## **Review Article**



# **Cancer immune resistance: can theories converge?**

## Rongze Lu<sup>1</sup>, Tolga Turan<sup>2</sup>, Josue Samayoa<sup>2</sup> and Francesco M. Marincola<sup>2</sup>

<sup>1</sup>AbbVie Cellular Molecular Oncology, 1500 Seaport Boulevard, Redwood City, CA 94063, U.S.A.; <sup>2</sup>AbbVie Immune Oncology Discovery, Redwood City, CA, U.S.A. **Correspondence:** Rongze Lu (rongze.lu@abbvie.com)

Immune oncology (IO) is challenged to expand its usefulness to a broader range of cancers. A second generation of IO agents acting beyond the realm of Checkpoint Inhibitor Therapy (CIT) is sought with the intent of turning immune-resistant cancers into appealing IO targets. The published literature proposes a profusion of models to explain cancer immune resistance to CIT that largely outnumber the immune landscapes and corresponding resistance mechanisms. In spite of the complex and contradicting models suggested to explain refractoriness to CIT, the identification of prevailing mechanisms and their targeting may not be as daunting as it at first appears. Here, we suggest that cancer cells go through a conserved evolutionary bottleneck facing a Two-Option Choice to evade recognition by the immune competent host: they can either adopt a clean oncogenic process devoid of immunogenic stimuli (immune-silent tumors) or display an entropic biology prone to immune recognition (immune-active tumors) but resilient to rejection thanks to the recruitment of compensatory immune suppressive processes. Strategies aimed at enhancing the effectiveness of CIT will be different according to the immune landscape targeted.

Immune oncology (IO) is urged to expand the usefulness of Checkpoint Inhibitor Therapy (CIT) to a broader range of refractory cancers [1-3]. In spite of the variety of models proposed to explain cancer immune resistance, the identification of prevailing mechanisms and their targeting may not be as daunting as it at first appears [4] as long as answers are sought following a well-designed and systematic strategy; to quote Jonas Salk: 'the answer to biological problems preexists, it is the question that needs to be discovered [5]. A survey of open access data from The Cancer Genome Atlas (TCGA) comprising four histotypes (breast, lung, colon carcinoma and melanoma) indicated that cancer cells go through a conserved evolutionary bottleneck facing a Two-Option Choice (TOC) to evade immune recognition by the immune competent host: they can either adopt a clean oncogenic process devoid of immunogenic stimuli (immune-silent tumors) or display an entropic biology prone to immune recognition (immune-active tumors) but resilient to rejection thanks to the recruitment of compensatory immune suppressive processes [3]. We refer to the first option as Primary Immune Resistance (PIRes) and to the second as Compensatory Immune Resistance (CIRes). These two landscapes may influence refractoriness to CIT through entirely distinct mechanisms. In addition, Secondary Immune Resistance (SIRes) may ensue as an escape mechanism following originally successful treatment. Finally, we refer to False-Immune Resistance in those cases in which treatment could not be completed due to limiting toxicity.

To explain the distinct landscapes and the respective reasons determining immune refractoriness to CIT, a wealth of observational and/or experimental models has been advocated that largely outnumber the three phenotypes of human cancer (Table 1). The current series presents some of the salient models that propose a targetable mechanism regulating the growth of cancer in the immune competent host, primarily focusing on immune regulatory control of cancer within the immune-active landscapes. However, this review will lean toward the discussion of potential strategies to immune convert silent tumors into immune-active ones, therefore, offering a window of opportunity for IO agents that would otherwise be unlikely to affect an immune-silent environment where innate resistance dominates.

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	References	ICR group
WNT/β–Catenin	[6,7]	Depleted
MAPK hypothesis	[8]	Depleted
Immunogenic cell death	[9,10,11]	Active
Regulatory T cells	[12,13]	Active
IL23-Th17 axis	[14–18]	Active
Myeloid suppressor cells	[19]	Active
PI3K-γ signature	[20–24]	Depleted
IDO/NOS signature	[25–27]	Ubiquitous
SGK1 signature	[28,29]	Depleted
Shc1 signature	[30]	Depleted
Barrier molecules	[31,32]	Depleted
Mesenchymal transition	[33–35]	Depleted
Cancer-associated fibroblasts	[36–40]	Ubiquitous
TAM receptor tyrosine kinases	[41–45]	Active
Hypoxia/adenosine suppression	[46,47]	Active
TREX1 clearance of cytosolic DNA	[48–50]	NA
Checkpoint cluster	[51,52]	Active
Oncogene addiction	[53,54]	Depleted
Epigenetic regulation	[55–58]	Depleted
Regulatory B cells	[59]	Active
NF-KB activation	[60]	

Table 1 Salient models explaining cancer immune landscapes
and pertinent literature

We recently proposed a theory that unifies current models of cancer immune resistance into a Theory of Everything (TOE) assigning each of them to a specific immune landscape according to their transcriptional expression pattern [3]. This conclusion was based on a survey of two open access datasets comprising ~3000 cases of breast cancer [3,8,61]. A nomenclature was proposed to define cancers according to their immune contexture ranking them according to the transcriptional expression of genes associated with Immune-mediated Tissue-specific Destruction (ITD). ITD is a conserved mechanism responsible for destructive flares of autoimmunity, acute allograft rejection, and graft-versus-host disease, clearance of pathogeninfected cells and rejection of cancer [62,63]. A signature representative of the ITD was selected from a larger set of interferon (IFN)-y-induced transcripts named the Immunologic Constant of Rejection (ICR) [62]. The ICR bears both predictive and prognostic implications within a continuum of anticancer immune surveillance [64] and includes four functional categories: CXCR3/CCR5 chemokines (CXCL9, CXCL10 and CCL5), Th1 signaling (IFNG, IL12B, TBX21, CD8A, STAT1, IRF1 and CD8B), effector (GNLY, PRF1, GZMA, GZMB and GZMH) and immune regulatory (CD274, CTLA4, FOXP3, IDO1 and PDCD1) functions. The expression of the 20 representative genes is highly correlated with the extended ICR signature that includes  $\sim 500$  genes [62,63,65]. It has been conclusively shown that responsiveness to CIT is observed almost exclusively in the immune-active landscape and is predetermined by a conducive microenvironment [35,66,67]. However, while the immune-active landscape is a prerequisite, it is not sufficient alone to predict immune response.

This concept was described originally by our group in 2002 in the context of other types of immunotherapy, including response to antigen-specific vaccination administered in combination with systemic interleukin-2 [68], and subsequently validated in the context of systemic interleukin-2 administration [69] and the adoptive transfer of tumor-infiltrating lymphocytes [70]. Therefore, immune responsiveness is promiscuous to treatment and it is multifactorial with the tumor microenvironment, playing a permissive but not exclusive role [71].



The 20-gene ICR signatures bear strong analogy with other IFN- $\gamma$ -dependent signatures predictive of immune responsiveness to interleukin-2-based therapies [68–70] and CIT [66]. Thus, we used these 20 genes as surrogate biomarkers to define immune landscapes more or less likely to be susceptible to CIT. The expression pattern of the 20 ICR genes defined four cancer immune landscapes that were segregated from ICR1 to ICR4 according to the crescendo of expression of ICR transcripts. ICR1 and ICR2 represent various degrees of immune depletion, while ICR3 and ICR4 demonstrate rising levels of expression of ICR genes. For the purpose of discussion, the four landscapes were conflated conceptually into immune-silent or immune-active clusters.

Subsequently, a selection of transcripts reported in association with various cancer immune resistance models was collated into a signature meant to unify within a single study the hallmarks of cancer immune biology (Table 1). We refer to this collection as the TOE signatures of resistance (sRes) and used it to define the geographical distribution of each model within immune landscapes.

We observed that most transcripts representative of immune regulatory mechanisms tightly correlated in expression with the ICR and TIS (tumor inflammation signature), suggesting that immune suppression goes hand-in-hand with immune activation [3].

Based on this analysis of breast cancer data, we hypothesized that immune-silent tumors evolve by employing a strictly essential interface of interactions with the host's stromal cells that exclude immune cell recognition. This may be due to the development of a cancer cell cycle that avoids Immunogenic Cell Death (ICD) or by the downstream induction of biochemical or mechanical barriers that hamper immune infiltration. Thus, these 'clean' tumors evolve through the promotion of cancer cells that adopt refined growth mechanisms reduced to the bare necessities of life. Indeed, similar observations could be corroborated by the analysis of another three cancer histotypes including lung, colon carcinomas and melanoma (Figure 1).

This hypothesis is corroborated by the observation that these tumors (1) are transcriptionally dormant compared with the immune-active ones and (2) bear low prevalence of mutations in oncogenes suggesting a more orderly growth process [8]. It is, therefore, reasonable to suppose that clean tumor growth is dependent on a stepwise oncogenic mechanism that avoids immune recognition [72–74]. Thus, we propose that the natural history of cancer is shaped at the cross-road of two biologies by a 'TOC' or Hobson's predicament: (1) immunogenic tumors can only survive in the host when immune suppressive mechanisms balance the reaction of the host and (2) silent tumors can grow undisturbed.

Here, we suggest that interference of a clean oncogenic pathway may result not only in cancer cell death but also in the disruption of its biology leading to less pristine processes conducive to ICD and allowing, therefore, a window of opportunity for IO agents [9-11,75-95].

This concept is based on the premise that cancer is fundamentally a cell biology problem with cancer cells orchestrating and directing their surroundings. Therefore, efforts aiming at altering the tumor microenvironment should primarily be directed toward the disruption of intrinsic cancer cell processes. A best example of the central role played by cancer cells in determining their surroundings is the Patient-Derived Xenograft (PDX) model; after three passages in immune-deficient mice, the mouse stromal cells completely replace the human, yet the original architecture of the cancer is maintained [96–98]. A second premise is that the immune environment of cancer is driven by a cascade of innate mechanisms (first signal), while the adaptive immune response requires signals initiated by the innate immune system that inform about the origin of the antigen and the type of response to be induced as described by Charles A. Janeway and by Polly Matzinger's Danger Model [83,99–108].

The leading role played by ICD in driving the immune landscape of cancer is counteracted by lines of thought that promote priming of adaptive immune responses by non-self-antigens (neo-antigens) generated by the translation of missense mutations into novel protein domains. This hypothesis is based on several experimental [109,110] as well as the clinical observation that cancers with high mutational burden are more frequently associated with the immune-active landscape and consequently with responsiveness to CIT [33,98,109]. This concept has been, however, questioned by recent observations by our [8] and others' groups [111]. Moreover, basic understanding of immunologic processes confutes the primary role that adaptive immunity plays in the rejection of cancer in the absence of a first initiating signal [99–104,107]. The conditionality of adaptive immune responses is suggested by experimental evidence that they are not an essential requirement for the rejection of cancer as exemplified by the transferrable anticancer innate immunity model [112–115] and by oncotropic virus-mediated immune rejection of xenografts in immune-deficient mice [116,117]. From the clinical standpoint, the secondary role played by adaptive immunity could also explain







Mapping gene signatures according to their expression in different immune landscapes of breast (BRCA), lung (LUAD) and colon (COAD) carcinoma, and melanoma (SKCM); in red are signatures selectively expressed in the immune-active (ICR4) landscape, in blue those selectively expressed in the immune-silent (ICR1) landscape and in white those that are ubiquitously expressed independent of immune landscape.

the paradoxical observation that vaccines aimed at priming adaptive immune responses can consistently elicit systemic immunity, which, however, does not correlate with tumor rejection [118,119]. It could be postulated that because of the adjuvants used in vaccine administration, at the site of vaccination the afferent loop of the adaptive immune response can be initiated stimulating chemoattraction and antigen presentation. However, at the tumor site, in the absence of a strong innate immunity-mediated chemoattraction, the efferent loop languishes mostly because of lack of trafficking on vaccine-induced cancer-specific T cells to the target tissue. It should also be pointed out that seminal studies done on the effectiveness of tumor-infiltrating lymphocytes demonstrated that their homing at the tumor site is necessary, though not sufficient, to induce tumor regression, emphasizing the critical role that chemoattraction plays in immune responsiveness [120]. In turn, chemoattraction of circulating T cells is tightly dependent on the expression of CXCR3- and CCR5-binding chemokines that are expressed in response to innate immune activation as a component of the ICR signature. Finally, it has been recently shown that the intra-lesional injection of oncolytic virus can turn an immune-silent tumor into an immunogenic one with activation of innate signals that secondarily attracts adaptive immune responses [121].

Thus, we believe that the prospect for IO therapy is to segregate future efforts according to immune landscapes and respective cause for refractoriness to CIT. It is likely that cancers displaying immune-activated landscapes and associated CIRes will benefit from combination of various IO agents that could shift the balance in favor of immune-effector over immune regulatory mechanisms. On the other hand, silent cancers will need to be primed to stir ICD and subsequent recruitment of innate and adaptive immune cells that could become suitable targets for IO agents including CIT.



## Summary

- Immune suppression goes hand-in-hand with immune activation.
- Immune-active tumors include almost exclusively all the immune regulatory mechanisms to counterbalance their immunogenicity.
- Immune-silent tumors are enriched of signatures that reflect activation of oncogenic mechanisms and exclude immune regulatory mechanism.
- Human cancers go through a conserved evolutionary bottleneck facing a two-option choice to evade immune recognition by the immune competent host: they either adopt a clean oncogenic process devoid of immunogenic stimuli or display an entropic biology prone to immune recognition but resilient to rejection thanks to the recruitment of compensatory immune suppressive processes.
- Immunotherapy agents including check point inhibitors work only on the immune-active tumors enriched of immune regulatory mechanisms.
- Immune-silent tumors need to be targeted with agents that can disrupt their lean biology and induce immunogenic cell death.

#### Abbreviations

CIRes, compensatory immune resistance; CIT, checkpoint inhibitor therapy; ICD, immunogenic cell death; ICR, Immunologic Constant of Rejection; IFN, interferon; IO, immune oncology; ITD, immune-mediated tissue-specific destruction; TCGA, The Cancer Genome Atlas; TLR, Toll-like receptor; TOC, Two-Option Choice; TOE, Theory of Everything.

#### **Competing Interests**

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### References

- 1 Emens, L.A., Ascierto, P.A., Darcy, P.K., Demaria, S., Eggermont, A.M.M., Redmond, W.L. et al. (2017) Cancer immunotherapy: Opportunities and challenges in the rapidly evolving clinical landscape. *Eur. J. Cancer* **81**, 116–129 https://doi.org/10.1016/j.ejca.2017.01.035
- 2 Ascierto, P.A. and McArthur, G.A. (2017) Checkpoint inhibitors in melanoma and early phase development in solid tumors: what's the future? *J. Transl. Med.* **15**, 173 https://doi.org/10.1186/s12967-017-1278-5
- 3 Turan, T., Kannan, D., Patel, M., Barnes, M.J., Tanlimco, S.G., Lu, R. et al. (2017) Immune oncology, immune responsiveness and the theory of everything. *J. Immunother. Cancer* In press
- 4 Rees, J. (2002) Complex disease and the new clinical sciences. Science 296, 698–700 https://doi.org/10.1126/science.296.5568.698
- 5 Salk, J. (1969) Immunological paradoxes: theoretical considerations in the rejection or retention of grafts, tumors, and normal tissue. *Ann. N.Y. Acad. Sci.* **164**, 365–380 https://doi.org/10.1111/j.1749-6632.1969.tb14051.x
- 6 Spranger, S., Bao, R. and Gajewski, T.F. (2015) Melanoma-intrinsic β-catenin signalling prevents anti-tumour immunity. *Nature* **523**, 231–235 https://doi.org/10.1038/nature14404
- Corrales, L., Matson, V., Flood, B., Spranger, S. and Gajewski, T.F. (2017) Innate immune signaling and regulation in cancer immunotherapy. *Cell Res.* 27, 96–108 https://doi.org/10.1038/cr.2016.149
- 8 Hendrickx, W., Simeone, I., Anjum, S., Mokrab, Y., Bertucci, F., Finetti, P. et al. (2017) Identification of genetic determinants of breast cancer immune phenotypes by integrative genome-scale analysis. *Oncoimmunology* **6**, e1253654 https://doi.org/10.1080/2162402X.2016.1253654
- 9 Galluzzi, L., Buqué, A., Kepp, O., Zitvogel, L. and Kroemer, G. (2017) Immunogenic cell death in cancer and infectious disease. *Nat. Rev. Immunol.* **17**, 97–111 https://doi.org/10.1038/nri.2016.107
- 10 Wegner, K.W., Saleh, D. and Degterev, A. (2017) Complex pathologic roles of RIPK1 and RIPK3: moving beyond necroptosis. *Trends Pharmacol. Sci.* **38**, 202–225 https://doi.org/10.1016/j.tips.2016.12.005
- 11 Matt, S. and Hofmann, T.G. (2016) The DNA damage-induced cell death response: a roadmap to kill cancer cells. *Cell Mol. Life Sci.* **73**, 2829–2850 https://doi.org/10.1007/s00018-016-2130-4



- 12 Abd Al Samid, M., Chaudhary, B., Khaled, Y.S., Ammori, B.J. and Elkord, E. (2016) Combining FoxP3 and Helios with GARP/LAP markers can identify expanded Treg subsets in cancer patients. *Oncotarget* 7, 14083–14094 https://doi.org/10.18632/oncotarget.7334
- 13 Elkord, E., Abd Al Samid, M. and Chaudhary, B. (2015) Helios, and not FoxP3, is the marker of activated Tregs expressing GARP/LAP. Oncotarget 6, 20026–20036 https://doi.org/10.18632/oncotarget.4771
- 14 Tosolini, M., Kirilovsky, A., Mlecnik, B., Fredriksen, T., Mauger, S., Bindea, G. et al. (2011) Clinical impact of different classes of infiltrating T cytotoxic and helper cells (Th1, th2, treg, th17) in patients with colorectal cancer. *Cancer Res.* 71, 1263–1271 https://doi.org/10.1158/0008-5472. CAN-10-2907
- 15 Coccia, M., Harrison, O.J., Schiering, C., Asquith, M.J., Becher, B., Powrie, F. et al. (2012) IL-1β mediates chronic intestinal inflammation by promoting the accumulation of IL-17A secreting innate lymphoid cells and CD4<sup>+</sup> Th17 cells. *J. Exp. Med.* **209**, 1595–1609 https://doi.org/10.1084/jem. 20111453
- 16 Kirchberger, S., Royston, D.J., Boulard, O., Thornton, E., Franchini, F., Szabady, R.L. et al. (2013) Innate lymphoid cells sustain colon cancer through production of interleukin-22 in a mouse model. J. Exp. Med. 210, 917–931 https://doi.org/10.1084/jem.20122308
- 17 Alinejad, V., Dolati, S., Motallebnezhad, M. and Yousefi, M. (2017) The role of IL17B-IL17RB signaling pathway in breast cancer. *Biomed. Pharmacother.* 88, 795–803 https://doi.org/10.1016/j.biopha.2017.01.120
- 18 Ngiow, S.F., Teng, M.W.L. and Smyth, M.J. (2013) A balance of interleukin-12 and -23 in cancer. *Trends Immunol.* **34**, 548–555 https://doi.org/10. 1016/j.it.2013.07.004
- 19 Munn, D.H. and Bronte, V. (2016) Immune suppressive mechanisms in the tumor microenvironment. *Curr. Opin. Immunol.* **39**, 1–6 https://doi.org/10. 1016/j.coi.2015.10.009
- 20 De Henau, O., Rausch, M., Winkler, D., Campesato, L.F., Liu, C., Cymerman, D.H. et al. (2016) Overcoming resistance to checkpoint blockade therapy by targeting Pl3Kγ in myeloid cells. *Nature* **539**, 443–447 https://doi.org/10.1038/nature20554
- 21 Kaneda, M.M., Messer, K.S., Ralainirina, N., Li, H., Leem, C.J., Gorjestani, S. et al. (2016) PI3Kγ is a molecular switch that controls immune suppression. *Nature* 539, 437–442 https://doi.org/10.1038/nature19834
- 22 Daragmeh, J., Barriah, W., Saad, B. and Zaid, H. (2016) Analysis of PI3K pathway components in human cancers. Oncol. Lett. 11, 2913–2918 PMID: 27073576
- 23 Liu, P., Cheng, H., Roberts, T.M. and Zhao, J.J. (2009) Targeting the phosphoinositide 3-kinase pathway in cancer. Nat. Rev. Drug Discov. 8, 627–644 https://doi.org/10.1038/nrd2926
- 24 Karlsson, E., Veenstra, C., Emin, S., Dutta, C., Pérez-Tenorio, G., Nordenskjöld, B. et al. (2015) Loss of protein tyrosine phosphatase, non-receptor type 2 is associated with activation of AKT and tamoxifen resistance in breast cancer. *Breast Cancer Res. Treat.* **153**, 31–40 https://doi.org/10.1007/ s10549-015-3516-y
- 25 Munn, D.H. and Mellor, A.L. (2016) IDO in the tumor microenvironment: inflammation, counter-regulation, and tolerance. *Trends Immunol.* **37**, 193–207 https://doi.org/10.1016/j.it.2016.01.002
- 26 Liu, Q., Tomei, S., Ascierto, M.L., De Giorgi, V., Bedognetti, D., Dai, C. et al. (2014) Melanoma NOS1 expression promotes dysfunctional IFN signaling. J. Clin. Invest. 124, 2147–2159 https://doi.org/10.1172/JCI69611
- 27 Mondanelli, G., Ugel, S., Grohmann, U. and Bronte, V. (2017) The immune regulation in cancer by the amino acid metabolizing enzymes ARG and IDO. *Curr. Opin. Pharmacol.* 35, 30–39 https://doi.org/10.1016/j.coph.2017.05.002
- 28 Di Cristofano, A. (2017) SGK1: the dark side of PI3K signaling. Curr. Top. Dev. Biol. 123, 49–71 https://doi.org/10.1016/bs.ctdb.2016.11.006
- 29 Xiaobo, Y., Qiang, L., Xiong, Q., Zheng, R., Jianhua, Z., Zhifeng, L. et al. (2016) Serum and glucocorticoid kinase 1 promoted the growth and migration of non-small cell lung cancer cells. *Gene* 576(1 Pt 2), 339–346 https://doi.org/10.1016/j.gene.2015.10.072
- 30 Ahn, R., Sabourin, V., Bolt, A.M., Hébert, S., Totten, S., De Jay, N. et al. (2017) The Shc1 adaptor simultaneously balances Stat1 and Stat3 activity to promote breast cancer immune suppression. *Nat. Commun.* 8, 14638 https://doi.org/10.1038/ncomms14638
- 31 Salerno, E.P., Bedognetti, D., Mauldin, I.S., Deacon, D.H., Shea, S.M., Pinczewski, J. et al. (2016) Human melanomas and ovarian cancers overexpressing mechanical barrier molecule genes lack immune signatures and have increased patient mortality risk. *Oncoimmunology* 5, e1240857 https://doi.org/10.1080/2162402X.2016.1240857
- 32 Buckanovich, R.J., Facciabene, A., Kim, S., Benencia, F., Sasaroli, D., Balint, K. et al. (2008) Endothelin B receptor mediates the endothelial barrier to T cell homing to tumors and disables immune therapy. *Nat. Med.* **14**, 28–36 https://doi.org/10.1038/nm1699
- 33 Hugo, W., Zaretsky, J.M., Sun, L., Song, C., Moreno, B.H., Hu-Lieskovan, S. et al. (2016) Genomic and transcriptomic features of response to anti-PD-1 therapy in metastatic melanoma. *Cell* **165**, 35–44 https://doi.org/10.1016/j.cell.2016.02.065
- 34 Shields, B.D., Mahmoud, F., Taylor, E.M., Byrum, S.D., Sengupta, D., Koss, B. et al. (2017) Indicators of responsiveness to immune checkpoint inhibitors. Sci. Rep. 7, 807 https://doi.org/10.1038/s41598-017-01000-2
- 35 Herbst, R.S., Soria, J.-C., Kowanetz, M., Fine, G.D., Hamid, O., Gordon, M.S. et al. (2014) Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature* **515**, 563–567 https://doi.org/10.1038/nature14011
- 36 Feig, C., Jones, J.O., Kraman, M., Wells, R.J.B., Deonarine, A., Chan, D.S. et al. (2013) Targeting CXCL12 from FAP-expressing carcinoma-associated fibroblasts synergizes with anti-PD-L1 immunotherapy in pancreatic cancer. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 20212–20217 https://doi.org/10.1073/ pnas.1320318110
- 37 Kraman, M., Bambrough, P.J., Arnold, J.N., Roberts, E.W., Magiera, L., Jones, J.O. et al. (2010) Suppression of antitumor immunity by stromal cells expressing fibroblast activation protein-α. Science **330**, 827–830 https://doi.org/10.1126/science.1195300
- 38 Öhlund, D., Elyada, E. and Tuveson, D. (2014) Fibroblast heterogeneity in the cancer wound. J. Exp. Med. 211, 1503–1523 https://doi.org/10.1084/ jem.20140692
- 39 Ohlund, D., Handly-Santana, A., Biffi, G., Elyada, E., Almeida, A.S., Ponz-Sarvise, M. et al. (2017) Distinct populations of inflammatory fibroblasts and myofibroblasts in pancreatic cancer. J. Exp. Med. 214, 579–596 PMID: 28232471
- 40 Özdemir, B.C., Pentcheva-Hoang, T., Carstens, J.L., Zheng, X., Wu, C.-C., Simpson, T.R. et al. (2014) Depletion of carcinoma-associated fibroblasts and fibrosis induces immunosuppression and accelerates pancreas cancer with reduced survival. *Cancer Cell* 25, 719–734 https://doi.org/10.1016/j.ccr.2014.04.005



- 41 Crittenden, M.R., Baird, J., Friedman, D., Savage, T., Uhde, L., Alice, A. et al. (2016) Mertk on tumor macrophages is a therapeutic target to prevent tumor recurrence following radiation therapy. *Oncotarget* 7, 78653–78666 PMID: 27602953
- 42 Akalu, Y.T., Rothlin, C.V. and Ghosh, S. (2017) TAM receptor tyrosine kinases as emerging targets of innate immune checkpoint blockade for cancer therapy. *Immunol. Rev.* 276, 165–177 https://doi.org/10.1111/imr.12522
- 43 Zhang, B., Fang, L., Wu, H.-M., Ding, P.-S., Xu, K. and Liu, R.-Y. (2016) Mer receptor tyrosine kinase negatively regulates lipoteichoic acid-induced inflammatory response via PI3K/Akt and SOCS3. *Mol. Immunol.* **76**, 98–107 https://doi.org/10.1016/j.molimm.2016.06.016
- 44 Grabiec, A.M. and Hussell, T. (2016) The role of airway macrophages in apoptotic cell clearance following acute and chronic lung inflammation. *Semin. Immunopathol.* **38**, 409–423 https://doi.org/10.1007/s00281-016-0555-3
- 45 Crittenden, M.R., Cottam, B., Savage, T., Nguyen, C., Newell, P. and Gough, M.J. (2012) Expression of NF-κB p50 in tumor stroma limits the control of tumors by radiation therapy. *PLoS ONE* **7**, e39295 https://doi.org/10.1371/journal.pone.0039295
- 46 Hatfield, S.M. and Sitkovsky, M. (2016) A2a adenosine receptor antagonists to weaken the hypoxia-HIF-1α driven immunosuppression and improve immunotherapies of cancer. *Curr. Opin. Pharmacol.* **29**, 90–96 https://doi.org/10.1016/j.coph.2016.06.009
- 47 Hu, C.-J., Wang, L.-Y., Chodosh, L.A., Keith, B. and Simon, M.C. (2003) Differential roles of hypoxia-inducible factor 1α (HIF-1α) and HIF-2α in hypoxic gene regulation. *Mol. Cell Biol.* **23**, 9361–9374 https://doi.org/10.1128/MCB.23.24.9361-9374.2003
- 48 Demaria, S., Golden, E.B., Formenti, S.C. (2015) Role of local radiation therapy in cancer immunotherapy. JAMA Oncol. 1, 1325–1332 https://doi.org/ 10.1001/jamaoncol.2015.2756
- 49 Vanpouille-Box, C., Diamond, J.M., Pilones, K.A., Zavadil, J., Babb, J.S., Formenti, S.C. et al. (2015) TGFbeta is a master regulator of radiation therapyinduced antitumor immunity. *Cancer Res.* **75**, 2232–2242 https://doi.org/10.1158/0008-5472.CAN-14-3511
- 50 Vanpouille-Box, C., Alard, A., Aryankalayil, M.J., Sarfraz, Y., Diamond, J.M., Schneider, R.J. et al. (2017) DNA exonuclease Trex1 regulates radiotherapy-induced tumour immunogenicity. *Nat. Commun.* **8**, 15618 https://doi.org/10.1038/ncomms15618
- 51 Benci, J.L., Xu, B., Qiu, Y., Wu, T.J., Dada, H., Twyman-Saint Victor, C. et al. (2016) Tumor interferon signaling regulates a multigenic resistance program to immune checkpoint blockade. *Cell* **167**, 1540–1554.e12 https://doi.org/10.1016/j.cell.2016.11.022
- 52 Koyama, S., Akbay, E.A., Li, Y.Y., Herter-Sprie, G.S., Buczkowski, K.A., Richards, W.G. et al. (2016) Adaptive resistance to therapeutic PD-1 blockade is associated with upregulation of alternative immune checkpoints. *Nat. Commun.* **7**, 10501 https://doi.org/10.1038/ncomms10501
- 53 Gainor, J.F., Shaw, A.T., Sequist, L.V., Fu, X., Azzoli, C.G., Piotrowska, Z. et al. (2016) EGFR mutations and ALK rearrangements are associated with low response rates to PD-1 pathway blockade in non-small cell lung cancer: a retrospective analysis. *Clin. Cancer Res.* **22**, 4585–4593 https://doi.org/10. 1158/1078-0432.CCR-15-3101
- 54 Akbay, E.A., Koyama, S., Carretero, J., Altabef, A., Tchaicha, J.H., Christensen, C.L. et al. (2013) Activation of the PD-1 pathway contributes to immune escape in EGFR-driven lung tumors. *Cancer Discov.* **3**, 1355–1363 https://doi.org/10.1158/2159-8290.CD-13-0310
- 55 Place, A.E., Jin Huh, S. and Polyak, K. (2011) The microenvironment in breast cancer progression: biology and implications for treatment. *Breast Cancer Res.* **13**, 227 https://doi.org/10.1186/bcr2912
- 56 Adeegbe, D.O., Liu, Y., Lizotte, P.H., Kamihara, Y., Aref, A.R., Almonte, C. et al. (2017) Synergistic immunostimulatory effects and therapeutic benefit of combined histone deacetylase and bromodomain inhibition in non-small cell lung cancer. *Cancer Discov.* **7**, 852–867 PMID:28408401
- 57 Guerriero, J.L., Sotayo, A., Ponichtera, H.E., Castrillon, J.A., Pourzia, A.L., Schad, S. et al. (2017) Class Ila HDAC inhibition reduces breast tumours and metastases through anti-tumour macrophages. *Nature* **543**, 428–432 https://doi.org/10.1038/nature21409
- 58 Mondino, A., Vella, G. and Icardi, L. (2017) Targeting the tumor and its associated stroma: One and one can make three in adoptive T cell therapy of solid tumors. *Cytokine Growth Factor Rev.* **36**, 57–65 PMID: 28629762
- 59 Saleh, S.M.I., Bertos, N., Gruosso, T., Gigoux, M., Souleimanova, M., Zhao, H. et al. (2017) Identification of interacting stromal axes in triple-negative breast cancer. *Cancer Res.* 77, 4673–4683 https://doi.org/10.1158/0008-5472.CAN-16-3427
- 60 Hopewell, E.L., Zhao, W., Fulp, W.J., Bronk, C.C., Lopez, A.S., Massengill, M. et al. (2013) Lung tumor NF-κB signaling promotes T cell-mediated immune surveillance. J. Clin. Invest. 123, 2509–2522 https://doi.org/10.1172/JCI67250
- 61 Miller, L.D., Chou, J.A., Black, M.A., Print, C., Chifman, J., Alistar, A. et al. (2016) Immunogenic subtypes of breast cancer delineated by gene classifiers of immune responsiveness. *Cancer Immunol. Res.* **4**, 600–610 https://doi.org/10.1158/2326-6066.CIR-15-0149
- 62 Wang, E., Worschech, A. and Marincola, F.M. (2008) The immunologic constant of rejection. *Trends Immunol.* **29**, 256–262 https://doi.org/10.1016/j.it. 2008.03.002
- 63 Spivey, T.L., Uccellini, L., Ascierto, M.L., Zoppoli, G., De Giorgi, V., Delogu, L.G. et al. (2011) Gene expression profiling in acute allograft rejection: challenging the immunologic constant of rejection hypothesis. *J. Transl. Med.* **9**, 174 https://doi.org/10.1186/1479-5876-9-174
- 64 Galon, J., Angell, H.K., Bedognetti, D. and Marincola, F.M. (2013) The continuum of cancer immunosurveillance: prognostic, predictive, and mechanistic signatures. *Immunity* **39**, 11–26 https://doi.org/10.1016/j.immuni.2013.07.008
- 65 Panelli, M.C., Stashower, M.E., Slade, H.B., Smith, K., Norwood, C., Abati, A. et al. (2007) Sequential gene profiling of basal cell carcinomas treated with imiquimod in a placebo-controlled study defines the requirements for tissue rejection. *Genome Biol.* **8**, R8 https://doi.org/10.1186/gb-2007-8-1-r8
- 66 Ayers, M., Lunceford, J., Nebozhyn, M., Murphy, E., Loboda, A., Kaufman, D.R. et al. (2017) IFN-γ-related mRNA profile predicts clinical response to PD-1 blockade. J. Clin. Invest. 127, 2930–2940 PMID: 28650338
- 67 Turneh, P.C., Harview, C.L., Yearley, J.H., Shintaku, I.P., Taylor, E.J.M., Robert, L. et al. (2014) PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* **515**, 568–571 https://doi.org/10.1038/nature13954
- 68 Wang, E., Miller, L.D., Ohnmacht, G.A., Mocellin, S., Perez-Diez, A., Petersen, D. et al. (2002) Prospective molecular profiling of melanoma metastases suggests classifiers of immune responsiveness. *Cancer Res.* **62**, 3581–3586 PMID: 12097256
- 69 Weiss, G.R., Grosh, W.W., Chianese-Bullock, K.A., Zhao, Y., Liu, H., Slingluff, Jr, C.L. et al. (2011) Molecular insights on the peripheral and intratumoral effects of systemic high-dose rlL-2 (aldesleukin) administration for the treatment of metastatic melanoma. *Clin. Cancer Res.* 17, 7440–7450 https://doi.org/10.1158/1078-0432.CCR-11-1650
- 70 Bedognetti, D., Spivey, T.L., Zhao, Y., Uccellini, L., Tomei, S., Dudley, M.E. et al. (2013) CXCR3/CCR5 pathways in metastatic melanoma patients treated with adoptive therapy and interleukin-2. Br. J. Cancer 109, 2412–2423 https://doi.org/10.1038/bjc.2013.557
- 71 Wang, E., Uccellini, L. and Marincola, F.M. (2012) A genetic inference on cancer immune responsiveness. *Oncoimmunology* **1**, 520–525 https://doi.org/ 10.4161/onci.19531



- 72 Spranger, S. and Gajewski, T.F. (2015) A new paradigm for tumor immune escape: β-catenin-driven immune exclusion. *J. Immunother. Cancer* **3**, 43 https://doi.org/10.1186/s40425-015-0089-6
- 73 Spranger, S., Sivan, A., Corrales, L. and Gajewski, T.F. (2016) Tumor and host factors controlling antitumor immunity and efficacy of cancer immunotherapy. Adv. Immunol. 130, 75–93 https://doi.org/10.1016/bs.ai.2015.12.003
- 74 Sweis, R.F., Spranger, S., Bao, R., Paner, G.P., Stadler, W.M., Steinberg, G. et al. (2016) Molecular drivers of the non-T-cell-inflamed tumor microenvironment in urothelial bladder cancer. *Cancer Immunol. Res.* 4, 563–568 https://doi.org/10.1158/2326-6066.CIR-15-0274
- 75 Galluzzi, L., Buqué, A., Kepp, O., Zitvogel, L. and Kroemer, G. (2015) Immunological effects of conventional chemotherapy and targeted anticancer agents. *Cancer Cell* 28, 690–714 https://doi.org/10.1016/j.ccell.2015.10.012
- 76 Galluzzi, L., Vacchelli, E., Bravo-San Pedro, J.M., Buqué, A., Senovilla, L., Baracco, E.E. et al. (2014) Classification of current anticancer immunotherapies. *Oncotarget* 5, 12472–12508 https://doi.org/10.18632/oncotarget.2998
- 77 Garg, A.D., Galluzzi, L., Apetoh, L., Baert, T., Birge, R.B., Bravo-San Pedro, J.M. et al. (2015) Molecular and translational classifications of DAMPs in immunogenic cell death. *Front. Immunol.* **6**, 588 PMID: 26635802
- 78 Hannani, D., Sistigu, A., Kepp, O., Galluzzi, L., Kroemer, G. and Zitvogel, L. (2011) Prerequisites for the antitumor vaccine-like effect of chemotherapy and radiotherapy. *Cancer J.* 17, 351–358 https://doi.org/10.1097/PP0.0b013e3182325d4d
- 79 Kepp, O., Galluzzi, L., Giordanetto, F., Tesniere, A., Vitale, I., Martins, I. et al. (2009) Disruption of the PP1/GADD34 complex induces calreticulin exposure. Cell Cycle 8, 3971–3977 https://doi.org/10.4161/cc.8.23.10191
- 80 Kepp, O., Galluzzi, L., Martins, I., Schlemmer, F., Adjemian, S., Michaud, M. et al. (2011) Molecular determinants of immunogenic cell death elicited by anticancer chemotherapy. *Cancer Metastasis Rev.* **30**, 61–69 https://doi.org/10.1007/s10555-011-9273-4
- 81 Kepp, O., Galluzzi, L., Zitvogel, L. and Kroemer, G. (2010) Pyroptosis a cell death modality of its kind? Eur. J. Immunol. 40, 627–630 https://doi.org/10.1002/eji.200940160
- 82 Kepp, O., Senovilla, L., Vitale, I., Vacchelli, E., Adjemian, S., Agostinis, P. et al. (2014) Consensus guidelines for the detection of immunogenic cell death. Oncoimmunology 3, e955691 https://doi.org/10.4161/21624011.2014.955691
- 83 Kroemer, G., Galluzzi, L., Kepp, O. and Zitvogel, L. (2013) Immunogenic cell death in cancer therapy. Annu. Rev. Immunol. 31, 51–72 https://doi.org/10.1146/annurev-immunol-032712-100008
- 84 Ma, Y., Adjemian, S., Mattarollo, S.R., Yamazaki, T., Aymeric, L., Yang, H. et al. (2013) Anticancer chemotherapy-induced intratumoral recruitment and differentiation of antigen-presenting cells. *Immunity* **38**, 729–741 https://doi.org/10.1016/j.immuni.2013.03.003
- 85 Ma, Y., Adjemian, S., Yang, H., Catani, J.P.P., Hannani, D., Martins, I. et al. (2013) ATP-dependent recruitment, survival and differentiation of dendritic cell precursors in the tumor bed after anticancer chemotherapy. *Oncoimmunology* 2, e24568 https://doi.org/10.4161/onci.24568
- 86 Ma, Y., Yamazaki, T., Yang, H., Kepp, O., Galluzzi, L., Zitvogel, L. et al. (2013) Tumor necrosis factor is dispensable for the success of immunogenic anticancer chemotherapy. *Oncoimmunology* **2**, e24786 https://doi.org/10.4161/onci.24786
- 87 Martins, I., Kepp, O., Galluzzi, L., Senovilla, L., Schlemmer, F., Adjemian, S. et al. (2010) Surface-exposed calreticulin in the interaction between dying cells and phagocytes. *Ann. N.Y. Acad. Sci.* **1209**, 77–82 https://doi.org/10.1111/j.1749-6632.2010.05740.x
- 88 Martins, I., Michaud, M., Sukkurwala, A.Q., Adjemian, S., Ma, Y., Shen, S. et al. (2012) Premortem autophagy determines the immunogenicity of chemotherapy-induced cancer cell death. Autophagy 8, 413–415 https://doi.org/10.4161/auto.19009
- 89 Martins, I., Wang, Y., Michaud, M., Ma, Y., Sukkurwala, A.Q., Shen, S. et al. (2014) Molecular mechanisms of ATP secretion during immunogenic cell death. *Cell Death Differ.* 21, 79–91 https://doi.org/10.1038/cdd.2013.75
- 90 Menger, L., Vacchelli, E., Adjemian, S., Martins, I., Ma, Y., Shen, S. et al. (2012) Cardiac glycosides exert anticancer effects by inducing immunogenic cell death. *Sci. Transl. Med.* 4, 143ra99 https://doi.org/10.1126/scitranslmed.3003807
- 91 Michaud, M., Martins, I., Sukkurwala, A.Q., Adjemian, S., Ma, Y., Pellegatti, P. et al. (2011) Autophagy-dependent anticancer immune responses induced by chemotherapeutic agents in mice. *Science* 334, 1573–1577 https://doi.org/10.1126/science.1208347
- 92 Sukkurvala, A.Q., Adjemian, S., Senovilla, L., Michaud, M., Spaggiari, S., Vacchelli, E. et al. (2014) Screening of novel immunogenic cell death inducers within the NCI mechanistic diversity set. *Oncoimmunology* 3, e28473 https://doi.org/10.4161/onci.28473
- 93 Sukkurwala, A.Q., Martins, I., Wang, Y., Schlemmer, F., Ruckenstuhl, C., Durchschlag, M. et al. (2014) Immunogenic calreticulin exposure occurs through a phylogenetically conserved stress pathway involving the chemokine CXCL8. *Cell Death Differ*, **21**, 59–68 https://doi.org/10.1038/cdd.2013.73
- 94 Zitvogel, L., Galluzzi, L., Kepp, O., Smyth, M.J. and Kroemer, G. (2015) Type I interferons in anticancer immunity. *Nat. Rev. Immunol.* **15**, 405–414 https://doi.org/10.1038/nri3845
- 95 Zitvogel, L., Kepp, O., Galluzzi, L. and Kroemer, G. (2012) Inflammasomes in carcinogenesis and anticancer immune responses. Nat. Immunol. 13, 343–351 https://doi.org/10.1038/ni.2224
- 96 Maykel, J., Liu, J.H., Li, H., Shultz, L.D., Greiner, D.L. and Houghton, J. (2014) NOD-scidll2rg tm1Wjl and NOD-Rag1 null ll2rg tm1Wjl: a model for stromal cell-tumor cell interaction for human colon cancer. *Dig. Dis. Sci.* 59, 1169–1179 https://doi.org/10.1007/s10620-014-3168-5
- 97 Schietinger, A., Philip, M. and Schreiber, H. (2008) Specificity in cancer immunotherapy. Semin. Immunol. 20, 276–285 https://doi.org/10.1016/j. smim.2008.07.001
- 98 Blankenstein, T., Leisegang, M., Uckert, W. and Schreiber, H. (2015) Targeting cancer-specific mutations by T cell receptor gene therapy. *Curr. Opin. Immunol.* **33**, 112–119 https://doi.org/10.1016/j.coi.2015.02.005
- 99 Janeway, Jr, C.A. (2013) Pillars article: approaching the asymptote? Evolution and revolution in immunology. J. Immunol. 191, 4475–4487 PMID: 24141854
- 100 Janeway, Jr, C.A. (1989) Approaching the asymptote? Evolution and revolution in immunology. *Cold Spring Harb. Symp. Quant. Biol.* **54** (Pt 1), 1–13 https://doi.org/10.1101/SQB.1989.054.01.003
- 101 Janeway, Jr, C.A. and Medzhitov, R. (1998) Introduction: the role of innate immunity in the adaptive immune response. *Semin. Immunol.* **10**, 349–350 https://doi.org/10.1006/smim.1998.0142
- 102 Medzhitov, R. and Janeway, Jr, C.A. (1997) Innate immunity: the virtues of a nonclonal system of recognition. *Cell* **91**, 295–298 https://doi.org/10. 1016/S0092-8674(00)80412-2
- 103 Medzhitov, R. and Janeway, Jr, C.A. (1997) Innate immunity: impact on the adaptive immune response. *Curr. Opin. Immunol.* **9**, 4–9 https://doi.org/10. 1016/S0952-7915(97)80152-5



- 104 Fuchs, E.J. and Matzinger, P. (1996) Is cancer dangerous to the immune system? Semin. Immunol. 8, 271–280 https://doi.org/10.1006/smim.1996. 0035
- 105 Fuchs, E.J. and Matzinger, P. (1992) B cells turn off virgin but not memory T cells. *Science* **258**, 1156–1159 https://doi.org/10.1126/science. 1439825
- 106 Fuchs, E.J., Ridge, J.P. and Matzinger, P. (1996) Response: immunological tolerance. *Science* 272, 1406–1408 https://doi.org/10.1126/science.272. 5267.1406
- 107 Matzinger, P. (2002) An innate sense of danger. Ann. N.Y. Acad. Sci. 961, 341-342 https://doi.org/10.1111/j.1749-6632.2002.tb03118.x
- 108 Matzinger, P. (1998) An innate sense of danger. Semin. Immunol. 10, 399-415 https://doi.org/10.1006/smim.1998.0143
- 109 Gubin, M.M., Zhang, X., Schuster, H., Caron, E., Ward, J.P., Noguchi, T. et al. (2014) Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature* **515**, 577–581 https://doi.org/10.1038/nature13988
- 110 Ward, J.P., Gubin, M.M. and Schreiber, R.D. (2016) The role of neoantigens in naturally occurring and therapeutically induced immune responses to cancer. *Adv. Immunol.* **130**, 25–74 https://doi.org/10.1016/bs.ai.2016.01.001
- 111 Spranger, S., Luke, J.J., Bao, R., Zha, Y., Hernandez, K.M., Li, Y. et al. (2016) Density of immunogenic antigens does not explain the presence or absence of the T-cell-inflamed tumor microenvironment in melanoma. *Proc. Natl. Acad. Sci. U.S.A.* **113**, E7759–E7E68 https://doi.org/10.1073/pnas. 1609376113
- 112 Cui, Z., Willingham, M.C., Hicks, A.M., Alexander-Miller, M.A., Howard, T.D., Hawkins, G.A. et al. (2003) Spontaneous regression of advanced cancer: identification of a unique genetically determined, age-dependent trait in mice. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 6682–6687 https://doi.org/10.1073/ pnas.1031601100
- 113 Hicks, A.M., Riedlinger, G., Willingham, M.C., Alexander-Miller, M.A., Von Kap-Herr, C., Pettenati, M.J. et al. (2006) Transferable anticancer innate immunity in spontaneous regression/complete resistance mice. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 7753–7758 https://doi.org/10.1073/pnas. 0602382103
- 114 Hicks, A.M., Willingham, M.C., Du, W., Pang, C.S., Old, L.J. and Cui, Z. (2006) Effector mechanisms of the anti-cancer immune responses of macrophages in SR/CR mice. *Cancer Immun.* 6, 11 PMID: 17073402
- 115 Riedlinger, G., Adams, J., Stehle, Jr, J.R., Blanks, M.J., Sanders, A.M., Hicks, A.M. et al. (2010) The spectrum of resistance in SR/CR mice: the critical role of chemoattraction in the cancer/leukocyte interaction. *BMC Cancer* **10**, 179 https://doi.org/10.1186/1471-2407-10-179
- 116 Worschech, A., Chen, N., Yu, Y.A., Zhang, Q., Pos, Z., Weibel, S. et al. (2009) Systemic treatment of xenografts with vaccinia virus GLV-1h68 reveals the immunologic facet of oncolytic therapy. *BMC Genomics* **10**, 301 https://doi.org/10.1186/1471-2164-10-301
- 117 Worschech, A., Haddad, D., Stroncek, D.F., Wang, E., Marincola, F.M. and Szalay, A.A. (2009) The immunologic aspects of poxvirus oncolytic therapy. *Cancer Immunol. Immunother.* 58, 1355–1362 https://doi.org/10.1007/s00262-009-0686-7
- 118 Lee, K.H., Panelli, M.C., Kim, C.J., Riker, A.I., Bettinotti, M.P., Roden, M.M. et al. (1998) Functional dissociation between local and systemic immune response during anti-melanoma peptide vaccination. J. Immunol. **161**, 4183–4194 PMID: 9780192
- 119 Lee, K.H., Wang, E., Nielsen, M.B., Wunderlich, J., Migueles, S., Connors, M. et al. (1999) Increased vaccine-specific T cell frequency after peptide-based vaccination correlates with increased susceptibility to in vitro stimulation but does not lead to tumor regression. *J. Immunol.* **163**, 6292–6300 PMID: 10570323
- 120 Pockaj, B.A., Sherry, R.M., Wei, J.P., Yannelli, J.R., Carter, C.S., Leitman, S.F. et al. (1994) Localization of 111indium-labeled tumor infiltrating lymphocytes to tumor in patients receiving adoptive immunotherapy. Augmentation with cyclophosphamide and correlation with response. *Cancer* **73**, 1731–1737 PMID: 8156501
- 121 Ribas, A., Dummer, R., Puzanov, I., VanderWalde, A., Andtbacka, R.H.I., Michielin, O. et al. (2017) Oncolytic virotherapy promotes intratumoral T cell infiltration and improves anti-PD-1 immunotherapy. *Cell* **170**, 1109–1119.e10 https://doi.org/10.1016/j.cell.2017.08.027