

Mitochondrial genomes of the headstanders *Megaleporinus muyscorum* (Steindachner 1900) and *Megaleporinus obtusidens* (Valenciennes 1837), (Characiformes, Anostomidae)

Melissa Rincón-Sandoval^a  Ricardo Betancur-R^b and Javier A. Maldonado-Ocampo^a 

^aLaboratorio de Ictiología, Unidad de Ecología y Sistemática -UNESIS-, Departamento de Biología, Facultad de Ciencias, Pontificia Universidad Javeriana, Bogotá, Colombia; ^bDepartment of Biology, University of Puerto Rico – Río Piedras, San Juan, Puerto Rico

ABSTRACT

We report two mitochondrial genomes of headstanders, derived from target capture and Illumina sequencing (HiSeq 2500 PE100). One trans-Andean species *Megaleporinus muyscorum* (mitochondrial consensus genome of 25 individuals) from Colombia and one cis-Andean species *M. obtusidens* from Argentina. Regarding *M. muyscorum*, mitochondrial genome has 13 protein-coding genes, 1 D-loop, 2 ribosomal RNAs, 21 transfer RNAs, and is 14,434 bp in length, for *M. obtusidens* mitochondrial genome has 13 protein-coding genes, 2 ribosomal RNAs, 22 transfer RNAs, and is 15,546 bp in length.

ARTICLE HISTORY

Received 9 April 2018
Accepted 2 May 2018

KEYWORDS

Headstanders; mitochondrial genomes; target capture

The family Anostomidae is composed by primary freshwater species that occur in South America, commonly known as headstanders that are broadly distributed on both sides of the Andes (Nelson 1994). This family comprises 14 genera and 156 species (Eschmeyer et al. 2018). The species belonging to the genus *Megaleporinus* are used in fisheries and aquaculture and although these species are in a category of least concern according to CITES, it is necessary to follow up the populations and this is one of the reasons why the information of the mitochondrial genome of these species will be of great importance given that have not been published so far.


In this work, 25 individuals in total of *Megaleporinus muyscorum* were collected in field at Atrato, Cauca, Magdalena, San Jorge, and Sinu rivers basins (five individuals per basin) in Colombia and a *M. obtusidens* tissue was from Iguazu (Argentina) belonging to the STRI collection (STRI-2487); for sampling see Appendix S1 (Supporting information). Total DNA was obtained from 20 mg of muscle with the BioSprint 96 from QIAGEN[®] at The Smithsonian Tropical Research Institute (STRI). Following extraction, we quantified DNA extracts using a Qubit fluorometer (Life Technologies, Inc, Carlsbad, CA) and samples were sent to MYcroarray, Inc (Ann Arbor, MI) for a targeted sequencing approach. Probes were targeted (>200 base pairs) using target capture and Illumina sequencing (HiSeq 2500 PE100). The capture target design is described in detail by Arcila et al. (2017). Although probes were not designed to target mtDNA genes, we tested whether the raw Illumina data contained mtDNA genomes.

We mapped reads from each sample to this most relative available mitochondrial genome of reference *M. elongatus* (GenBank NC034281) using SAMTOOLS (v1.3.1; Li et al. 2009). Then, we winnowed the resulting SAM to only those read alignments hitting the highest-hit references for each locus that were selected for downstream analysis. After that, we removed PCR duplicates converting the SAM to BAM and used the SAMTOOLS rmdup program. The final rmdup.bam file that contained all contigs, was imported to GENEIOUS (v8.1.9; Kearsse et al. 2012). Contigs were assembled by mapping sequences against the master reference using the ‘high sensitivity/medium’ algorithm with five iterations and ends were trimmed using defaults. A consensus sequence per each species was extracted from the multiple alignment, and were cleaned posteriorly and visually inspected to detect stop codons. The mtDNA genome was annotated using MitoAnnotator (Iwasaki et al. 2013).

The mitochondrial consensus genome of *M. muyscorum* (Genbank MH286914) is complete, with 13 protein-coding genes, 1 D-loop, 2 ribosomal RNAs, and 21 transfer RNAs (we couldn’t recover through capture 1 tRNA) for a total length of 14434 bp. For *M. obtusidens* (Genbank MH286915) we couldn’t recover D-loop, this genome has 13 protein-coding genes, 2 ribosomal RNAs, and 22 transfer RNAs for a total length of 15546 bp. Base compositions are typical for vertebrate mitochondrial genomes with 29% A, 25.8% C, 17.9% G, 27.3% T for *M. muyscorum* and 28.5% A, 27.9% C, 17.5% G, 26.1% T for *M. obtusidens*.

We reconstructed a phylogenetic tree based on the complete mitochondrial genome of 26 individuals belonging to

CONTACT Javier A. Maldonado-Ocampo  maldonadoj@javeriana.edu.co  Laboratorio de Ictiología, Unidad de Ecología y Sistemática -UNESIS-, Departamento de Biología, Facultad de Ciencias, Pontificia Universidad Javeriana, Bogotá, Colombia

 Supplemental data for this article can be accessed [here](#).

© 2018 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

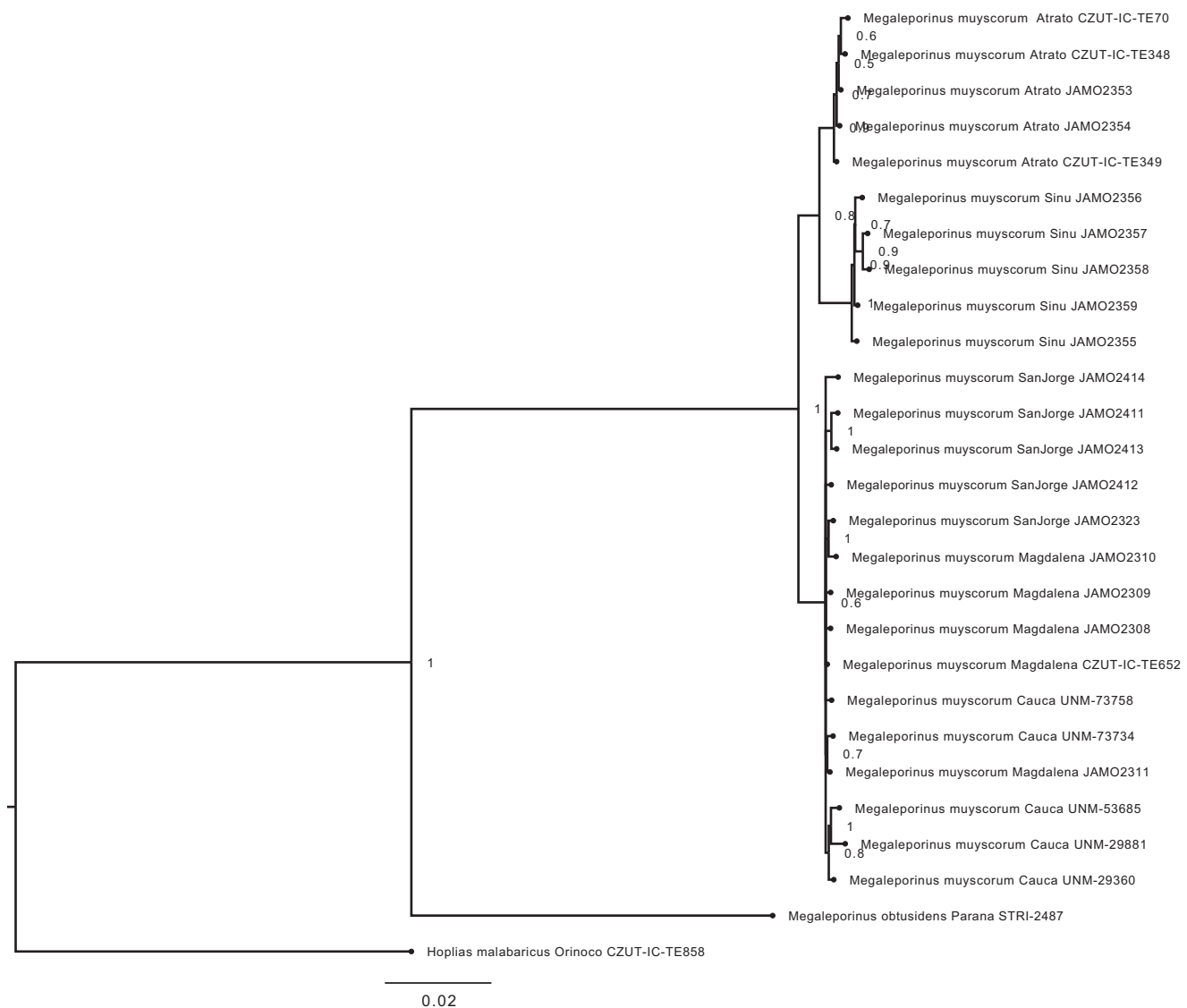


Figure 1. Molecular phylogeny of headstanders *Megaleporinus muyscorum* and *Megaleporinus obtusidens*, based on complete mitogenome. Support values at each node are Bayesian posterior probabilities. Branch label include information about sampled basin and tissue availability (JAMO: Pontificia Universidad Javeriana; CZUT-IC-TE: Universidad del Tolima; UNM: Universidad Nacional de Colombia ? Medellin; STRI: Smithsonian Tropical Research Institute).

the genus *Megaleporinus*. The Bayesian tree was performed using MrBayes (v3.2.6; Huelsenbeck and Ronquist 2001; Ronquist and Huelsenbeck 2003) for 8 MCMC runs, 10 million of generations, and sampled every 1000. Cis-andean species *M. obtusidens* and *H. malabaricus* are both the sister group of the trans-Andean species; three remarkable cluster are depicted in Figure 1. Atrato, Sinu, and the other one comprise the individuals belonging to the Magdalena basin.

Acknowledgements

The authors would like to thank Universidad del Tolima (Colombia), Universidad Nacional de Colombia -Medellin- (Colombia) and Smithsonian Tropical Research Institute (Panama) for helping providing tissue samples and in field work.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the Division of Research of the Pontificia Universidad Javeriana (Grant number: 00006714) to J.A.M.O., and by National Science Foundation (NSF) grants [DEB-147184, DEB- 1541491] to R.B.R. M.R.S was supported by a fellowship from Colciencias, through grant [567-2012]. This work was funded by Departamento Administrativo de Ciencia, Tecnología e Innovación.

ORCID

Melissa Rincón-Sandoval  <http://orcid.org/0000-0002-0200-5775>
 Javier A. Maldonado-Ocampo  <http://orcid.org/0000-0003-3024-237X>

References

Arcila D, Orti G, Vari R, Armbruster JW, Stiassny MLJ, Ko KD, Sabaj MH, Lundberg J, Revell LJ, Betancur-R R. 2017. Genome-wide interrogation advances resolution of recalcitrant groups in the tree of life. *Nature Ecology & Evolution*. 1:0020.

- Eschmeyer WN, Fricke R, van der Laan R, eds. 2018. Catalog of fishes: Anostomidae; [accessed 15 March 2018]. (<http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>).
- Huelsenbeck JP, Ronquist F. 2001. MRBAYES: Bayesian inference of phylogeny. *Bioinformatics*. 17:754–755.
- Iwasaki W, Fukunaga T, Isagozawa R, Yamada K, Maeda Y, Satoh TP, Sado T, Mabuchi K, Takeshima H, Miya M, Nishida M. 2013. MitoFish and MitoAnnotator: a mitochondrial genome database of fish with an accurate and automatic annotation pipeline. *Mol Biol Evol*. 30:2531–2540.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, et al. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics*. 28: 1647–1649.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, et al. 2009. Genome project data processing subgroup (2009) the sequence alignment/map format and SAMtools. *Bioinformatics*. 25:2078–2079.
- Nelson JS. 1994. *Fishes of the world*. 3rd ed. New York: John Wiley & Sons, Inc.; p. 600.
- Ronquist F, Huelsenbeck JP. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics*. 19:1572–1574.