatitis B: a 5-year open-label follow-up study. *Lancet.* 2013;381:468–475.
- Chang TT, Liaw YF, Wu SS, et al. Long-term entecavir therapy results in the reversal of fibrosis/cirrhosis and continued histological improvement in patients with chronic hepatitis B. *Hepatology*. 2010;52:886–893.
- Poynard T, McHutchison J, Manns M, et al. Impact of pegylated interferon alfa-2b and ribavirin on liver fibrosis in

patients with chronic hepatitis C. *Gastroenterology*. 2002;122: 1303–1313.

- Pockros PJ, Hamzeh FM, Martin P, et al. Histologic outcomes in hepatitis C-infected patients with varying degrees of virologic response to interferon-based treatments. *Hepatology*. 2010;52: 1193–1200.
- Manne V, Akhtar E, Saab S. Cirrhosis regression in patients with viral hepatitis B and C: a systematic review. J Clin Gastroenterol. 2014;48:e76–e84.
- Bunton GL, Cameron R. Regeneration of liver after biliary cirrhosis. Ann N Y Acad Sci. 1963;111:412–421.
- Falkmer S, Samuelson G, Sjolin S. Penicillamine-induced normalization of clinical signs, and liver morphology and histochemistry in a case of Wilson's disease. *Pediatrics*. 1970;45: 260–268.
- Grand RJ, Vawter GF. Juvenile Wilson disease: histologic and functional studies during penicillamine therapy. *J Pediatr.* 1975;87: 1161–1170.
- Mezey E, Imbembo AL. Hepatic collagen proline hydroxylase activity in hepatic disease following jejunoileal bypass for morbid obesity. *Surgery*. 1978;83:345–353.
- Pradhan AM, Bhave SA, Joshi VV, et al. Reversal of Indian childhood cirrhosis by D-penicillamine therapy. J Pediatr Gastroenterol Nutr. 1995;20:28–35.
- Dufour JF, DeLellis R, Kaplan MM. Reversibility of hepatic fibrosis in autoimmune hepatitis. *Ann Intern Med.* 1997;127: 981–985.
- Kaplan MM, DeLellis RA, Wolfe HJ. Sustained biochemical and histologic remission of primary biliary cirrhosis in response to medical treatment. *Ann Intern Med.* 1997;126:682–688.
- Kershenobich D, Vargas F, Garcia-Tsao G, et al. Colchicine in the treatment of cirrhosis of the liver. N Engl J Med. 1988;318: 1709–1713.
- Knauer CM, Gamble CN, Monroe LS. The reversal of hemochromatotic cirrhosis by multiple phlebotomies: report of a case. *Gastroenterology*. 1965;49:667–671.
- Powell LW, Kerr JF. Reversal of "cirrhosis" in idiopathic haemochromatosis following long-term intensive venesection therapy. *Australas Ann Med.* 1970;19:54–57.
- Duffy TJ, Meister L. Hemochromatosis. report of a case followed for seven years with repeated phlebotomies and liver biopsies. *Am J Med.* 1963;35:434–438.
- 32. Mader JH. Idiopathic hemochromatosis treated by repeated phlebotomies; report of two cases. *J Indiana State Med Assoc*. 1964;57:33–39.
- Blackburn CR, Mcguinness AE, Kaldor I. Removal of excess body iron in haemochromatosis by repeated venesection. *Australas Ann Med.* 1953;2:202–205.
- Falize L, Guillygomarc'h A, Perrin M, et al. Reversibility of hepatic fibrosis in treated genetic hemochromatosis: a study of 36 cases. *Hepatology*. 2006;44:472–477.
- Wanless IR, Wong F, Blendis LM, et al. Hepatic and portal vein thrombosis in cirrhosis: possible role in development of parenchymal extinction and portal hypertension. *Hepatology*. 1995;21:1238–1247.
- Wanless IR. The role of vascular injury and congestion in the pathogenesis of cirrhosis: the congestive escalator and the parenchymal extinction sequence. *Curr Hepatol Rep.* 2020;19: 40–53.
- Schaffner F, Popper H. Capillarization of the hepatic sinusoids in man. *Gastroenterology*. 1963;4:239–242.
- Taguchi K, Asano G. Neovascularization of pericellular fibrosis in alcoholic liver disease. *Acta Pathol Jpn*. 1988;38:615–626.
- 39. Guido M, Sarcognato S, Russo FP, et al. Focus on histological abnormalities of intrahepatic vasculature in chronic viral hepatitis. *Liver Int.* 2018;38:1770–1776.
- Popper H. What Are The Major Types of Hepatic Cirrhosis? Controversy in Internal Medicine. Philadelphia, PA: Saunders. 223–243.
- Nakanuma Y, Hoso M, Sasaki M, et al. Histopathology of the liver in non-cirrhotic portal hypertension of unknown aetilogy. *Histopathology*. 1996;28:195–204.

- 42. Grilli E, Galati V, Petrosillo N, et al. Incomplete septal cirrhosis after high-dose methylprednisolone therapy and regression of liver injury. *Liver Int.* 2015;35:674–676.
- Schinoni MI, Andrade Z, de Freitas LA, et al. Incomplete septal cirrhosis: an enigmatic disease. *Liver Int*. 2004;24:452–456.
- 44. Wang G, Wang Q, Liang N, et al. Oncogenic driver genes and tumor microenvironment determine the type of liver cancer. *Cell Death Dis.* 2020;11:313.
- 45. Seo HR. Roles of tumor microenvironment in hepatocelluar carcinoma. *Curr Cancer Ther Rev.* 2015;11:82–93.
- Capece D, Fischietti M, Verzella D, et al. The inflammatory microenvironment in hepatocellular carcinoma: a pivotal role for tumor-associated macrophages. *Biomed Res Int*. 2013;2013:187204.
- Santhakumar C, Gane EJ, Liu K, et al. Current perspectives on the tumor microenvironment in hepatocellular carcinoma. *Hepatol Int.* 2020;14:947–957.
- Vito A, El-Sayes N, Mossman K. Hypoxia-driven immune escape in the tumor microenvironment. *Cells*. 2020;9:992.
- Lin CA, Chang LL, Zhu H, et al. Hypoxic microenvironment and hepatocellular carcinoma treatment. *Hepatoma Res.* 2018; 4:26.
- Hamaguchi T, Iizuka N, Tsunedomi R, et al. Glycolysis module activated by hypoxia-inducible factor lalpha is related to the aggressive phenotype of hepatocellular carcinoma. *Int J Oncol.* 2008;33:725–731.
- Dai XY, Zhuang LH, Wang DD, et al. Nuclear translocation and activation of YAP by hypoxia contributes to the chemoresistance of SN38 in hepatocellular carcinoma cells. *Oncotarget*. 2016;7:6933–6947.
- Niu B, Hann H-W. Hepatitis B Virus–Related Hepatocellular Carcinoma: Carcinogenesis, Prevention, and Treatment, Updates in Liver Cancer, Hesham Mohamed Abdeldayem, IntechOpen. 2017.
- Levrero M, Zucman-Rossi J. Mechanisms of HBV-induced hepatocellular carcinoma. J Hepatol. 2016;64(suppl 1):S84–S101.
- Moriya K, Yotsuyanagi H, Shintani Y, et al. Hepatitis C virus core protein induces hepatic steatosis in transgenicmice. J Gen Virol. 1997;78(pt 7):1527–1531.
- 55. Shintani Y, Fujie H, Miyoshi H, et al. Hepatitis C virus infection and diabetes: direct involvement of the virus in the development of insulin resistance. *Gastroenterology*. 2004;126:840–848.
- Moriya K, Fujie H, Shintani Y, et al. The core protein of hepatitis C virus induces hepatocellular carcinoma in transgenic mice. *Nat Med.* 1998;4:1065–1067.
- Kawamura H, Govindarajan S, Aswad F, et al. HCV core expression in hepatocytes protects against autoimmune liver injury and promotes liver regeneration in mice. *Hepatology*. 2006;44:936–944.
- Nakamoto S, Imazeki F, Fukai K, et al. Association between mutations in the core region of hepatitis C virus genotype 1 and hepatocellular carcinoma development. J Hepatol. 2009;52:72–78.
- Sakata K, Hara M, Terada T, et al. HCV NS3 protease enhances liver fibrosis via binding to and activating TGF-beta type I receptor. *Sci Rep.* 2013;3:3243.
- Appel N, Zayas M, Miller S, et al. Essential role of domain III of nonstructural protein 5A for hepatitis C virus infectious particle assembly. *PLoS Pathog.* 2008;4:e1000035.
- Enomoto N, Sakuma I, Asahina Y, et al. Mutations in the nonstructural protein 5A gene and response to interferon in patients with chronic hepatitis C virus 1b infection. *N Engl J Med.* 1996;334:77–81.
- 62. Gale MJ Jr, Korth MJ, Tang NM, et al. Evidence that hepatitis C virus resistance to interferon is mediated through repression of the PKR protein kinase by the nonstructural 5A protein. *Virology*. 1997;230:217–227.
- 63. Wang AG, Lee DS, Moon HB, et al. Non-structural 5A protein of hepatitis C virus induces a range of liver pathology in transgenic mice. *J Pathol.* 2009;219:253–262.
- Majumder M, Ghosh AK, Steele R, et al. Hepatitis C virus NS5A protein impairs TNF-mediated hepatic apoptosis, but not by an anti-FAS antibody, in transgenic mice. *Virology*. 2002;294:94–105.

- Fuchs BC, Hoshida Y, Fujii T, et al. Epidermal growth factor receptor inhibition attenuates liver fibrosis and development of hepatocellular carcinoma. *Hepatology*. 2014;59:1577–1590.
- Hoshida Y, Villanueva A, Kobayashi M, et al. Gene expression in fixed tissues and outcome in hepatocellular carcinoma. *N Engl J Med.* 2008;359:1995–2004.
- Hoshida Y, Villanueva A, Sangiovanni A, et al. Prognostic gene expression signature for patients with hepatitis C-related early-stage cirrhosis. *Gastroenterology*. 2013;144:1024–1030.
- Nakagawa S, Wei L, Song WM, et al. Molecular liver cancer prevention in cirrhosis by organ transcriptome analysis and lysophosphatidic acid pathway inhibition. *Cancer Cell*. 2016;30: 879–890.
- Ninio L, Nissani A, Meirson T, et al. Hepatitis C virus enhances the invasiveness of hepatocellular carcinoma via EGFR-mediated invadopodia formation and activation. *Cells*. 2019;8:1395.
- Devhare PB, Sasaki R, Shrivastava S, et al. Exosomemediated intercellular communication between hepatitis C virus-infected hepatocytes and hepatic stellate cells. *J Virol.* 2017;91:e02225–e02316.
- 71. Llovet JM, Zucman-Rossi J, Pikarsky E, et al. Hepatocellular carcinoma. *Nat Rev Dis Primers*. 2016;2:16018.
- He G, Yu GY, Temkin V, et al. Hepatocyte IKKbeta/NFkappaB inhibits tumor promotion and progression by preventing oxidative stress-driven STAT3 activation. *Cancer Cell*. 2010;17:286–297.
- Fabregat I, Caballero-Diaz D. Transforming growth factorbeta-induced cell plasticity in liver fibrosis and hepatocarcinogenesis. *Front Oncol.* 2018;8:357.
- 74. Yang L, Inokuchi S, Roh YS, et al. Transforming growth factor-beta signaling in hepatocytes promotes hepatic fibrosis and carcinogenesis in mice with hepatocyte-specific deletion of TAK1. *Gastroenterology*. 2013;144:1042.e1–1054.e4.
- Kwon YC, Sasaki R, Meyer K, et al. Hepatitis C virus core protein modulates endoglin (CD105) signaling pathway for liver pathogenesis. J Virol. 2017;91:e01235–e01317.
- Yang LY, Lu WQ, Huang GW, et al. Correlation between CD105 expression and postoperative recurrence and metastasis of hepatocellular carcinoma. *BMC Cancer*. 2006;6:110.
- Mee CJ, Farquhar MJ, Harris HJ, et al. Hepatitis C virus infection reduces hepatocellular polarity in a vascular endothelial growth factor-dependent manner. *Gastroenterology*. 2010; 138:1134–1142.
- Schoenleber SJ, Kurtz DM, Talwalkar JA, et al. Prognostic role of vascular endothelial growth factor in hepatocellular carcinoma: systematic review and meta-analysis. *Br J Cancer*. 2009;100:1385–1392.
- Mazzanti R, Messerini L, Monsacchi L, et al. Chronic viral hepatitis induced by hepatitis C but not hepatitis B virus infection correlates with increased liver angiogenesis. *Hepatology*. 1997; 25:229–234.
- Noureddin M, Vipani A, Bresee C, et al. NASH leading cause of liver transplant in women: updated analysis of indications for liver transplant and ethnic and gender variances. *Am J Gastroenterol.* 2018;113:1649–1659.
- Younossi Z, Anstee QM, Marietti M, et al. Global burden of NAFLD and NASH: trends, predictions, risk factors and prevention. *Nat Rev Gastroenterol Hepatol.* 2018;15:11–20.
- Hotamisligil GS. Inflammation, metaflammation and immunometabolic disorders. *Nature*. 2017;542:177–185.
- Peiseler M, Tacke F. Inflammatory mechanisms underlying nonalcoholic steatohepatitis and the transition to hepatocellular carcinoma. *Cancers.* 2021;13:730.

- Nakagawa H, Hayata Y, Kawamura S, et al. Lipid metabolic reprogramming in hepatocellular carcinoma. *Cancers (Basel)*. 2018;10:447.
- Fellows IW, Stewart M, Jeffcoate WJ, et al. Hepatocellular carcinoma in primary haemochromatosis in the absence of cirrhosis. *Gut.* 1988;29:1603–1606.
- Deugnier YM, Guyader D, Crantock L, et al. Primary liver cancer in genetic hemochromatosis: a clinical, pathological, and pathogenetic study of 54 cases. *Gastroenterology*. 1993;104:228–234.
- Goh J, Callagy G, McEntee G, et al. Hepatocellular carcinoma arising in the absence of cirrhosis in genetic haemochromatosis: three case reports and review of literature. *Eur J Gastroenterol Hepatol.* 1999;11:915–919.
- Kohler HH, Hohler T, Kusel U, et al. Hepatocellular carcinoma in a patient with hereditary hemochromatosis and noncirrhotic liver: a case report. *Pathol Res Pract*. 1999;195:509–513.
- Britto MR, Thomas LA, Balaratnam N, et al. Hepatocellular carcinoma arising in non-cirrhotic liver in genetic haemochromatosis. *Scand J Gastroenterol.* 2000;35:889–893.
- Blumberg RS, Chopra S, Ibrahim R, et al. Primary hepatocellular carcinoma in idiopathic hemochromatosis after reversal of cirrhosis. *Gastroenterology*. 1988;95:1399–1402.
- Kisseleva T, Brenner D. Molecular and cellular mechanisms of liver fibrosis and its regression. *Nat Rev Gastroenterol Hepatol.* 2021;18:151–166.
- Selicean S, Wang C, Guixé-Muntet S, et al. Regression of portal hypertension: underlying mechanisms and therapeutic strategies. *Hepatol Int.* 2021;15:36–50.
- Kisseleva T, Cong M, Paik Y, et al. Myofibroblasts revert to an inactive phenotype during regression of liver fibrosis. *Proc Natl Acad Sci USA*. 2012;109:9448–9453.
- Yang L, Kwon J, Popov Y, et al. Vascular endothelial growth factor promotes fibrosis resolution and repair in mice. *Gastroenterology*. 2014;146:1339.e1–1350.e1.
- Villa E, Camma C, Marietta M, et al. Enoxaparin prevents portal vein thrombosis and liver decompensation in patients with advanced cirrhosis. *Gastroenterology*. 2012;143:1253–1260.e4.
- Schepis F, Villa E. Thrombophilic genetic risk factors for liver fibrosis: to screen or not to screen? J Hepatol. 2015;63:1311–1313.
- Tripathi DM, Vilaseca M, Lafoz E, et al. Simvastatin prevents progression of acute on chronic liver failure in rats with cirrhosis and portal hypertension. *Gastroenterology*. 2018;155:1564–1577.
- Meireles CZ, Pasarin M, Lozano JJ, et al. Simvastatin attenuates liver injury in rodents with biliary cirrhosis submitted to hemorrhage/resuscitation. *Shock*. 2017;47:370–377.
- Turco L, Villanueva C, La Mura V, et al. Lowering portal pressure improves outcomes of patients with cirrhosis, with or without ascites: a meta-analysis. *Clin Gastroenterol Hepatol.* 2020;18:313–327.e6.
- Lim YS, Han S, Heo NY, et al. Mortality, liver transplantation, and hepatocellular carcinoma among patients with chronic hepatitis B treated with entecavir vs lamivudine. *Gastroenterology*. 2014;147:152–161.
- 101. Kim JH, Sinn DH, Kang W, et al. Low-level viremia and the increased risk of hepatocellular carcinoma in patients receiving entecavir treatment. *Hepatology*. 2017;66:335–343.
- 102. Bruden DJT, McMahon BJ, Townshend-Bulson L, et al. Risk of end-stage liver disease, hepatocellular carcinoma, and liverrelated death by fibrosis stage in the hepatitis C Alaska Cohort. *Hepatology*. 2017;66:37–45.
- 103. Papatheodoridis GV, Chan HL, Hansen BE, et al. Risk of hepatocellular carcinoma in chronic hepatitis B: assessment and modification with current antiviral therapy. *J Hepatol.* 2015;62:956–967.