

Review

Beyond Bone Loss: A Biology Perspective on Osteoporosis Pathogenesis, Multi-Omics Approaches, and Interconnected Mechanisms

Yixin Zhao ¹, Jihan Wang ², Lijuan Xu ¹, Haofeng Xu ¹, Yu Yan ¹, Heping Zhao ¹ and Yuzhu Yan ^{1,*} 

¹ Clinical Laboratory of Honghui Hospital, Xi'an Jiaotong University, Xi'an 710054, China; 727042677@163.com (Y.Z.)

² Yan'an Medical College, Yan'an University, Yan'an 716000, China

* Correspondence: yanyuzhu2020@163.com

Abstract: Osteoporosis is a systemic bone disorder characterized by decreased bone mass and deteriorated microarchitecture, leading to an increased risk of fractures. Recent studies have revealed that its pathogenesis involves complex biological processes beyond bone remodeling, including oxidative stress, chronic inflammation, cellular senescence, osteoimmunology, gut microbiota alterations, and epigenetic modifications. Oxidative stress disrupts bone homeostasis by promoting excessive free radical production and osteoclast activity. Chronic inflammation and the accumulation of senescent cells impair skeletal repair mechanisms. Advances in osteoimmunology have highlighted the critical role of immune–bone crosstalk in regulating bone resorption and formation. Moreover, the gut–bone axis, mediated by microbial metabolites, influences bone metabolism through immune and endocrine pathways. Epigenetic changes, such as DNA methylation and histone modification, contribute to gene–environment interactions, affecting disease progression. Multi-omics approaches (genomics, proteomics, and metabolomics) systematically identify molecular networks and comorbid links with diabetes/cardiovascular diseases, revealing pathological feedback loops that exacerbate bone loss. In conclusion, osteoporosis pathogenesis extends beyond bone remodeling to encompass systemic inflammation, immunometabolic dysregulation, and gut microbiota–host interactions. Future research should focus on integrating multi-omics biomarkers with targeted therapies to advance precision medicine strategies for osteoporosis prevention and treatment.

Keywords: osteoporosis pathogenesis; bone remodeling; osteoimmunology; epigenetics; multi-omics



check for updates

Academic Editor: Elisa Belluzzi

Received: 2 April 2025

Revised: 9 June 2025

Accepted: 10 June 2025

Published: 12 June 2025

Citation: Zhao, Y.; Wang, J.; Xu, L.;

Xu, H.; Yan, Y.; Zhao, H.; Yan, Y.

Beyond Bone Loss: A Biology

Perspective on Osteoporosis

Pathogenesis, Multi-Omics

Approaches, and Interconnected

Mechanisms. *Biomedicines* **2025**, *13*,1443. [https://doi.org/10.3390/](https://doi.org/10.3390/biomedicines13061443)[biomedicines13061443](https://doi.org/10.3390/biomedicines13061443)**Copyright:** © 2025 by the authors.

Licensee MDPI, Basel, Switzerland.

This article is an open access article

distributed under the terms and

conditions of the Creative Commons

Attribution (CC BY) license

[\(https://creativecommons.org/](https://creativecommons.org/licenses/by/4.0/)[licenses/by/4.0/\).](https://creativecommons.org/licenses/by/4.0/)

1. Introduction

Osteoporosis, a prevalent skeletal disorder characterized by reduced bone density and compromised architecture, significantly increases fracture risk and global disease burden [1]. When bone mineral density (BMD) falls 2.5 standard deviations below the mean for healthy young adults (T-score ≤ -2.5), osteoporosis remains underdiagnosed, with its true prevalence often being inferred from fracture rates [1]. Globally, osteoporotic fractures account for 0.83% of non-communicable disease burden, with notable prevalence in aging populations: 10.3% in U.S. adults ≥ 50 years and 33.49% in middle-aged and elderly Chinese adults [2,3]. In the UK, there is a similar burden of osteoporosis: the authors of an epidemiological study hypothesized that one in two women and one in five men aged over 50 years will suffer an osteoporotic fracture [4]. The authors of a meta-analysis

concluded that the global prevalence of osteoporosis is approximately 18.3% in the age range of 15–105 years, where the prevalence rate in women is 23.1%, compared with 11.7% in men [5]. In a previous study, it was estimated that the number of people worldwide at high risk of osteoporotic fracture will double from 2010 to 2040. Asia has the highest proportion in the world, followed by Europe [6]. In addition, geographical location also has an impact on osteoporosis fractures: studies have shown that the probability of hip fractures is lower in low-latitude countries, possibly because of prolonged sunlight exposure [7].

Osteoporosis is inherently multifactorial, arising from complex interactions between genetic predisposition, environmental factors, hormonal changes, metabolic dysregulation, and cellular signaling pathways. This complexity necessitates holistic investigative approaches. Advancements in omics technologies are revolutionizing our understanding of osteoporosis. Genome-wide association studies (GWASs) allow for the identification of susceptibility genes, while transcriptomics, particularly single-cell approaches, reveals cellular heterogeneity and signaling pathways in bone remodeling. Proteomics uncovers key proteins linked to BMD, offering insights into diagnostic markers and therapeutic targets. Metabolomics explores metabolic shifts in calcium, phosphorus, and energy balance, providing potential biomarkers for early detection. Emerging fields such as epigenomics and microbiome research allow for the further elucidation of gene–environment interactions and gut–bone axis dynamics. Multi-omics integration is poised to propel osteoporosis management into the era of precision medicine, enabling early warning, precise typing, and targeted therapies. Therefore, the use of omics methods can provide a more comprehensive understanding, treatment, and prevention of osteoporosis. Genomics has confirmed the impact of variations in genes such as collagen type I alpha1 (*COL1A1*) and receptor-related protein 5 (*LRP5*) on bone health through candidate gene association studies. GWASs identify numerous risk loci related to osteoporosis. With RNA sequencing (RNA-seq), non-coding RNAseq, single-cell RNA sequencing (ScRNA-seq), and other proteomic research methods, not only can the key transcription factors of osteoporosis be identified; the differentially expressed genes of osteoporosis can also be identified to determine the etiology. Commonly used research methods in proteomics include TMT proteomics, liquid chromatography–tandem mass spectrometry (LC–MS/MS), Matrix-Assisted Laser Desorption/Ionization Time-of-Flight/Time-of-Flight Mass Spectrometry (MALDI-TOF/TOF), etc. Proteomics helps us to find osteogenic and osteoclastic markers, including Alkaline Phosphatase (ALP), osterix (OSX), the C-terminal telopeptide of type I X-linked collagen (CTX), etc. Meanwhile, the proteomic analysis of plasma, exosomes, and various body fluids allows one to more easily obtain specimens for the diagnosis of osteoporosis. Mass spectrometry analysis is a common research method in metabolomics that has revealed that osteoporosis is related to amino acid metabolism, lipid metabolism, energy metabolism, etc. Multi-omics analysis allows for the integration of the research results of multiple omics, which are then systematically analyzed. At the same time, Machine Learning can be used to analyze vast and complex datasets [8].

Osteoporosis frequently co-occurs with cardiovascular, respiratory, and other systemic diseases, highlighting its role in multicomorbidity networks [9]. Understanding these comorbidities is crucial to developing preventive strategies and therapeutic interventions and reducing the socioeconomic burden.

This review synthesizes the latest omics-driven insights into osteoporosis pathogenesis, explores its comorbidities, and identifies future research directions to advance clinical applications and personalized management.

2. Pathogenesis

Through osteogenic differentiation, osteoblasts synthesize extracellular matrix proteins and become osteocytes, comprising more than 90% of all bone cells, embedded in the mineralized bone matrix. Osteoclasts resorb the bone matrix by adhering to the bone [10]. Osteogenic differentiation is influenced by a variety of signaling pathways, including Wnt, bone morphogenetic proteins (BMPs), transforming growth factor- β (TGF- β), hedgehog, parathyroid hormone (PTH), fibroblast growth factors (FGFs), Notch, and others. The key transcription factors involved include Runt-related transcription factor 2 (*Runx2*), osterix (*Osx*), β -catenin, activating transcription factor 4 (*Atf4*), etc. [11]. Osteoclasts are differentiated from hematopoietic stem cells (HSCs), a process which is stimulated by monocyte/macrophage colony-stimulating factor (M-CSF) and the activation of the receptor activator of nuclear factor kappa B (RANK) with its ligand (RANKL). Osteoprotegerin (OPG) is a soluble receptor that competes with RANK to bind RANKL, thereby limiting osteoclastogenesis [12]. Osteoporosis is caused by a breakdown in the balance between bone resorption and bone formation [12]. Thus, the disruption of the cells, molecules, and signals involved in bone formation and resorption may lead to osteoporosis. Given that osteoporosis is a multifactorial disease, we summarized several pathologic mechanisms.

2.1. Endocrinology

Estrogen deficiency is recognized as the predominant etiological factor in postmenopausal osteoporosis. The expression of estrogen receptors (ERs), including the ER α and ER β isoforms, in osteoblasts, osteoclasts, and osteocytes underscores the multifaceted role of estrogen in bone homeostasis. Estrogen signaling exerts anabolic effects on bone through diverse molecular pathways. Specifically, estrogen, by binding to ER α , can augment Wnt/ β -catenin and BMP signaling to increase osteogenesis [13]. Additionally, estrogen extends the life span of osteoblasts by inhibiting apoptosis [14]. Schiavi et al. demonstrated that estrogen receptor inhibition significantly reduces fibronectin expression (FN1), impairing matrix production in the absence of mechanical stimulation [15].

Estrogen also modulates osteoclast activity. In estrogen-deficient murine models, elevated levels of pro-inflammatory cytokines, including tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1), and interleukin-6 (IL-6), drive osteoclast maturation and bone resorption [16]. Estrogen suppresses the RANKL- and M-CSF-induced differentiation of myelomonocytic precursors into osteoclasts [17] and enhances osteoblast-derived TGF- β production, which promotes osteoclast apoptosis [18]. Osteocytes, which are mechanosensitive cells, exhibit altered responsiveness to mechanical stimuli under estrogen regulation. Estrogen modulates fluid flow-induced intracellular calcium oscillations in osteocytes, potentially influencing their differentiation [19]. Furthermore, osteocytes regulate osteoclast activity via RANKL production [20].

Estrogen glucocorticoids exhibit dose-dependent effects on bone metabolism. At physiological levels, glucocorticoids stimulate osteoblasts to secrete Wnt ligands (e.g., Wnt7b, Wnt10b, and Wnt9a), promoting the osteogenic differentiation of mesenchymal stem cells (MSCs) [21]. Osteoblast-specific glucocorticoid knockout models exhibit delayed calvarial bone development and impaired mineralization [22]. However, excessive glucocorticoid exposure upregulates peroxisome proliferator-activated receptor gamma receptor 2 (PPAR γ 2), shifting osteogenic differentiation toward adipogenesis [23]. Glucocorticoids inhibit Wnt16 expression in osteoblasts and attenuate the Wnt3a-mediated activation of β -catenin signaling [24,25]. They also upregulate sclerostin (*Sost*) expression in osteocytes, further suppressing the Wnt pathway [23]. Glucocorticoids enhance osteoclastogenesis by increasing M-CSF and RANKL expression while inhibiting OPG [26]. Paradoxically, they prolong osteoclast life span and induce osteocyte apoptosis [22]. Ad-

ditionally, glucocorticoids stimulate osteocytes to produce Wnt antagonists, including sclerostin and Dickkopf-related protein 1 (DKK1) [23].

Estrogen glucocorticoid hyperthyroidism and hyperparathyroidism are well-established causes of secondary osteoporosis. Thyroxine is critical to bone development and the maintenance of adult bone mass [27]. It accelerates bone turnover, shortening the remodeling cycle [28]. The authors of a two-sample Mendelian randomization analysis identified a causal relationship between hyperthyroidism and osteoporosis in European populations [29]. Thyroid hormone (T3) stimulates osteoblastic bone marrow stromal cells and osteoblast cell lines to produce IL-6 and IL-8, which promote osteoclastogenesis [30]. Despite advances in understanding the thyroid hormone's role in bone turnover, its precise molecular mechanisms remain elusive. Hyperparathyroidism activates osteoclast activity, primarily affecting cortical bone, as evidenced by reduced BMD in the distal forearm and hip [31]. Trabecular bone may also be impacted, although the underlying mechanisms warrant further investigation [32]. These hormones change bone mass by influencing osteogenesis or osteogenesis (Figure 1).

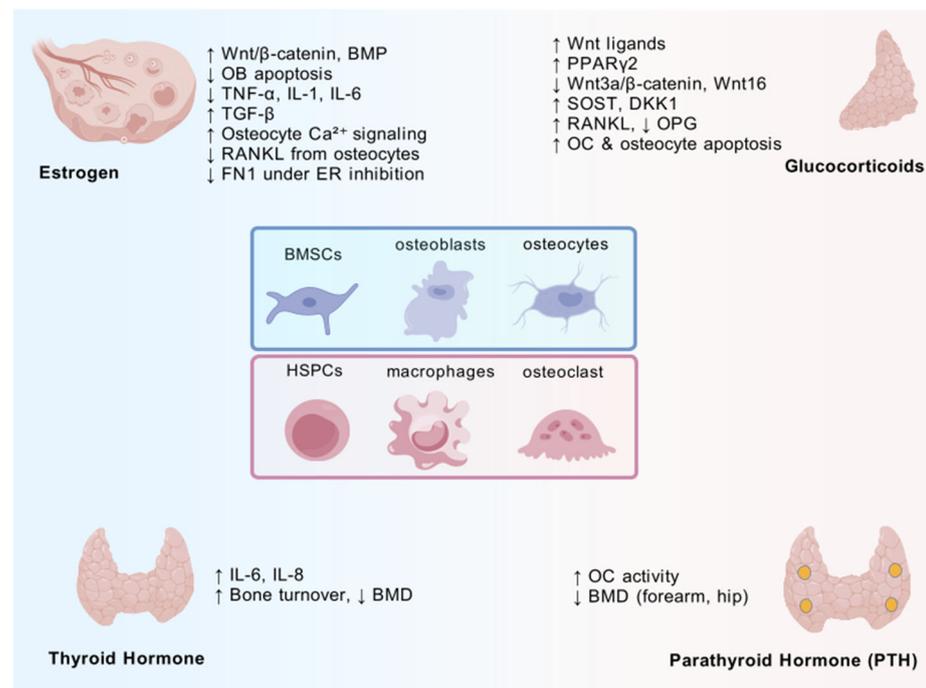


Figure 1. Hormonal regulation of bone cells contributing to osteoporosis. Estrogen deficiency, glucocorticoid excess, thyroid hormone excess, and elevated parathyroid hormone disrupt bone remodeling via cell-type-specific mechanisms. Key pathways and literature references are annotated. Created with biogdp.com [33].

2.2. Oxidative Stress and Inflammation

Oxidative stress and inflammation are central to the pathogenesis of osteoporosis, driving bone loss through their interplay and independent mechanisms. Oxidative stress, characterized by excessive reactive oxygen species (ROS) and reactive nitrogen species (RNS) [34], arises from mitochondrial dysfunction and cellular oxygen metabolism [35]. This process impairs bone formation by reducing osteoblastogenesis, increasing osteoblast and osteocyte apoptosis, and enhancing osteoclastogenesis [34]. Pathological conditions such as postmenopause [36], aging [37], hyperglycemia [38], and elevated fatty acids [39] exacerbate ROS accumulation, further compromising bone health.

Hydrogen peroxide (H₂O₂), a model for oxidative stress, inhibits the osteogenic differentiation of bone marrow stromal cells (BMSCs) by suppressing Wnt signaling and osteogenic markers while inducing osteoblast apoptosis [40,41]. The PI3K/AKT/mTOR

and JNK pathways are critical mediators of oxidative stress-induced osteoblast dysfunction [42,43]. Conversely, ROS promote osteoclastogenesis by activating NF- κ B and upregulating osteoclast markers such as c-Fos and NFATc1 [44]. The Nrf2/Keap1/ARE pathway counteracts ROS, decreasing bone resorption and oxidative damage [45]. Targeting these pathways with antioxidants offers therapeutic potential [17].

Inflammation, particularly chronic low-grade inflammation in aging, and oxidative stress exert the synergistic effect of exacerbating osteoporosis [46]. ROS amplify pro-inflammatory signaling, while inflammation-induced mitochondrial dysfunction further increases oxidative stress [47,48]. Key cytokines, including TNF- α , IL-1, IL-6, and IL-17, drive osteoclastogenesis by activating the NF- κ B and MAPK pathways, increasing RANKL and M-CSF and suppressing OPG [36]. IL-1 and IL-18 further enhance RANKL expression, while TNF- α inhibits Wnt signaling via DKK1 [36]. These findings highlight the dual role of inflammation in directly promoting bone resorption and indirectly amplifying oxidative stress.

Targeting oxidative stress and inflammation represents a promising strategy for osteoporosis treatment. Antioxidants that activate the Nrf2 pathway or reduce ROS production can mitigate bone loss [49]. Similarly, anti-inflammatory agents that inhibit TNF- α , IL-1, or IL-17 signaling may restore bone homeostasis [36]. Combined therapies addressing both pathways could offer synergistic benefits, particularly in aging and postmenopausal osteoporosis.

2.3. Cellular Senescence

Cellular senescence, marked by irreversible growth arrest and the senescence-associated secretory phenotype (SASP), is a key driver of aging-related osteoporosis [50]. The SASP, comprising pro-inflammatory cytokines (e.g., IL-6 and IL-8) and immunomodulatory factors, disrupts bone homeostasis by promoting inflammation and impairing remodeling [51]. In the aging-bone microenvironment, senescent osteocytes, myeloid cells, and other bone-resident cells accumulate, exacerbating bone loss through SASP-mediated mechanisms [52,53].

Senescent cells disrupt bone homeostasis by enhancing osteoclast activity and suppressing osteoblast function. The selective elimination of senescent cells or the inhibition of the SASP preserves trabecular and cortical bone mass, highlighting their role in age-related osteoporosis [54]. Aging shifts the differentiation of BMSCs from osteogenic to adipogenic, as evidenced by the reduced expression of osteogenic markers (*Runx2*, *Dlx5*, collagen, and osteocalcin) and increased adipogenic regulator PPAR- γ 2 levels [17]. This imbalance is driven by diminished TGF- β activity and altered BMP signaling, favoring adipogenesis over osteogenesis [55]. Targeting this transition offers a potential therapeutic avenue for osteoporosis.

Mitochondrial dysfunction and impaired mitophagy underpin cellular senescence. Senescent cells exhibit elevated ROS production, accelerating telomere shortening and promoting SASP-driven inflammation [56–58]. Sirtuin-3 (Sirt3), a mitochondrial regulator, mitigates BMSC senescence by enhancing mitochondrial function and mitophagy, offering a promising strategy to counteract bone loss [59,60]. Senolytics, SASP inhibitors, and mitochondrial modulators (e.g., Sirt3 activators) represent novel interventions for osteoporosis. These approaches target senescent cells and their inflammatory effects, restoring bone homeostasis in aging populations.

We can observe that oxidative stress, inflammation, and senescence interact with each other. Mutually influencing factors are briefly described below (Figure 2).

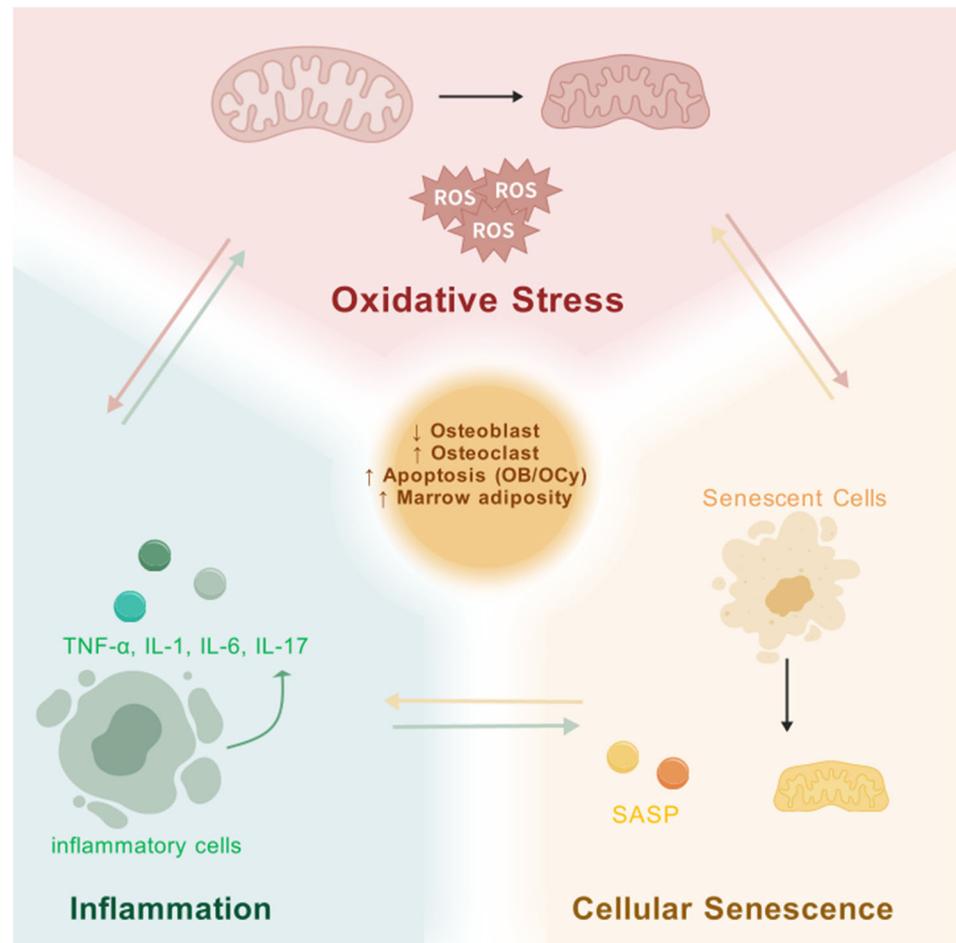


Figure 2. Interplay of Oxidative Stress, Inflammation, and Cellular Senescence in Osteoporosis. These interconnected processes synergistically impair bone homeostasis by enhancing osteoclast activity, suppressing osteoblast function, and promoting bone marrow adiposity. Mitochondrial dysfunction and ROS amplification link all three mechanisms, representing promising therapeutic targets. Created with biogdp.com [33].

2.4. Osteoimmunology in Osteoporosis: The Interplay Between Bone and Immune Cells

Osteoimmunology explores the intricate crosstalk between bone biology and immunology, emphasizing the role of immune cells in bone remodeling. Lymphocytes, particularly T and B cells, are critical mediators in this process, influencing bone homeostasis through cytokine production and interactions with the RANK/RANKL/OPG axis.

Resting T cells maintain bone mass by supporting OPG production, as evidenced by osteoporosis in T-cell-deficient nude mice [61]. However, activated T cells promote osteoclastogenesis by secreting pro-osteoclastic cytokines such as RANKL and TNF- α . Among the T-cell subsets, Th17 cells are the most potent pro-osteoclastogenic CD4⁺ cells, producing IL-17A, IL-17F, IL-21, and IL-22 [62]. IL-17 drives osteoclastogenesis by inducing RANK expression [17], while Th17 cells also secrete IL-1, IL-6, TNF, and IFN- γ , further enhancing osteoclast activity [63]. Th17 cells are implicated in various bone diseases, highlighting their pathological significance [62]. Natural killer T (NKT) cells contribute to osteoclastogenesis by producing IFN- γ , which activates macrophages and T cells to secrete TNF- α , and by directly secreting RANKL and M-CSF [64]. In contrast, CD8⁺ T cells exert a bone-protective role by secreting OPG alongside RANKL [65], while regulatory T (Treg) cells inhibit osteoclastogenesis by suppressing RANKL and M-CSF production, thereby increasing bone volume [66].

B cells play a dual role in bone remodeling. In postmenopausal osteoporosis, B cells exhibit increased surface RANKL expression, contributing to bone loss. Mice lacking RANKL in B cells are partially protected from ovariectomy-induced osteoporosis [67]. Conversely, B-cell knockout (KO) mice develop osteoporosis due to reduced OPG levels, underscoring B cells as a primary source of OPG under physiological conditions [68]. The ability of B cells to regulate the RANKL/OPG balance offers a novel perspective on the pathogenesis of osteoporosis.

Targeting immune cell-mediated pathways, such as Th17 cell activity or the RANK/RANKL/OPG axis, represents a promising strategy for osteoporosis treatment. Modulating Treg cell function or enhancing B-cell-derived OPG production could restore bone homeostasis, particularly in postmenopausal osteoporosis.

2.5. Gut Microbiome

The gut microbiome (GM) is a key regulator of bone metabolism, influencing the “microbiota–skeletal” axis through nutrient absorption, immune modulation, and microbial metabolites [69]. Germ-free (GF) mice exhibit increased bone mass, highlighting the GM’s role in bone homeostasis [70]. Interventions such as probiotics and antibiotics can modulate bone mass by altering GM composition [71,72]. The GM enhances the absorption of essential minerals (Ca, Mg, and P) critical to bone mineralization [73]. The microbial fermentation of dietary fiber produces short-chain fatty acids (SCFAs), which improve calcium absorption and promote osteogenic differentiation while inhibiting osteoclast activity [74]. The GM modulates the mucosal immune system, influencing bone remodeling via the GM–immune–bone axis [75]. For example, *Bacillus clausii* enhances Treg cell activity, while SCFAs, particularly butyrate, promote Treg cell generation [76]. Parathyroid hormone (PTH) combined with butyrate induces Treg cells, stimulating CD8+ T cells to produce the osteogenic factor Wnt10b [77]. However, in hyperparathyroidism, PTH expands pro-osteoclastic TNF+ T and Th17 cells, driving bone loss [78]. Targeting the GM through dietary interventions (e.g., prebiotics and probiotics) or microbial metabolites (e.g., SCFAs) offers promising strategies for osteoporosis prevention and treatment.

2.6. Epigenetic Regulation

Epigenetics plays a pivotal role in bone homeostasis, regulating osteoblast and osteoclast differentiation through DNA methylation and histone modifications [79]. Aberrant epigenetic changes are increasingly implicated in osteoporosis pathogenesis, offering new avenues for therapeutic intervention.

DNA methylation patterns in osteoporosis patients reveal key insights into bone biology. In whole-blood analyses, differential methylation in genes such as *ABLIM2*, *CDKL5*, *RHOJ*, *PDCD1*, and *ZNF267*, with hypermethylation in *ABLIM2*, *CDKL5*, *RHOJ*, and *PDCD1* and hypomethylation in *ZNF267* were identified [80]. The hypermethylation of *BMP2* downregulates osteoblast markers, while *RUNX2* and *SP7* exhibit reduced methylation during osteoblastic differentiation [80]. Osteogenic lineage-specific genes (*Dlx5*, *Runx2*, *Bglap*, and *Osterix*) are demethylated in osteogenic differentiation, as is osteocalcin (OCN), promoting osteogenesis [81,82]. In osteoporotic fractures, lower *RANKL* promoter methylation and higher *OPG* methylation correlate with increased *RANKL* expression and decreased *OPG* expression, which enhance bone resorption [83].

Histone modifications, including acetylation, methylation, phosphorylation, ubiquitylation, and SUMOylation, regulate gene expression in bone metabolism [84]. The inhibition of EHMT2-mediated H3K27 methylation suppresses RANKL-induced osteoclast differentiation [85]. KDM5A reduces H3K4me3 at the *Runx2* promoters, inhibiting BMP2-induced osteogenesis, while Ash11 enriches H3K4me3 at the *Osx*, *Runx2*, *Hoxa10*, and *Sox9* promot-

ers to promote osteogenesis [86]. GCN5 and PCAF enhance osteogenic differentiation by acetylating H3K9 at the Wnt (*Wnt1*, *Wnt6*, *Wnt10a*, and *Wnt10b*) and BMP pathway (*BMP2*, *BMP4*, *SMAD1*, *BMPI1B*, and *RUNX2*) gene promoters, respectively [87,88].

In addition, there are also interrelationships among osteoimmunology, the gut microbiome, and epigenetics. The relevant mechanism and connection are presented in Figure 3. Beyond these, osteoporosis is influenced by genetic, nutritional, and lifestyle factors. Genetic factors account for up to 85% of peak bone mass variance, with parental fractures predicting offspring fracture risk [89]. Vitamin D supplementation reduces bone turnover and increases BMD [90]. Gut microbiota-derived short-chain fatty acids (SCFAs), such as butyrate, promote Treg cell generation, which modulates bone metabolism and inflammatory balance [91]. Epigenetic modifications also affect plasma 25-hydroxyvitamin D3 [25(OH)D] levels and cellular senescence in bone cells [92].

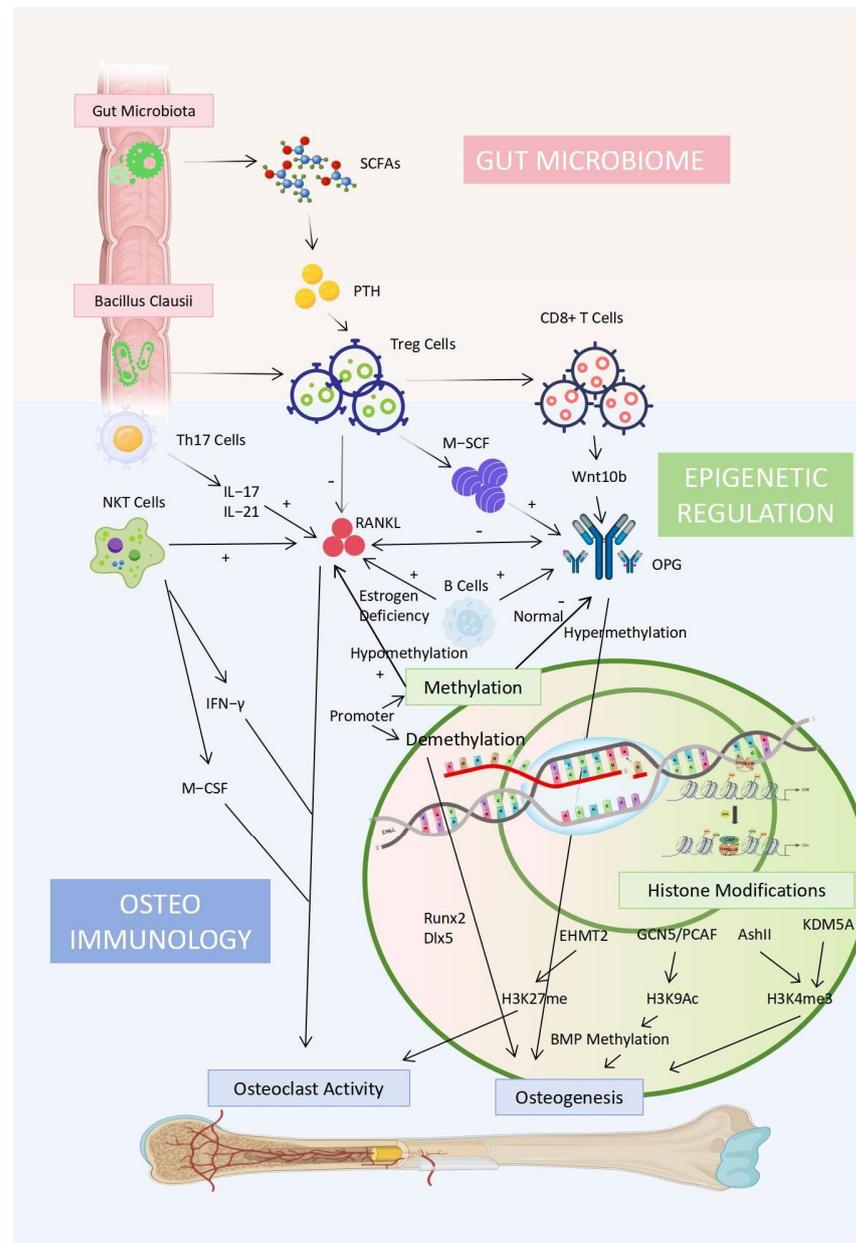


Figure 3. Interplay between osteoimmunology, gut microbiome, and epigenetic regulation in osteoporosis. This diagram summarizes the key mechanisms linking immune responses, gut microbiota, and epigenetic modifications in the regulation of bone metabolism.

3. Omics of Osteoporosis

3.1. Genomics

Osteoporosis is a multifactorial disease with a strong genetic component, as evidenced by the 60–80% heritability of BMD [93]. GWASs have allowed for the identification of over 1100 loci influencing BMD, though the causal genes for many remain elusive [94]. Key genetic pathways, including the Wnt/ β -catenin and RANKL/RANK/OPG signaling cascades, are central to bone homeostasis and remodeling. Understanding these genetic underpinnings offers significant potential for diagnosis, risk stratification, and targeted therapies.

Type I collagen, encoded by *COL1A1* and *COL1A2*, is a major component of the bone matrix. Mutations in these genes, such as the *COL1A1* Sp1 polymorphism (rs1800012), are associated with reduced BMD and increased fracture risk [95]. Rare missense mutations in *COL1A2* (p.Gly496Ala and p.Gly703Ser) have also been linked to osteoporosis [96].

The *LRP5* gene, a coreceptor in the Wnt signaling pathway, is critical to bone formation. Loss-of-function mutations, such as V667M and A1330V, are associated with reduced BMD, vertebral fractures, and early-onset osteoporosis [97–99].

The estrogen receptor gene *ESR1* regulates bone mass through estrogen signaling. Polymorphisms such as rs9340799 (XbaI) are associated with BMD variations and fracture risk in postmenopausal women, though findings across populations are inconsistent [100,101].

SOST encodes sclerostin, a Wnt pathway inhibitor. SNPs such as rs1513670 and rs7220711 are linked to low-trauma fractures and BMD [102]. The RANKL/RANK/OPG pathway, regulated by *TNFRSF11A* and *TNFRSF11B*, influences osteoclast activity. SNPs in these genes (e.g., rs3018362) are associated with BMD and osteoporosis risk [103].

GWASs have not only allowed for the identification of genetic risks but have also facilitated the discovery of therapeutic targets. Five of the eight anti-osteoporosis drugs in clinical use or trials, including denosumab and sclerostin inhibitors, were directly identified with GWASs [104]. Genetic support for drug mechanisms doubles the likelihood of clinical trial success, underscoring the translational potential of genomics [105]. GWASs enable the identification of high-risk individuals and personalized treatment strategies. For instance, genetic loci associated with medication-related osteonecrosis of the jaw (MRONJ) and atypical femoral fractures in bisphosphonate users have been identified, highlighting the utility of genetic screening in mitigating adverse effects [106,107].

3.2. Transcriptomics

Transcriptomics, the study of gene expression differences across biological states, has evolved from microarrays to RNA sequencing, offering unprecedented insights into osteoporosis pathogenesis [108]. By analyzing transcriptional profiles, the key genes and regulatory networks involved in bone formation and resorption have been identified, providing novel therapeutic targets and diagnostic biomarkers [93].

Transcriptomic studies highlight the roles of *RUNX2*, *RANKL/OPG*, and *SOST* in bone turnover disorders. For instance, male idiopathic osteoporosis patients exhibit reduced expression of *WNT10B*, *RUNX2*, *RANKL*, and *SOST* in iliac crest biopsies compared with healthy controls [109]. Reduced *RUNX2*, *SP7*, and *SOST* expression has also been observed in osteoporotic femoral neck and head tissues [110]. *RUNX2*, a master transcription factor for osteoblast differentiation, regulates critical signaling pathways, including Wnt, FGF, and hedgehog, to promote osteoblast proliferation and lineage commitment [111]. Similarly, *Sp7* (Osterix), a zinc finger transcription factor, is essential to osteoblast differentiation and bone formation [112]. Conversely, increased *RANKL* expression and elevated *RANKL/OPG* ratios, biomarkers of bone resorption, are associated with osteopenia in postmenopausal women [113].

MicroRNAs (miRNAs) and long non-coding RNAs (lncRNAs) are emerging as critical epigenetic regulators of bone metabolism. miRNAs, such as miR-185, miR-139-5p, and miR-433-3p, modulate osteoblast differentiation by targeting Wnt/ β -catenin signaling components (e.g., *CTNNB1* and *FZD4*) [114–116]. miR-146a downregulation increases *OPG*, *Wnt2*, and β -catenin expression, inhibiting osteoporosis in ovariectomized rats [117]. Conversely, miR-183 and miR-451a suppress osteoblastogenesis by targeting *Smad4* and *Bmp6*, respectively [118,119].

lncRNAs, transcripts exceeding 200 nucleotides, also regulate bone metabolism. For example, lncRNA *H19* promotes osteoblast differentiation by inhibiting *Dkk4* and activating Wnt signaling [120]. *MSC-AS1* and *MALAT1* enhance osteogenic differentiation, while *CASC11* is upregulated in osteoporosis and promote osteoblast apoptosis or recurrence [121–123]. These findings underscore the potential of non-coding RNAs as diagnostic markers and therapeutic targets.

scRNA-seq provides a high-resolution view of gene expression at the cellular level, revealing unique transcriptional profiles in BMSCs, osteoblasts, and osteoclasts [93]. For instance, *CRIP1* expression is reduced in osteoporosis, and its overexpression rescues bone loss by enhancing osteogenic differentiation [124]. *LRRc17* knockdown ameliorates ovariectomy-induced bone loss, while *STRA6* promotes adipogenic over osteogenic differentiation in osteoporotic MSCs [125,126]. scRNA-seq has also allowed for the identification of distinct osteoblast subpopulations with unique gene expression profiles, offering new insights into bone cell heterogeneity [127].

Although scRNA-seq can reveal cellular heterogeneity within tissues, it remains unclear how cells interact and organize spatially. Spatial transcriptomics investigates cellular interactions within tissues while preserving their native spatial context [128]. Xue et al. found that SSPCs expressing high levels of platelet-derived growth factor receptor β (PDGFR β) and Ly6a/Sca-1 are highly enriched within the cortical bone tissue, primarily along the outer periosteal surface. This specific enrichment zone constitutes a critical niche for fracture healing [129]. Using spatial transcriptomic profiling, Jiang et al. demonstrated that the highly metastatic, triple-negative breast cancer cell line MDA-MB-231 disrupts bone homeostasis and inhibits fracture healing within key callus regions, including the hard callus, soft callus, and fibrous interzone [130]. Wang et al. applied scRNA-seq and spatial transcriptomics to primary human femoral head tissue cells. Their analysis revealed that within the inflammatory microenvironment of osteoporosis, macrophages promote osteoclastogenesis through the RETN–Cyclase-Associated Protein 1 (CAP1) complex signaling axis [131]. Furthermore, spatial transcriptomic studies have revealed mechanisms underlying progenitor zonation during embryonic osteochondral development. [132]. Currently, spatial transcriptomics techniques have been predominantly applied to osteoarthritis research; however, their application in osteoporosis studies remains limited and requires further advancement [133].

3.3. Proteomics

Proteomics, which has evolved from traditional techniques to high-throughput methods such as mass spectrometry and protein pathway arrays, complements genomics and transcriptomics by providing a comprehensive view of molecular signaling pathways [134]. This approach has allowed for the identification of novel diagnostic and prognostic biomarkers, advancing personalized precision medicine in osteoporosis. In proteomic studies, diverse specimens, including bone tissue, serum/plasma, exosomes, and bone-related cells, are utilized to elucidate disease mechanisms and therapeutic targets.

Bone tissue proteomics offers a direct reflection of osteoporosis pathology. Tandem mass tag (TMT) analysis in ovariectomized (OVX) rats revealed 91 upregulated and

42 downregulated proteins, including increased transferrin receptor (TFR1 and TFRC) and decreased ceruloplasmin (Cp) and BMP-2 [135]. REG γ expression is notably reduced in osteoporotic patients and OVX mouse models [136]. The proteomic profiling of femoral heads from postmenopausal women identified 53 downregulated and 22 upregulated proteins, with GSTP1, LAMP2, COPB1, and RAB5B being implicated in osteoporosis with iron accumulation [137]. Despite its clinical relevance, bone tissue is challenging to obtain and preserve, limiting its widespread use.

Serum and plasma proteomics provide minimally invasive, cost-effective screening options. Common bone turnover markers, such as ALP, procollagen type I N-terminal propeptide (P1NP), and C-terminal telopeptide of type I collagen (CTX), are widely used in clinical practice [8]. The authors of prospective studies have identified 22 serum proteins, including PHLA, SHBG, and APOA1, significantly correlated with BMD [138]. Vitamin D-binding protein (VDBP) levels are inversely associated with BMD [139], while RYR1, APOA1, and FETB are upregulated in osteoporotic postmenopausal women [140]. Novel plasma proteins, such as ASAHL, component C7, and tetranectin, have also been linked to BMD variations [141,142]. Peptide fragments, such as ITIH4, are downregulated in patients with high bone turnover, suggesting their role in osteoclast activity [143].

The proteomic analysis of bone-related cells, though technically challenging, offers insights into cell-targeted therapies. MSCs exhibit upregulated FBLN2 and NPR3 during osteogenic differentiation [144], and high-glucose conditions reduce osteogenesis in MSCs, with annexin A7 and fumarate hydratase having been identified as upregulated proteins [145]. Osteocyte-derived extracellular vesicles (YO-EVs) enriched with tropomyosin-1 (TPM1) enhance matrix stiffness and osteogenesis, highlighting their therapeutic potential for senile osteoporosis [146]. Differential protein expression in circulating monocytes, such as Ras Suppressor-1 (RSU1), superoxide dismutase 2 (SOD2), and glutathione peroxidase-1 (GPX1), contributes to osteoclastogenesis and BMD variation [93]. Annexin A2 (ANXA2) is upregulated in the monocytes of individuals with low BMD and is associated with hip fractures [147,148].

The extracellular bone matrix, comprising collagen and non-collagenous proteins (NCPs), plays a critical role in bone structure and function. Altered posttranslational modifications, such as increased collagen I deamidation, are linked to age-dependent fracture risk [149]. Disrupted glycosylation further compromises bone quality. Proteins such as PLS3, involved in bone mineralization under mechanical stimulation, and AnxA6, critical to extracellular matrix (ECM) mineralization, have emerged as key players in bone homeostasis [150,151]. These findings underscore the potential of bone matrix proteomics in the development of novel therapeutic strategies.

3.4. Metabolomics

Metabolomics, the study of the end products of cellular metabolism, provides critical insights into osteoporosis by elucidating disruptions in amino acid, lipid, and energy metabolism [152]. These metabolic changes are closely linked to bone remodeling and have emerged as potential biomarkers and therapeutic targets.

Postmenopausal osteoporosis is associated with disrupted amino acid metabolism. Studies in Japanese women revealed significantly lower levels of glycyl-glycine and cystine, alongside elevated hydroxyproline, in groups with low BMD. Hydroxyproline, a collagen degradation product, serves as a marker of osteoclast-mediated bone resorption [153]. Elevated glutamine and altered levels of taurine, β -alanine, and 5-hydroxycaproic acid have also been identified as potential biomarkers in diverse populations [154,155]. In European-ancestry women, γ -aminobutanoate, threonine, cysteine, taurine, and glutamic acid were significantly associated with BMD [156].

Osteoblasts and osteoclasts require substantial energy for bone remodeling. Estrogen deficiency, as in oophorectomy, increases insulin resistance and disrupts glucose metabolism, leading to elevated glucose and lactate levels [157]. Osteoclast bone resorption is energy-intensive, relying on glycolysis and oxidative phosphorylation for ATP production [158]. Disturbances in the tricarboxylic acid (TCA) cycle, such as reduced citric acid and α -ketoglutaric acid, are observed in osteoporosis and diabetes-related bone disorders [8]. Bone also regulates systemic energy metabolism through hormone secretion, creating a feedback loop between bone and energy homeostasis [159].

Lipid metabolism plays a pivotal role in bone health, with increased bone marrow fat content being associated with bone loss [8]. Elevated triglycerides (TGs) and cholesteryl esters, alongside reduced sphingomyelin, are observed in osteoporotic models [160,161]. Lipidomic studies reveal robust changes in fatty acyls, glycerolipids, and glycerophospholipids in osteoporosis [162]. Oxidized lipids and cholesterol imbalance disrupt bone homeostasis by promoting adipogenesis over osteogenesis via PPAR γ signaling and inhibiting Wnt- β -catenin pathways [163]. Targeting lipid regulation offers therapeutic potential for osteoporosis [8].

Beyond metabolomics, epigenetic studies highlight the role of DNA methylation in osteoporosis. For instance, Wnt3a-induced osteogenesis is regulated by promoter methylation status [164], and RXRA methylation influences childhood bone mass [165]. The gut microbiome, a key regulator of nutrition, metabolism, and immunity, impacts bone health through short-chain fatty acids (SCFAs), immune modulation, and miRNA regulation [166]. Probiotics and microbiome diversity restoration have shown promise in preventing postmenopausal osteoporosis [167]. The relevant omics research on osteoporosis is presented in Figure 4.

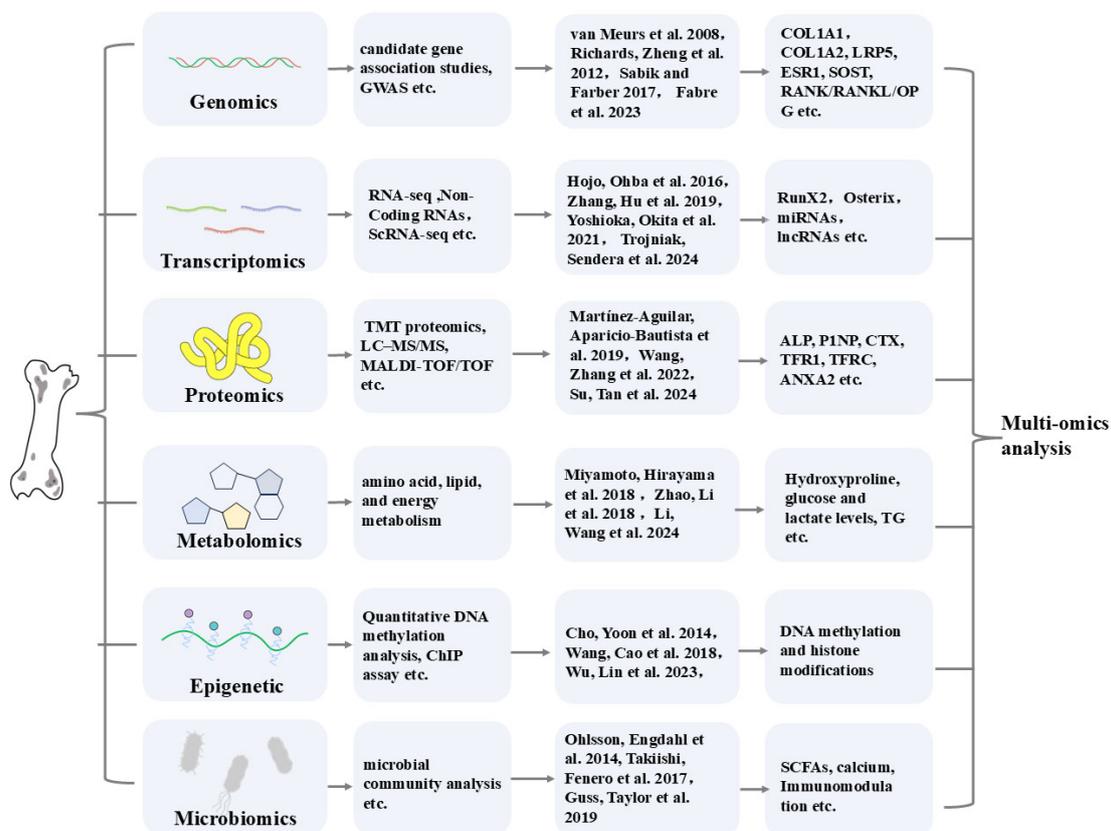


Figure 4. Omics analysis of osteoporosis biomarkers [8,71,72,80,83,96,98,99,104,112,114,121,127,135,137,139,153,162,164,166].

Multi-omics analysis integrates genomic, transcriptomic, methylomic, and metabolomic data to identify biomarkers and biological pathways associated with osteoporosis. For example, Qiu et al. identified osteoporosis biomarkers (*FADS2* and *ADRA2A*) and pathways linked to BMD variation [168]. Single-cell and transcriptome analyses revealed neutrophil genes (*DND1* and *HIRA*) as crucial players in osteoporosis development [169]. The authors of a multi-omics analysis of gut microbiota (GM) and metabolites found that the enzymes related to purine and tryptophan metabolism are involved in the progression of OP. They found that these enzymes are from Lachnospiraceae_NK4A136_group, Blautia, Rs-E47_termite_group, UCG-009, and Clostridia_UCG-014 based on gut microbiomics. In a recent study, the authors found that *USP6NL*, *SELENOT*, and *TAF1A* play an important regulatory role in the development of osteoporosis and can be used as potential therapeutic targets through the integration of Mendelian randomization, single-cell RNA sequencing, and bioinformatic analyses [170]. Multi-omics approaches account for genetic, environmental, and lifestyle factors, advancing precision medicine [171].

4. Osteoporosis and Comorbidities

Emerging evidence highlights the complex interplay between osteoporosis and chronic conditions such as cardiovascular diseases and immune disorders, mediated by shared pathways including chronic inflammation, oxidative stress, and immunometabolic crosstalk. These interactions not only exacerbate bone fragility but also position osteoporosis as a sentinel indicator of multisystem aging and dysfunction.

4.1. Cardiovascular Disease

Epidemiological evidence consistently demonstrates a significant association between osteoporosis and cardiovascular disease (CVD). Studies have shown that postmenopausal women with osteopenia or osteoporosis exhibit higher coronary calcium scores, a marker of atherosclerosis, than controls [172]. Furthermore, lower BMD has been strongly linked to an increased risk of coronary artery stenosis [173]. Elevated homocysteine levels, which negatively correlate with BMD, further underscore this relationship [174]. These findings collectively suggest that osteoporosis and CVD share a bidirectional pathophysiological connection.

The interplay between osteoporosis and CVD is underpinned by shared risk factors, including estrogen deficiency, aging, sedentary lifestyle, and diabetes [173]. At the molecular level, cytokines and signaling pathways such as BMPs, OPG, and Wnt signaling play dual roles in bone metabolism and cardiovascular health. BMPs, known for promoting osteogenic differentiation, also contribute to vascular calcification through pro-inflammatory and pro-oxidative effects, particularly in hyperhomocysteinemia [175]. Similarly, OPG, which inhibits osteoclastic bone resorption, is paradoxically associated with atherosclerosis severity, though it may protect against vascular calcification in certain contexts [176]. Wnt signaling, crucial to bone formation, also exacerbates vascular calcification via the upregulation of *Msx-2* and the downregulation of the Wnt inhibitor *Dkk1* [177]. Pro-inflammatory cytokines such as IL-6 and TNF- α , which drive bone resorption, are also pivotal to the development of atherosclerosis, highlighting the inflammatory axis as a common pathway [173].

Calcium balance is essential to maintaining bone health, with inadequate intake increasing the risk of osteoporosis and fractures. While dietary calcium positively correlates with BMD and reduces bone loss [178], excessive calcium supplementation has been linked to increased coronary artery calcification (CAC) and elevated cardiovascular mortality [179]. Notably, dietary calcium intake appears safer than supplementation in

terms of cardiovascular risk [180]. This dichotomy underscores the need for individualized calcium management in osteoporosis patients.

Pharmacological interventions for osteoporosis also carry cardiovascular implications. Romosozumab, which activates Wnt signaling to enhance bone formation, has been associated with serious cardiovascular adverse events [181]. Conversely, bisphosphonates may reduce atherosclerosis and vascular calcification, though they pose a short-term risk of atrial fibrillation [182]. Thus, the selection of anti-osteoporosis therapies must carefully balance bone health benefits with cardiovascular safety.

4.2. Respiratory Diseases

Clinical evidence underscores a significant association between respiratory diseases and osteoporosis, with patients exhibiting a heightened risk of osteopenia and osteoporotic fractures [183]. Chronic obstructive pulmonary disease (COPD), in particular, is strongly linked to osteoporosis, with disease severity correlating with increased bone loss [184]. The pathogenesis of respiratory disease-induced osteoporosis is multifactorial, involving chronic hypoxia and prolonged use of corticosteroids [185].

Dyspnea, a hallmark of respiratory diseases, leads to chronic hypoxia, a key driver of osteoporosis [186]. Hypoxia-induced reactive oxygen species (ROS) accumulation triggers oxidative stress and inflammation, both implicated in bone resorption [187]. Hypoxia-inducible factor-1 alpha (HIF-1 α) plays a dual role in this process. On one hand, HIF-1 α protects bone cells by inhibiting mitochondrial apoptosis and promoting angiogenesis, essential to bone formation [188]. On the other hand, HIF-1 α stimulates osteoclast differentiation via the upregulation of RANKL and NFATc1, exacerbating bone resorption [189]. Thus, maintaining physiological levels of HIF-1 α is critical to bone homeostasis.

Corticosteroids, a mainstay in respiratory disease management, significantly impact bone health. While inhaled corticosteroids are effective in controlling symptoms and preventing disease exacerbations, their long-term use has been associated with an increased fracture risk in COPD patients [190]. However, conflicting evidence suggests that fractures may be more closely tied to the underlying disease pathology rather than corticosteroid use itself [191]. Oral corticosteroids, often prescribed for severe respiratory conditions, pose a greater threat, with bone loss being the most pronounced within the first 3–6 months of treatment [192]. These findings highlight the need for careful consideration of corticosteroid regimens to minimize adverse skeletal effects.

Emerging evidence suggests that osteoporosis may conversely exacerbate respiratory disease outcomes. Longitudinal studies have demonstrated that reduced femoral neck bone mineral density (FNBMD) is associated with increased COPD mortality, with men experiencing a higher risk than women [193]. These findings emphasize the bidirectional nature of the relationship, underscoring the importance of addressing bone health in respiratory disease management.

4.3. Osteoarthritis

The relationship between osteoarthritis (OA) and osteoporosis (OP) has been a subject of debate. Early studies indicated that OA was associated with higher bone density, with Radin and Rose, in 1986, proposing that OA strengthens subchondral bone, thereby increasing its resistance to deformation [194]. This hypothesis was supported by findings that OA patients tend to develop fragility fractures later than those without OA [195]. However, more recent research has yielded conflicting results regarding the connection between OA and OP. Some studies indicate that higher bone density may reduce OA progression risk [196], while others suggest that genetically predicted low BMD may increase OA susceptibility [197]. In contrast, separate findings suggest that OP might

reduce OA incidence and that genetic predisposition to OP negatively correlates with knee OA [196]. A meta-analysis concluded that OA neither protects against OP nor increases its risk beyond that of age- and sex-matched controls, with discrepancies likely due to disease location and progression stage [198,199].

Despite their apparent differences, OA and OP share several pathophysiological mechanisms. Inflammation is a major contributing factor to both conditions. Pro-inflammatory cytokines such as IL-1 β , TNF- α , and IL-6 contribute to both OP-related bone loss and OA-associated cartilage degradation [194]. Age-related chronic inflammation (“inflammaging”) further exacerbates these processes by promoting oxidative stress and joint deterioration [200]. Estrogen deficiency is another common factor, as it disrupts cartilage metabolism, accelerates OA progression, and weakens muscle strength, thereby reducing joint stability [201]. At the genetic level, the Wnt signaling pathway plays a role in both OA cartilage development and OP-related bone metabolism [202]. Additionally, gut microbiota-derived inflammatory mediators may contribute to low-grade systemic inflammation, further promoting OA progression [203].

Osteopontin (OPN) plays a pivotal role in both bone and joint health by regulating the activity of chondrocytes, synoviocytes, osteoclasts, and osteoblasts [204]. Elevated OPN levels in plasma, synovial fluid, and cartilage correlate with OA severity and progression [93]. Similarly, OPN is a recognized risk factor for postmenopausal OP and serves as an early diagnostic biomarker for the disease [204]. OPN promotes inflammation and contributes to OA and OP progression by stimulating the production of TNF- α and IL-6, further linking these two conditions [152].

Recognizing the interplay between OA and OP is crucial to optimizing treatment. OA patients require strategies to maintain bone mass and prevent fractures, while OP patients should prioritize joint stability and the early detection of OA symptoms [205]. Beyond conventional approaches such as exercise, calcium, and vitamin D supplementation, several pharmacological interventions show promise for addressing both conditions: Bisphosphonates, widely used for OP, also reduce subchondral bone turnover, mitigate bone loss, and preserve cartilage integrity in OA [206]. Denosumab inhibits NF- κ B signaling, reducing chondrocyte apoptosis, subchondral bone remodeling, and cartilage degradation, suggesting its potential role in slowing OA progression [207]. Tocotrienols (TTs), a form of vitamin E, have demonstrated benefits in animal models by improving bone density, reducing oxidative stress and inflammation, and protecting cartilage from degradation [208].

4.4. Tumor

The relationship between cancer and osteoporosis has garnered increasing attention in recent years. Cancer and its treatments, particularly chemotherapy, radiation, and hormone therapy, not only affect bone density directly but also exacerbate osteoporosis by altering bone metabolism. In hormone-dependent cancers such as breast cancer, reduced estrogen levels increase bone resorption, contributing to osteoporosis [209]. Chemotherapy and radiotherapy, commonly used to treat cancer, further accelerate osteoporosis by directly damaging bone cells, especially osteoblasts and osteoclasts [210].

Beyond treatment-related effects, certain tumors, particularly non-small-cell lung cancer (NSCLC), can induce osteoporosis by secreting bone resorption factors or osteoclast-promoting signals [211]. For example, elevated osteopontin (OPN) expression in NSCLC patients is linked to the development of bone metastases [212]. Studies indicate that reducing OPN expression may help inhibit osteoclast differentiation [213], suggesting that tumor cells promote osteoporosis by altering bone metabolism and may worsen bone destruction via metastasis.

Bone metastasis often results in osteolytic or osteoblastic lesions, severely impacting patients' quality of life [214]. This process involves complex interactions between tumor cells and the bone microenvironment, disrupting normal bone remodeling. Verbruggen et al. [215] identified a novel mechanism in breast and prostate cancers whereby osteocytes inhibit tumor cell proliferation through TNF- α secretion. However, tumor cells counteract this inhibition by secreting TGF- β , creating a feedback loop that accelerates metastasis. Such feedback loops are critical to understanding how tumor–osteocyte interactions promote metastasis.

Dumanskiy et al. [216] found that highly metastatic tumors, such as breast and prostate cancer, exacerbate bone metabolism disorders by secreting bone resorption-promoting factors such as OPN and RANKL. Similarly, metabolic reprogramming in the tumor–bone microenvironment has been proposed as a key driver of cancer progression. Whitburn and Edwards [217] reviewed these metabolic adaptations, noting that osteolytic and osteoblastic lesions in breast and prostate cancer are associated with the dysregulation of bone resorption and formation. The altered microenvironment, due to osteoblast and osteoclast metabolism, supports tumor cell survival and growth.

Fan et al. [218] explored osteoclast–cancer cell metabolic crosstalk and its role in resistance to PARP inhibitors in bone metastasis. They found that glutamine metabolism is crucial to cancer cell survival under treatment stress. Moreover, Verbruggen et al. [219] showed that osteocyte paracrine signaling can inhibit tumor cell proliferation and invasion. However, this effect was reversed in breast cancer cells when osteocytes were subjected to mechanical stimulation, emphasizing the role of the mechanical microenvironment in bone metastasis.

The activation of immune cells has also been implicated in bone homeostasis disruption, particularly in cancer patients. In postmenopausal women with low estrogen levels, a low-grade chronic inflammatory state further intensifies bone resorption and accelerates osteoporosis [220].

In summary, the metabolic impact of tumor cells on osteoporosis involves both direct factors from the tumor and the side effects of cancer treatment. Tumor cells promote osteoporosis by secreting cytokines that damage bone cells and alter immune system function. Additionally, hormonal changes, immune cell activation, metabolic reprogramming, and mechanical factors induced by cancer therapies exacerbate bone metabolism disorders, further contributing to the development and progression of osteoporosis.

4.5. Neurological Disorders and Their Association with Osteoporosis

The increasing prevalence of neurodegenerative diseases, such as Alzheimer's disease (AD) and Parkinson's disease (PD), among the elderly has highlighted their strong correlation with osteoporosis. The coexistence of these conditions poses significant clinical challenges, as neurological disorders can accelerate osteoporosis through multiple mechanisms.

One major contributing factor is the reduced physical activity in patients with AD and PD due to motor dysfunction and cognitive decline, leading to decreased bone strength and increased fracture risk [221]. Beyond immobility, these diseases influence bone metabolism through the brain–bone axis. In AD, the accumulation of amyloid- β and tau protein, key hallmarks of neurodegeneration, has been implicated in osteoporosis development by modulating neuropeptide Y (NPY) expression, which impacts bone density and the trabecular architecture [222]. Similarly, dopamine deficiency in PD disrupts neurotransmitter balance, negatively affecting bone metabolism and increasing fracture susceptibility [223].

Recent studies indicate that brain-derived extracellular vesicles (BEVs) play a crucial role in mediating these effects. BEVs, which carry proteins and RNA, can travel through

the circulatory system and influence osteoblast and osteoclast function, ultimately leading to bone loss [224]. Moreover, the sympathetic nervous system is a key regulator of bone homeostasis. Its activation promotes bone resorption by stimulating osteoclast activity while inhibiting osteoblast differentiation. Neuropeptide Y signaling has been identified as a mediator of this process, shifting mesenchymal stem cell differentiation toward adipogenesis at the expense of bone formation [225]. Notably, pharmacological sympathetic blockade has been shown to preserve bone density and improve the trabecular microstructure [226].

Inflammatory pathways further link neurodegeneration and osteoporosis. In AD, elevated levels of pro-inflammatory molecules such as Prokineticin 2 (PROK2) and colony-stimulating factor 3 (CSF3) have been associated with both neuroinflammation and bone loss, suggesting a shared pathological mechanism between these diseases [227].

Conversely, osteoporosis may also influence neurological function via the bone–brain axis. Epidemiological studies indicate that osteoporosis often coexists with cognitive impairment, with fractures accelerating cognitive decline in elderly patients [228]. Bone-derived molecules such as osteocalcin not only regulate bone metabolism but also play essential roles in cognition and memory preservation [229]. Emerging research highlights the potential neuroprotective effects of extracellular vesicles secreted by young osteocytes, which may modulate AD-related pathways and support brain health [230]. Moreover, the dysregulation of the AKT signaling pathway, a key modulator of glucose metabolism, has been implicated in both osteoporosis and neurodegeneration, further reinforcing their interconnected pathophysiology [231].

The management of osteoporosis in patients with neurological disorders presents significant challenges. Many central nervous system medications, including antidepressants, antipsychotics, and antiepileptics, have been shown to adversely affect bone metabolism, exacerbating osteoporosis progression [232]. Conversely, standard osteoporosis treatments, such as bisphosphonates, may influence neurological function, particularly in neurodegenerative conditions [233]. Vitamin D supplementation has demonstrated dual benefits, improving both cognitive function and BMD in these patients, underscoring its potential as a therapeutic intervention [234].

Addressing the bidirectional relationship between neurological disorders and osteoporosis requires an integrated treatment strategy that optimizes bone and brain health while minimizing adverse effects. Future research should be focused on targeted therapies that simultaneously mitigate neurodegeneration and bone loss to improve patient outcomes.

5. Conclusions

Osteoporosis is a multifactorial disease, resulting from the combined effects of genetic, metabolic, inflammatory, and environmental factors. Epigenetics and the microbiome also play a role in bone mass regulation. The primary mechanisms involve an osteogenic–osteoclastic imbalance, which disrupts bone metabolism and facilitates the progression of osteoporosis. These etiological factors are interconnected. For instance, inflammation and oxidative stress are significant contributors to senescence and aging phenotypes [235], with aging manifestations including oxidative stress [236,237]. The authors of a previous study mentioned an oxidative stress–inflammation–aging trinity to clarify their interrelationships and highlight their synergistic effect on advancing osteoporosis [238]. In addition, the “microbiota–skeletal” axis can also influence other pathological mechanisms and contribute to bone remodeling, as the gut microbiota can regulate the immune system and produce Treg cells, which have been proven to play a role in bone remodeling. Additionally, PTH can not only act as a hormone to regulate bone development but also work in conjunction with intestinal microbiota to jointly stimulate the production of Treg cells. In recent years, it has been proved that SCFAs, especially butyrate

and propionate, could be used as potent histone deacetylases (HDACs) [239], which can repress the function of Runx2 in differentiating osteoblasts [240]. Finally, although not proven to be related to osteoporosis, senescence, and inflammation have been shown to regulate epigenetic modifications. The degree of DNA methylation varies with age, as do the epigenetic modifications of senescent cells [241]. Similarly to the expression of the pro-inflammatory cytokine IL-6, the expression of DNA methyltransferase (DNMT), which is a DNA-modifying enzyme, can be enhanced [242]; DNMT3A upregulation might contribute to SOD2 hypermethylation and increased ROS generation, which may enhance oxidative stress [243].

The advent of omics technologies has revolutionized the understanding of osteoporosis, enabling the identification of susceptibility genes and the differentiation patterns of osteoblasts and osteoclasts and diagnostic metabolites and therapeutic targets. Additionally, exploring the comorbidity mechanisms (described in Figure 5) between osteoporosis and other diseases offers novel insights and treatment strategies, while addressing comorbidities helps break the vicious cycle of mutual disease influence. Leveraging omics, particularly emerging technologies, facilitates the development of personalized diagnostic markers and fosters interdisciplinary collaboration across orthopedics, endocrinology, microbiology, and genetics, further advancing osteoporosis research and therapy.

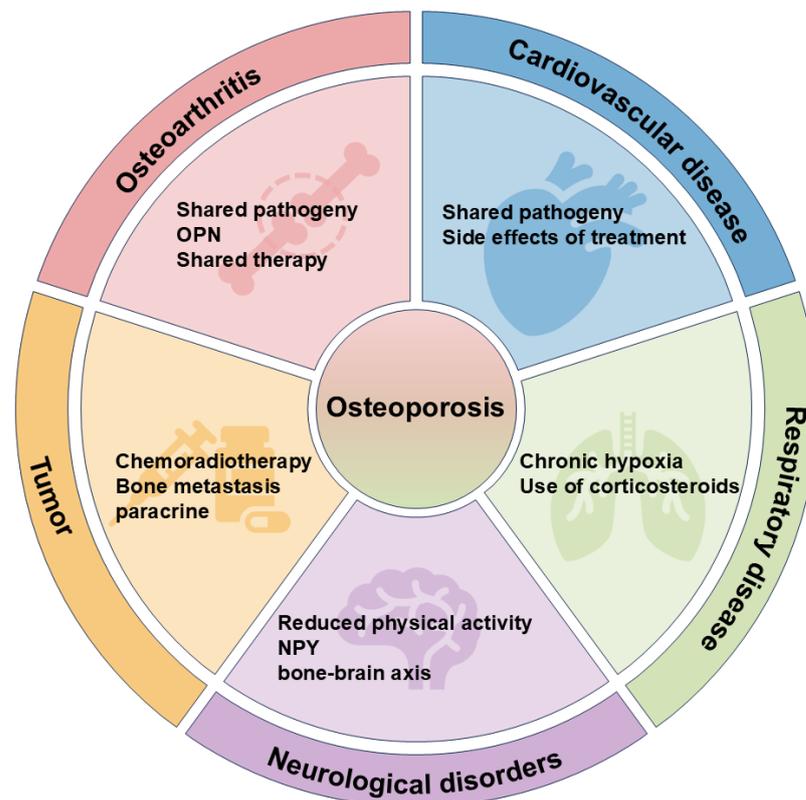


Figure 5. The comorbidity mechanism of osteoporosis with other diseases.

6. Future Perspectives

Addressing these multifaceted challenges necessitates focused future efforts. Key priorities include leveraging multi-omics integration and longitudinal studies powered by AI to decipher causal pathways and dynamic interactions over time. Unraveling the precise mechanisms of the microbiome–skeletal axis, resolving the paradoxical roles of metabolites like SCFAs, and exploring targeted epigenetic editing strategies (e.g., CRISPR, DNMT/HDAC modulators) are crucial for understanding and manipulating bone biology. Furthermore, exploiting shared pathological mechanisms with comorbidities (Figure 5),

such as chronic inflammation, offers novel therapeutic avenues to disrupt disease cycles. The ultimate translation lies in precision medicine: developing integrated diagnostic panels (genetic, epigenetic, microbiome, metabolic) for early risk stratification and personalized interventions, alongside biomarkers predicting treatment response. Overcoming these challenges through robust interdisciplinary collaboration is paramount to harnessing converging insights and advancing toward predictive, preventive, and personalized osteoporosis management.

Author Contributions: Y.Z. was responsible for the investigation in this study and drafted the original manuscript. Y.Y. (Yuzhu Yan) and H.X. mainly contributed to the conceptualization of the study, funding acquisition, and project administration, while also participating in the review and editing of the manuscript. J.W. and L.X. made significant contributions to the review and editing of the manuscript. Y.Y. (Yu Yan) and H.Z. jointly undertook funding acquisition, project administration, and supervision, and also contributed to the review and editing of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Project of Xi'an Science and Technology (No. 22YXYJ0041) and the Youth Cultivation Project of Xi'an Health Commission (No. 2025qn08).

Conflicts of Interest: The authors do not have any competing interests regarding this study.

References

1. Johnston, C.B.; Dagar, M. Osteoporosis in Older Adults. *Med. Clin. N. Am.* **2020**, *104*, 873–884. [[CrossRef](#)]
2. Wright, N.C.; Looker, A.C.; Saag, K.G.; Curtis, J.R.; Delzell, E.S.; Randall, S.; Dawson-Hughes, B. The recent prevalence of osteoporosis and low bone mass in the United States based on bone mineral density at the femoral neck or lumbar spine. *J. Bone Miner. Res.* **2014**, *29*, 2520–2526. [[CrossRef](#)] [[PubMed](#)]
3. Wang, J.; Shu, B.; Tang, D.Z.; Li, C.G.; Xie, X.W.; Jiang, L.J.; Jiang, X.B.; Chen, B.L.; Lin, X.C.; Wei, X.; et al. The prevalence of osteoporosis in China, a community based cohort study of osteoporosis. *Front. Public Health* **2023**, *11*, 1084005. [[CrossRef](#)]
4. Clynes, M.A.; Harvey, N.C.; Curtis, E.M.; Fuggle, N.R.; Dennison, E.M.; Cooper, C. The epidemiology of osteoporosis. *Br. Med. Bull.* **2020**, *133*, 105–117. [[CrossRef](#)] [[PubMed](#)]
5. Salari, N.; Ghasemi, H.; Mohammadi, L.; Behzadi, M.H.; Rabieenia, E.; Shohaimi, S.; Mohammadi, M. The global prevalence of osteoporosis in the world: A comprehensive systematic review and meta-analysis. *J. Orthop. Surg. Res.* **2021**, *16*, 609. [[CrossRef](#)]
6. Odén, A.; McCloskey, E.V.; Kanis, J.A.; Harvey, N.C.; Johansson, H. Burden of high fracture probability worldwide: Secular increases 2010–2040. *Osteoporos. Int.* **2015**, *26*, 2243–2248. [[CrossRef](#)]
7. Dong, Y.; Zhang, Y.; Song, K.; Kang, H.; Ye, D.; Li, F. What was the Epidemiology and Global Burden of Disease of Hip Fractures From 1990 to 2019? Results From and Additional Analysis of the Global Burden of Disease Study 2019. *Clin. Orthop. Relat. Res.* **2023**, *481*, 1209–1220. [[CrossRef](#)] [[PubMed](#)]
8. Li, Q.; Wang, J.; Zhao, C. From Genomics to Metabolomics: Molecular Insights into Osteoporosis for Enhanced Diagnostic and Therapeutic Approaches. *Biomedicines* **2024**, *12*, 2389. [[CrossRef](#)]
9. Nuño-Solinis, R.; Rodríguez-Pereira, C.; Alonso-Morán, E.; Orueta, J.F. Comorbidity and healthcare expenditure in women with osteoporosis living in the basque country (Spain). *J. Osteoporos.* **2014**, *2014*, 205954. [[CrossRef](#)]
10. Noh, J.Y.; Yang, Y.; Jung, H. Molecular Mechanisms and Emerging Therapeutics for Osteoporosis. *Int. J. Mol. Sci.* **2020**, *21*, 7623. [[CrossRef](#)]
11. Zhu, S.; Chen, W.; Masson, A.; Li, Y.P. Cell signaling and transcriptional regulation of osteoblast lineage commitment, differentiation, bone formation, and homeostasis. *Cell Discov.* **2024**, *10*, 71. [[CrossRef](#)] [[PubMed](#)]
12. Teitelbaum, S.L. Bone resorption by osteoclasts. *Science* **2000**, *289*, 1504–1508. [[CrossRef](#)] [[PubMed](#)]
13. Cheng, C.H.; Chen, L.R.; Chen, K.H. Osteoporosis Due to Hormone Imbalance: An Overview of the Effects of Estrogen Deficiency and Glucocorticoid Overuse on Bone Turnover. *Int. J. Mol. Sci.* **2022**, *23*, 1376. [[CrossRef](#)] [[PubMed](#)]
14. Wang, Y.; Mei, R.; Hao, S.; Luo, P.; Wang, P.; Almatari, Y.; Guo, L.; Guo, L. Up-regulation of SIRT1 induced by 17beta-estradiol promotes autophagy and inhibits apoptosis in osteoblasts. *Aging* **2021**, *13*, 23652–23671. [[CrossRef](#)] [[PubMed](#)]
15. Schiavi, J.; Fodera, D.M.; Brennan, M.A.; McNamara, L.M. Estrogen depletion alters osteogenic differentiation and matrix production by osteoblasts in vitro. *Exp. Cell Res.* **2021**, *408*, 112814. [[CrossRef](#)]
16. Vrachnis, N.; Zygouris, D.; Vrachnis, D.; Antonakopoulos, N.; Fotiou, A.; Panagopoulos, P.; Kolialexi, A.; Pappa, K.; Mastorakos, G.; Iliodromiti, Z. Effects of Hormone Therapy and Flavonoids Capable on Reversal of Menopausal Immune Senescence. *Nutrients* **2021**, *13*, 2363. [[CrossRef](#)]

17. Zhivodernikov, I.V.; Kirichenko, T.V.; Markina, Y.V.; Postnov, A.Y.; Markin, A.M. Molecular and Cellular Mechanisms of Osteoporosis. *Int. J. Mol. Sci.* **2023**, *24*, 15772. [[CrossRef](#)]
18. Eastell, R.; O'Neill, T.W.; Hofbauer, L.C.; Langdahl, B.; Reid, I.R.; Gold, D.T.; Cummings, S.R. Postmenopausal osteoporosis. *Nat. Rev. Dis. Primers* **2016**, *2*, 16069. [[CrossRef](#)]
19. Deepak, V.; Kayastha, P.; McNamara, L.M. Estrogen deficiency attenuates fluid flow-induced [Ca(2+)]_i oscillations and mechanoresponsiveness of MLO-Y4 osteocytes. *FASEB J.* **2017**, *31*, 3027–3039. [[CrossRef](#)]
20. Miura, T.; Etani, Y.; Noguchi, T.; Hirao, M.; Takami, K.; Goshima, A.; Kurihara, T.; Fukuda, Y.; Ochiai, N.; Kanamoto, T.; et al. Igaratimod suppresses sclerostin and receptor activator of NF-κB ligand production via the extracellular signal-regulated kinase/early growth response protein 1/tumor necrosis factor alpha pathway in osteocytes and ameliorates disuse osteoporosis in mice. *Bone* **2024**, *181*, 117026. [[CrossRef](#)]
21. Bensreti, H.; Alhamad, D.W.; Gonzalez, A.M.; Pizarro-Mondesir, M.; Bollag, W.B.; Isales, C.M.; McGee-Lawrence, M.E. Update on the Role of Glucocorticoid Signaling in Osteoblasts and Bone Marrow Adipocytes During Aging. *Curr. Osteoporos. Rep.* **2023**, *21*, 32–44. [[CrossRef](#)] [[PubMed](#)]
22. Hachemi, Y.; Rapp, A.E.; Picke, A.K.; Weidinger, G.; Ignatius, A.; Tuckermann, J. Molecular mechanisms of glucocorticoids on skeleton and bone regeneration after fracture. *J. Mol. Endocrinol.* **2018**, *61*, R75–R90. [[CrossRef](#)] [[PubMed](#)]
23. Compston, J. Glucocorticoid-induced osteoporosis: An update. *Endocrine* **2018**, *61*, 7–16. [[CrossRef](#)] [[PubMed](#)]
24. Hildebrandt, S.; Baschant, U.; Thiele, S.; Tuckermann, J.; Hofbauer, L.C.; Rauner, M. Glucocorticoids suppress *Wnt16* expression in osteoblasts in vitro and in vivo. *Sci. Rep.* **2018**, *8*, 8711. [[CrossRef](#)]
25. Ohnaka, K.; Tanabe, M.; Kawate, H.; Nawata, H.; Takayanagi, R. Glucocorticoid suppresses the canonical Wnt signal in cultured human osteoblasts. *Biochem. Biophys. Res. Commun.* **2005**, *329*, 177–181. [[CrossRef](#)]
26. Wang, L.T.; Chen, L.R.; Chen, K.H. Hormone-Related and Drug-Induced Osteoporosis: A Cellular and Molecular Overview. *Int. J. Mol. Sci.* **2023**, *24*, 5814. [[CrossRef](#)]
27. Branstetter, R.M., IV; Islam, R.K.; Toups, C.A.; Parra, A.N.; Lee, Z.; Ahmadzadeh, S.; Varrassi, G.; Shekoohi, S.; Kaye, A.D. Mechanisms and Treatment Options for Hyperthyroid-Induced Osteoporosis: A Narrative Review. *Cureus* **2023**, *15*, e48798. [[CrossRef](#)]
28. Bassett, J.H.; Williams, G.R. Critical role of the hypothalamic-pituitary-thyroid axis in bone. *Bone* **2008**, *43*, 418–426. [[CrossRef](#)]
29. Qi, W.; Wang, D.; Hong, Y.; Yao, J.; Wang, H.; Zhu, L.; Pan, H. Investigating the causal relationship between thyroid dysfunction diseases and osteoporosis: A two-sample Mendelian randomization analysis. *Sci. Rep.* **2024**, *14*, 12784. [[CrossRef](#)]
30. Bassett, J.H.; Williams, G.R. Role of Thyroid Hormones in Skeletal Development and Bone Maintenance. *Endocr. Rev.* **2016**, *37*, 135–187. [[CrossRef](#)]
31. Rosen, C.J. Endocrine disorders and osteoporosis. *Curr. Opin. Rheumatol.* **1997**, *9*, 355–361. [[CrossRef](#)] [[PubMed](#)]
32. Jones, A.R.; Simons, K.; Harvey, S.; Grill, V. Bone Mineral Density Compared to Trabecular Bone Score in Primary Hyperparathyroidism. *J. Clin. Med.* **2022**, *11*, 330. [[CrossRef](#)] [[PubMed](#)]
33. Jiang, S.; Li, H.; Zhang, L.; Mu, W.; Zhang, Y.; Chen, T.; Wu, J.; Tang, H.; Zheng, S.; Liu, Y.; et al. Generic Diagramming Platform (GDP): A comprehensive database of high-quality biomedical graphics. *Nucleic Acids Res.* **2025**, *53*, D1670–D1676. [[CrossRef](#)]
34. Kimball, J.S.; Johnson, J.P.; Carlson, D.A. Oxidative Stress and Osteoporosis. *J. Bone Jt. Surg. Am. Vol.* **2021**, *103*, 1451–1461. [[CrossRef](#)] [[PubMed](#)]
35. Palma, F.R.; Gantner, B.N.; Sakiyama, M.J.; Kayzuka, C.; Shukla, S.; Lacchini, R.; Cunniff, B.; Bonini, M.G. ROS production by mitochondria: Function or dysfunction? *Oncogene* **2024**, *43*, 295–303. [[CrossRef](#)]
36. Iantomasi, T.; Romagnoli, C.; Palmi, G.; Donati, S.; Falsetti, I.; Miglietta, F.; Aurilia, C.; Marini, F.; Giusti, F.; Brandi, M.L. Oxidative Stress and Inflammation in Osteoporosis: Molecular Mechanisms Involved and the Relationship with microRNAs. *Int. J. Mol. Sci.* **2023**, *24*, 3772. [[CrossRef](#)]
37. Chen, W.M.; Chiang, J.C.; Lin, Y.C.; Lin, Y.N.; Chuang, P.Y.; Chang, Y.C.; Chen, C.C.; Wu, K.Y.; Hsieh, J.C.; Chen, S.K.; et al. Lysophosphatidic acid receptor LPA(3) prevents oxidative stress and cellular senescence in Hutchinson-Gilford progeria syndrome. *Aging Cell* **2020**, *19*, e13064. [[CrossRef](#)]
38. Bhatti, J.S.; Sehrawat, A.; Mishra, J.; Sidhu, I.S.; Navik, U.; Khullar, N.; Kumar, S.; Bhatti, G.K.; Reddy, P.H. Oxidative stress in the pathophysiology of type 2 diabetes and related complications: Current therapeutic strategies and future perspectives. *Free Radic. Biol. Med.* **2022**, *184*, 114–134. [[CrossRef](#)]
39. Zuo, G.; Zhang, T.; Huang, L.; Araujo, C.; Peng, J.; Travis, Z.; Okadab, T.; Ocak, U.; Zhang, G.; Tang, J.; et al. Corrigendum to "Activation of TGR5 with INT-777 attenuates oxidative stress and neuronal apoptosis via cAMP/PKCε/ALDH2 pathway after subarachnoid hemorrhage in rats" [Free Radic. Biol. Med. (2019 Nov 1) 143 441-453]. *Free Radic. Biol. Med.* **2024**, *216*, 78–79. [[CrossRef](#)]
40. Jin, W.; Zhu, X.; Yao, F.; Xu, X.; Chen, X.; Luo, Z.; Zhao, D.; Li, X.; Leng, X.; Sun, L. Cytoprotective effect of Fufang Lurong Jiangu capsule against hydrogen peroxide-induced oxidative stress in bone marrow stromal cell-derived osteoblasts through the Nrf2/HO-1 signaling pathway. *Biomed. Pharmacother.* **2020**, *121*, 109676. [[CrossRef](#)]

41. Sharma, A.R.; Sharma, G.; Lee, Y.H.; Chakraborty, C.; Lee, S.S.; Seo, E.M. Sodium Selenite Promotes Osteoblast Differentiation via The WNT/ β -Catenin Signaling Pathway. *Cell J.* **2022**, *24*, 309–315. [[PubMed](#)]
42. Yang, K.; Pei, L.; Zhou, S.; Tao, L.; Zhu, Y. Metformin attenuates H₂O₂-induced osteoblast apoptosis by regulating SIRT3 via the PI3K/AKT pathway. *Exp. Ther. Med.* **2021**, *22*, 1316. [[CrossRef](#)] [[PubMed](#)]
43. Li, X.; Lin, H.; Zhang, X.; Jaspers, R.T.; Yu, Q.; Ji, Y.; Forouzanfar, T.; Wang, D.; Huang, S.; Wu, G. Notoginsenoside R1 attenuates oxidative stress-induced osteoblast dysfunction through JNK signalling pathway. *J. Cell. Mol. Med.* **2021**, *25*, 11278–11289. [[CrossRef](#)]
44. Wang, N.; Hao, Y.; Fu, L. Trimethylamine-N-Oxide Promotes Osteoclast Differentiation and Bone Loss via Activating ROS-Dependent NF- κ B Signaling Pathway. *Nutrients* **2022**, *14*, 3955. [[CrossRef](#)]
45. Zhao, Y.; Wang, C.; Qiu, F.; Liu, J.; Xie, Y.; Lin, Z.; He, J.; Chen, J. Trimethylamine-N-oxide promotes osteoclast differentiation and oxidative stress by activating NF- κ B pathway. *Aging* **2024**, *16*, 9251–9263. [[CrossRef](#)]
46. Goodnough, L.H.; Goodman, S.B. Relationship of Aging, Inflammation, and Skeletal Stem Cells and Their Effects on Fracture Repair. *Curr. Osteoporos. Rep.* **2022**, *20*, 320–325. [[CrossRef](#)]
47. Bhatt, S.; Nagappa, A.N.; Patil, C.R. Role of oxidative stress in depression. *Drug Discov. Today* **2020**, *25*, 1270–1276. [[CrossRef](#)]
48. Sies, H.; Berndt, C.; Jones, D.P. Oxidative Stress. *Annu. Rev. Biochem.* **2017**, *86*, 715–748. [[CrossRef](#)] [[PubMed](#)]
49. Wang, G.; Ma, C.; Chen, K.; Wang, Z.; Qiu, H.; Chen, D.; He, J.; Zhang, C.; Guo, D.; Lai, B.; et al. Cycloastragenol Attenuates Osteoclastogenesis and Bone Loss by Targeting RANKL-Induced Nrf2/Keap1/ARE, NF- κ B, Calcium, and NFATc1 Pathways. *Front. Pharmacol.* **2021**, *12*, 810322. [[CrossRef](#)]
50. Birch, J.; Gil, J. Senescence and the SASP: Many therapeutic avenues. *Genes Dev.* **2020**, *34*, 1565–1576. [[CrossRef](#)]
51. Khosla, S.; Farr, J.N.; Tchkonja, T.; Kirkland, J.L. The role of cellular senescence in ageing and endocrine disease. *Nat. Rev. Endocrinol.* **2020**, *16*, 263–275. [[CrossRef](#)] [[PubMed](#)]
52. Piemontese, M.; Almeida, M.; Robling, A.G.; Kim, H.N.; Xiong, J.; Thostenson, J.D.; Weinstein, R.S.; Manolagas, S.C.; O'Brien, C.A.; Jilka, R.L. Old age causes de novo intracortical bone remodeling and porosity in mice. *JCI Insight* **2017**, *2*, e93771. [[CrossRef](#)] [[PubMed](#)]
53. Farr, J.N.; Fraser, D.G.; Wang, H.; Jaehn, K.; Ogrodnik, M.B.; Weivoda, M.M.; Drake, M.T.; Tchkonja, T.; LeBrasseur, N.K.; Kirkland, J.L.; et al. Identification of Senescent Cells in the Bone Microenvironment. *J. Bone Miner. Res.* **2016**, *31*, 1920–1929. [[CrossRef](#)]
54. Farr, J.N.; Xu, M.; Weivoda, M.M.; Monroe, D.G.; Fraser, D.G.; Onken, J.L.; Negley, B.A.; Sfeir, J.G.; Ogrodnik, M.B.; Hachfeld, C.M.; et al. Targeting cellular senescence prevents age-related bone loss in mice. *Nat. Med.* **2017**, *23*, 1072–1079. [[CrossRef](#)]
55. Moerman, E.J.; Teng, K.; Lipschitz, D.A.; Lecka-Czernik, B. Aging activates adipogenic and suppresses osteogenic programs in mesenchymal marrow stroma/stem cells: The role of PPAR- γ 2 transcription factor and TGF- β /BMP signaling pathways. *Aging Cell* **2004**, *3*, 379–389. [[CrossRef](#)]
56. Habiballa, L.; Salmonowicz, H.; Passos, J.F. Mitochondria and cellular senescence: Implications for musculoskeletal ageing. *Free Radic. Biol. Med.* **2019**, *132*, 3–10. [[CrossRef](#)]
57. Petersen, S.; Saretzki, G.; von Zglinicki, T. Preferential accumulation of single-stranded regions in telomeres of human fibroblasts. *Exp. Cell Res.* **1998**, *239*, 152–160. [[CrossRef](#)]
58. Birch, J.; Passos, J.F. Targeting the SASP to combat ageing: Mitochondria as possible intracellular allies? *BioEssays* **2017**, *39*, 1600235. [[CrossRef](#)] [[PubMed](#)]
59. Guo, Y.; Jia, X.; Cui, Y.; Song, Y.; Wang, S.; Geng, Y.; Li, R.; Gao, W.; Fu, D. Sirt3-mediated mitophagy regulates AGEs-induced BMSCs senescence and senile osteoporosis. *Redox Biol.* **2021**, *41*, 101915. [[CrossRef](#)]
60. Liu, F.; Yuan, L.; Li, L.; Yang, J.; Liu, J.; Chen, Y.; Zhang, J.; Lu, Y.; Yuan, Y.; Cheng, J. S-sulphydration of SIRT3 combats BMSC senescence and ameliorates osteoporosis via stabilizing heterochromatic and mitochondrial homeostasis. *Pharmacol. Res.* **2023**, *192*, 106788. [[CrossRef](#)]
61. Zou, L.; Barnett, B.; Safah, H.; Larussa, V.F.; Evdemon-Hogan, M.; Mottram, P.; Wei, S.; David, O.; Curiel, T.J.; Zou, W. Bone marrow is a reservoir for CD4⁺CD25⁺ regulatory T cells that traffic through CXCL12/CXCR4 signals. *Cancer Res.* **2004**, *64*, 8451–8455. [[CrossRef](#)] [[PubMed](#)]
62. Fischer, V.; Haffner-Luntzer, M. Interaction between bone and immune cells: Implications for postmenopausal osteoporosis. *Semin. Cell Dev. Biol.* **2022**, *123*, 14–21. [[CrossRef](#)] [[PubMed](#)]
63. Wu, D.; Cline-Smith, A.; Shashkova, E.; Perla, A.; Katyal, A.; Aurora, R. T-Cell Mediated Inflammation in Postmenopausal Osteoporosis. *Front. Immunol.* **2021**, *12*, 687551. [[CrossRef](#)]
64. Srivastava, R.K.; Dar, H.Y.; Mishra, P.K. Immunoporosis: Immunology of Osteoporosis-Role of T Cells. *Front. Immunol.* **2018**, *9*, 657. [[CrossRef](#)]
65. Choi, Y.; Woo, K.M.; Ko, S.H.; Lee, Y.J.; Park, S.J.; Kim, H.M.; Kwon, B.S. Osteoclastogenesis is enhanced by activated B cells but suppressed by activated CD8⁺ T cells. *Eur. J. Immunol.* **2001**, *31*, 2179–2188. [[CrossRef](#)]
66. Bozec, A.; Zaiss, M.M. T Regulatory Cells in Bone Remodelling. *Curr. Osteoporos. Rep.* **2017**, *15*, 121–125. [[CrossRef](#)]

67. Föger-Samwald, U.; Dovjak, P.; Azizi-Semrad, U.; Kersch-Schindl, K.; Pietschmann, P. Osteoporosis: Pathophysiology and therapeutic options. *EXCLI J.* **2020**, *19*, 1017–1037. [[PubMed](#)]
68. Pietschmann, P.; Mechtcheriakova, D.; Meshcheryakova, A.; Föger-Samwald, U.; Ellinger, I. Immunology of Osteoporosis: A Mini-Review. *Gerontology* **2016**, *62*, 128–137. [[CrossRef](#)]
69. Behera, J.; Ison, J.; Tyagi, S.C.; Tyagi, N. The role of gut microbiota in bone homeostasis. *Bone* **2020**, *135*, 115317. [[CrossRef](#)]
70. Sjögren, K.; Engdahl, C.; Henning, P.; Lerner, U.H.; Tremaroli, V.; Lagerquist, M.K.; Bäckhed, F.; Ohlsson, C. The gut microbiota regulates bone mass in mice. *J. Bone Miner. Res.* **2012**, *27*, 1357–1367. [[CrossRef](#)]
71. Guss, J.D.; Taylor, E.; Rouse, Z.; Roubert, S.; Higgins, C.H.; Thomas, C.J.; Baker, S.P.; Vashishth, D.; Donnelly, E.; Shea, M.K.; et al. The microbial metagenome and bone tissue composition in mice with microbiome-induced reductions in bone strength. *Bone* **2019**, *127*, 146–154. [[CrossRef](#)] [[PubMed](#)]
72. Ohlsson, C.; Engdahl, C.; Fåk, F.; Andersson, A.; Windahl, S.H.; Farman, H.H.; Movérare-Skrtic, S.; Islander, U.; Sjögren, K. Probiotics protect mice from ovariectomy-induced cortical bone loss. *PLoS ONE* **2014**, *9*, e92368. [[CrossRef](#)]
73. Rodrigues, F.C.; Castro, A.S.; Rodrigues, V.C.; Fernandes, S.A.; Fontes, E.A.; de Oliveira, T.T.; Martino, H.S.; de Lucas Fortes Ferreira, C.L. Yacon flour and *Bifidobacterium longum* modulate bone health in rats. *J. Med. Food* **2012**, *15*, 664–670. [[CrossRef](#)]
74. Lucas, S.; Omata, Y.; Hofmann, J.; Böttcher, M.; Iljazovic, A.; Sarter, K.; Albrecht, O.; Schulz, O.; Krishnacoumar, B.; Krönke, G.; et al. Short-chain fatty acids regulate systemic bone mass and protect from pathological bone loss. *Nat. Commun.* **2018**, *9*, 55. [[CrossRef](#)]
75. Pacifici, R. Bone Remodeling and the Microbiome. *Cold Spring Harb. Perspect. Med.* **2018**, *8*, a031203. [[CrossRef](#)]
76. Dar, H.Y.; Pal, S.; Shukla, P.; Mishra, P.K.; Tomar, G.B.; Chattopadhyay, N.; Srivastava, R.K. *Bacillus clausii* inhibits bone loss by skewing Treg-Th17 cell equilibrium in postmenopausal osteoporotic mice model. *Nutrition* **2018**, *54*, 118–128. [[CrossRef](#)] [[PubMed](#)]
77. Li, J.Y.; Yu, M.; Pal, S.; Tyagi, A.M.; Dar, H.; Adams, J.; Weitzmann, M.N.; Jones, R.M.; Pacifici, R. Parathyroid hormone-dependent bone formation requires butyrate production by intestinal microbiota. *J. Clin. Investig.* **2020**, *130*, 1767–1781. [[CrossRef](#)] [[PubMed](#)]
78. Yu, M.; Malik Tyagi, A.; Li, J.Y.; Adams, J.; Denning, T.L.; Weitzmann, M.N.; Jones, R.M.; Pacifici, R. PTH induces bone loss via microbial-dependent expansion of intestinal TNF⁺ T cells and Th17 cells. *Nat. Commun.* **2020**, *11*, 468. [[CrossRef](#)]
79. Letarouilly, J.G.; Broux, O.; Clabaut, A. New insights into the epigenetics of osteoporosis. *Genomics* **2019**, *111*, 793–798. [[CrossRef](#)]
80. Wu, Y.L.; Lin, Z.J.; Li, C.C.; Lin, X.; Shan, S.K.; Guo, B.; Zheng, M.H.; Li, F.; Yuan, L.Q.; Li, Z.H. Epigenetic regulation in metabolic diseases: Mechanisms and advances in clinical study. *Signal Transduct. Target. Ther.* **2023**, *8*, 98. [[CrossRef](#)]
81. Kim, K.T.; Lee, Y.S.; Han, I. The Role of Epigenomics in Osteoporosis and Osteoporotic Vertebral Fracture. *Int. J. Mol. Sci.* **2020**, *21*, 9455. [[CrossRef](#)] [[PubMed](#)]
82. Qadir, A.; Liang, S.; Wu, Z.; Chen, Z.; Hu, L.; Qian, A. Senile Osteoporosis: The Involvement of Differentiation and Senescence of Bone Marrow Stromal Cells. *Int. J. Mol. Sci.* **2020**, *21*, 349. [[CrossRef](#)] [[PubMed](#)]
83. Wang, P.; Cao, Y.; Zhan, D.; Wang, D.; Wang, B.; Liu, Y.; Li, G.; He, W.; Wang, H.; Xu, L. Influence of DNA methylation on the expression of OPG/RANKL in primary osteoporosis. *Int. J. Med. Sci.* **2018**, *15*, 1480–1485. [[CrossRef](#)] [[PubMed](#)]
84. Wang, R.; Wang, Y.; Zhu, L.; Liu, Y.; Li, W. Epigenetic Regulation in Mesenchymal Stem Cell Aging and Differentiation and Osteoporosis. *Stem Cells Int.* **2020**, *2020*, 8836258. [[CrossRef](#)] [[PubMed](#)]
85. Kim, K.; Shin, Y.; Kim, J.; Ulmer, T.S.; An, W. H3K27me1 is essential for MMP-9-dependent H3N-terminal tail proteolysis during osteoclastogenesis. *Epigenetics Chromatin* **2018**, *11*, 23. [[CrossRef](#)]
86. Astleford, K.; Campbell, E.; Norton, A.; Mansky, K.C. Epigenetic Regulators Involved in Osteoclast Differentiation. *Int. J. Mol. Sci.* **2020**, *21*, 7080. [[CrossRef](#)]
87. Jing, H.; Su, X.; Gao, B.; Shuai, Y.; Chen, J.; Deng, Z.; Liao, L.; Jin, Y. Epigenetic inhibition of Wnt pathway suppresses osteogenic differentiation of BMSCs during osteoporosis. *Cell Death Dis.* **2018**, *9*, 176. [[CrossRef](#)]
88. Zhang, P.; Liu, Y.; Jin, C.; Zhang, M.; Lv, L.; Zhang, X.; Liu, H.; Zhou, Y. Histone H3K9 Acetyltransferase PCAF Is Essential for Osteogenic Differentiation Through Bone Morphogenetic Protein Signaling and May Be Involved in Osteoporosis. *Stem Cells* **2016**, *34*, 2332–2341. [[CrossRef](#)]
89. Sabri, S.A.; Chavarria, J.C.; Ackert-Bicknell, C.; Swanson, C.; Burger, E. Osteoporosis: An Update on Screening, Diagnosis, Evaluation, and Treatment. *Orthopedics* **2023**, *46*, e20–e26. [[CrossRef](#)]
90. Lips, P.; van Schoor, N.M. The effect of vitamin D on bone and osteoporosis. *Best Pract. Res. Clin. Endocrinol. Metab.* **2011**, *25*, 585–591. [[CrossRef](#)]
91. Ding, K.; Hua, F.; Ding, W. Gut Microbiome and Osteoporosis. *Aging Dis.* **2020**, *11*, 438–447. [[CrossRef](#)]
92. Bahrami, A.; Sadeghnia, H.R.; Tabatabaeizadeh, S.A.; Bahrami-Taghanaki, H.; Behboodi, N.; Esmaeili, H.; Ferns, G.A.; Mobarhan, M.G.; Avan, A. Genetic and epigenetic factors influencing vitamin D status. *J. Cell. Physiol.* **2018**, *233*, 4033–4043. [[CrossRef](#)]
93. Yang, T.L.; Shen, H.; Liu, A.; Dong, S.S.; Zhang, L.; Deng, F.Y.; Zhao, Q.; Deng, H.W. A road map for understanding molecular and genetic determinants of osteoporosis. *Nat. Rev. Endocrinol.* **2020**, *16*, 91–103. [[CrossRef](#)]

94. Lovšin, N.; Zupan, J.; Marc, J. Genetic effects on bone health. *Curr. Opin. Clin. Nutr. Metab. Care* **2018**, *21*, 233–239. [[CrossRef](#)] [[PubMed](#)]
95. Moradifard, S.; Hoseinbeyki, M.; Emam, M.M.; Parchiniparchin, F.; Ebrahimi-Rad, M. Association of the Sp1 binding site and -1997 promoter variations in COL1A1 with osteoporosis risk: The application of meta-analysis and bioinformatics approaches offers a new perspective for future research. *Mutat. Res. Rev. Mutat. Res.* **2020**, *786*, 108339. [[CrossRef](#)]
96. Sabik, O.L.; Farber, C.R. Using GWAS to identify novel therapeutic targets for osteoporosis. *Transl. Res.* **2017**, *181*, 15–26. [[CrossRef](#)] [[PubMed](#)]
97. Gong, Y.; Slee, R.B.; Fukai, N.; Rawadi, G.; Roman-Roman, S.; Reginato, A.M.; Wang, H.; Cundy, T.; Glorieux, F.H.; Lev, D.; et al. LDL receptor-related protein 5 (LRP5) affects bone accrual and eye development. *Cell* **2001**, *107*, 513–523. [[CrossRef](#)]
98. van Meurs, J.B.; Trikalinos, T.A.; Ralston, S.H.; Balcells, S.; Brandi, M.L.; Brixen, K.; Kiel, D.P.; Langdahl, B.L.; Lips, P.; Ljunggren, O.; et al. Large-scale analysis of association between *LRP5* and *LRP6* variants and osteoporosis. *JAMA* **2008**, *299*, 1277–1290. [[CrossRef](#)] [[PubMed](#)]
99. Fabre, S.; Bourmaud, M.; Mabilieu, G.; Goulet, R.; Couturier, A.; Dentel, A.; Picaud, S.; Funck-Brentano, T.; Collet, C.; Cohen-Solal, M. *Lrp5* p.Val667Met Variant Compromises Bone Mineral Density and Matrix Properties in Osteoporosis. *JBMR Plus* **2023**, *7*, e10741. [[CrossRef](#)]
100. Rojano-Mejía, D.; Coral-Vázquez, R.M.; Coronel, A.; Cortes-Espinosa, L.; del Carmen Aguirre-García, M.; Valencia-Villalvazo, E.Y.; Canto, P. Relation of the estrogen receptor and vitamin D receptor polymorphisms with bone mineral density in postmenopausal Mexican-mestizo women. *Gene* **2014**, *537*, 10–14. [[CrossRef](#)]
101. Mondockova, V.; Adamkovicova, M.; Lukacova, M.; Grosskopf, B.; Babosova, R.; Galbavy, D.; Martiniakova, M.; Omelka, R. The estrogen receptor 1 gene affects bone mineral density and osteoporosis treatment efficiency in Slovak postmenopausal women. *BMC Med. Genet.* **2018**, *19*, 174. [[CrossRef](#)] [[PubMed](#)]
102. Styrkarsdottir, U.; Halldorsson, B.V.; Gretarsdottir, S.; Gudbjartsson, D.F.; Walters, G.B.; Ingvarsson, T.; Jonsdottir, T.; Saemundsdottir, J.; Snorraddottir, S.; Center, J.R.; et al. New sequence variants associated with bone mineral density. *Nat. Genet.* **2009**, *41*, 15–17. [[CrossRef](#)]
103. Casas-Avila, L.; Cruz-Arenas, E.; Ponce-de-León-Suárez, V.; Sánchez-Bringas, G.; Olivares-Bañuelos, B.; Chávez-Heres, T.; Valdés-Flores, M. High risk of lumbar spine osteoporosis with the RANK rs3018362 polymorphism. *Gynecol. Endocrinol.* **2019**, *35*, 981–984. [[CrossRef](#)] [[PubMed](#)]
104. Richards, J.B.; Zheng, H.F.; Spector, T.D. Genetics of osteoporosis from genome-wide association studies: Advances and challenges. *Nat. Rev. Genet.* **2012**, *13*, 576–588. [[CrossRef](#)]
105. Nelson, M.R.; Tipney, H.; Painter, J.L.; Shen, J.; Nicoletti, P.; Shen, Y.; Floratos, A.; Sham, P.C.; Li, M.J.; Wang, J.; et al. The support of human genetic evidence for approved drug indications. *Nat. Genet.* **2015**, *47*, 856–860. [[CrossRef](#)] [[PubMed](#)]
106. Lamy, O.; Stoll, D.; Aubry-Rozier, B.; Rodriguez, E.G. Stopping Denosumab. *Curr. Osteoporos. Rep.* **2019**, *17*, 8–15. [[CrossRef](#)]
107. Adler, R.A. Treating osteoporosis in patients with atypical femoral fracture. *J. Bone Miner. Res.* **2024**, *39*, 1711–1715. [[CrossRef](#)]
108. Casamassimi, A.; Federico, A.; Rienzo, M.; Esposito, S.; Ciccodicola, A. Transcriptome Profiling in Human Diseases: New Advances and Perspectives. *Int. J. Mol. Sci.* **2017**, *18*, 1652. [[CrossRef](#)]
109. Chandra, A.; Rajawat, J. Skeletal Aging and Osteoporosis: Mechanisms and Therapeutics. *Int. J. Mol. Sci.* **2021**, *22*, 3553. [[CrossRef](#)]
110. Föger-Samwald, U.; Patsch, J.M.; Schamall, D.; Alaghebandan, A.; Deutschmann, J.; Salem, S.; Mousavi, M.; Pietschmann, P. Molecular evidence of osteoblast dysfunction in elderly men with osteoporotic hip fractures. *Exp. Gerontol.* **2014**, *57*, 114–121. [[CrossRef](#)]
111. Komori, T. Whole Aspect of Runx2 Functions in Skeletal Development. *Int. J. Mol. Sci.* **2022**, *23*, 5776. [[CrossRef](#)] [[PubMed](#)]
112. Hojo, H.; Ohba, S.; He, X.; Lai, L.P.; McMahon, A.P. Sp7/Osterix Is Restricted to Bone-Forming Vertebrates where It Acts as a Dlx Co-factor in Osteoblast Specification. *Dev. Cell* **2016**, *37*, 238–253. [[CrossRef](#)]
113. Azizieh, F.Y.; Shehab, D.; Jarallah, K.A.; Gupta, R.; Raghupathy, R. Circulatory Levels of RANKL, OPG, and Oxidative Stress Markers in Postmenopausal Women With Normal or Low Bone Mineral Density. *Biomark. Insights* **2019**, *14*, 1177271919843825. [[CrossRef](#)] [[PubMed](#)]
114. Trojniak, J.; Sendera, A.; Banaś-Zabczyk, A.; Kopańska, M. The MicroRNAs in the Pathophysiology of Osteoporosis. *Int. J. Mol. Sci.* **2024**, *25*, 6240. [[CrossRef](#)]
115. Long, H.; Sun, B.; Cheng, L.; Zhao, S.; Zhu, Y.; Zhao, R.; Zhu, J. miR-139-5p Represses BMSC Osteogenesis via Targeting Wnt/ β -Catenin Signaling Pathway. *DNA Cell Biol.* **2017**, *36*, 715–724. [[CrossRef](#)] [[PubMed](#)]
116. Tang, X.; Lin, J.; Wang, G.; Lu, J. MicroRNA-433-3p promotes osteoblast differentiation through targeting DKK1 expression. *PLoS ONE* **2017**, *12*, e0179860. [[CrossRef](#)]
117. Liu, H.; Yue, X.; Zhang, G. Downregulation of miR-146a inhibits osteoporosis in the jaws of ovariectomized rats by regulating the Wnt/ β -catenin signaling pathway. *Int. J. Mol. Med.* **2021**, *47*, 6. [[CrossRef](#)]

118. Qin, X.B.; Wen, K.; Wu, X.X.; Yao, Z.J. MiR-183 regulates the differentiation of osteoblasts in the development of osteoporosis by targeting Smad4. *Acta Histochem.* **2021**, *123*, 151786. [[CrossRef](#)]
119. Lu, X.D.; Han, W.X.; Liu, Y.X. Suppression of miR-451a accelerates osteogenic differentiation and inhibits bone loss via Bmp6 signaling during osteoporosis. *Biomed. Pharmacother.* **2019**, *120*, 109378. [[CrossRef](#)]
120. Li, B.; Liu, J.; Zhao, J.; Ma, J.X.; Jia, H.B.; Zhang, Y.; Xing, G.S.; Ma, X.L. LncRNA-H19 Modulates Wnt/ β -catenin Signaling by Targeting Dkk4 in Hindlimb Unloaded Rat. *Orthop. Surg.* **2017**, *9*, 319–327. [[CrossRef](#)]
121. Zhang, N.; Hu, X.; He, S.; Ding, W.; Wang, F.; Zhao, Y.; Huang, Z. LncRNA MSC-AS1 promotes osteogenic differentiation and alleviates osteoporosis through sponging microRNA-140-5p to upregulate BMP2. *Biochem. Biophys. Res. Commun.* **2019**, *519*, 790–796. [[CrossRef](#)]
122. Zhao, Y.; Ning, J.; Teng, H.; Deng, Y.; Sheldon, M.; Shi, L.; Martinez, C.; Zhang, J.; Tian, A.; Sun, Y.; et al. Long noncoding RNA Malat1 protects against osteoporosis and bone metastasis. *Nat. Commun.* **2024**, *15*, 2384. [[CrossRef](#)]
123. Yu, H.; Zhou, W.; Yan, W.; Xu, Z.; Xie, Y.; Zhang, P. LncRNA CASC11 is upregulated in postmenopausal osteoporosis and is correlated with TNF- α . *Clin. Interv. Aging* **2019**, *14*, 1663–1669. [[CrossRef](#)] [[PubMed](#)]
124. Chen, R.; Jin, Y.; Lian, R.; Yang, J.; Liao, Z.; Jin, Y.; Deng, Z.; Feng, S.; Feng, Z.; Wei, Y.; et al. CRIP1 regulates osteogenic differentiation of bone marrow stromal cells and pre-osteoblasts via the Wnt signaling pathway. *Biochem. Biophys. Res. Commun.* **2024**, *727*, 150277. [[CrossRef](#)]
125. Liu, F.; Yuan, Y.; Bai, L.; Yuan, L.; Li, L.; Liu, J.; Chen, Y.; Lu, Y.; Cheng, J.; Zhang, J. LRRc17 controls BMSC senescence via mitophagy and inhibits the therapeutic effect of BMSCs on ovariectomy-induced bone loss. *Redox Biol.* **2021**, *43*, 101963. [[CrossRef](#)] [[PubMed](#)]
126. Song, I.; Choi, Y.J.; Jin, Y.; Kim, J.W.; Koh, J.T.; Ji, H.M.; Jeong, S.Y.; Won, Y.Y.; Kim, W.; Chung, Y.S. STRA6 as a possible candidate gene for pathogenesis of osteoporosis from RNA-seq analysis of human mesenchymal stem cells. *Mol. Med. Rep.* **2017**, *16*, 4075–4081. [[CrossRef](#)] [[PubMed](#)]
127. Yoshioka, H.; Okita, S.; Nakano, M.; Minamizaki, T.; Nubukiyo, A.; Sotomaru, Y.; Bonnelye, E.; Kozai, K.; Tanimoto, K.; Aubin, J.E.; et al. Single-Cell RNA-Sequencing Reveals the Breadth of Osteoblast Heterogeneity. *JBMR Plus* **2021**, *5*, e10496. [[CrossRef](#)]
128. Chen, T.Y.; You, L.; Hardillo, J.A.U.; Chien, M.P. Spatial Transcriptomic Technologies. *Cells* **2023**, *12*, 2042. [[CrossRef](#)]
129. Xiao, X.; Juan, C.; Drennon, T.; Uyttingco, C.R.; Vishlaghi, N.; Sokolowski, D.; Xu, L.; Levi, B.; Sammarco, M.C.; Tower, R.J. Spatial transcriptomic interrogation of the murine bone marrow signaling landscape. *Bone Res.* **2023**, *11*, 59. [[CrossRef](#)]
130. Jiang, W.; Caruana, D.L.; Back, J.; Lee, F.Y. Unique Spatial Transcriptomic Profiling of the Murine Femoral Fracture Callus: A Preliminary Report. *Cells* **2024**, *13*, 522. [[CrossRef](#)]
131. Wang, Y.; Wang, Q.; Xu, Q.; Li, J.; Zhao, F. Single-cell RNA sequencing analysis dissected the osteo-immunology microenvironment and revealed key regulators in osteoporosis. *Int. Immunopharmacol.* **2022**, *113*, 109302. [[CrossRef](#)] [[PubMed](#)]
132. To, K.; Fei, L.; Pett, J.P.; Roberts, K.; Blain, R.; Polański, K.; Li, T.; Yayon, N.; He, P.; Xu, C.; et al. A multi-omic atlas of human embryonic skeletal development. *Nature* **2024**, *635*, 657–667. [[CrossRef](#)] [[PubMed](#)]
133. Zhuang, H.; Lin, Y.; Lin, C.; Zheng, M.; Li, Y.; Yao, X.; Xu, Y. Transcriptome sequencing-based analysis of the molecular mechanism underlying the effect of lncRNA AC003090.1 on osteoporosis. *J. Orthop. Surg. Res.* **2025**, *20*, 346. [[CrossRef](#)]
134. Cui, M.; Cheng, C.; Zhang, L. High-throughput proteomics: A methodological mini-review. *Lab. Investig. J. Tech. Methods Pathol.* **2022**, *102*, 1170–1181. [[CrossRef](#)] [[PubMed](#)]
135. Su, H.; Tan, G.; Guo, W.; Yu, J.S.; Xu, Z.; Zhuang, R.; Xue, H. Discovery of potential ferroptosis and osteoporosis biomarkers through TMT proteomics and bioinformatics analysis. *Biomed. Eng. Online* **2024**, *23*, 120. [[CrossRef](#)]
136. Du, Y.; Chen, H.; Zhou, L.; Guo, Q.; Gong, S.; Feng, S.; Guan, Q.; Shi, P.; Lv, T.; Guo, Y.; et al. REG γ is essential to maintain bone homeostasis by degrading TRAF6, preventing osteoporosis. *Proc. Natl. Acad. Sci. USA* **2024**, *121*, e2405265121. [[CrossRef](#)]
137. Wang, A.; Zhang, H.; Li, G.; Chen, B.; Li, J.; Zhang, T.; Liu, B.; Cao, Z.; Liu, G.; Jia, P.; et al. Deciphering core proteins of osteoporosis with iron accumulation by proteomics in human bone. *Front. Endocrinol.* **2022**, *13*, 961903. [[CrossRef](#)]
138. Xu, J.; Cai, X.; Miao, Z.; Yan, Y.; Chen, D.; Yang, Z.X.; Yue, L.; Hu, W.; Zhuo, L.; Wang, J.T.; et al. Proteome-wide profiling reveals dysregulated molecular features and accelerated aging in osteoporosis: A 9.8-year prospective study. *Aging Cell* **2024**, *23*, e14035. [[CrossRef](#)]
139. Martínez-Aguilar, M.M.; Aparicio-Bautista, D.I.; Ramírez-Salazar, E.G.; Reyes-Grajeda, J.P.; De la Cruz-Montoya, A.H.; Antuna-Puente, B.; Hidalgo-Bravo, A.; Rivera-Paredes, B.; Ramírez-Palacios, P.; Quiterio, M.; et al. Serum Proteomic Analysis Reveals Vitamin D-Binding Protein (VDBP) as a Potential Biomarker for Low Bone Mineral Density in Mexican Postmenopausal Women. *Nutrients* **2019**, *11*, 2853. [[CrossRef](#)]
140. Aparicio-Bautista, D.I.; Becerra-Cervera, A.; Rivera-Paredes, B.; Aguilar-Ordoñez, I.; Ríos-Castro, E.; Reyes-Grajeda, J.P.; Salmerón, J.; Hidalgo-Bravo, A.; Velázquez-Cruz, R. Label-free quantitative proteomics in serum reveals candidate biomarkers associated with low bone mineral density in Mexican postmenopausal women. *Geroscience* **2024**, *46*, 2177–2195. [[CrossRef](#)]

141. Liang, X.; Du, Y.; Wen, Y.; Liu, L.; Li, P.; Zhao, Y.; Ding, M.; Cheng, B.; Cheng, S.; Ma, M.; et al. Assessing the Genetic Correlations Between Blood Plasma Proteins and Osteoporosis: A Polygenic Risk Score Analysis. *Calcif. Tissue Int.* **2019**, *104*, 171–181. [[CrossRef](#)] [[PubMed](#)]
142. Sasaki, K.; Ozasa, Y.; Iba, K.; Wada, T.; Imai, S.; Matsumoto, K.; Sohma, H.; Aoshima, M.; Yamashita, T.; Kokai, Y. Significant increase of plasma tetranectin in ovx mice as defined by proteomics analysis. *J. Orthop. Sci.* **2014**, *19*, 809–819. [[CrossRef](#)] [[PubMed](#)]
143. Wang, J.; Xue, M.; Hu, Y.; Li, J.; Li, Z.; Wang, Y. Proteomic Insights into Osteoporosis: Unraveling Diagnostic Markers of and Therapeutic Targets for the Metabolic Bone Disease. *Biomolecules* **2024**, *14*, 554. [[CrossRef](#)]
144. Liu, J.; He, S.; Ma, B.; Li, X.; Wang, Y.; Xiong, J. TMT-based quantitative proteomic analysis revealed that FBLN2 and NPR3 are involved in the early osteogenic differentiation of mesenchymal stem cells (MSCs). *Aging* **2023**, *15*, 7637–7654. [[CrossRef](#)]
145. Aswamenakul, K.; Klakklai, P.; Pannengpetch, S.; Tawonsawatruk, T.; Isarankura-Na-Ayudhya, C.; Roytrakul, S.; Nantasenam, C.; Supokawej, A. Proteomic study of in vitro osteogenic differentiation of mesenchymal stem cells in high glucose condition. *Mol. Biol. Rep.* **2020**, *47*, 7505–7516. [[CrossRef](#)]
146. Wang, Z.X.; Lin, X.; Cao, J.; Liu, Y.W.; Luo, Z.W.; Rao, S.S.; Wang, Q.; Wang, Y.Y.; Chen, C.Y.; Zhu, G.Q.; et al. Young osteocyte-derived extracellular vesicles facilitate osteogenesis by transferring tropomyosin-1. *J. Nanobiotechnol.* **2024**, *22*, 208. [[CrossRef](#)]
147. Deng, F.Y.; Lei, S.F.; Zhang, Y.; Zhang, Y.L.; Zheng, Y.P.; Zhang, L.S.; Pan, R.; Wang, L.; Tian, Q.; Shen, H.; et al. Peripheral blood monocyte-expressed ANXA2 gene is involved in pathogenesis of osteoporosis in humans. *Mol. Cell. Proteom. MCP* **2011**, *10*, M111.011700. [[CrossRef](#)]
148. Zhou, X.; Wu, L.F.; Wang, W.Y.; Lu, X.; Jiang, Z.H.; Zhang, Y.H.; Jiang, D.H.; Jiang, J.N.; Gao, H.Q.; Lei, S.F.; et al. Anxa2 attenuates osteoblast growth and is associated with hip BMD and osteoporotic fracture in Chinese elderly. *PLoS ONE* **2018**, *13*, e0194781. [[CrossRef](#)] [[PubMed](#)]
149. Creecy, A.; Brown, K.L.; Rose, K.L.; Voziyan, P.; Nyman, J.S. Post-translational modifications in collagen type I of bone in a mouse model of aging. *Bone* **2021**, *143*, 115763. [[CrossRef](#)]
150. Chin, S.M.; Unnold-Cofre, C.; Naismith, T.; Jansen, S. The actin-bundling protein, PLS3, is part of the mechanoresponsive machinery that regulates osteoblast mineralization. *Front. Cell Dev. Biol.* **2023**, *11*, 1141738. [[CrossRef](#)]
151. Veschi, E.A.; Bolean, M.; Strzelecka-Kiliszek, A.; Bandorowicz-Pikula, J.; Pikula, S.; Granjon, T.; Mebarek, S.; Magne, D.; Ramos, A.P.; Rosato, N.; et al. Localization of Annexin A6 in Matrix Vesicles During Physiological Mineralization. *Int. J. Mol. Sci.* **2020**, *21*, 1367. [[CrossRef](#)] [[PubMed](#)]
152. Zhao, Z.; Cai, Z.; Chen, A.; Cai, M.; Yang, K. Application of metabolomics in osteoporosis research. *Front. Endocrinol.* **2022**, *13*, 993253. [[CrossRef](#)]
153. Miyamoto, T.; Hirayama, A.; Sato, Y.; Koboyashi, T.; Katsuyama, E.; Kanagawa, H.; Fujie, A.; Morita, M.; Watanabe, R.; Tando, T.; et al. Metabolomics-based profiles predictive of low bone mass in menopausal women. *Bone Rep.* **2018**, *9*, 11–18. [[CrossRef](#)] [[PubMed](#)]
154. You, Y.S.; Lin, C.Y.; Liang, H.J.; Lee, S.H.; Tsai, K.S.; Chiou, J.M.; Chen, Y.C.; Tsao, C.K.; Chen, J.H. Association between the metabolome and low bone mineral density in Taiwanese women determined by ¹H NMR spectroscopy. *J. Bone Miner. Res.* **2014**, *29*, 212–222. [[CrossRef](#)]
155. Yu, L.; Qi, H.; An, G.; Bao, J.; Ma, B.; Zhu, J.; Ouyang, G.; Zhang, P.; Fan, H.; Zhang, Q. Association between metabolic profiles in urine and bone mineral density of pre- and postmenopausal Chinese women. *Menopause* **2019**, *26*, 94–102. [[CrossRef](#)] [[PubMed](#)]
156. Zhao, Q.; Shen, H.; Su, K.J.; Zhang, J.G.; Tian, Q.; Zhao, L.J.; Qiu, C.; Zhang, Q.; Garrett, T.J.; Liu, J.; et al. Metabolomic profiles associated with bone mineral density in US Caucasian women. *Nutr. Metab.* **2018**, *15*, 57. [[CrossRef](#)]
157. Lv, H.; Jiang, F.; Guan, D.; Lu, C.; Guo, B.; Chan, C.; Peng, S.; Liu, B.; Guo, W.; Zhu, H.; et al. Metabolomics and Its Application in the Development of Discovering Biomarkers for Osteoporosis Research. *Int. J. Mol. Sci.* **2016**, *17*, 2018. [[CrossRef](#)]
158. Da, W.; Tao, L.; Zhu, Y. The Role of Osteoclast Energy Metabolism in the Occurrence and Development of Osteoporosis. *Front. Endocrinol.* **2021**, *12*, 675385. [[CrossRef](#)]
159. Confavreux, C.B. Bone: From a reservoir of minerals to a regulator of energy metabolism. *Kidney Int.* **2011**, *79* (Suppl. 121), S14–S19. [[CrossRef](#)]
160. Kan, B.; Zhao, Q.; Wang, L.; Xue, S.; Cai, H.; Yang, S. Association between lipid biomarkers and osteoporosis: A cross-sectional study. *BMC Musculoskelet. Disord.* **2021**, *22*, 759. [[CrossRef](#)]
161. During, A.; Coutel, X.; Bertheaume, N.; Penel, G.; Olejnik, C. Long Term Ovariectomy-Induced Osteoporosis is Associated with High Stearoyl-CoA Desaturase Indexes in Rat Femur. *Calcif. Tissue Int.* **2020**, *106*, 315–324. [[CrossRef](#)] [[PubMed](#)]
162. Zhao, H.; Li, X.; Zhang, D.; Chen, H.; Chao, Y.; Wu, K.; Dong, X.; Su, J. Integrative Bone Metabolomics-Lipidomics Strategy for Pathological Mechanism of Postmenopausal Osteoporosis Mouse Model. *Sci. Rep.* **2018**, *8*, 16456. [[CrossRef](#)]
163. Tian, L.; Yu, X. Lipid metabolism disorders and bone dysfunction—Interrelated and mutually regulated (review). *Mol. Med. Rep.* **2015**, *12*, 783–794. [[CrossRef](#)] [[PubMed](#)]

164. Cho, Y.D.; Yoon, W.J.; Kim, W.J.; Woo, K.M.; Baek, J.H.; Lee, G.; Ku, Y.; van Wijnen, A.J.; Ryoo, H.M. Epigenetic modifications and canonical wntless/int-1 class (WNT) signaling enable trans-differentiation of nonosteogenic cells into osteoblasts. *J. Biol. Chem.* **2014**, *289*, 20120–20128. [[CrossRef](#)]
165. Harvey, N.C.; Sheppard, A.; Godfrey, K.M.; McLean, C.; Garratt, E.; Ntani, G.; Davies, L.; Murray, R.; Inskip, H.M.; Gluckman, P.D.; et al. Childhood bone mineral content is associated with methylation status of the RXRA promoter at birth. *J. Bone Miner. Res.* **2014**, *29*, 600–607. [[CrossRef](#)]
166. Takiishi, T.; Fenero, C.I.M.; Câmara, N.O.S. Intestinal barrier and gut microbiota: Shaping our immune responses throughout life. *Tissue Barriers* **2017**, *5*, e1373208. [[CrossRef](#)]
167. Xu, X.; Jia, X.; Mo, L.; Liu, C.; Zheng, L.; Yuan, Q.; Zhou, X. Intestinal microbiota: A potential target for the treatment of postmenopausal osteoporosis. *Bone Res.* **2017**, *5*, 17046. [[CrossRef](#)]
168. Qiu, C.; Yu, F.; Su, K.; Zhao, Q.; Zhang, L.; Xu, C.; Hu, W.; Wang, Z.; Zhao, L.; Tian, Q.; et al. Multi-omics Data Integration for Identifying Osteoporosis Biomarkers and Their Biological Interaction and Causal Mechanisms. *iScience* **2020**, *23*, 100847. [[CrossRef](#)] [[PubMed](#)]
169. Zhang, B.; Pei, Z.; Tian, A.; He, W.; Sun, C.; Hao, T.; Ariben, J.; Li, S.; Wu, L.; Yang, X.; et al. Multi-omics Analysis to Identify Key Immune Genes for Osteoporosis based on Machine Learning and Single-cell Analysis. *Orthop. Surg.* **2024**, *16*, 2803–2820. [[CrossRef](#)]
170. Li, Q.; Guo, R.; Wu, Z.; Zhao, C.; Shen, C. Key genes linking gut microbiota, immune cells, and osteoporosis: A multi-omics approach. *Microb. Pathog.* **2025**, *202*, 107412. [[CrossRef](#)]
171. Babu, M.; Snyder, M. Multi-Omics Profiling for Health. *Mol. Cell. Proteom. MCP* **2023**, *22*, 100561. [[CrossRef](#)] [[PubMed](#)]
172. Barengolts, E.I.; Berman, M.; Kukreja, S.C.; Kouznetsova, T.; Lin, C.; Chomka, E.V. Osteoporosis and coronary atherosclerosis in asymptomatic postmenopausal women. *Calcif. Tissue Int.* **1998**, *62*, 209–213. [[CrossRef](#)]
173. Laroche, M.; Pécourneau, V.; Blain, H.; Breuil, V.; Chapurlat, R.; Cortet, B.; Sutter, B.; Degboe, Y. Osteoporosis and ischemic cardiovascular disease. *Jt. Bone Spine* **2017**, *84*, 427–432. [[CrossRef](#)]
174. Hu, X.; Ma, S.; Chen, L.; Tian, C.; Wang, W. Association between osteoporosis and cardiovascular disease in elderly people: Evidence from a retrospective study. *PeerJ* **2023**, *11*, e16546. [[CrossRef](#)] [[PubMed](#)]
175. Lampropoulos, C.E.; Papaioannou, I.; D’Cruz, D.P. Osteoporosis—a risk factor for cardiovascular disease? *Nat. Rev. Rheumatol.* **2012**, *8*, 587–598. [[CrossRef](#)] [[PubMed](#)]
176. Deligiorgi, M.V.; Panayiotidis, M.I.; Siasos, G.; Trafalis, D.T. Osteoporosis Entwined with Cardiovascular Disease: The Implication of Osteoprotegerin and the Example of Statins. *Curr. Med. Chem.* **2021**, *28*, 1443–1467. [[CrossRef](#)]
177. Shao, J.S.; Cheng, S.L.; Pingsterhaus, J.M.; Charlton-Kachigian, N.; Loewy, A.P.; Towler, D.A. Msx2 promotes cardiovascular calcification by activating paracrine Wnt signals. *J. Clin. Invest.* **2005**, *115*, 1210–1220. [[CrossRef](#)]
178. Kim, K.M.; Choi, S.H.; Lim, S.; Moon, J.H.; Kim, J.H.; Kim, S.W.; Jang, H.C.; Shin, C.S. Interactions between dietary calcium intake and bone mineral density or bone geometry in a low calcium intake population (KNHANES IV 2008–2010). *J. Clin. Endocrinol. Metab.* **2014**, *99*, 2409–2417. [[CrossRef](#)]
179. Anderson, J.J.; Kruszka, B.; Delaney, J.A.; He, K.; Burke, G.L.; Alonso, A.; Bild, D.E.; Budoff, M.; Michos, E.D. Calcium Intake From Diet and Supplements and the Risk of Coronary Artery Calcification and its Progression Among Older Adults: 10-Year Follow-up of the Multi-Ethnic Study of Atherosclerosis (MESA). *J. Am. Heart Assoc.* **2016**, *5*, e003815. [[CrossRef](#)]
180. Kong, S.H.; Kim, J.H.; Hong, A.R.; Cho, N.H.; Shin, C.S. Dietary calcium intake and risk of cardiovascular disease, stroke, and fracture in a population with low calcium intake. *Am. J. Clin. Nutr.* **2017**, *106*, 27–34. [[CrossRef](#)]
181. Wu, D.; Li, L.; Wen, Z.; Wang, G. Romosozumab in osteoporosis: Yesterday, today and tomorrow. *J. Transl. Med.* **2023**, *21*, 668. [[CrossRef](#)] [[PubMed](#)]
182. Fuggle, N.R.; Cooper, C.; Harvey, N.C.; Al-Daghri, N.; Brandi, M.L.; Bruyere, O.; Cano, A.; Dennison, E.M.; Diez-Perez, A.; Kaufman, J.M.; et al. Assessment of Cardiovascular Safety of Anti-Osteoporosis Drugs. *Drugs* **2020**, *80*, 1537–1552. [[CrossRef](#)]
183. Ma, Y.; Qiu, S.; Zhou, R. Osteoporosis in Patients With Respiratory Diseases. *Front. Physiol.* **2022**, *13*, 939253. [[CrossRef](#)]
184. Bitar, A.N.; Sulaiman, S.A.S.; Ali, I.; Khan, A.H. Prevalence, risk assessment, and predictors of osteoporosis among chronic obstructive pulmonary disease patients. *J. Adv. Pharm. Technol. Res.* **2021**, *12*, 395–401. [[CrossRef](#)] [[PubMed](#)]
185. Sood, V.; Rogers, L.; Khurana, S. Managing Corticosteroid-Related Comorbidities in Severe Asthma. *Chest* **2021**, *160*, 1614–1623. [[CrossRef](#)]
186. Evans, R.A.; Morgan, M.D. The systemic nature of chronic lung disease. *Clin. Chest Med.* **2014**, *35*, 283–293. [[CrossRef](#)]
187. Okoye, C.N.; Koren, S.A.; Wojtovich, A.P. Mitochondrial complex I ROS production and redox signaling in hypoxia. *Redox Biol.* **2023**, *67*, 102926. [[CrossRef](#)] [[PubMed](#)]
188. Xu, K.; Lu, C.; Ren, X.; Wang, J.; Xu, P.; Zhang, Y. Overexpression of HIF-1 α enhances the protective effect of mitophagy on steroid-induced osteocytes apoptosis. *Environ. Toxicol.* **2021**, *36*, 2123–2137. [[CrossRef](#)]
189. Xiao, C.; Bai, G.; Du, Y.; Jiang, H.; Yu, X. Association of high HIF-1 α levels in serous periodontitis with external root resorption by the NFATc1 pathway. *J. Mol. Histol.* **2020**, *51*, 649–658. [[CrossRef](#)]

190. Caramori, G.; Ruggeri, P.; Arpinelli, F.; Salvi, L.; Girbino, G. Long-term use of inhaled glucocorticoids in patients with stable chronic obstructive pulmonary disease and risk of bone fractures: A narrative review of the literature. *Int. J. Chronic Obstr. Pulm. Dis.* **2019**, *14*, 1085–1097. [[CrossRef](#)]
191. Zhang, N.; Fan, X.; Zhang, Y.; Xu, N.; Li, L. Risk of Fracture and Osteoporosis in Patients With COPD and Inhaled Corticosteroids Treatment. *Respir. Care* **2023**, *68*, 1719–1727. [[CrossRef](#)] [[PubMed](#)]
192. Buckley, L.; Guyatt, G.; Fink, H.A.; Cannon, M.; Grossman, J.; Hansen, K.E.; Humphrey, M.B.; Lane, N.E.; Magrey, M.; Miller, M.; et al. 2017 American College of Rheumatology Guideline for the Prevention and Treatment of Glucocorticoid-Induced Osteoporosis. *Arthritis Rheumatol.* **2017**, *69*, 1521–1537. [[CrossRef](#)]
193. Rodríguez-Gómez, I.; Gray, S.R.; Ho, F.K.; Petermann-Rocha, F.; Welsh, P.; Cleland, J.; Iliodromiti, S.; Ara, I.; Pell, J.; Sattar, N.; et al. Osteoporosis and Its Association With Cardiovascular Disease, Respiratory Disease, and Cancer: Findings From the UK Biobank Prospective Cohort Study. *Mayo Clin. Proc.* **2022**, *97*, 110–121. [[CrossRef](#)] [[PubMed](#)]
194. Radin, E.L.; Rose, R.M. Role of subchondral bone in the initiation and progression of cartilage damage. *Clin. Orthop. Relat. Res.* **1986**, *213*, 34–40. [[CrossRef](#)]
195. Ziemian, S.N.; Ayobami, O.O.; Rooney, A.M.; Kelly, N.H.; Holyoak, D.T.; Ross, F.P.; van der Meulen, M.C.H. Low bone mass resulting from impaired estrogen signaling in bone increases severity of load-induced osteoarthritis in female mice. *Bone* **2021**, *152*, 116071. [[CrossRef](#)] [[PubMed](#)]
196. Lin, L.; Luo, P.; Yang, M.; Wang, J.; Hou, W.; Xu, P. Causal relationship between osteoporosis and osteoarthritis: A two-sample Mendelian randomized study. *Front. Endocrinol.* **2022**, *13*, 1011246. [[CrossRef](#)]
197. Qu, Y.; Chen, S.; Han, M.; Gu, Z.; Zhang, Y.; Fan, T.; Zeng, M.; Ruan, G.; Cao, P.; Yang, Q.; et al. Osteoporosis and osteoarthritis: A bi-directional Mendelian randomization study. *Arthritis Res. Ther.* **2023**, *25*, 242. [[CrossRef](#)]
198. Kim, D.; Pirshahid, A.A.; Li, Y.; Varghese, T.; Pope, J.E. Prevalence of osteoporosis in osteoarthritis: A systematic review and meta-analysis. *Osteoporos. Int.* **2022**, *33*, 1687–1693. [[CrossRef](#)]
199. Im, G.I.; Kim, M.K. The relationship between osteoarthritis and osteoporosis. *J. Bone Miner. Metab.* **2014**, *32*, 101–109. [[CrossRef](#)]
200. Motta, F.; Barone, E.; Sica, A.; Selmi, C. Inflammaging and Osteoarthritis. *Clin. Rev. Allergy Immunol.* **2023**, *64*, 222–238. [[CrossRef](#)]
201. Ikeda, K.; Horie-Inoue, K.; Inoue, S. Functions of estrogen and estrogen receptor signaling on skeletal muscle. *J. Steroid Biochem. Mol. Biol.* **2019**, *191*, 105375. [[CrossRef](#)] [[PubMed](#)]
202. Velasco, J.; Zarrabeitia, M.T.; Prieto, J.R.; Perez-Castrillon, J.L.; Perez-Aguilar, M.D.; Perez-Núñez, M.I.; Sañudo, C.; Hernandez-Elena, J.; Calvo, I.; Ortiz, F.; et al. Wnt pathway genes in osteoporosis and osteoarthritis: Differential expression and genetic association study. *Osteoporos. Int.* **2010**, *21*, 109–118. [[CrossRef](#)] [[PubMed](#)]
203. Longo, U.G.; Lalli, A.; Bandini, B.; de Sire, R.; Angeletti, S.; Lustig, S.; Ammendolia, A.; Budhiparama, N.C.; de Sire, A. Role of the Gut Microbiota in Osteoarthritis, Rheumatoid Arthritis, and Spondylarthritis: An Update on the Gut-Joint Axis. *Int. J. Mol. Sci.* **2024**, *25*, 3242. [[CrossRef](#)]
204. Bai, R.J.; Li, Y.S.; Zhang, F.J. Osteopontin, a bridge links osteoarthritis and osteoporosis. *Front. Endocrinol.* **2022**, *13*, 1012508. [[CrossRef](#)] [[PubMed](#)]
205. Huang, K.; Cai, H. The interplay between osteoarthritis and osteoporosis: Mechanisms, implications, and treatment considerations—A narrative review. *Exp. Gerontol.* **2024**, *197*, 112614. [[CrossRef](#)]
206. Eriksen, E.F.; Shabestari, M.; Ghouri, A.; Conaghan, P.G. Bisphosphonates as a treatment modality in osteoarthritis. *Bone* **2021**, *143*, 115352. [[CrossRef](#)]
207. Shangguan, L.; Ding, M.; Wang, Y.; Xu, H.; Liao, B. Denosumab ameliorates osteoarthritis by protecting cartilage against degradation and modulating subchondral bone remodeling. *Regen. Ther.* **2024**, *27*, 181–190. [[CrossRef](#)]
208. Mohamad, N.V.; Ima-Nirwana, S.; Chin, K.Y. Self-emulsified annatto tocotrienol improves bone histomorphometric parameters in a rat model of oestrogen deficiency through suppression of skeletal sclerostin level and RANKL/OPG ratio. *Int. J. Med. Sci.* **2021**, *18*, 3665–3673. [[CrossRef](#)]
209. Hamood, R.; Hamood, H.; Merhasin, I.; Keinan-Boker, L. Hormone therapy and osteoporosis in breast cancer survivors: Assessment of risk and adherence to screening recommendations. *Osteoporos. Int.* **2019**, *30*, 187–200. [[CrossRef](#)]
210. Meyer, C.; Brockmueller, A.; Buhrmann, C.; Shakibaei, M. Prevention and Co-Management of Breast Cancer-Related Osteoporosis Using Resveratrol. *Nutrients* **2024**, *16*, 708. [[CrossRef](#)]
211. Ebstein, E.; Brocard, P.; Soussi, G.; Khoury, R.; Forien, M.; Khalil, A.; Vauchier, C.; Juge, P.A.; Léger, B.; Ottaviani, S.; et al. Burden of comorbidities: Osteoporotic vertebral fracture during non-small cell lung cancer—The BONE study. *Eur. J. Cancer* **2024**, *200*, 113604. [[CrossRef](#)] [[PubMed](#)]
212. Shi, L.; Hou, J.; Wang, L.; Fu, H.; Zhang, Y.; Song, Y.; Wang, X. Regulatory roles of osteopontin in human lung cancer cell epithelial-to-mesenchymal transitions and responses. *Clin. Transl. Med.* **2021**, *11*, e486. [[CrossRef](#)]
213. Guo, J.; Tong, C.Y.; Shi, J.G.; Li, X.J.; Chen, X.Q. Deletion of osteopontin in non-small cell lung cancer cells affects bone metabolism by regulating miR-34c/Notch1 axis: A clue to bone metastasis. *Eur. J. Histochem. EJH* **2023**, *67*, 3631. [[CrossRef](#)]

214. Macedo, F.; Ladeira, K.; Pinho, F.; Saraiva, N.; Bonito, N.; Pinto, L.; Goncalves, F. Bone Metastases: An Overview. *Oncol. Rev.* **2017**, *11*, 321.
215. Verbruggen, S.W.; Nolan, J.; Duffy, M.P.; Pearce, O.M.T.; Jacobs, C.R.; Knight, M.M. A Novel Primary Cilium-Mediated Mechanism Through which Osteocytes Regulate Metastatic Behavior of Both Breast and Prostate Cancer Cells. *Adv. Sci.* **2024**, *11*, e2305842. [[CrossRef](#)]
216. Dumanskiy, Y.V.; Syniachenko, O.V.; Stepko, P.A.; Taktashov, G.S.; Chernyshova, O.Y.; Stoliarova, O.Y. The state of bone metabolism in lung cancer patients. *Exp. Oncol.* **2018**, *40*, 136–139. [[CrossRef](#)] [[PubMed](#)]
217. Whitburn, J.; Edwards, C.M. Metabolism in the Tumour-Bone Microenvironment. *Curr. Osteoporos. Rep.* **2021**, *19*, 494–499. [[CrossRef](#)] [[PubMed](#)]
218. Fan, H.; Xu, Z.; Yao, K.; Zheng, B.; Zhang, Y.; Wang, X.; Zhang, T.; Li, X.; Hu, H.; Yue, B.; et al. Osteoclast Cancer Cell Metabolic Cross-talk Confers PARP Inhibitor Resistance in Bone Metastatic Breast Cancer. *Cancer Res.* **2024**, *84*, 449–467. [[CrossRef](#)]
219. Verbruggen, S.W.; Thompson, C.L.; Duffy, M.P.; Lunetto, S.; Nolan, J.; Pearce, O.M.T.; Jacobs, C.R.; Knight, M.M. Mechanical Stimulation Modulates Osteocyte Regulation of Cancer Cell Phenotype. *Cancers* **2021**, *13*, 2906. [[CrossRef](#)]
220. Cline-Smith, A.; Axelbaum, A.; Shashkova, E.; Chakraborty, M.; Sanford, J.; Panesar, P.; Peterson, M.; Cox, L.; Baldan, A.; Veis, D.; et al. Ovariectomy Activates Chronic Low-Grade Inflammation Mediated by Memory T Cells, Which Promotes Osteoporosis in Mice. *J. Bone Miner. Res.* **2020**, *35*, 1174–1187. [[CrossRef](#)]
221. Kwon, M.J.; Kim, J.H.; Kim, J.H.; Cho, S.J.; Nam, E.S.; Choi, H.G. The Occurrence of Alzheimer’s Disease and Parkinson’s Disease in Individuals With Osteoporosis: A Longitudinal Follow-Up Study Using a National Health Screening Database in Korea. *Front. Aging Neurosci.* **2021**, *13*, 786337. [[CrossRef](#)] [[PubMed](#)]
222. Dengler-Crish, C.M.; Eleftheriou, F. Shared mechanisms: Osteoporosis and Alzheimer’s disease? *Aging* **2019**, *11*, 1317–1318. [[CrossRef](#)] [[PubMed](#)]
223. van den Bos, F.; Speelman, A.D.; Samson, M.; Munneke, M.; Bloem, B.R.; Verhaar, H.J. Parkinson’s disease and osteoporosis. *Age Ageing* **2013**, *42*, 156–162. [[CrossRef](#)] [[PubMed](#)]
224. Liu, X.; Chen, C.; Jiang, Y.; Wan, M.; Jiao, B.; Liao, X.; Rao, S.; Hong, C.; Yang, Q.; Zhu, Y.; et al. Brain-derived extracellular vesicles promote bone-fat imbalance in Alzheimer’s disease. *Int. J. Biol. Sci.* **2023**, *19*, 2409–2427. [[CrossRef](#)]
225. He, J.Y.; Jiang, L.S.; Dai, L.Y. The roles of the sympathetic nervous system in osteoporotic diseases: A review of experimental and clinical studies. *Ageing Res. Rev.* **2011**, *10*, 253–263. [[CrossRef](#)]
226. Togari, A.; Arai, M. Pharmacological topics of bone metabolism: The physiological function of the sympathetic nervous system in modulating bone resorption. *J. Pharmacol. Sci.* **2008**, *106*, 542–546. [[CrossRef](#)]
227. Zhang, W.; Zhang, Y.; Hu, N.; Wang, A. Alzheimer’s disease-associated inflammatory pathways might contribute to osteoporosis through the interaction between PROK2 and CSF3. *Front. Neurol.* **2022**, *13*, 990779. [[CrossRef](#)]
228. Lee, D.Y.; Na, D.L.; Seo, S.W.; Chin, J.; Lim, S.J.; Choi, D.; Min, Y.K.; Yoon, B.K. Association between cognitive impairment and bone mineral density in postmenopausal women. *Menopause* **2012**, *19*, 636–641. [[CrossRef](#)]
229. Zhang, Y.W.; Cao, M.M.; Li, Y.J.; Dai, G.C.; Lu, P.P.; Zhang, M.; Bai, L.Y.; Chen, X.X.; Zhang, C.; Shi, L.; et al. The regulative effect and repercussion of probiotics and prebiotics on osteoporosis: Involvement of brain-gut-bone axis. *Crit. Rev. Food Sci. Nutr.* **2023**, *63*, 7510–7528. [[CrossRef](#)]
230. Jiang, Y.L.; Wang, Z.X.; Liu, X.X.; Wan, M.D.; Liu, Y.W.; Jiao, B.; Liao, X.X.; Luo, Z.W.; Wang, Y.Y.; Hong, C.G.; et al. The Protective Effects of Osteocyte-Derived Extracellular Vesicles Against Alzheimer’s Disease Diminished with Aging. *Adv. Sci.* **2022**, *9*, e2105316. [[CrossRef](#)]
231. Fehsel, K.; Christl, J. Comorbidity of osteoporosis and Alzheimer’s disease: Is ‘AKT’-ing on cellular glucose uptake the missing link? *Ageing Res. Rev.* **2022**, *76*, 101592. [[CrossRef](#)]
232. Rice, J.N.; Gillett, C.B.; Malas, N.M. The Impact of Psychotropic Medications on Bone Health in Youth. *Curr. Psychiatry Rep.* **2018**, *20*, 104. [[CrossRef](#)]
233. Zameer, S.; Najmi, A.K.; Vohora, D.; Akhtar, M. Bisphosphonates: Future perspective for neurological disorders. *Pharmacol. Rep. PR* **2018**, *70*, 900–907. [[CrossRef](#)]
234. Iwamoto, J.; Sato, Y.; Tanaka, K.; Takeda, T.; Matsumoto, H. Prevention of hip fractures by exposure to sunlight and pharmacotherapy in patients with Alzheimer’s disease. *Aging Clin. Exp. Res.* **2009**, *21*, 277–281. [[CrossRef](#)] [[PubMed](#)]
235. Papaconstantinou, J. The Role of Signaling Pathways of Inflammation and Oxidative Stress in Development of Senescence and Aging Phenotypes in Cardiovascular Disease. *Cells* **2019**, *8*, 1383. [[CrossRef](#)] [[PubMed](#)]
236. López-Otín, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. *Cell* **2013**, *153*, 1194–1217. [[CrossRef](#)] [[PubMed](#)]
237. Masud, T.; Frost, M.; Ryg, J.; Matzen, L.; Ibsen, M.; Abrahamsen, B.; Brixen, K. Central nervous system medications and falls risk in men aged 60–75 years: The Study on Male Osteoporosis and Aging (SOMA). *Age Ageing* **2013**, *42*, 121–124. [[CrossRef](#)]
238. Ebert, T.; Tran, N.; Schurgers, L.; Stenvinkel, P.; Shiels, P.G. Ageing—Oxidative stress, PTMs and disease. *Mol. Asp. Med.* **2022**, *86*, 101099. [[CrossRef](#)]

239. Li, L.; Zhao, S.; Xiang, T.; Feng, H.; Ma, L.; Fu, P. Epigenetic connection between gut microbiota-derived short-chain fatty acids and chromatin histone modification in kidney diseases. *Chin. Med. J.* **2022**, *135*, 1692–1694. [[CrossRef](#)]
240. Kang, J.S.; Alliston, T.; Delston, R.; Derynck, R. Repression of Runx2 function by TGF-beta through recruitment of class II histone deacetylases by Smad3. *EMBO J.* **2005**, *24*, 2543–2555. [[CrossRef](#)]
241. Sikora, E.; Bielak-Zmijewska, A.; Dudkowska, M.; Krzystyniak, A.; Mosieniak, G.; Wesierska, M.; Wlodarczyk, J. Cellular Senescence in Brain Aging. *Front. Aging Neurosci.* **2021**, *13*, 646924. [[CrossRef](#)] [[PubMed](#)]
242. Kong, X.; Gong, Z.; Zhang, L.; Sun, X.; Ou, Z.; Xu, B.; Huang, J.; Long, D.; He, X.; Lin, X.; et al. JAK2/STAT3 signaling mediates IL-6-inhibited neurogenesis of neural stem cells through DNA demethylation/methylation. *Brain Behav. Immun.* **2019**, *79*, 159–173. [[CrossRef](#)] [[PubMed](#)]
243. Jung, Y.D.; Park, S.K.; Kang, D.; Hwang, S.; Kang, M.H.; Hong, S.W.; Moon, J.H.; Shin, J.S.; Jin, D.H.; You, D.; et al. Epigenetic regulation of miR-29a/miR-30c/DNMT3A axis controls SOD2 and mitochondrial oxidative stress in human mesenchymal stem cells. *Redox Biol.* **2020**, *37*, 101716. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.