



# **Syndromic Inherited Retinal Diseases: Genetic, Clinical and Diagnostic Aspects**

## Yasmin Tatour and Tamar Ben-Yosef \*

Ruth & Bruce Rappaport Faculty of Medicine, Technion-Israel Institute of Technology, Haifa 31096, Israel; yasmin.t.90@gmail.com

\* Correspondence: benyosef@technion.ac.il; Tel.: +972-4-829-5228

Received: 9 September 2020; Accepted: 1 October 2020; Published: 2 October 2020



Abstract: Inherited retinal diseases (IRDs), which are among the most common genetic diseases in humans, define a clinically and genetically heterogeneous group of disorders. Over 80 forms of syndromic IRDs have been described. Approximately 200 genes are associated with these syndromes. The majority of syndromic IRDs are recessively inherited and rare. Many, although not all, syndromic IRDs can be classified into one of two major disease groups: inborn errors of metabolism and ciliopathies. Besides the retina, the systems and organs most commonly involved in syndromic IRDs are the central nervous system, ophthalmic extra-retinal tissues, ear, skeleton, kidney and the cardiovascular system. Due to the high degree of phenotypic variability and phenotypic overlap found in syndromic IRDs, correct diagnosis based on phenotypic features alone may be challenging and sometimes misleading. Therefore, genetic testing has become the benchmark for the diagnosis and management of patients with these conditions, as it complements the clinical findings and facilitates an accurate clinical diagnosis and treatment.

Keywords: retina; inherited retinal diseases; syndrome

## 1. Introduction

The retina is a multi-layered sensory tissue that lines the back of the eye. Its main function is the transduction of light energy into an electrical potential change, via a process known as phototransduction. The light-sensitive elements of the retina are the photoreceptor cells. The retina contains two types of photoreceptors, rods and cones. Rods (approximately 120 million in the human eye) are in charge of night vision, while cones (6 to 7 million in the human eye) are in charge of visual acuity and color vision. The highest cone concentration is found in the central region of the retina, known as the macula. Photoreceptors are highly compartmentalized cells, with the nucleus and other cellular organs located in the inner segment (IS), while the entire phototransduction machinery is included in the outer segment (OS).

Inherited retinal diseases (IRDs), which are among the most common genetic diseases in humans, define a clinically heterogeneous group of disorders, which cause visual loss due to improper development, dysfunction or premature death of the retinal photoreceptors [1]. IRDs are distinguished by several factors, including the type and location of affected cells and the timing of disease onset. The most common form of IRD is retinitis pigmentosa (RP) (also known as rod–cone dystrophy) [2]. Other IRD forms include cone/cone–rod dystrophy (CD/CRD) [3]; Leber congenital amaurosis (LCA) [4]; macular dystrophy (MD); and achromatopsia (rod monochromatism) [5], among others.

IRD is also one of the most genetically heterogeneous groups of disorders in humans, with over 260 genes identified to date (RetNet at https://sph.uth.edu/retnet/). It can be inherited as autosomal recessive (AR), autosomal dominant (AD) or X-linked (XL). Mitochondrial and digenic modes of inheritance have also been described. While in most cases of IRD the disease is limited to the eye

(non-syndromic), over 80 forms of syndromic IRD have been described. Approximately 200 genes are associated with these syndromes (Table 1). In some cases of syndromic IRD, the retinal disease may be the presenting symptom and other systemic findings evolve during childhood, puberty or later on in life. In other cases, the first identifiable symptom of the syndrome is non-ocular and the retinal phenotype is revealed only later in life.

The topic of systemic diseases associated with IRDs has been reviewed before, including the description of some of these syndromes [6]. In the current review, we provide a comprehensive summary of the vast majority of syndromic IRD forms reported to date, for which the underlying gene/s have been identified (as listed in OMIM-Online Mendelian Inheritance in Man, https://www.ncbi. nlm.nih.gov/omim, and reported in the literature). We discuss different aspects, including the marked genetic heterogeneity of some of these syndromes, phenotypic overlap and diagnostic approaches.

Syndrome (MIM/Reference)	Gene	Inheritance *	Main Ocular Phenotypes <sup>#</sup>	Main Extra-Ocular Phenotypes <sup>¶</sup>
Abetalipoproteinemia; ABL (#200100)	МТТР	AR	RP	Fat malabsorption, neurodegeneration, acanthocytosis
Aicardi Syndrome; AIC (#304050)	Xp22 abnormalities	XLD	Chorioretinopathy, OA, microphthalmia, optic nerve coloboma, cataract	Callosal agenesis, PGR, microcephaly, ID, skeletal anomalies, neoplasia
Alagille Syndrome 1; ALGS1 (#118450)	JAG1	AD	Iris stromal hypoplasia, posterior embryotoxon, microcornea, anomalous optic disc, peripapillary retinal depigmentation, chorioretinopathy	Liver disease, skeletal and renal involvement, characteristic facial features, ID, FTT
Alport Syndrome 1; ATS1 (#3010150)	COL4A5	XLD	Fleck retinopathy, cataract, myopia, corneal abnormalities	HL, renal disease
Alstrom Syndrome; ALMS (#203800)	ALMS1	AR	CRD, MD, cataract	DD, SS, obesity, HL, cardiac, skeletal, hepatic, renal and endocrine involvement
Alpha-Methylacyl-CoA Racemase Deficiency; AMACRD (#614307)	AMACR	AR	RP	Neurodegeneration
Autoimmune Polyendocrine Syndrome, Type I, with or without Reversible Metaphyseal Dysplasia; APS1 (#240300)	AIRE	AD, AR	RP, keratopathy, keratoconjunctivitis	Multiple autoantibodies, anemia, hepatic, gastrointestinal, dental, skin, hair and endocrine involvement, hypogonadism
Bardet–Biedl Syndrome; BBS (#209900, #615981, #600151, #615982, #615983, #605231, #615984, #615985, #615986, #615987, #615988, #615989, #615990, #615991, 615992, #615993, #615994, #615995, #615996, #617119, #617406) [7]	BBS1, BBS2, ARL6, BBS4, BBS5, MKKS, BBS7, TTC8, PTHB1, BBS10, TRIM32, BBS12, MKS1, CEP290, WDPCP, SDCCAG8, LZTFL1, BBIP1, IFT27, IFT74, C80RF37, CEP164	AR	RP, strabismus, cataract	ID, SS, obesity, hypogonadism, renal disease, polydactyly
Cerebellar Atrophy with Pigmentary Retinopathy [8]	MSTO1	AR	RD	Cerebellar atrophy, ID, PGR
Congenital Disorder of Glycosylation; CDG (#212065, #617082, #613861, #608799, #300896)	PMM2, NUS1, DHDDS, DPM1, SLC35A2	AR	RP	FTT, microcephaly, ID, neurodegeneration, cardiac, hepatic, gastrointestinal, renal and hematological involvement

Table 1. Syndromic inherited retinal diseases (IRDs).

Syndrome (MIM/Reference)	Gene	Inheritance *	Main Ocular Phenotypes <sup>#</sup>	Main Extra-Ocular Phenotypes <sup>¶</sup>
Congenital Disorder of Glycosylation with Defective Fucosylation 2; CDGF2 (#618324)	FCSK	AR	MD, OA, strabismus	FTT, ID, hypotonia, neurodegeneration, gastrointestinal anomalie
Cranioectodermal Dysplasia 4; CED4 (#614378)	WDR19	AR	RP	Skeletal anomalies, SS, respiratory, hepatic and renal involvement
Ceroid Lipofuscinosis, Neuronal; CLN (#256730, #204500, #204200, #256731, #601780, #610951, #600143, #610127, #614706)	PPT1, TPP1, CLN3, CLN5, CLN6, MFSD8, CLN8, CTSD, GRN	AR	RP, CRD, OA	Microcephaly, ID, neurodegeneration
Cohen Syndrome; COH1 (#216550)	VPS13B	AR	RD, OA, strabismus, high myopia	ID, DD, microcephaly, SS obesity, skeletal, cardiac, hematological and endocrine involvement
Coenzyme Q10 Deficiency, Primary, 1; COQ10D1 (#607426)	COQ2	AR	RP	ID, cerebellar atrophy, HI cardiac, hepatic, renal and muscular involvement
Combined Oxidative Phosphorylation Deficiency 29; COXPD29 (#616811)	TXN2	AR	RD, OA	Microcephaly, hypotonia DD, ID, neurodegeneration
Charcot–Marie–Tooth Disease, X-linked recessive, 5; CMTX5 (#311070)	PRPS1	XLR	RP, OA	Peripheral neuropathy, H
Cone–Rod Dystrophy and Hearing Loss 1; CRDHL1 (#617236)	CEP78	AR	CRD	HL
Cockayne Syndrome; CS (#216400, #133540)	ERCC8, ERCC6	AR	RD, OA, cataract, strabismus	IUGR, PGR, microcephal ID, neurodegeneration, HL, renal, skeletal and skin involvement
Cystinosis, Nephropathic; CTNS (#219800, #219900)	CTNS	AR	RD, corneal crystals	Renal disease, neurodegeneration, skeletal and endocrine anomalies
Danon Disease (#300257)	LAMP2	XLD	RD	Cardiac disease, myopath ID
Diabetes and Deafness, Maternally Inherited; MIDD (#520000)	MTTL1, MTTE, MTTK, mitochondrial DNA rearrangements	Mi	RD, MD, ophthalmoplegia	HL, cardiac and neurological anomalies, diabetes mellitus
Dyskeratosis Congenita, Autosomal Dominant 3; DKCA3 (#613990)	TINF2	AD	RD, blockage of lacrimal ducts	IUGR, SS, microcephaly, ID, HL, respiratory, skin skeletal and hematologica involvement, neoplasia
Hypobetalipoproteinemia, Familial, 1; FHBL1 (#615558)	АРОВ	AR	RP	Fat malabsorption, neurodegeneration, acanthocytosis
Hypobetalipoproteinemia, Acanthocytosis, Retinitis Pigmentosa and Pallidal Degeneration; HARP (#607236)	PANK2	AR	RP	Fat malabsorption, neurodegeneration, acanthocytosis
Hypotrichosis, Congenital, with Juvenile Macular Dystrophy; HJMD (#601553)	CDH3	AR	MD	Hypotrichosis
Hermansky–Pudlak Syndrome; HPS (#614072, #614073, #614077)	HPS3, HPS4, BLOC1S3	AR	Hypopigmentation of retina and choroid, foveal hypoplasia, nystagmus, iris transillumination	Skin and hair hypopigmentation, bleeding diathesis

Syndrome	Gene	Inheritance *	Main Ocular	Main Extra-Ocular	
(MIM/Reference)	Gene	Inneritance	Phenotypes #	Phenotypes <sup>¶</sup>	
Hyper-IgD Syndrome; HIDS (#260920)	MVK	AR	RP	Hematological anomalies, gastrointestinal and skeletal involvement, periodic fever	
Hyperoxaluria, Primary, Type I; HP1 (#259900)	AGXT	AR	RD, OA	Renal disease, dental, cardiovascular and skin involvement, peripheral neuropathy	
Intellectual Developmental Disorder and Retinitis Pigmentosa; IDDRP (#618195)	SCAPER	AR	RP, MD, cataract	ID, skeletal abnormalities, male sterility	
Jalili Syndrome (#217080)	CNNM4	AR	CRD	Amelogenesis imperfecta	
Joubert Syndrome; JBTS (#213300, #608091, #608629, #610188, #610688, #611560, #612291, #612285, #614464, #614465, #614844, #614970, #615636, #615665, #616781, #617121, #617562, #617622, #618161, #300804)	INPP5E, TMEM216, AHI1, CEP290, TMEM67, RPGRIP1L, ARL13B, CC2D2A, CEP41, TMEM138, ZNF423, TMEM231, CSPP1, PDE6D, CEP104, MKS1, TMEM107, ARMC9, ARL3	AR	RD, chorioretinal coloboma, optic nerve coloboma, microphthalmia, oculomotor apraxia, esotropia, ptosis	Brain structural anomalies, FTT, macrocephaly, ID, neurodegeneration, genitourinary, hepatic, respiratory and skeletal involvement	
	OFD1	XLR	_		
Kearns–Sayre Syndrome; KSS (#530000)	Mitochondrial DNA deletions	Mi	RD, ophthalmoplegia	SS, microcephaly, neurodegeneration, cardiac, renal and endocrine involvement	
Laurence–Moon Syndrome; LNMS (#245800)	PNPLA6	AR	Chorioretinal degeneration	ID, neurodegeneration, genitourinary abnormalities	
Leber Congenital Amaurosis with Early-Onset Deafness; LCAEOD (#617879)	TUBB4B	AD	LCA	HL	
Lipodystrophy, familial partial, type7; FPLD7 (#606721)	CAV1	AD	RD, cataract	Lack of facial fat, orthostatic hypotension, neurological and skin involvement	
Methylmalonic Aciduria and Homocystinuria, cblC type; MAHCC (#277400)	ММАСНС	AR	RP, CRD	FTT, microcephaly, ID, neurodegeneration, renal and hematological involvement	
Mevalonic Aciduria; MEVA (#610377)	MVK	AR	RP, OA, cataract	FTT, DD, neurodegeneration, spleen, hepatic, skeletal, skin and hematological involvement	
Microcephaly and Chorioretinopathy, autosomal recessive; MCCRP (#251270, #616171, #616335)	TUBGCP6, PLK4, TUBGCP4	AR	Chorioretinopathy, OA, microphthalmia, microcornea, cataract	IUGR, microcephaly, brain structural anomalies, DD, ID, neurodegeneration, SS	
Microcephaly with or without Chorioretinopathy, Lymphedema or Mental Retardation; MCLMR (#152950)	KIF11	AD	Chorioretinopathy, myopia, hypermetropia, corneal opacity, microcornea, microphthalmia, cataract	Microcephaly, ID, neurodegeneration, lymphedema	
Microphthalmia, Syndromic 5; MCOPS5 (#610125)	OTX2	AD	RD, microphthalmia, anophthalmia, optic nerve hypoplasia or agenesis, microcornea, cataract	Brain structural anomalies, hypotonia, pituitary dysfunction, DD, SS, cleft palate, abnormal genitalia, joint laxity	

		lable 1. Cont.			
Syndrome (MIM/Reference)	Gene	Inheritance *	Main Ocular Phenotypes <sup>#</sup>	Main Extra-Ocular Phenotypes <sup>¶</sup>	
Mitochondrial Complex II Deficiency (#252011)	SDHA, SDHD, SDHAF1	AR	RD, OA, ptosis, ophthalmoplegia	SS, cardiac, skeletal, muscular and neurological involvement	
Mitochondrial Complex IV Deficiency (#220110)	APOPT1, COA3, COX6A2, COX6B1, COX8A, COX10, COX14, COX20, PET100, TACO1	AR	RD, OA, ptosis	FTT, brain structural anomalies, ID, HL, cardiac, respiratory, hepatic, renal and muscular involvement	
Mucolipidosis III alpha/beta; MLIII A/B (#252600)	GNPTAB	AR	RD, corneal clouding	Neurodegeneration, ID, SS, coarse facies, skeletal, cardiac and skin involvement	
Mucolipidosis IV; ML4 (#252650)	MCOLN1	AR	RD, OA, corneal disease, strabismus	Microcephaly, ID, neurodegeneration	
Mucopolysaccharidosis; MPS (#309900, #252930, #607014,	IDS	XLR	RP, ptosis, corneal – clouding	Neurodegeneration, ID, SS, coarse facies, HL, skeletal, cardiac, respiratory,	
#253000, #253010)	HGSNAT, IDUA, GALN5, GLB1	AR		hepatic, gastrointestinal and skin involvement	
Nephronophthisis 15; NPHP15 (#614845)	CEP164	AR	LCA	Renal disease	
Neurodegeneration with Brain Iron Accumulation 1; NBIA1 (#234200)	PANK2	AR	RD, OA, eyelid apraxia	Neurodegeneration, gastrointestinal, skeletal, skin and muscular involvement	
Neuropathy, Ataxia and Retinitis Pigmentosa; NARP (#551500)	MTATP6	Mi	RP	Neurodegeneration, ataxia	
Norrie Disease; ND (#310600)	NDP	XLR	Retinal dysgenesis, retinal dysplasia, OA, microphthalmia, vitreous atrophy, corneal opacities, iris atrophy, cataract	HL, ID, neurodegeneration	
Oculoauricular Syndrome; OCACS (#612109)	HMX1	AR	RP, microphthalmia, microcornea, cataract, microphakia, sclerocornea, increased intraocular pressure	External ear abnormalities	
Orofaciodigital Syndrome XVI; OFD16 (#617563)	TMEM107	AR	RD, oculomotor apraxia, ptosis	Facial anomalies, breathing abnormalities, polydactyly, hypotonia, ID, neurological anomalies	
Oliver–McFarlane Syndrome; OMCS (#275400)	PNPLA6	AR	Chorioretinopathy, OA	SS, ID, neurodegeneration, obesity, male external genitalia abnormalities, endocrine anomalies	
Peroxisomal Acyl-CoA Oxidase Deficiency (#264470)	ACOX1	AR	RD, OA, strabismus	Neurodegeneration, ID, HL, liver disease	
Peroxisome Biogenesis Disorder; PBD (#214100, #614866, #601539, #234580, #614879, #266510)	PEX1, PEX2, PEX5, PEX6, PEX7, PEX12	AR	RD, OA, corneal clouding, cataract	FTT, neurodegeneration, ID, HL, dental, cardiac, hepatic, genitourinary and skeletal involvement	
Posterior Column Ataxia with Retinitis Pigmentosa; AXPC1 (#609033)	FLVCR1	AR	RP, OA	Posterior column ataxia, neurodegeneration, gastrointestinal and skeletal involvement	
Polyneuropathy, Hearing Loss, Ataxia, Retinitis Pigmentosa and Cataract; PHARC (#612674)	ABHD12	AR	RP, OA, cataract	Ataxia, neurodegeneration, HL	

Syndrome (MIM/Reference)	Gene	Inheritance *	Main Ocular Phenotypes <sup>#</sup>	Main Extra-Ocular Phenotypes <sup>¶</sup>
Pseudoxanthoma Elasticum; PXE (#264800)	ABCC6	AR	RD, MD, choroidal neovascularization	Skin lesions, cardiovascular disease, gastrointestinal and genitourinary involvement
Refsum Disease, classic (#266500)	РНҮН	AR	RP	Neurodegeneration, ataxia, HL, anosmia, cardiac, skeletal and skin involvement
Retinal Dystrophy, Iris Coloboma and Comedogenic Acne Syndrome; RDCCAS (#615147)	RPB4	AR	RD, coloboma of the iris, displacement of the pupil, microcornea, cataract	Comedogenic acne
Retinal Dystrophy and Iris Coloboma with or without Cataract; RDICC (#616722)	MIR204	AD	RD, coloboma of the iris, congenital cataract	
Retinal Dystrophy, Juvenile Cataracts and Short Stature Syndrome; RDJCSS (#616108)	RDH11	AR	RD, juvenile cataracts	SS, DD, ID, dental anomalies
Retinal Dystrophy and Obesity; RDOB (#616188)	TUB	AR	RD	Obesity
Revesz Syndrome (#268130)	TINF2	AD	RD	IUGR, brain structural anomalies, neurodegeneration, ID, aplastic anemia, skin, hair and nail abnormalities
Retinitis Pigmentosa–Deafness Syndrome (#500004)	MTTS2	Mi	RP	HL
Retinitis Pigmentosa and Erythrocytic Microcytosis; RPEM (#616959)	TRNT1	AR	RP	Erythrocytic microcytosis and additional hematologic abnormalities
Retinitis Pigmentosa, Hypopituitarism, Nephronophtisis and mild Skeletal Dysplasia; RHYNS (#602152)	TMEM67	AR	RP	Hypopituitarism, renal disease, skeletal anomalies HL
Retinitis Pigmentosa 82 with or without Situs Inversus; RP82 (#615434)	ARL2BP	AR	RP	Situs inversus, male infertility
Retinitis Pigmentosa with or without Skeletal Anomalies; RPSKA (#250410)	CWC27	AR	RP	SS, skeletal anomalies, ID
Retinitis Pigmentosa, X-linked and Sinorespiratory Infections, with or without Deafness (#300455)	RPGR	XL	RP	Recurrent respiratory infections, HL
Senior–Løken Syndrome; SLSN (#266900, #606996, #609254, #610189, #613615, #616307, #616629)	NPHP1, NPHP4, IQCB1, CEP290, SDCCAG8, WDR19, TRAF3IP1	AR	RP, LCA	Renal disease
Short Stature, Hearing Loss, Retinitis Pigmentosa and Distinctive Facies; SHRF (#617763)	EXOSC2	AR	RP, corneal dystrophy, glaucoma, strabismus	SS, facial anomalies, HL, neurodegeneration, DD, ID
Sideroblastic Anemia with B-cell Immunodeficiency, Periodic Fevers and Developmental Delay; SIFD (#616084)	TRNT1	AR	RP	Sideroblastic anemia, immunodeficiency, growth retardation, DD, periodic fever, HL, neurological, cardiac and renal involvement

Table 1. Cont.

Syndrome (MIM/Reference)	Gene	Inheritance *	Main Ocular Phenotypes <sup>#</sup>	Main Extra-Ocular Phenotypes <sup>¶</sup>
Spondylometaphyseal Dysplasia with Cone–Rod Dystrophy; SMDCRD (#608940)	PCYT1A	AR	CRD	Skeletal anomalies, PGR
Spondylometaphyseal Dysplasia, Axial; SMDAX (#602271)	CFAP410	AR	RP, CRD, OA	Skeletal anomalies, respiratory disease, reduced sperm motility
Short-Rib Thoracic Dysplasia 9 with or without Polydactyly; SRTD9 (#266920)	IFT140	AR	RP	Skeletal anomalies, renal disease, ID
Thiamine-Responsive Megaloblastic Anemia Syndrome; TRMA (#249270)	SLC19A2	AR	OA, RD	Megaloblastic anemia, diabetes mellitus, HL
Usher Syndrome; USH (#276900, #276904, #601067, #602083, #606943, #614869, #276901, #605472, #611383, #276902, #614504)	MYO7A, USH1C, CDH23, PCDH15, USH1G, CIB2, USH2A, ADGRV1, WHRN, CLRN1, HARS1	AR	RP	HL, vestibular dysfunction
Wolfram Syndrome 1, WFS1 (#222300)	WFS1	AR	OA, RD	Diabetes mellitus, diabetes insipidus, HL, neurodegeneration, genitourinary and neurologic involvement
White–Sutton Syndrome, WHSUS (#616364)	POGZ	AD	RP, OA, cortical blindness	DD, characteristic facial features, hypotonia, HL, joint laxity, gastrointestinal anomalies
Xeroderma Pigmentosum, group B; XPB (#610651)	ERCC3	AR	RD, OA, micropathalmia	Neoplasia, skin anomalies, SS, microcephaly, HL, ID, brain structural anomalies, neurodegeneration

\* AD, autosomal dominant; AR, autosomal recessive; Mi, mitochondrial; XL, X-linked; XLD, X-linked dominant; XLR, X-linked recessive. # CRD, cone–rod dystrophy; LCA, Leber congenital amaurosis; MD, macular dystrophy; OA, optic atrophy; RD, retinal dystrophy; RP, retinitis pigmentosa. ¶ DD, developmental delay; FTT, failure to thrive; HL, hearing loss; ID, intellectual disability; IUGR, intrauterine growth restriction; PGR, postnatal growth retardation; SS, short stature.

#### 2. Syndromic IRD Types

The majority of syndromic IRDs are recessively inherited and rare. Many, although not all, syndromic IRDs can be classified into one of two major disease groups: inborn errors of metabolism (IEM) and ciliopathies.

IEMs are genetic disorders leading to failure of carbohydrate metabolism, protein metabolism, fatty acid oxidation or glycogen storage. Many IEMs present with neurologic symptoms [9]. The retina develops from an embryonic forebrain pouch and is considered an extension of the brain. Therefore, neurodegeneration resulting from IEMs often involves retinal degeneration (RD) as well. Major forms of syndromic IRD that belong to the IEM group include congenital disorders of glycosylation (CDG) [10], neuronal ceroid lipofuscinoses (CLNs) [11], mucopolysaccharidoses (MPSs) [12], peroxisomal diseases [13] and more (Table 1).

Ciliopathies are a group of genetic diseases caused by mutations in genes associated with the structure and function of primary cilia. Primary cilia function as signaling hubs that sense environmental cues and are pivotal for organ development and function, and for tissue homeostasis. By their nature, cilia defects are usually pleiotropic, affecting more than one system [14]. Photoreceptor OSs are highly modified primary sensory cilia. The proximal end of the OS is linked to the cell body (i.e., the IS) via a connecting cilium which is structurally homologous to the transition zone of primary cilia [15]. Consequently, retinal pathogenesis is a common finding in ciliopathies. Other organs which are commonly affected in ciliopathies are the central nervous system (CNS), kidney, liver, skeleton and inner ear. Major forms of syndromic IRD that belong to the ciliopathy group include Bardet–Biedl

Syndrome (BBS) [16], Joubert Syndrome (JBTS) [17], Usher Syndrome (USH) [18], Senior–Løken Syndrome (SLN) [19] and Alstrom Syndrome (ALMS) [20] (Table 1).

#### 3. Genetic Heterogeneity in Syndromic IRDs

Over 80 forms of syndromic IRD have been described (Table 1). Most of these syndromes are caused by a single gene. However, 14 of 81 (17%) of the syndromes listed in Table 1 are genetically heterogeneous, and some of them are associated with multiple causative genes. The most genetically heterogeneous forms of syndromic IRD are three recessively inherited ciliopathies: BBS, JBTS and USH. The protein products of the genes associated with each one of these ciliopathies tend to form multi-protein complexes in the retina and in additional tissues, thus explaining the similar phenotypes caused by mutations in each of these genes.

BBS (prevalence of about 1/125,000) is characterized by a combination of RP, postaxial polydactyly (and other skeletal abnormalities), hypogonadism, renal disease, intellectual disability (ID) and truncal obesity [16]. Twenty-one causative genes have been reported to date (OMIM) (Table 1). Their protein products are involved in lipid homeostasis, intraflagellar transport, establishing planar cell polarity, regulation of intracellular trafficking and centrosomal functions. Eight of these genes encode for subunits of a protein complex, the BBSome, which is integral in ciliary as well as intracellular trafficking [21]. In the retina, the BBSome is required for photoreceptor OS formation and maintenance [22], as well as for retinal synaptogenesis [23].

JBTS (prevalence of 1/55,000–1/200,000) is characterized by a peculiar midbrain–hindbrain malformation, known as the molar tooth sign. The neurological presentation of JBTS includes hypotonia that evolves into ataxia, developmental delay, abnormal eye movements and neonatal breathing abnormalities. This picture is often associated with variable multiorgan involvement, mainly of the retina, kidney and liver [17]. RD has been reported in 38% of patients [24]. To date, 36 causative genes have been identified, all encoding for proteins expressed in the primary cilium or its apparatus (OMIM). Mutations in 20 of these genes (listed in Table 1) have been specifically associated with RD and additional ocular abnormalities (such as nystagmus and oculomotor apraxia). Ocular abnormalities have also been reported in patients with mutations in most other JBTS genes. However, since RD was not specifically reported in these patients, these genes are not listed in Table 1. Given the marked phenotypic heterogeneity found in JBTS patients, it is very likely that a retinal phenotype will be associated with these genes in the future, as additional patients are discovered.

USH (prevalence of 1–4/25,000) is characterized by the combination of RP and sensorineural hearing loss (HL). Based on the severity and progression of HL, age at onset of RP and the presence or absence of vestibular impairment, the majority of USH cases can be classified into one of three clinical subtypes (USH1-3). Eleven USH genes have been identified to date (OMIM) (Table 1). Their protein products are associated with a wide range of functions, including actin-binding molecular motors, cell adhesion, scaffolding and cellular trafficking. USH proteins form complexes and function cooperatively in neurosensory cells of both the retina and the inner ear (reviewed in [18,25]).

### 4. Phenotypic Overlap in Syndromic IRDs

When referring to syndromic IRD, phenotypic overlap is a common phenomenon, which can be divided into three groups, as detailed below:

#### 4.1. Phenotypic Overlap between Different IRD Syndromes

Many types of syndromic IRD have a multi-systemic nature. Certain organs are commonly involved in syndromic IRDs. Specifically, CNS involvement (usually manifested as ID) is found in 68% of IRD syndromes (Table 1 and Figure 1), and over 80 genes are associated with the combination of IRD and ID [26] (Table 1). In addition to ID, the most common findings in syndromic IRD are extra-retinal eye abnormalities and ear, skeletal, renal and cardiovascular involvement (Figure 1). Most of these syndromes are phenotypically heterogeneous, with many patients exhibiting only some

of the phenotypic features. These factors lead to a marked phenotypic overlap between different syndromes, and to a diagnostic challenge. For example, the combination of RD, ID, renal disease and skeletal abnormalities is found in numerous forms of syndromic IRD, including BBS, JBTS and ALMS, among others (Table 1). The combination of retinal abnormalities and HL as prominent symptoms is found in USH, as well as in CRD and HL 1 syndrome [27], Leber congenital amaurosis with early-onset deafness syndrome [28], Norrie disease [29], peroxisome biogenesis disorders, Refsum disease [30] and more (Table 1). These overlaps may often lead to diagnostic mistakes [27,31,32].

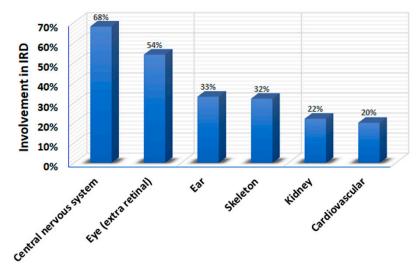


Figure 1. Systems and organs most commonly involved in syndromic IRDs.

#### 4.2. Syndromic Versus Non-Syndromic IRD Caused by the Same Genes

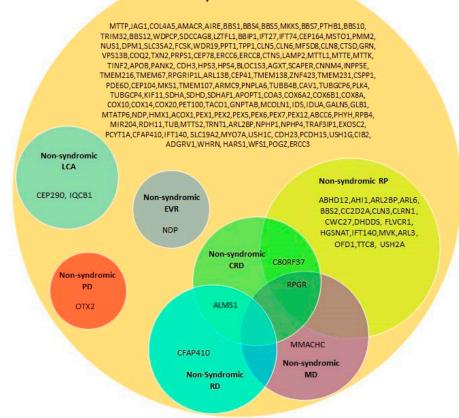
Twenty-eight of the genes listed in Table 1 can cause both syndromic and non-syndromic IRD (Table 2 and Figure 2). In general, milder hypomorphic mutations in these genes are associated with non-syndromic IRD, while null mutations lead to the involvement of additional tissues. In addition, the involvement of additional genetic and environmental factors in the determination of the final phenotypic outcome cannot be excluded. A prominent example is the *USH2A* gene. Mutations in this gene are the most common cause of USH, and specifically of USH2 (RP with congenital, mild to moderate sensorineural HL) [33]. Moreover, *USH2A* variants are also one of the commonest causes of AR non-syndromic RP worldwide [34–36]. It appears that the specific combination of *USH2A* variants determines whether one has USH2 or non-syndromic RP [37–39]. In addition, RD is more severe in patients with *USH2A*-related USH2 than in patients with *USH2A*-related non-syndromic RP. However, the reason is not completely understood [38].

Gene	Syndromic IRD	Non-Syndromic IRD (MIM)	Reference
ABHD12	PHARC	arRP	[40]
AHI1	JBTS3	arRP	[41]
ALMS1	ALMS	arCRD, arEORD	[42]
ARL2BP	RP with situs inversus	arRP (#615434)	[43,44]
ARL3	JBTS35	adRP (#618173)	[45]
ARL6	BBS3	arRP (#613575)	[46]
BBS2	BBS2	arRP (#616562)	[47]
C8ORF37	BBS21	arCRD, arRP (#614500)	[48]
CC2D2A	JBTS9, MKS6	arRP	[49]
CEP290	BBS14, JBTS5, MKS4, SLSN6	arLCA (#611755)	[50]
CFAP410	SMDAX	arRD with or without macular staphyloma (#617547)	[51,52]

Gene	Syndromic IRD	Non-Syndromic IRD (MIM)	Reference
CLN3	CLN3	arRP	[53]
CLRN1	USH3A	arRP (#614180)	[54]
CWC27	RPSKA	arRP (#250410)	[55]
DHDDS	CDG1BB	arRP (#613861)	[56,57]
FLVCR1	PCARP	arRP	[58]
HGSNAT	MPS3C	arRP (#616544)	[59]
IFT140	SRTD9 with/without polydactyly	arRP (#617781)	[60]
IQCB1	SLSN5	arLCA	[61]
MFSD8	CLN7	arMD (#616170), arRD	[62]
MMACHC	MAHCC	arMD	[63]
MVK	HIDS, MEVA	arRP	[64]
NDP	ND	XL EVR (#305390)	[65]
OFD1	JBTS10	XL RP (#300424)	[66]
OTX2	RD with pituitary dysfunction	adPD (#610125)	[67]
RPGR	RP, sinorespiratory infections and deafness	XL CRD (#304020), XL MD (#300834), XL RP (#300029)	[68]
TTC8	BBS8	arRP (#613464)	[69]
USH2A	USH2A	arRP (#613809)	[70]

ALMS: Alstrom syndrome; ar: autosomal recessive; ad: autosomal dominant; BBS: Bardet–Biedl syndrome; CDG: congenital disorder of glycosylation; CLN: ceroid lipofuscinosis neuronal; CRD: cone–rod dystrophy; EORD: early-onset retinal degeneration; EVR: exudative vitreoretinopathy; HIDS: hyper-IgD syndrome; JBTS: Joubert syndrome; LCA: Leber congenital amaurosis; MAHCC: methylmalonic aciduria and homocystinuria, cblC type; MD: macular dystrophy; MEVA: mevalonic aciduria; MKS: Meckel syndrome; MPS: mucopolysaccharidosis; ND: Norrie disease; PCARP: posterior column ataxia with retinitis pigmentosa; PD: pattern dystrophy; PHARC: polyneuropathy, hearing loss, ataxia, retinitis pigmentosa and cataract; RD: retinal dystrophy; RP: retinitis pigmentosa; RPSKA: retinitis pigmentosa with skeletal anomalies; SLSN: Senior–Løken syndrome; SMDAX: spondylometaphyseal dysplasia axial; SRTD: short-rib thoracic dysplasia; USH: Usher syndrome; XL: X-linked.

#### Syndromic IRD



**Figure 2.** A Venn diagram showing the involvement of syndromic IRD genes in non-syndromic IRD phenotypes. CRD: cone–rod dystrophy; EVR: exudative vitreoretinopathy; LCA: Leber congenital amaurosis; MD: macular dystrophy; PD: pattern dystrophy; RD: retinal dystrophy; RP: retinitis pigmentosa.

#### 4.3. Co-Existence of Non-Syndromic IRD and Additional Non-Ocular Diseases

IRD is one of the most genetically heterogeneous groups of disorders in humans, and most cases of IRD are non-syndromic. Non-syndromic IRD may coincide with other genetic (and non-genetic) rare conditions, leading to a clinical suspicion or diagnosis of a syndrome. For example, co-occurrence of non-syndromic RP and non-syndromic HL in a family may appear as USH [71].

#### 5. Diagnostic Challenges

Due to the high degree of phenotypic variability and phenotypic overlap found in syndromic IRD, as described above, correct diagnosis based on phenotypic features alone may be challenging and sometimes misleading. Therefore, genetic testing has become the benchmark for the diagnosis and management of patients with these conditions, as it complements the clinical findings and facilitates an accurate clinical diagnosis. Establishing a correct diagnosis is important for both the patients and their family members, for multiple reasons: it enables the understanding of the natural history course, and the prediction of disease prognosis; it aids in tailoring correct follow-up and treatment, including potential gene-targeted therapies [72]; it leads to a reduction in disease prevalence, by genetic screening and counseling in high-risk populations; it allows the patients to pursue prenatal counseling and reproductive planning; and it enables identification of novel disease genes and mechanisms.

The existence of common founder mutations in certain populations allows for quick and efficient mutation screening in affected individuals, based on the relevant phenotype and ethnic background. This is performed by PCR-based DNA amplification and Sanger sequencing, or by specifically designed assays. Some examples are common USH3A-, USH1F-, ML4- and BBS2-causative mutations found in the Ashkenazi Jewish population [73–76]; and USH3A- and MKS1-causative mutations found in the Finnish population [77,78]. Nevertheless, for most syndromic IRD patients worldwide, this strategy is not effective.

Currently, the most efficient approach for genetic diagnosis in monogenic diseases, including IRDs, is next-generation sequencing (NGS). NGS technologies facilitate the screening of the entire genome (whole genome sequencing, WGS); of all protein-coding regions (whole exome sequencing, WES); or of protein-coding regions of pre-determined panels of genes (targeted NGS, T-NGS) [79,80]. Since protein-coding regions comprise only 1–2% of the entire genome while harboring over 85% of variants causing Mendelian disorders, WES is still considered as the method of choice for genetic analysis, in both clinical and research settings. However, worldwide diagnostic yields of IRD patients by WES only range between 60% and 70% [36,81,82]. The missing mutations can be divided into four groups: (1) mutations located within exons, but missed due to technical issues, e.g., lack of coverage; (2) mutations located within covered exons, but missed due to limitations in data analysis and interpretation; (3) non-coding variants that may affect gene expression, mRNA stability, splicing and more; and (4) structural variants, such as large deletions, duplications and inversions, which are missed by WES. The latter two may be identified by WGS [34].

#### 6. Summary and Conclusions

Over 80 forms of syndromic IRDs have been described, and approximately 200 causative genes identified. Due to the high degree of phenotypic variability and phenotypic overlap found in syndromic IRD, correct diagnosis based on phenotypic features alone is insufficient, and genetic testing has become the benchmark for the diagnosis and management of patients with these conditions. For most patients, molecular diagnosis should be based on NGS technologies. Currently, WES is the most popular approach for genetic analysis in patients with monogenic diseases, including IRDs. However, the continuous progress in both technical and bioinformatic aspects, as well as the reduction of costs, is already leading to a shift towards WGS as the method of choice.

**Author Contributions:** Conceptualization, T.B.-Y.; methodology, T.B.-Y.; validation, Y.T. and T.B.-Y.; formal analysis, Y.T.; investigation, Y.T. and T.B.-Y.; resources, T.B.-Y.; data curation, Y.T. and T.B.-Y.; writing—original draft preparation, T.B.-Y.; writing—review and editing, T.B.-Y.; supervision, T.B.-Y.; funding acquisition, T.B.-Y. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Israel Science Foundation (grant number 525/19).

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

## References

- Duncan, J.L.; Pierce, E.A.; Laster, A.M.; Daiger, S.P.; Birch, D.G.; Ash, J.D.; Iannaccone, A.; Flannery, J.G.; Sahel, J.A.; Zack, D.J.; et al. Inherited Retinal Degenerations: Current Landscape and Knowledge Gaps. *Transl. Vis. Sci. Technol.* 2018, 7, 6. [CrossRef] [PubMed]
- Verbakel, S.K.; van Huet, R.A.C.; Boon, C.J.F.; den Hollander, A.I.; Collin, R.W.J.; Klaver, C.C.W.; Hoyng, C.B.; Roepman, R.; Klevering, B.J. Non-syndromic retinitis pigmentosa. *Prog. Retin. Eye Res.* 2018, 66, 157–186. [CrossRef] [PubMed]
- 3. Thiadens, A.A.; Phan, T.M.; Zekveld-Vroon, R.C.; Leroy, B.P.; van den Born, L.I.; Hoyng, C.B.; Klaver, C.C.; Roosing, S.; Pott, J.W.; van Schooneveld, M.J.; et al. Clinical course, genetic etiology, and visual outcome in cone and cone-rod dystrophy. *Ophthalmology* **2012**, *119*, 819–826. [CrossRef] [PubMed]
- Kumaran, N.; Moore, A.T.; Weleber, R.G.; Michaelides, M. Leber congenital amaurosis/early-onset severe retinal dystrophy: Clinical features, molecular genetics and therapeutic interventions. *Br. J. Ophthalmol.* 2017, 101, 1147–1154. [CrossRef] [PubMed]
- 5. Tsang, S.H.; Sharma, T. Rod Monochromatism (Achromatopsia). Adv. Exp. Med. Biol. 2018, 1085, 119–123.
- 6. Werdich, X.Q.; Place, E.M.; Pierce, E.A. Systemic diseases associated with retinal dystrophies. *Semin. Ophthalmol.* **2014**, *29*, 319–328. [CrossRef]
- Shamseldin, H.E.; Shaheen, R.; Ewida, N.; Bubshait, D.K.; Alkuraya, H.; Almardawi, E.; Howaidi, A.; Sabr, Y.; Abdalla, E.M.; Alfaifi, A.Y.; et al. The morbid genome of ciliopathies: An update. *Genet. Med.* 2020, 22, 1051–1060. [CrossRef]
- 8. Iwama, K.; Takaori, T.; Fukushima, A.; Tohyama, J.; Ishiyama, A.; Ohba, C.; Mitsuhashi, S.; Miyatake, S.; Takata, A.; Miyake, N.; et al. Novel recessive mutations in MSTO1 cause cerebellar atrophy with pigmentary retinopathy. *J. Hum. Genet.* **2018**, *63*, 263–270. [CrossRef]
- 9. Ferreira, C.R.; van Karnebeek, C.D.M. Inborn errors of metabolism. *Handb. Clin. Neurol.* 2019, 162, 449–481.
- Freeze, H.H.; Schachter, H.; Kinoshita, T. Genetic Disorders of Glycosylation. In *Essentials of Glycobiology*, 3rd ed.; Varki, A., Cummings, R.D., Esko, J.D., Stanley, P., Hart, G.W., Aebi, M., Darvill, A.G., Kinoshita, T., Packer, N.H., Prestegard, J.H., et al., Eds.; Cold Spring Harbor Laboratory Press: New York, NY, USA, 2017; Chapter 45.
- 11. Nita, D.A.; Mole, S.E.; Minassian, B.A. Neuronal ceroid lipofuscinoses. *Epileptic Disord*. **2016**, *18*, 73–88. [CrossRef]
- 12. Muenzer, J. Overview of the mucopolysaccharidoses. *Rheumatology (Oxford)* **2011**, *50* (Suppl. 5), v4–v12. [CrossRef]
- 13. Imanaka, T. Biogenesis and Function of Peroxisomes in Human Disease with a Focus on the ABC Transporter. *Biol. Pharm. Bull.* **2019**, *42*, 649–665. [CrossRef] [PubMed]
- 14. Sreekumar, V.; Norris, D.P. Cilia and development. *Curr. Opin. Genet. Dev.* **2019**, *56*, 15–21. [CrossRef] [PubMed]
- 15. May-Simera, H.; Nagel-Wolfrum, K.; Wolfrum, U. Cilia—The sensory antennae in the eye. *Prog. Retin. Eye Res.* **2017**, *60*, 144–180. [CrossRef]
- 16. Tsang, S.H.; Aycinena, A.R.P.; Sharma, T. Ciliopathy: Bardet-Biedl Syndrome. *Adv. Exp. Med. Biol.* **2018**, 1085, 171–174. [PubMed]
- 17. Valente, E.M.; Dallapiccola, B.; Bertini, E. Joubert syndrome and related disorders. *Handb. Clin. Neurol.* **2013**, *113*, 1879–1888. [PubMed]
- 18. Geleoc, G.G.S.; El-Amraoui, A. Disease mechanisms and gene therapy for Usher syndrome. *Hear. Res.* **2020**, 394, 107932. [CrossRef] [PubMed]

- Tsang, S.H.; Aycinena, A.R.P.; Sharma, T. Ciliopathy: Senior-Loken Syndrome. *Adv. Exp. Med. Biol.* 2018, 1085, 175–178. [PubMed]
- 20. Tsang, S.H.; Aycinena, A.R.P.; Sharma, T. Ciliopathy: Alstrom Syndrome. *Adv. Exp. Med. Biol.* 2018, 1085, 179–180. [PubMed]
- 21. Petriman, N.A.; Lorentzen, E. Moving proteins along in the cilium. Elife 2020, 9, e55254. [CrossRef]
- 22. Hsu, Y.; Garrison, J.E.; Kim, G.; Schmitz, A.R.; Searby, C.C.; Zhang, Q.; Datta, P.; Nishimura, D.Y.; Seo, S.; Sheffield, V.C. BBSome function is required for both the morphogenesis and maintenance of the photoreceptor outer segment. *PLoS Genet.* **2017**, *13*, e1007057. [CrossRef]
- 23. Hsu, Y.; Garrison, J.E.; Seo, S.; Sheffield, V.C. The absence of BBSome function decreases synaptogenesis and causes ectopic synapse formation in the retina. *Sci. Rep.* **2020**, *10*, 8321. [CrossRef]
- 24. Wang, S.F.; Kowal, T.J.; Ning, K.; Koo, E.B.; Wu, A.Y.; Mahajan, V.B.; Sun, Y. Review of Ocular Manifestations of Joubert Syndrome. *Genes* **2018**, *9*, 605. [CrossRef] [PubMed]
- 25. El-Amraoui, A.; Petit, C. The retinal phenotype of Usher syndrome: Pathophysiological insights from animal models. *Comptes Rendus Biol.* **2014**, 337, 167–177. [CrossRef] [PubMed]
- Yang, X.R.; Benson, M.D.; MacDonald, I.M.; Innes, A.M. A diagnostic approach to syndromic retinal dystrophies with intellectual disability. *Am. J. Med. Genet. C Semin. Med. Genet.* 2020, 184, 538–570. [CrossRef] [PubMed]
- Namburi, P.; Ratnapriya, R.; Khateb, S.; Lazar, C.H.; Kinarty, Y.; Obolensky, A.; Erdinest, I.; Marks-Ohana, D.; Pras, E.; Ben-Yosef, T.; et al. Bi-allelic Truncating Mutations in CEP78, Encoding Centrosomal Protein 78, Cause Cone-Rod Degeneration with Sensorineural Hearing Loss. *Am. J. Hum. Genet.* 2016, *99*, 777–784. [CrossRef]
- Luscan, R.; Mechaussier, S.; Paul, A.; Tian, G.; Gerard, X.; Defoort-Dellhemmes, S.; Loundon, N.; Audo, I.; Bonnin, S.; LeGargasson, J.F.; et al. Mutations in TUBB4B Cause a Distinctive Sensorineural Disease. *Am. J. Hum. Genet.* 2017, 101, 1006–1012. [CrossRef]
- 29. Sims, K.B. NDP-Related Retinopathies. In *GeneReviews*<sup>®</sup>; Adam, M.P., Ardinger, H.H., Pagon, R.A., Wallace, S.E., Bean, L.J., Stephens, K., Amemiya, A., Eds.; University of Washington: Seattle, WA, USA, 2014.
- 30. Tsang, S.H.; Sharma, T. Inborn Errors of Metabolism: Refsum Disease. *Adv. Exp. Med. Biol.* **2018**, 1085, 191–192.
- 31. Raas-Rothschild, A.; Wanders, R.J.; Mooijer, P.A.; Gootjes, J.; Waterham, H.R.; Gutman, A.; Suzuki, Y.; Shimozawa, N.; Kondo, N.; Eshel, G.; et al. A PEX6-defective peroxisomal biogenesis disorder with severe phenotype in an infant, versus mild phenotype resembling Usher syndrome in the affected parents. *Am. J. Hum. Genet.* 2002, 70, 1062–1068. [CrossRef]
- Smith, C.E.; Poulter, J.A.; Levin, A.V.; Capasso, J.E.; Price, S.; Ben-Yosef, T.; Sharony, R.; Newman, W.G.; Shore, R.C.; Brookes, S.J.; et al. Spectrum of PEX1 and PEX6 variants in Heimler syndrome. *Eur. J. Hum. Genet.* 2016, 24, 1565–1571. [CrossRef]
- Le Quesne Stabej, P.; Saihan, Z.; Rangesh, N.; Steele-Stallard, H.B.; Ambrose, J.; Coffey, A.; Emmerson, J.; Haralambous, E.; Hughes, Y.; Steel, K.P.; et al. Comprehensive sequence analysis of nine Usher syndrome genes in the UK National Collaborative Usher Study. *J. Med. Genet.* 2012, 49, 27–36. [CrossRef] [PubMed]
- Carss, K.J.; Arno, G.; Erwood, M.; Stephens, J.; Sanchis-Juan, A.; Hull, S.; Megy, K.; Grozeva, D.; Dewhurst, E.; Malka, S.; et al. Comprehensive Rare Variant Analysis via Whole-Genome Sequencing to Determine the Molecular Pathology of Inherited Retinal Disease. *Am. J. Hum. Genet.* 2017, *100*, 75–90. [CrossRef] [PubMed]
- Dockery, A.; Stephenson, K.; Keegan, D.; Wynne, N.; Silvestri, G.; Humphries, P.; Kenna, P.F.; Carrigan, M.; Farrar, G.J. Target 5000: Target Capture Sequencing for Inherited Retinal Degenerations. *Genes* 2017, *8*, 304. [CrossRef] [PubMed]
- 36. Sharon, D.; Ben-Yosef, T.; Goldenberg-Cohen, N.; Pras, E.; Gradstein, L.; Soudry, S.; Mezer, E.; Zur, D.; Abbasi, A.H.; Zeitz, C.; et al. A nationwide genetic analysis of inherited retinal diseases in Israel as assessed by the Israeli inherited retinal disease consortium (IIRDC). *Hum. Mutat.* 2019, *41*, 140–149. [CrossRef] [PubMed]
- 37. Lenassi, E.; Vincent, A.; Li, Z.; Saihan, Z.; Coffey, A.J.; Steele-Stallard, H.B.; Moore, A.T.; Steel, K.P.; Luxon, L.M.; Heon, E.; et al. A detailed clinical and molecular survey of subjects with nonsyndromic USH2A retinopathy reveals an allelic hierarchy of disease-causing variants. *Eur. J. Hum. Genet.* 2015, 23, 1318–1327. [CrossRef] [PubMed]

- 38. Pierrache, L.H.; Hartel, B.P.; van Wijk, E.; Meester-Smoor, M.A.; Cremers, F.P.; de Baere, E.; de Zaeytijd, J.; van Schooneveld, M.J.; Cremers, C.W.; Dagnelie, G.; et al. Visual Prognosis in USH2A-Associated Retinitis Pigmentosa Is Worse for Patients with Usher Syndrome Type IIa Than for Those with Nonsyndromic Retinitis Pigmentosa. *Ophthalmology* 2016, *123*, 1151–1160. [CrossRef] [PubMed]
- Sengillo, J.D.; Cabral, T.; Schuerch, K.; Duong, J.; Lee, W.; Boudreault, K.; Xu, Y.; Justus, S.; Sparrow, J.R.; Mahajan, V.B.; et al. Electroretinography Reveals Difference in Cone Function between Syndromic and Nonsyndromic USH2A Patients. *Sci. Rep.* 2017, *7*, 11170. [CrossRef]
- Nishiguchi, K.M.; Avila-Fernandez, A.; van Huet, R.A.; Corton, M.; Perez-Carro, R.; Martin-Garrido, E.; Lopez-Molina, M.I.; Blanco-Kelly, F.; Hoefsloot, L.H.; van Zelst-Stams, W.A.; et al. Exome sequencing extends the phenotypic spectrum for ABHD12 mutations: From syndromic to nonsyndromic retinal degeneration. *Ophthalmology* 2014, 121, 1620–1627. [CrossRef]
- 41. Nguyen, T.T.; Hull, S.; Roepman, R.; van den Born, L.I.; Oud, M.M.; de Vrieze, E.; Hetterschijt, L.; Letteboer, S.J.F.; van Beersum, S.E.C.; Blokland, E.A.; et al. Missense mutations in the WD40 domain of AHI1 cause non-syndromic retinitis pigmentosa. *J. Med. Genet.* **2017**, *54*, 624–632. [CrossRef]
- 42. Aldrees, A.; Abdelkader, E.; Al-Habboubi, H.; Alrwebah, H.; Rahbeeni, Z.; Schatz, P. Non-syndromic retinal dystrophy associated with homozygous mutations in the ALMS1 gene. *Ophthalmic Genet.* **2019**, *40*, 77–79. [CrossRef]
- 43. Audo, I.; El Shamieh, S.; Mejecase, C.; Michiels, C.; Demontant, V.; Antonio, A.; Condroyer, C.; Boyard, F.; Letexier, M.; Saraiva, J.P.; et al. ARL2BP mutations account for 0.1% of autosomal recessive rod-cone dystrophies with the report of a novel splice variant. *Clin. Genet.* **2017**, *92*, 109–111. [PubMed]
- Davidson, A.E.; Schwarz, N.; Zelinger, L.; Stern-Schneider, G.; Shoemark, A.; Spitzbarth, B.; Gross, M.; Laxer, U.; Sosna, J.; Sergouniotis, P.I.; et al. Mutations in ARL2BP, encoding ADP-ribosylation-factor-like 2 binding protein, cause autosomal-recessive retinitis pigmentosa. *Am. J. Hum. Genet.* 2013, *93*, 321–329. [CrossRef] [PubMed]
- 45. Holtan, J.P.; Teigen, K.; Aukrust, I.; Bragadottir, R.; Houge, G. Dominant ARL3-related retinitis pigmentosa. *Ophthalmic Genet.* **2019**, 40, 124–128. [CrossRef] [PubMed]
- 46. Aldahmesh, M.A.; Safieh, L.A.; Alkuraya, H.; Al-Rajhi, A.; Shamseldin, H.; Hashem, M.; Alzahrani, F.; Khan, A.O.; Alqahtani, F.; Rahbeeni, Z.; et al. Molecular characterization of retinitis pigmentosa in Saudi Arabia. *Mol. Vis.* **2009**, *15*, 2464–2469.
- 47. Shevach, E.; Ali, M.; Mizrahi-Meissonnier, L.; McKibbin, M.; El-Asrag, M.; Watson, C.M.; Inglehearn, C.F.; Ben-Yosef, T.; Blumenfeld, A.; Jalas, C.; et al. Association Between Missense Mutations in the BBS2 Gene and Nonsyndromic Retinitis Pigmentosa. *JAMA Ophthalmol.* **2015**, *133*, 312–318. [CrossRef]
- Khan, A.O.; Decker, E.; Bachmann, N.; Bolz, H.J.; Bergmann, C. C8orf37 is mutated in Bardet-Biedl syndrome and constitutes a locus allelic to non-syndromic retinal dystrophies. *Ophthalmic Genet.* 2016, 37, 290–293. [CrossRef]
- Mejecase, C.; Hummel, A.; Mohand-Said, S.; Andrieu, C.; El Shamieh, S.; Antonio, A.; Condroyer, C.; Boyard, F.; Foussard, M.; Blanchard, S.; et al. Whole exome sequencing resolves complex phenotype and identifies CC2D2A mutations underlying non-syndromic rod-cone dystrophy. *Clin. Genet.* 2019, *95*, 329–333. [CrossRef]
- 50. den Hollander, A.I.; Koenekoop, R.K.; Yzer, S.; Lopez, I.; Arends, M.L.; Voesenek, K.E.; Zonneveld, M.N.; Strom, T.M.; Meitinger, T.; Brunner, H.G.; et al. Mutations in the CEP290 (NPHP6) gene are a frequent cause of Leber congenital amaurosis. *Am. J. Hum. Genet.* **2006**, *79*, 556–561. [CrossRef]
- 51. Khan, A.O.; Eisenberger, T.; Nagel-Wolfrum, K.; Wolfrum, U.; Bolz, H.J. C21orf2 is mutated in recessive early-onset retinal dystrophy with macular staphyloma and encodes a protein that localises to the photoreceptor primary cilium. *Br. J. Ophthalmol.* **2015**, *99*, 1725–1731. [CrossRef]
- 52. Suga, A.; Mizota, A.; Kato, M.; Kuniyoshi, K.; Yoshitake, K.; Sultan, W.; Yamazaki, M.; Shimomura, Y.; Ikeo, K.; Tsunoda, K.; et al. Identification of Novel Mutations in the LRR-Cap Domain of C21orf2 in Japanese Patients With Retinitis Pigmentosa and Cone-Rod Dystrophy. *Investig. Ophthalmol. Vis. Sci.* 2016, 57, 4255–4263. [CrossRef]
- 53. Ku, C.A.; Hull, S.; Arno, G.; Vincent, A.; Carss, K.; Kayton, R.; Weeks, D.; Anderson, G.W.; Geraets, R.; Parker, C.; et al. Detailed Clinical Phenotype and Molecular Genetic Findings in CLN3-Associated Isolated Retinal Degeneration. *JAMA Ophthalmol.* **2017**, *135*, 749–760. [CrossRef] [PubMed]

- 54. Khan, M.I.; Kersten, F.F.; Azam, M.; Collin, R.W.; Hussain, A.; Shah, S.T.; Keunen, J.E.; Kremer, H.; Cremers, F.P.; Qamar, R.; et al. CLRN1 mutations cause nonsyndromic retinitis pigmentosa. *Ophthalmology* **2011**, *118*, 1444–1448. [CrossRef] [PubMed]
- 55. Xu, M.; Xie, Y.A.; Abouzeid, H.; Gordon, C.T.; Fiorentino, A.; Sun, Z.; Lehman, A.; Osman, I.S.; Dharmat, R.; Riveiro-Alvarez, R.; et al. Mutations in the Spliceosome Component CWC27 Cause Retinal Degeneration with or without Additional Developmental Anomalies. *Am. J. Hum. Genet.* **2017**, *100*, 592–604. [CrossRef] [PubMed]
- Lam, B.L.; Zuchner, S.L.; Dallman, J.; Wen, R.; Alfonso, E.C.; Vance, J.M.; Pericak-Vance, M.A. Mutation K42E in dehydrodolichol diphosphate synthase (DHDDS) causes recessive retinitis pigmentosa. *Adv. Exp. Med. Biol.* 2014, 801, 165–170.
- 57. Zelinger, L.; Banin, E.; Obolensky, A.; Mizrahi-Meissonnier, L.; Beryozkin, A.; Bandah-Rozenfeld, D.; Frenkel, S.; Ben-Yosef, T.; Merin, S.; Schwartz, S.B.; et al. A missense mutation in DHDDS, encoding dehydrodolichyl diphosphate synthase, is associated with autosomal-recessive retinitis pigmentosa in Ashkenazi Jews. *Am. J. Hum. Genet.* **2011**, *88*, 207–215. [CrossRef]
- 58. Kuehlewein, L.; Schols, L.; Llavona, P.; Grimm, A.; Biskup, S.; Zrenner, E.; Kohl, S. Phenotypic spectrum of autosomal recessive retinitis pigmentosa without posterior column ataxia caused by mutations in the FLVCR1 gene. *Graefes Arch. Clin. Exp. Ophthalmol.* **2019**, 257, 629–638. [CrossRef]
- Haer-Wigman, L.; Newman, H.; Leibu, R.; Bax, N.M.; Baris, H.N.; Rizel, L.; Banin, E.; Massarweh, A.; Roosing, S.; Lefeber, D.J.; et al. Non-syndromic retinitis pigmentosa due to mutations in the mucopolysaccharidosis type IIIC gene, heparan-alpha-glucosaminide N-acetyltransferase (HGSNAT). *Hum. Mol. Genet.* 2015, 24, 3742–3751. [CrossRef]
- 60. Xu, M.; Yang, L.; Wang, F.; Li, H.; Wang, X.; Wang, W.; Ge, Z.; Wang, K.; Zhao, L.; Li, H.; et al. Mutations in human IFT140 cause non-syndromic retinal degeneration. *Hum. Genet.* **2015**, *134*, 1069–1078. [CrossRef]
- 61. Stone, E.M.; Cideciyan, A.V.; Aleman, T.S.; Scheetz, T.E.; Sumaroka, A.; Ehlinger, M.A.; Schwartz, S.B.; Fishman, G.A.; Traboulsi, E.I.; Lam, B.L.; et al. Variations in NPHP5 in patients with nonsyndromic leber congenital amaurosis and Senior-Loken syndrome. *Arch. Ophthalmol.* **2011**, *129*, 81–87. [CrossRef]
- Khan, K.N.; El-Asrag, M.E.; Ku, C.A.; Holder, G.E.; McKibbin, M.; Arno, G.; Poulter, J.A.; Carss, K.; Bommireddy, T.; Bagheri, S.; et al. Specific Alleles of CLN7/MFSD8, a Protein That Localizes to Photoreceptor Synaptic Terminals, Cause a Spectrum of Nonsyndromic Retinal Dystrophy. *Investig. Ophthalmol. Vis. Sci.* 2017, 58, 2906–2914. [CrossRef]
- Collison, F.T.; Xie, Y.A.; Gambin, T.; Jhangiani, S.; Muzny, D.; Gibbs, R.; Lupski, J.R.; Fishman, G.A.; Allikmets, R. Whole Exome Sequencing Identifies an Adult-Onset Case of Methylmalonic Aciduria and Homocystinuria Type C (cblC) with Non-Syndromic Bull's Eye Maculopathy. *Ophthalmic Genet.* 2015, 36, 270–275. [CrossRef] [PubMed]
- Siemiatkowska, A.M.; van den Born, L.I.; van Hagen, P.M.; Stoffels, M.; Neveling, K.; Henkes, A.; Kipping-Geertsema, M.; Hoefsloot, L.H.; Hoyng, C.B.; Simon, A.; et al. Mutations in the mevalonate kinase (MVK) gene cause nonsyndromic retinitis pigmentosa. *Ophthalmology* 2013, *120*, 2697–2705. [CrossRef] [PubMed]
- Chen, Z.Y.; Battinelli, E.M.; Fielder, A.; Bundey, S.; Sims, K.; Breakefield, X.O.; Craig, I.W. A mutation in the Norrie disease gene (NDP) associated with X-linked familial exudative vitreoretinopathy. *Nat. Genet.* 1993, 5, 180–183. [CrossRef] [PubMed]
- Webb, T.R.; Parfitt, D.A.; Gardner, J.C.; Martinez, A.; Bevilacqua, D.; Davidson, A.E.; Zito, I.; Thiselton, D.L.; Ressa, J.H.; Apergi, M.; et al. Deep intronic mutation in OFD1, identified by targeted genomic next-generation sequencing, causes a severe form of X-linked retinitis pigmentosa (RP23). *Hum. Mol. Genet.* 2012, 21, 3647–3654. [CrossRef]
- 67. Vincent, A.; Forster, N.; Maynes, J.T.; Paton, T.A.; Billingsley, G.; Roslin, N.M.; Ali, A.; Sutherland, J.; Wright, T.; Westall, C.A.; et al. OTX2 mutations cause autosomal dominant pattern dystrophy of the retinal pigment epithelium. *J. Med. Genet.* **2014**, *51*, 797–805. [CrossRef]
- Tee, J.J.; Smith, A.J.; Hardcastle, A.J.; Michaelides, M. RPGR-associated retinopathy: Clinical features, molecular genetics, animal models and therapeutic options. *Br. J. Ophthalmol.* 2016, 100, 1022–1027. [CrossRef]

- 69. Riazuddin, S.A.; Iqbal, M.; Wang, Y.; Masuda, T.; Chen, Y.; Bowne, S.; Sullivan, L.S.; Waseem, N.H.; Bhattacharya, S.; Daiger, S.P.; et al. A splice-site mutation in a retina-specific exon of BBS8 causes nonsyndromic retinitis pigmentosa. *Am. J. Hum. Genet.* **2010**, *86*, 805–812. [CrossRef]
- 70. Rivolta, C.; Sweklo, E.A.; Berson, E.L.; Dryja, T.P. Missense mutation in the USH2A gene: Association with recessive retinitis pigmentosa without hearing loss. *Am. J. Hum. Genet.* **2000**, *66*, 1975–1978. [CrossRef]
- 71. Ehrenberg, M.; Weiss, S.; Orenstein, N.; Goldenberg-Cohen, N.; Ben-Yosef, T. The co-occurrence of rare non-ocular phenotypes in patients with inherited retinal degenerations. *Mol. Vis.* **2019**, *25*, 691–702.
- Ku, C.A.; Pennesi, M.E. The new landscape of retinal gene therapy. Am. J. Med. Genet. C Semin. Med. Genet. 2020, 184, 846–859. [CrossRef]
- Bach, G.; Webb, M.B.; Bargal, R.; Zeigler, M.; Ekstein, J. The frequency of mucolipidosis type IV in the Ashkenazi Jewish population and the identification of 3 novel MCOLN1 mutations. *Hum. Mutat.* 2005, 26, 591. [CrossRef] [PubMed]
- 74. Ben-Yosef, T.; Ness, S.L.; Madeo, A.C.; Bar-Lev, A.; Wolfman, J.H.; Ahmed, Z.M.; Desnick, R.J.; Willner, J.P.; Avraham, K.B.; Ostrer, H.; et al. A mutation of PCDH15 among Ashkenazi Jews with the type 1 Usher syndrome. *N. Engl. J. Med.* **2003**, *348*, 1664–1670. [CrossRef] [PubMed]
- 75. Fedick, A.; Jalas, C.; Abeliovich, D.; Krakinovsky, Y.; Ekstein, J.; Ekstein, A.; Treff, N.R. Carrier frequency of two BBS2 mutations in the Ashkenazi population. *Clin. Genet.* **2014**, *85*, 578–582. [CrossRef] [PubMed]
- 76. Ness, S.L.; Ben-Yosef, T.; Bar-Lev, A.; Madeo, A.C.; Brewer, C.C.; Avraham, K.B.; Kornreich, R.; Desnick, R.J.; Willner, J.P.; Friedman, T.B.; et al. Genetic homogeneity and phenotypic variability among Ashkenazi Jews with Usher syndrome type III. *J. Med. Genet.* 2003, 40, 767–772. [CrossRef] [PubMed]
- 77. Joensuu, T.; Hamalainen, R.; Yuan, B.; Johnson, C.; Tegelberg, S.; Gasparini, P.; Zelante, L.; Pirvola, U.; Pakarinen, L.; Lehesjoki, A.E.; et al. Mutations in a novel gene with transmembrane domains underlie Usher syndrome type 3. *Am. J. Hum. Genet.* **2001**, *69*, 673–684. [CrossRef]
- 78. Kyttala, M.; Tallila, J.; Salonen, R.; Kopra, O.; Kohlschmidt, N.; Paavola-Sakki, P.; Peltonen, L.; Kestila, M. MKS1, encoding a component of the flagellar apparatus basal body proteome, is mutated in Meckel syndrome. *Nat. Genet.* 2006, *38*, 155–157. [CrossRef]
- 79. Branham, K.; Schlegel, D.; Fahim, A.T.; Jayasundera, K.T. Genetic testing for inherited retinal degenerations: Triumphs and tribulations. *Am. J. Med. Genet. C Semin. Med. Genet.* **2020**, *184*, 571–577. [CrossRef]
- Mansfield, B.C.; Yerxa, B.R.; Branham, K.H. Implementation of a registry and open access genetic testing program for inherited retinal diseases within a non-profit foundation. *Am. J. Med. Genet. C Semin. Med. Genet.* 2020, 184, 838–845. [CrossRef]
- Stone, E.M.; Andorf, J.L.; Whitmore, S.S.; DeLuca, A.P.; Giacalone, J.C.; Streb, L.M.; Braun, T.A.; Mullins, R.F.; Scheetz, T.E.; Sheffield, V.C.; et al. Clinically Focused Molecular Investigation of 1000 Consecutive Families with Inherited Retinal Disease. *Ophthalmology* 2017, *124*, 1314–1331. [CrossRef]
- 82. Haer-Wigman, L.; van Zelst-Stams, W.A.; Pfundt, R.; van den Born, L.I.; Klaver, C.C.; Verheij, J.B.; Hoyng, C.B.; Breuning, M.H.; Boon, C.J.; Kievit, A.J.; et al. Diagnostic exome sequencing in 266 Dutch patients with visual impairment. *Eur. J. Hum. Genet.* **2017**, *25*, 591–599. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).