



# Draft Genome Sequences of 27 *Salmonella enterica* Serovar Schwarzengrund Isolates from Clinical Sources

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**ABSTRACT** Twenty-seven *Salmonella enterica* serovar Schwarzengrund isolates from clinical sources were sequenced as part of a larger study to examine phenotypic and genotypic characteristics. The majority of the sequenced strains were isolated from human stool ( $n = 20$ ) followed by urine ( $n = 3$ ) and blood ( $n = 2$ ). Four isolate sequences contained plasmids of known incompatibility groups.

*Salmonella enterica* serovar Schwarzengrund strains can cause salmonellosis in humans and infections in other animal species (1). The spread of multidrug-resistant *S. Schwarzengrund* from imported food products to humans has been reported (1). *S. Schwarzengrund* strains isolated from imported food products have been found to display a high level of resistance to fluoroquinolones (2). Furthermore, the production of extended-spectrum  $\beta$ -lactamase (ESBL), including carbapenemase (KPC-2), by *S. Schwarzengrund* has been found in different countries, which concerns observers (3–5). ESBL genes are often encoded on plasmids (3, 4, 6), which can facilitate their rapid transfer among Gram-negative pathogens, and this has created a major public health concern. Therefore, monitoring antimicrobial resistance (AMR) dynamics and characterization of mobile genetic elements, including plasmids of *S. Schwarzengrund* isolated from domestic and imported foods, food animals, and humans, is important. In the present study, whole-genome sequencing analyses of *S. Schwarzengrund* isolates from clinical sources will facilitate the study of AMR and resistance transmission.

Twenty-seven *S. Schwarzengrund* isolates from clinical sources were collected from the Minnesota Department of Health (Saint Paul, MN) and sequenced at the Division of Microbiology, National Center for Toxicological Research, U.S. Food and Drug Administration (Jefferson, AR). Epidemiological information for these isolates is listed in Table 1.

Stool samples were plated on Hektoen enteric agar, salmonella-shigella agar, and enrichment broths (Becton, Dickinson and Company [BD], Franklin Lakes, NJ). The plate or broth was then incubated at 35°C for 18 to 24 h. Colonies that were suspiciously lactose negative and positive for H<sub>2</sub>S were subsequently inoculated in motility-indole-lysine agar, triple sugar iron agar, and a urea agar plate (BD). Identification was confirmed using matrix-assisted laser desorption ionization–time of flight mass (MALDI-TOF) spectrometry. Likewise, blood and other clinical isolates were identified using standard microbiological procedures.

The total bacterial DNA was extracted with a DNeasy blood and tissue kit (Qiagen, Valencia, CA), and DNA sequencing libraries were constructed with the Nextera XT DNA library preparation kit (Illumina, San Diego, CA). Samples were multiplexed with a unique combination of two indexes of the Nextera XT index kit. Whole-genome sequencing (WGS) reactions were carried out on an Illumina MiSeq instrument with a 2 × 300 paired-end format (7). Trimming and *de novo* assembly were performed with

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**TABLE 1** Whole-genome sequencing analyses of *Salmonella* Schwarzengrund isolates from clinical sources

Strain no.	Source of isolate	Yr of isolation	No. of sequence reads	$N_{50}$ value (bp)	No. of contigs	Assembly size (bp)	No. of CDSs	G+C content (%)	GenBank accession no.
MDH-1	Blood	2003	4,813,548	255,417	61	4,771,771	4,845	52.2	<a href="#">QZET00000000</a>
MDH-2	Stool	2003	3,729,214	340,162	56	4,775,162	4,828	52.2	<a href="#">QZES00000000</a>
MDH-3	Stool	2004	3,706,472	112,364	100	4,684,005	4,717	52.1	<a href="#">QZER00000000</a>
MDH-4	Stool	2004	3,221,282	167,201	80	4,674,736	4,715	52.1	<a href="#">QZEQ00000000</a>
MDH-5	Stool	2004	4,754,358	149,628	118	4,715,784	4,767	52.0	<a href="#">QZEP00000000</a>
MDH-6	Stool	2005	3,945,558	138,815	101	4,774,615	4,854	52.2	<a href="#">QZEO00000000</a>
MDH-7	Stool	2005	4,316,768	93,969	128	4,775,144	4,853	52.2	<a href="#">QZEN00000000</a>
MDH-8	Stool	2006	2,438,762	91,707	158	5,117,666	5,291	51.6	<a href="#">QZEM00000000</a>
MDH-9	Stool	2007	3,737,938	85,408	119	4,700,238	4,787	52.0	<a href="#">QZEL00000000</a>
MDH-10	Stool	2007	4,576,284	102,545	106	4,693,974	4,766	52.0	<a href="#">QZEK00000000</a>
MDH-11	Stool	2008	4,129,638	389,856	43	4,722,650	4,752	52.2	<a href="#">QZFD00000000</a>
MDH-12	Stool	2009	4,399,374	230,080	54	4,706,223	4,745	52.1	<a href="#">QZFE00000000</a>
MDH-13	Stool	2010	4,764,064	310,035	49	4,739,724	4,806	52.1	<a href="#">QZFF00000000</a>
MDH-14	Blood	2011	3,208,474	312,411	51	4,767,578	4,829	52.2	<a href="#">QZFG00000000</a>
MDH-15	Stool	2011	2,124,138	177,815	85	4,766,481	4,829	52.2	<a href="#">QZMP00000000</a>
MDH-16	Stool	2012	1,892,712	153,861	67	4,800,611	4,894	52.0	<a href="#">QZFH00000000</a>
MDH-17	Stool	2012	1,532,044	123,253	100	4,802,467	4,863	52.0	<a href="#">QZFI00000000</a>
MDH-18	Gallbladder	2013	3,806,536	75,786	162	4,881,277	5,005	52.1	<a href="#">QZFJ00000000</a>
MDH-19	Urine	2013	2,291,464	60,132	201	4,775,923	4,879	52.2	<a href="#">QZFK00000000</a>
MDH-20	Subhepatic aspirate	2013	2,936,934	69,988	168	4,888,146	5,028	52.1	<a href="#">QZFL00000000</a>
MDH-21	Stool	2014	2,343,572	57,255	195	4,688,259	4,773	52.1	<a href="#">QZFM00000000</a>
MDH-22	Stool	2014	2,023,560	59,303	176	4,687,016	4,762	52.1	<a href="#">QZMQ00000000</a>
MDH-23	Urine	2014	1,872,516	52,555	195	4,690,085	4,776	52.1	<a href="#">QZFN00000000</a>
MDH-24	Stool	2014	2,607,656	53,183	197	4,682,121	4,730	52.2	<a href="#">QZMR00000000</a>
MDH-25	Stool	2015	2,424,548	35,368	313	4,778,730	4,868	52.1	<a href="#">QZFO00000000</a>
MDH-26	Urine	2016	3,795,856	36,628	274	4,807,879	4,944	52.0	<a href="#">QZFP00000000</a>
MDH-27	Stool	2016	2,566,758	54,067	225	4,807,042	4,891	52.0	<a href="#">QZFQ00000000</a>

CLC Genomics Workbench (v. 9, Qiagen, Germantown, MD). Genome sequences from individual samples were examined simultaneously with individual annotation tools, such as Rapid Annotation using Subsystem Technology (RAST) (8) and Pathosystems Resource Integration Center (PATRIC) (9), to cross-examine the sequence data. Subsequently, sequences were submitted to the NCBI using the WGS submission portal for final annotation with the Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) (10) (Table 1). The number of contigs, assembly size, coding sequences (CDSs), and G+C contents of each sample included in Table 1 were annotated with PATRIC. Annotation performed with the PGAAP was used as the final annotation available in the NCBI. We applied default settings for the bioinformatic software tools used for sequence trimming, *de novo* assembly, and annotation of the sequences.

Among 27 *S. Schwarzengrund* clinical isolates, PlasmidFinder (11) analyses showed that four of the isolates contained plasmids of identified incompatibility (Inc) groups. MDH-8 contained the IncHI2 and IncHI2A plasmids, and MDH-18, MDH-20, and MDH-25 contained the IncI1 plasmid. ResFinder (12) analyses showed that MDH-8 contained the *aph(6)-Id*, *strA*, and *tet(B)* resistance genes; MDH-18 and MDH-20 contained the *aph(3')-Ia* and *bla<sub>CMY-2</sub>* genes; and MDH-25 contained the *aac(3)-VIa*, *aadA1*, and *sul1* genes.

**Data availability.** This whole-genome shotgun project is deposited at DDBJ/ENA/GenBank under the accession numbers listed in Table 1, and the SRA submission of raw data (FastQ format) is recorded under the accession number [PRJNA312617](#).

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