





## Complete Genome Sequence of a *Listeria monocytogenes* Strain with Antimicrobial Resistance Genes Isolated from Lettuce in Canada

Amit Mathews, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Lin Ru Wang, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Lin Ru Wang, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Mingsong Kang, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Mingsong Kang, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Mingsong Kang, Mingsong Kang, Donail Naushad, Marc-Olivier Duceppe, Mingsong Kang, Mingsong Kan

<sup>a</sup>Ottawa Laboratory-Fallowfield, Canadian Food Inspection Agency, Ottawa, Ontario, Canada <sup>b</sup>Greater Toronto Area Laboratory, Canadian Food Inspection Agency, Toronto, Ontario, Canada

**ABSTRACT** *Listeria monocytogenes*, a Gram-positive, rod-shaped, non-spore-forming bacterium, is an important foodborne bacterial pathogen for humans worldwide. Here, we report the complete genome sequence of a Canadian *Listeria monocytogenes* strain with an antimicrobial resistance (AMR) gene that was isolated from lettuce.

*I isteria monocytogenes*, a Gram-positive bacterium that is found in various environments, animals, and food, is one of the most important foodborne pathogens for humans worldwide due to its high mortality rate (1). We report the complete genome sequence of *L. monocytogenes* strain GTA-L258, which was isolated from lettuce taken in Canada in 2015. The strain was isolated by preenrichment in *Listeria* enrichment broth at 30°C for 24 h, selective enrichment in buffered *Listeria* enrichment broth with morpholinepropanesulfonic acid (MOPS) (MOPS-BLEB) at 35°C for 18 to 24 h, and isolation from preenrichment cells grown on Oxford agar at 35°C for 48 h and on RAPID'*L.mono* (RLM) medium at 35°C for 24 h, with confirmation by hemolysis on blood agar plates at 35°C for 24 h, motility testing on tryptose agar at 30°C for 24 h, and Vitek (bioMérieux, Canada) testing (2).

Genomic DNA (gDNA) was extracted from an overnight culture in brain heart infusion medium using a Maxwell 16-cell DNA purification kit (Promega, USA) for Illumina sequencing and a NanoBind CBB Big DNA kit (Circulomics, USA) for Nanopore sequencing and was quantified using a Qubit fluorometer (Thermo Fisher Scientific, USA). MiSeq sequencing was conducted by library preparation with the Nextera XT library preparation kit (Illumina, USA) followed by sequencing for 600-bp cycles, which produced 576,050 paired-end reads with 89× coverage. Nanopore sequencing was performed using the 1D native barcoding gDNA protocol (EXP-NBD104 and SQK-LSK109; Oxford Nanopore Technologies, UK) without shearing, followed by sequencing using a FLO-MIN106 (R9.4.1) flow cell on a MinION MK1C device, which produced 115,064 reads (average length, 15,598 bp; N<sub>50</sub>, 35.6 kb) with 466× coverage. Base calling was performed using Guppy v5.0.11 in super high accuracy mode, trimming using Porechop v0.2.3 (3), and filtration using Filtlong v0.2.1 (4). Assembly of the long reads was performed using Flye v2.7 (5), corrected using Medaka v1.4.4, and polished with Illumina MiSeq reads using a combination of NextPolish v1.4.0, ntEdit v1.3.5, and Polypolish v0.5.0 after trimming and filtering with fastp v0.23.2. The circularity and genome rotation using dnaA as the starting point were determined using the fixstart plugin from Circlator v1.5.5 (6). The sequencing coverage depth was determined using minimap2 v2.17 (7) and SAMtools v1.13 (8) for long reads and BWA v0.7.17 and SAMtools v1.13 for short reads. Gene prediction and annotation were performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.0 (9).

Antimicrobial resistance (AMR) genes were identified using ResFinder v4.1.5 (10) and RGI v5.2.0 (11). The plasmids were identified by mlplasmids v1.0.0 using *Listeria monocytogenes* as the species model (12). Prophage sequences were analyzed using PHASTER (13) (accessed

**Editor** David Rasko, University of Maryland School of Medicine

© Crown copyright 2022. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Amit Mathews, amit.mathews@inspection.gc.ca.

The authors declare no conflict of interest.

Received 8 April 2022 Accepted 15 May 2022 Published 6 June 2022

July 2022 Volume 11 Issue 7 10.1128/mra.00298-22

online on 31 January 2022). Default parameters for pipelines were used except where otherwise noted (https://github.com/OLF-Bioinformatics/nanopore).

The GTA-L258 complete genome contains a single chromosome (2,939,920 bp) harboring a *fosX* AMR gene, without identification of plasmids or intact prophages (PHASTER score of >90). The annotated genome contains 2,839 coding sequences (CDSs) and 67 tRNAs, with a genomic GC content of 37.89%, similar to 2,889 CDSs and a GC content of 37.88% on average for *L. monocytogenes* strains in the NCBI database (accessed 31 January 2022).

**Data availability.** The whole-genome sequence of GTA-L258 was deposited in GenBank under accession number CP092060. MinION base-called and MiSeq base-called fastq files are available in the NCBI Sequence Read Archive (SRA) under accession numbers SRR17965225 and SRR17965217, respectively.

## **ACKNOWLEDGMENTS**

This work was funded by the Canadian Food Inspection Agency.

We thank the Microbiology Laboratory of the Canadian Food Inspection Agency Greater Toronto Area Laboratory for technical support for bacterial isolation.

## **REFERENCES**

- Radoshevich L, Cossart P. 2018. Listeria monocytogenes: towards a complete picture of its physiology and pathogenesis. Nat Rev Microbiol 16:32–46. https://doi.org/10.1038/nrmicro.2017.126.
- Pagotto F, Hebert K, Farber J. 2011. Isolation of *Listeria monocytogenes* and other *Listeria* spp. from foods and environmental samples: MFHPB-30. *In* HPB methods for the microbiological analysis of foods, vol 2. The compendium of analytical methods. Health Canada, Ottawa, Canada.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. Microb Genom 3:e000132. https://doi.org/10.1099/mgen.0.000132.
- Kang M, Chmara J, Duceppe MO, Phipps-Todd B, Huang H. 2020. Complete genome sequence of a Canadian Klebsiella michiganensis strain, obtained using Oxford Nanopore Technologies sequencing. Microbiol Resour Announc 9:e00960-20. https://doi.org/10.1128/MRA.00960-20.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, errorprone reads using repeat graphs. Nat Biotechnol 37:540–546. https://doi .org/10.1038/s41587-019-0072-8.
- Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. Genome Biol 16:294. https://doi.org/10.1186/s13059-015-0849-0.
- Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. Bioinformatics 34:3094–3100. https://doi.org/10.1093/bioinformatics/bty191.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. Bioinformatics 25:2078–2079. https://doi.org/10.1093/bioinformatics/btp352.

- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/ nar/qkw569.
- Bortolaia V, Kaas RS, Ruppe E, Roberts MC, Schwarz S, Cattoir V, Philippon A, Allesoe RL, Rebelo AR, Florensa AF, Fagelhauer L, Chakraborty T, Neumann B, Werner G, Bender JK, Stingl K, Nguyen M, Coppens J, Xavier BB, Malhotra-Kumar S, Westh H, Pinholt M, Anjum MF, Duggett NA, Kempf I, Nykäsenoja S, Olkkola S, Wieczorek K, Amaro A, Clemente L, Mossong J, Losch S, Ragimbeau C, Lund O, Aarestrup FM. 2020. ResFinder 4.0 for predictions of phenotypes from genotypes. J Antimicrob Chemother 75:3491–3500. https://doi.org/10.1093/jac/dkaa345.
- 11. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen AV, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran HK, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F, Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistome surveillance with the Comprehensive Antibiotic Resistance Database. Nucleic Acids Res 48:D517–D525. https://doi.org/10.1093/nar/gkz935.
- Arredondo-Alonso S, Rogers MRC, Braat JC, Verschuuren TD, Top J, Corander J, Willems RJL, Schurch AC. 2018. mlplasmids: a user-friendly tool to predict plasmid- and chromosome-derived sequences for single species. Microb Genom 4:e000224. https://doi.org/10.1099/mgen.0.000224.
- Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. Nucleic Acids Res 44:W16–W21. https://doi.org/10.1093/nar/gkw387.

July 2022 Volume 11 Issue 7 10.1128/mra.00298-22