

Contents lists available at ScienceDirect

Data in Brief





Data Article

Agronomical and analytical trait data assessed in a set of quinoa genotypes growing in the UAE under different irrigation salinity conditions



Fatima Zahra Rezzouk^a, Mohammad Ahmed Shahid^b, Ismahane A. Elouafi^b, Bangwei Zhou^c, José L. Araus^a, Maria D. Serret^a,*

ARTICLE INFO

Article history: Received 30 April 2020 Revised 18 May 2020 Accepted 18 May 2020 Available online 30 May 2020

Keywords: Irrigation Isotopic composition Leaf pigments Mineral content Manuring, Quinoa Seed yield

ABSTRACT

The importance of quinoa has been emphasized considerably in the recent decades, as a highly nutritional crop seed that is tolerant to salinity and amenable to arid agronomical conditions. The focus of this paper is to provide raw and a supplemental data of the research article entitled "Agronomic performance of irrigated quinoa in desert areas: comparing different approaches for early assessment of salinity stress" [1], aiming to compare different approaches for early detection, at the genotypic and crop levels, of the effect of salinity caused by irrigation on the agronomic performance of this crop. A set of 20 genotypes was grown under drip irrigation in sandy soil, amended with manure, at the International Center for Biosaline Agriculture (UAE) for two weeks, after which half of the trial was submitted to irrigation with saline water and this was continued until crop maturity. After eight weeks of applying the two irrigation regimes, pigment contents were evaluated in fully expanded leaves. The

E-mail address: dserret@ub.edu (M.D. Serret).

^a Section of Plant Physiology, University of Barcelona, 08028 Barcelona, and AGROTECNIO (Center of Research in Agrotechnology), 25198 Lleida, Spain.

^b International Center for Biosaline Agriculture (ICBA), P.O. Box 14660, Dubai, U.A.E.

^cKey Laboratory of Vegetation Ecology, Ministry of Education, Institute of Grassland Science, Northeast Normal University, Changchun, China.

^{*} Corresponding author. Maria D. Serret, Section of Plant Physiology, University of Barcelona, 08028 Barcelona, and AGROTECNIO (Center of Research in Agrotechnology), 25198 Lleida, Spain.

same leaves were then harvested, dried and the stable carbon and nitrogen isotope compositions ($\delta^{13}\mathrm{C}$ and $\delta^{15}\mathrm{N}$) and the total nitrogen and carbon contents of the dry matter analyzed, together with ion concentrations. At maturity yield components were assessed and yield harvested. Data analysis demonstrated significant differences in genotypes response under each treatment, within all assessed parameters. The significant level was provided using the Tukey-b test on independent samples. The present dataset highlights the potential use of different approaches to crop phenotyping and monitoring decision making.

© 2020 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY license. (http://creativecommons.org/licenses/by/4.0/)

Specifications Table

Subject

Specific subject area

Type of data

How data were acquired

Agronomy and Crop Science

This dataset provides information comparing a wide range of approaches for early assessment of salinity stress in quinoa under irrigation and the negative

effect of excessive manuring.

Tables Figure

Leaf pigments were assessed using a portable leaf-clip sensor (Dualex, Dualex Force-A, Orsay, France). The Dualex sensor operates with a UV excitation beam at 357 nm, which corresponds to the maximum absorption for flavonoids, and a red reference beam at 650 nm, which corresponds to the maximum absorption for chlorophyll [2].

Stable isotopic composition of leaf dry matter were acquired by pulverizing dried leaf samples using a Mixer Mill (MM400, RETSCH GmbH, Germany) and subsampling approximately 1 mg of the pulverized material into tin capsules for further analysis using an elemental analyzer (Flash 1112 EA;

ThermoFinnigan, Schwerte, Germany) coupled with an isotope ratio mass spectrometer (Delta C IRMS, ThermoFinnigan), operating in continuous flow mode.

Soluble fraction was determined by subsampling 50 mg of the pulverized leaf material and suspending each sample with 1 mL of Milli-Q water in an Eppendorf tube (Eppendorf Scientific, Hamburg, Germany) for 20 min at about 5°C. The sample was then centrifuged at 12000 g for 5 min and at 5°C.

Afterwards, the supernatant containing the water-soluble fraction was pipetted into a new Eppendorf and heated at 100° C for 3 min to denature the proteins. Samples were centrifuged again (12000~g for 5 min at 5° C), and $100~\mu$ l of the resulting aliquot was placed in tin capsules and dried at 70° C for 2 hours. The soluble fraction of carbon and nitrogen isotope compositions was then determined in the same manner as the stable isotopic composition of the leaf dry matter.

Ion concentrations in leaves were obtained by acid-digesting and diluting 100 mg of each sample; then the solution was analyzed using an Inductively Coupled Plasma Emission Spectrometer (L3200RL, Perkin Elmer, Uberlingen, Germany).

Data format

Raw Analyzed

Parameters for data collection

Leaf pigment contents were determined around 8 weeks after the two irrigation treatments were imposed. Afterwards, the same leaves were washed with tap and distilled water, dried in an oven at 60°C for 48h, and ground to a fine powder for further ion and stable isotopic composition and total N and C analyses.

(continued on next page)

Description of data collection	Pigments were measured in 10 fully expanded leaves, selected from the central rows.
	At physiological maturity, 5 plants were selected from the central rows. Height was measured from the ground to the top of the inflorescence, and number of
	branches was recorded at different node positions. Number of inflorescences per plant was counted, and the length of 3 random inflorescences was
	averaged.
	Biomass and seed yield were assessed by manually harvesting the 5 plants from the middle row of each plot.
	Ion and stable isotopic composition were analyzed at the Scientific Facilities of the University of Barcelona
	Max, min and average temperature, and precipitation data were acquired from the meteorological station at ICBA.
Data source location	Institution: International Center for Biosaline Agriculture (ICBA) City: Dubai
	Country: The United Arab Emirates
	Latitude and longitude (and GPS coordinates) for collected samples/data: 25°05′49″ N and 55°23′25″E
Data accessibility	Repository name: Mendeley Data
,	DOI: 10.17632/r5ywtt8w39.1 (reserved but not active until publication)
	Direct URL to data:
	https://data.mendeley.com/datasets/r5ywtt8w39/draft?a=fb0d4661-eaf5-
	4781-80a5-0913bba85cb5
Related research article	Fatima Zahra Rezzouk, Mohammad Ahmed Shahid, Ismahane A. Elouafi,
	Bangwei Zhou, José L. Araus, Maria D. Serret, Agronomic performance of
	irrigated quinoa in desert areas: comparing different approaches for early assessment of salinity stress Agricultural Water Management

Data description

Supplemental tables displaying averaged values of yield components (supplemental table 1), ion concentrations (supplemental table 2), pigments (supplemental table 3), stables isotopes and their elemental analysis (supplemental table 4), of quinoa accessions grown under different irrigation treatments (fresh water and saline water), and genotypes (20 lines), exhibiting significant differences between treatments and among genotypes. Thus, means exhibiting different letters are significantly different (P < 0.05) by the post-hoc Tukey-b test on independent samples within each treatment (Fresh water and saline water). Values for accessions 10 and 18 under saline irrigation conditions are not included in the median separation because of the lack of replications. The distribution of climate parameters (maximum, minimum and average temperatures, and precipitation) during the quinoa growing period is displayed in supplemental Fig. 1.

For each trait, the values provided correspond to the three replicates per genotype and the two irrigation (fresh water and saline water) treatments. Assessed traits were: yield components (seed yield, biomass, plant height, branches, inflorescences, inflorescence length) at maturity, together with ion concentrations (sodium, phosphorus, potassium, calcium, magnesium concentrations and the K+/Na+, Ca²⁺/Na+ and Mg²⁺/Na+ ratios), leaf pigments (chlorophylls, flavonoids, anthocyanins and nitrogen balance index (NBI)), carbon and nitrogen concentrations on a dry matter basis, and carbon (δ^{13} C) and nitrogen (δ^{15} N) isotope composition in the dry matter and soluble fraction measured in fully expanded leaves 8 weeks after irrigation treatments were imposed are presented in the Raw data Tables 1, 2, 3 and 4.

Experimental Design, Materials, and Methods

Two field experiments were planted on November 19th, 2016. Quinoa seeds were sown by hand following a randomized complete block design with three replicates per genotype. Plot size

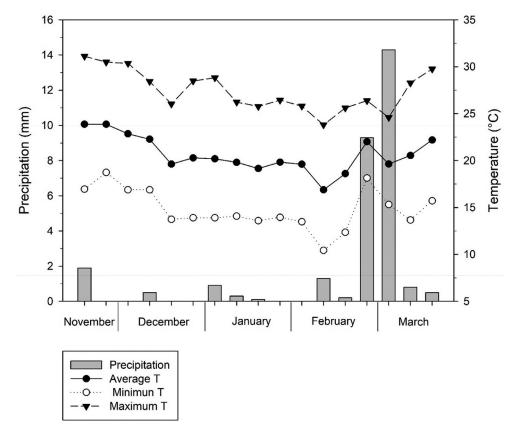


Fig. 1. Maximum, minimum and average temperature and precipitation during the quinoa growing period.

was 2×2 meters, with a plant-to-plant distance of 25 cm and 50 cm between rows, totaling 45 plants per plot (5×9) . During the two first weeks, both trials were supplied with fresh water drip-irrigation (1 dS m⁻¹) to avoid hindering germination. Then, two different treatments were imposed for the rest of the growing period to a) irrigation with fresh water and b) irrigation with saline water (15 dS m-1).

Eight weeks after treatments application, 10 fully expanded leaves were assessed randomly from the central rows of each plot in both trials, using a leaf pigment meter (Dualex). The same leaves were collected, dried, ground to a fine powder and analyzed for ion concentration determination using an Inductively Coupled Plasma Emission Spectrometer (ICPES), and stable isotope composition and elemental analysis determination, using an elemental analyzer coupled with an isotope ratio mass spectrometer (EA-IRMS).

At physiological maturity, yield components were assessed as described previously in Hussain et al. [3]: 5 plants were selected from the central rows. Height was measured from the ground to the top of inflorescence on the main stem. Similarly, the number of branches was recorded at different node positions of the main stem including basal branches. The number of inflorescences per plant was counted, and the length of 3 random inflorescences was averaged. Biomass and seed yield were assessed by manually harvesting the 5 plants from the middle row of each plot.

Table 1Average plant height, branches per plant, inflorescences per plant, inflorescence length, biomass and seed yield in the set of quinoa accessions grown under fresh water and saline irrigation treatments. Means exhibiting different letters are significantly different (P < 0.05) by the post-hoc Tukey-b test on independent samples within each treatment (fresh and saline water). Values for accessions 10 and 18 are presented but not included in the separation of means because of their poor agronomical performance, particularly under saline irrigation. Genotype numbers as detailed in Table 1.

Treatment	Genotype	Yield components Plant height (cm)	Branches plant ⁻¹	Inflorescences	Inflorescence length	Biomass (g m ⁻²)	Seed yield (g m ⁻²
				plant ⁻¹	(cm)		
Fresh water	1	102.6 ^{bc}	8.27 ^{ab}	7.60 ^{ab}	26.90 ^{cde}	2225 ^{ab}	504.8 ^a
	2	123.8 ^{ab}	7.67 ^{ab}	6.87 ^{ab}	39.90 ^b	1960 ^{ab}	459.3 ^a
	3	144.8a	4.60 ^b	4.00 ^b	50.83 ^a	2680 ^{ab}	392.2ab
	4	122.1 ^b	6.93 ^{ab}	5.93 ^{ab}	38.48 ^{bc}	2060 ^{ab}	400.7 ^{ab}
	5	88.2 ^{cd}	6.00 ^b	5.53 ^{ab}	27.90bcde	1427 ^b	297.0 ^{ab}
	6	88.9 ^{cd}	9.13 ^{ab}	7.87 ^{ab}	33.07 ^{bcde}	2460 ^{ab}	428.4ab
	7	100.6bcd	7.27 ^{ab}	6.53 ^{ab}	35.03 ^{bcd}	2000 ^{ab}	510.0a
	8	120.3 ^b	7.67 ^{ab}	5.20 ^{ab}	40.13b	1940 ^{ab}	544.3a
	9	114.9 ^b	7.47 ^{ab}	4.80 ^b	33.77 ^{bcde}	1400 b	401.3ab
	10	46.89-	7.13-	6.87-	17.99-	690-	42.15-
	11	111.7 ^{bc}	8.07 ^{ab}	6.33ab	34.77 ^{bcd}	2920ab	402.4ab
	12	117.9 ^b	11.07 ^a	9.07ª	39.07 ^{bc}	3080 ^{ab}	440.3ª
	13	107.8 ^{bc}	8.93 ^{ab}	8.13 ^{ab}	35.77 ^{bcd}	3440a	543.6a
	14	110.1 ^{bc}	8.27 ^{ab}	7.00 ^{ab}	33.37 ^{bcde}	2080 ^{ab}	363.3ab
	15	61.8 ^{ef}	7.80 ^{ab}	6.73 ^{ab}	23.97 ^{de}	2180 ^{ab}	323.4 ^{ab}
	16	53.8 ^f	6.33 ^{ab}	5.87 ^{ab}	22.20 ^e	1483 ^b	84.7 ^b
	17	111.3 ^{bc}	8.40 ^{ab}	7.33 ^{ab}	33.93bcde	2685 ^{ab}	632.4ª
	18	25,37-	5.73-	5.47-	10.53-	464-	50.8-
	19	104.5 ^{bc}	7.60 ^{ab}	7.13 ^{ab}	32.97bcde	2120 ^{ab}	503.0a
	20	77.2 ^{de}	8.73 ^{ab}	7.87 ^{ab}	29.37 ^{bcde}	1395 ^b	354.4 ^{ab}
Saline water	1	84.6 ^{bc}	6.73 ^b	6.07 ^b	25.53 ^{cdefg}	1487 ^{bc}	386.2 ^{abc}
Juline Water	2	85.7 ^{bc}	6.73 ^b	5.73 ^b	28.00 ^{bcdef}	1140 ^{bc}	187.3 ^{cd}
	3	115.1 ^a	6.07 ^b	5.33 ^b	39.53ª	1940 ^{abc}	221.5 ^{bcd}
	4	106.5ab	10.3 ^a	8.80a	35.67 ^{ab}	1700 ^{bc}	249.0 ^{abcd}
	5	78.5°	6.67 ^b	5.80 ^b	30.60 ^{bcde}	1410 ^{bc}	216.8 ^{bcd}
	6	80.9bc	5.40 ^b	5.40 ^b	28.88 ^{bcde}	1610 ^{bc}	442.4 ^a
	7	65.7 ^{cd}	4.80 ^b	4.60 ^b	27.37 ^{bcdefg}	1300 ^{bc}	286.9 ^{abcd}
	8	88.9bc	6.33 ^b	5.80 ^b	30.37 ^{bcde}	1620 ^{bc}	379.5 ^{abc}
	9	85.0 ^{bc}	6.07 ^b	5.60 ^b	31.40 ^{bcd}	1380 ^{bc}	289.8 ^{abcd}
	10	31.58-	2.90-	2.80-	14.5-	1300	203.0
	11	91.4 ^{bc}	7.53b	7.53 ^{ab}	33.80 ^{bcd}	3480a	416.1 ^{ab}
	12	47.9 ^d	6.47 ^b	6.13 ^b	24.57 ^{defg}	1707 ^{bc}	107.4 ^d
	13	83.9bc	6.13b	6.13b	32.50 ^{bcd}	2660 ^{abc}	281.8 ^{abcd}
	14	80.9 ^{bc}	6.80 ^b	6.53 ^{ab}	34.37 ^{abc}	2200 ^{abc}	380.1 ^{abc}
	15	52.7 d	5.27 ^b	4.67 ^b	21.80 ^{efg}	1242 ^{bc}	198.3 ^{cd}
	16	45.0 ^d	6.80 ^b	5.53b	18.90 ^g	2967 ^{ab}	226.8 ^{bcd}
	17	45.0° 82.6°	6.40 ^b	5.67 ^b	18.90° 27.87 ^{bcdef}	1715 ^{bc}	387.5 ^{abc}
	18	21.17	2.80 ⁻	2.67-	9.55 ⁻	1350-	51.8 ⁻
	19	66.2 ^{cd}	6.07 ^b	5.80 ^b	9.55 26.30 ^{cdefg}	1060°	280.1 ^{abcd}
	20	44.1 ^d	4.93b	4.60 ^b	19.28 ^{fg}	1060°	188.6 ^{bcd}
	20	44.1	4.93	4.00	19.28	1040	188.050

Table 2

Average sodium, phosphorus, potassium, calcium and magnesium concentrations and the K^+/Na^+ , Ca^{2+}/Na^+ and Mg^{2+}/Na^+ ratios in fully expanded leaves of quinoa accessions grown for eight weeks under different (fresh water and saline) irrigation treatments. Means exhibiting different letters are significantly different (P < 0.05) by the post-hoc Tukey-b test on independent samples within each treatment (fresh and saline water). Values for accession 18 under saline irrigation conditions are not included in the median separation because of the lack of replications. Genotype numbers as detailed in Table 1.

Treatment	Genotype	Ion concentration Na ⁺ (mmol.g ⁻¹)		$K^+(mmol.g^{-1})$	Ratios Ca ²⁺ (mmol.g ⁻¹)	Mg^{2+} (mmol.g $^{-1}$)	K ⁺ /Na ⁺	Ca ²⁺ /Na ⁺	Mg ²⁺ /Na ⁺
Fresh water	1	0.05 ^a	0.16 ^{bc}	1.61ª	0.48 ^{ef}	0.32 ^f	30.20 ^{ab}	8.99ª	5.95ª
	2	0.03 ^a	0.15 ^{bc}	1.65 ^a	0.49 ^{ef}	0.36 ^{ef}	64.06 ^{ab}	19.21a	13.92a
	3	0.04^{a}	0.18 ^{ab}	1.53 ^a	0.49 ^{ef}	0.43bcdef	45.69ab	14.37a	13.12a
	4	0.06^{a}	0.19 ^{ab}	1.57ª	0.59bcdef	0.42 ^{cdef}	44.17 ^{ab}	14.77 ^a	10.10 ^a
	5	0.07 ^a	0.09 ^{bc}	1.50 ^a	0.84 ^{ab}	0.62bc	23.41 ^{ab}	13.03a	9.57a
	6	0.14 ^a	0.05 ^c	2.03ª	0.78 ^{abcd}	0.42 ^{cdef}	16.72 ^{ab}	6.19 ^a	3.31a
	7	0.08^{a}	0.11bc	1.98ª	0.67 ^{bcdef}	0.61bc	25.90ab	8.49a	7.78 ^a
	8	0.05 ^a	0.18 ^{ab}	1.44 ^a	0.53 ^{def}	0.36 ^{ef}	29.51 ^{ab}	11.02ª	7.45 ^a
	9	0.05 ^a	0.17 ^{abc}	1.45 ^a	0.81 ^{abc}	0.52bcdef	34.66 ^{ab}	17.92ª	12.12a
	10	0.08^{a}	0.09 ^{bc}	1.95 ^a	0.70 ^{bcdef}	0.68 ^{ab}	25.78 ^{ab}	9.16 ^a	8.97ª
	11	0.04^{a}	0.17 ^{abc}	1.98ª	0.42 ^f	0.34 ^f	50.72 ^{ab}	10.83 ^a	8.75ª
	12	0.06a	0.14 ^{bc}	1.95 ^a	0.47 ^{ef}	0.37 ^{ef}	39.66ab	9.00a	6.98ª
	13	0.06a	0.14 ^{bc}	2.08ª	0.44 ^{ef}	0.34 ^f	49.37ab	9.91ª	7.52ª
	14	0.13 ^a	0.17 ^{abc}	1.94ª	0.47 ^{ef}	0.38 ^{def}	35.60 ^{ab}	8.14 ^a	6.31 ^a
	15	0.11 ^a	0.19 ^{ab}	1.55 ^a	0.57 ^{cdef}	0.44bcdef	21.10 ^{ab}	7.02ª	5.21 ^a
	16	0.17 ^a	0.11 ^{bc}	1.72ª	0.74abcde	0.57bcde	19.30 ^{ab}	7.37ª	5.43a
	17	0.03a	0.17 ^{abc}	1.91a	0.50 ^{ef}	0.32 ^f	81.62a	19.03a	12.56a
	18	0.11 ^a	0.29a	1.85 ^a	0.82abc	0.82ª	18.9ab	8.09a	8.12a
	19	0.11a	0.12bc	1.60a	0.96a	0.60bcd	16.30ab	10.50a	6.38a
	20	0.13 ^a	0.10 ^{bc}	1.6ª	0.79 ^{abcd}	0.54bcdef	12.75 ^b	6.26a	4.27a
Saline water	1	0.15 ^{ab}	0.13 ^{ab}	1.40bc	0.54ab	0.41 ab	11.67a	4.22ab	3.12ab
	2	0.06 ^b	0.14 ^{ab}	1.43 ^{abc}	0.46 ^{ab}	0.38b	26.30a	8.19 ^a	6.65 ^{ab}
	3	0.06 ^b	0.14 ^{ab}	1.39 ^{bc}	0.46 ^{ab}	0.45 ^{ab}	26.65ª	7.96 ^{ab}	7.99a
	4	0.29ab	0.16ab	1.29 ^c	0.61 ^{ab}	0.59ab	7.62ª	3.29ab	3.09ab
	5	0.20 ^{ab}	0.13 ^{ab}	1.38 ^{bc}	0.77ª	0.69ª	8.24a	4.39 ^{ab}	4.12 ^{ab}
	6	0.25 ^{ab}	0.05 ^b	1.95 ^{ab}	0.79 ^a	0.54 ^{ab}	11.85a	4.00 ^{ab}	2.71 ^{ab}
	7	0.20 ^{ab}	0.09b	1.64 ^{abc}	0.66 ^{ab}	0.66 ^{ab}	9.33a	3.74 ^{ab}	3.64 ^{ab}
	8	0.11 ^b	0.15 ^{ab}	1.49 ^{abc}	0.43 ^b	0.42 ^{ab}	14.04 ^a	3.97 ^{ab}	3.86 ^{ab}
	9	0.15 ^{ab}	0.12 ^{ab}	1.50 ^{abc}	0.66 ^{ab}	0.57 ^{ab}	11.11 ^a	5.05 ^{ab}	3.35 ^{ab}
10 11 12 13 14	10	0.12-	0.07-	1.46-	0.81-	0.78-	12.08-	6.70-	6.39-
	11	0.16 ^{ab}	0.10 ^b	1.87 ^{abc}	0.47 ^{ab}	0.41 ^{ab}	21.30a	4.34ab	2.14 ^{ab}
	12	0.47a	0.10 ^b	1.43 ^{abc}	0.59 ^{ab}	0.63ab	3.28ª	1.34 ^b	1.39 ^b
	13	0.20 ^{ab}	0.13 ^{ab}	1.63 ^{abc}	0.48 ^{ab}	0.46 ^{ab}	10.29 ^a	2.79 ^{ab}	2.61 ^{ab}
	14	0.24 ^{ab}	0.12 ^{ab}	1.65 ^{abc}	0.48 ^{ab}	0.45 ^{ab}	10.74 ^a	2.86 ^{ab}	2.54 ^{ab}
	15	0.14 ^{ab}	0.24a	1.55abc	0.50 ab	0.52ab	13.23a	4.20 ^{ab}	4.41 ^{ab}
	16	0.15 ^{ab}	0.10 ^b	1.76 ^{abc}	0.64 ab	0.57 ^{ab}	14.45 ^a	5.16 ^{ab}	4.55 ^{ab}
	17	0.11 ^b	0.15 ^{ab}	2.05 ^a	0.57 ^{ab}	0.54 ^{ab}	35.87ª	4.53 ^{ab}	7.19 ^{ab}
	18	0.37-	0.32-	1.56-	1.23-	1.18-	4.26-	3.34-	3.22-
	19	0.21 ^{ab}	0.12 ^{ab}	1.63 ^{abc}	0.77ª	0.67 ^{ab}	8.45a	3.97 ^{ab}	3.39 ^{ab}
	20	0.26 ^{ab}	0.08 ^b	1.56 ^{abc}	0.67 ^{ab}	0.59 ^{ab}	6.19 ^a	2.63 ^{ab}	2.28 ^{ab}

Table 3 Average chlorophyll, anthocyanin and flavonoid contents (arbitrary units) and the nitrogen balance index (NBI), of fully expanded leaves of in quinoa accessions grown for eight weeks under different (fresh water and saline) irrigation treatments. Means exhibiting different letters are significantly different (P < 0.05) by the post-hoc Tukey-b test on independent samples within each treatment (fresh and saline water). Values for accession 18 under saline irrigation conditions are not included in the median separation because of the lack of replications. Genotype numbers as detailed in Table 1.

Γreatment	Genotype	Pigments Chlorophyll	Anthocyanins	Flavonoids	NBI
Fresh water	1	29.34 ^{ab}	0.13 ^{ab}	1.44 ^{bcd}	20.99 ^{ab}
	2	30.84 ^{ab}	0.12 ^{ab}	1.54 ^{abc}	20.19 ^{ab}
	3	28.15 ^{ab}	0.12 ^{ab}	1.48 ^{abcd}	19.83ab
	4	31.75a	0.11 ^b	1.61 ^{ab}	19.91 ^{ab}
	5	28.69ab	0.12 ^{ab}	1.59 ^{ab}	18.29 ^{ab}
	6	27.67 ^{ab}	0.12 ^{ab}	1.26 ^d	22,36ª
	7	28.40 ^{ab}	0.12 ^{ab}	1.32 ^{cd}	24.84 ^a
	8	29.56ab	0.13 ^{ab}	1.57 ^{abc}	19.16 ^{ab}
	9	26.71 ^{ab}	0.14 ^{ab}	1.70 ^{ab}	16.11 ^{ab}
	10	29.89ab	0.12 ^{ab}	1.57 ^{abc}	19.22 ^{ab}
	11	25.27ab	0.15 ^a	1.74 ^a	14.70 ^{ab}
	12	28.96ab	0.13 ^{ab}	1.61 ^{ab}	18.13 ^{ab}
	13	28.89 ^{ab}	0.13 ^{ab}	1.64 ^{ab}	18.00 ^{ab}
	14	23.66 ^b	0.15 ^a	1.66 ^{ab}	14.26 ^b
	15	30.55 ^{ab}	0.13 0.13 ^{ab}	1.69 ^{ab}	18.11 ^{ab}
	16	26.48 ^{ab}	0.13 ^{ab}	0.49 ^{abcd}	18.24 ^{ab}
	17	29.27 ^{ab}	0.13 ^{ab}	1.66 ^{ab}	18.00 ^{ab}
	18	29.04 ^{ab}	0.12 ^{ab}	1.64 ^{ab}	18.01 ^{ab}
	19	32.63ª	0.12 0.13 ^{ab}	1.61 ^{ab}	20.35 ^{ab}
	20	29.70 ^{ab}	0.12 ^{ab}	1.59 ^{ab}	18.83 ^{ab}
Saline water	1	33.85 ^{ab}	0.12 b	1.55 ^{ab}	21.98 ^{abc}
Junie Water	2	35.46 ^{ab}	0.12 0.10 ^b	1.57 ^{ab}	22.76 ^{ab}
	3	35.33ab	0.11 ^b	1.70 ^{ab}	21.15 ^{abc}
	4	34.28 ^{ab}	0.11 ^b	1.61 ^{ab}	21.66 ^{abc}
	5	30.72 ^{ab}	0.13 ^{ab}	1.84 ^a	16.93bc
	6	34.44 ^{ab}	0.13 ^b	1.41 ^b	24.82a
	7	34.29 ^{ab}	0.11 ^b	1.40 ^b	25.08a
	8	31.25 ^{ab}	0.11 0.13 ^{ab}	1.80 ^a	17.64 ^{bc}
	9	35.43ab	0.13 ^b	1.81ª	19.78 ^{abc}
	10	35.68ab	0.11 ^b	1.59 ^{ab}	22.42 ^{abc}
	11	29.31 ^{ab}	0.13 ^{ab}	1.78a	16.63 ^{bc}
	12	27.40b	0.15 ^a	1.76 ^a	15.92°
	13	29.88ab	0.14 ^{ab}	1.76 ^a	17.24 ^{bc}
	14	27.90 ^b	0.14 ^{ab}	1.76 ^a	16.04 ^{bc}
	15	32.50 ^{ab}	0.14 ^{ab}	1.76 ^a	18.55 ^{abc}
	16	32.50 31.83 ^{ab}	0.12 ^b	1.62 ^{ab}	19.83 ^{abc}
	17	34.94 ^{ab}	0.12 ^b	1.62 ^{ab}	22.29 ^{abc}
	17	34.94 ⁻⁵ 19.04 ⁻	0.115	1.54-	12.90 ⁻
	18 19	19.04 36.95a	0.27 0.13 ^{ab}	1.54 1.78a	12.90 21.10 ^{abc}
	19 20	35.38 ^{ab}	0.13 ^{ab}	1.78 ^a 1.64 ^{ab}	21.10 ^{abc} 22.33 ^{abc}

Average, minimum and maximum temperature and precipitation data were obtained from the meteorological station of the International Center for Biosaline Agriculture (ICBA)

Raw data were analyzed using the statistical package SPSS (SPSS Inc.), using a multivariate analysis coupled with the post hoc test (Tukey-b) to assist differences between genotypes within each treatment.

Graphs were created using the SigmaPlot program 10.0 (SPSS Inc.).

Table 4Average carbon and nitrogen concentrations on a dry matter basis, and carbon (δ^{13} C) and nitrogen (δ^{15} N) isotope composition in the dry matter and soluble fraction of fully expanded leaves of quinoa accessions grown for eight weeks under different (fresh water and saline) irrigation treatments. Means exhibiting different letters are significantly different (P < 0.05) by the post-hoc Tukey-b test on independent samples within each treatment (control and salinity). Values for accession 18 under saline irrigation conditions are not included in the median separation because of the lack of replications. Genotype numbers as detailed in Table 1.

	El	emental analysis and stable isotopes (dry matter)				Stable isotopes (soluble	
Treatment	Genotype	C (%)	N (%)	$\delta^{13} {\sf C} \ (\%)$	δ^{15} N (‰)	fraction) δ^{13} C (‰)	δ^{15} N (‰)
Fresh water	1	37.01 ^{ab}	3.57 ^{ab}	-29.27ª	14.20 ^a	-30.74ª	10.06a
	2	38.36a	3.68 ^{ab}	-29.39a	13.04 ^a	-30.80a	8.75a
	3	38.00a	3.41 ^{ab}	-29.99a	13.59 ^a	-31.20a	10.39a
	4	38.01 ^a	3.86 ^{ab}	-29.61a	11.99 ^a	-30.66 ^a	10.86 ^a
	5	35.91 ^{ab}	3.13 ^{ab}	-29.52 ^a	13.37 ^a	-32.15 ^a	8.72 ^a
	6	35.61 ^{ab}	3.10 ^{ab}	-29.67a	11.52a	-30.67a	7.35ª
	7	35.74 ^{ab}	3.70 ^{ab}	-29.12a	13.61 ^a	-30.89a	10.52a
	8	37.49a	3.23 ^{ab}	-30.04a	13.58 ^a	-30.90a	11.54 ^a
	9	36.39ab	3.27 ^{ab}	-29.48a	13.25 ^a	-32.12a	10.36a
	10	35.76 ^{ab}	4.19 ^a	-28.70a	14.42a	-30.08a	12.67ª
	11	37.09 ^{ab}	3.14 ^{ab}	-29.01ª	15.02ª	-31.01ª	9.18ª
	12	37.76 ^a	3.81 ^{ab}	-29.25 ^a	15.08 ^a	-30.37 ^a	8.10 ^a
	13	37.58ª	3.77 ^{ab}	-28.99a	14.74 ^a	-30.62a	13.36 ^a
	14	35.55 ^{ab}	2.79 ^b	-28.61ª	12.52a	-30.92a	10.30a
	15	37.06 ^{ab}	3.68 ^{ab}	-29.50a	15.98a	-31.19 ^a	11.87ª
	16	35.61ab	3.44 ^{ab}	-29.44a	13.67a	-30.78a	9.88a
	17	37.50a	3.45 ^{ab}	-28.64a	15.51a	-30.83a	9.49a
	18	32.76 ^b	2.89b	-29.11a	11.46a	-30.74a	11.30a
	19	35.65 ^{ab}	3.56 ^{ab}	-28.70 ^a	15.04 ^a	-30.89 ^a	11.92ª
	20	36.04ab	3.37 ^{ab}	-28.77a	13.56a	-31.33a	7.50a
Saline water	1	35.64a	3.20a	-28.98a	11.00 ^a	-31.10a	7.64a
	2	37.14 ^a	3.48a	-29.18a	11.58a	-30.49a	10.07a
	3	37.20 ^a	3.24 ^a	-28.93a	11.89 ^a	-30.81a	8.20a
	4	35.38a	3.49a	-29.01a	9.43a	-30.88a	5.86a
	5	33.51a	2.56a	-29.37a	8.67a	-31.28a	3.69a
	6	33.48 ^a	3.13 ^a	-29.25a	7.50 ^a	-30.66a	7.09 ^a
	7	34.33a	3.56a	-28.68a	11.37a	-30.65a	8.85a
	8	36.42a	2.81a	-29.72a	11.25 ^a	-31.49a	8.27a
	9	34.27a	2.97a	-28.68a	8.46a	-31.38a	6.40a
	10	34.66-	3.98-	-28.54-	14.9-	-31.75-	13.78-
	11	32.64a	2.93a	-28.58a	11.92a	-30.18a	8.62a
	12	34.53a	3.43a	-28.71a	11.09 ^a	-30.84a	8.81a
	13	36.37a	3.55a	-28.57a	12.71 ^a	-30.40a	9.49a
	14	34.68a	2.96ª	-28.74a	11.34 ^a	-30.63a	6.73ª
	15	34.84ª	3.52a	-29.26a	14.61 ^a	-30.86a	11.19 ^a
	16	34.61 ^a	3.70 ^a	-28.95ª	15.09 ^a	-31.33a	10.23ª
	17	35.66a	3.69a	-28.57a	12.68ª	-30.08a	9.94ª
	18	28.23-	2.19-	-26.66	9.19-	-29.58-	7.53-
	19	33.58ª	3.19 ^a	-28.23ª	10.42a	-30.57ª	8.86ª
	20	34.53 ^a	3.31 ^a	-28.63 ^a	10.76 ^a	-31.15 ^a	7.82ª

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that have, or could be perceived to have, influenced the work reported in this article.

Acknowledgments

The participation of Jose L. Araus in this work was supported by a mobility fellowship from the "Salvador de Madariaga" program, "Ministerio de Educación, Cultura y Deporte", and the ICREA Academia, Generalitat de Catalunya, Spain. We also acknowledge the logistic support provided by ICBA to Maria D. Serret and Jose L. Araus.

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.dib.2020.105758.

References

- [1] F.Z. Rezzouk, M.A. Shahid, I.A. Elouafi, B. Zhou, J.L. Araus, M.D. Serret, Agronomic performance of irrigated quinoa in desert areas: comparing different approaches for early assessment of salinity stress, Agric. Water Manage (2020).
- [2] Z.G. Cerovic, G. Masdoumier, N. Ben Ghozlen, G. Latouche, A new optical leaf-clip meter for simultaneous non-destructive assessment of leaf chlorophyll and epidermal flavonoids, Physiol. Plant. 146 (2012) 251–260, doi:10.1111/j.1399-3054.2012.01639.x.
- [3] M.I. Hussain, A.J. Al-Dakheel, M.J. Reigosa, Genotypic differences in agro-physiological, biochemical and isotopic responses to salinity stress in quinoa (*Chenopodium quinoa* Willd.) plants: prospects for salinity tolerance and yield stability, Plant Physiol. Biochem 129 (2018) 411–420, doi:10.1016/j.plaphy.2018.06.023.