GENOME SEQUENCES





## Molecular Characterization of Southern African Territories 2 (SAT2) Serotype of Foot-and-Mouth Disease Virus from Nigeria in 2017 to 2018

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ABSTRACT This report describes the nucleotide sequences of eight Southern African Territories 2 (SAT2) serotype foot-and-mouth disease virus strains from 2017 to 2018 outbreaks in cattle in Nigeria. These viruses belong to topotype VII of SAT2 and were closely related to previous isolates from Nigeria and other West African countries.

Foot-and-mouth disease virus (FMDV), genus Aphthovirus, family Picornaviridae, causes a highly communicable disease of cloven-hoofed animals responsible for production losses and trade restrictions ([1,](#page-3-0) [2\)](#page-3-1). Seven serotypes (O, A, C, Asia 1, and South African Territories 1 [SAT1], 2, and 3) exist ([3\)](#page-3-2). Serotypes O, A, and SAT1, -2, and -3 have been reported in Nigeria [\(4](#page-3-3)[–](#page-3-4)[6\)](#page-3-5). Due to constant evolution of FMDV and porous borders in West Africa, monitoring of circulating FMDV is required in order to facilitate vaccine selection.

Here, epithelial tissues in BD universal transport system vials (VWR, Canada) were collected from FMD outbreaks in Plateau State (PL) and Bauchi State (BAU) in Nigeria in 2017 to 2018 and sent to the National Centre for Foreign Animal Disease (NCFAD), Canada. Tissue homogenates (10%) were prepared in a Precellys tissue grinding kit (ESBE Scientific, Canada) and clarified ([7](#page-3-6)). RNA was extracted using the MagMax viral RNA isolation kit (Life Technologies, Canada) ([8\)](#page-3-7) and tested for FMDV using real-time reverse transcription-PCR (rRT-PCR) as described previously [\(9](#page-3-8)). Near full-genome sequences of FMDV [\(Table 1\)](#page-0-0) were obtained

<span id="page-0-0"></span>TABLE 1 Accession numbers and identification of the FMDV SAT2 viruses from Nigeria in 2017 to 2018



<sup>*a*</sup> nt, nucleotides.

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<span id="page-1-0"></span>FIG 1 The evolutionary history was inferred by using the maximum likelihood method and the Hasegawa-Kishino-Yano model. The tree with the highest log likelihood (–9305.89) is shown. The percentage of trees in which the associated taxa clustered together is shown next (Continued on next page)

from rRT-PCR-positive samples using next-generation sequencing (NGS) ([10](#page-3-9)). RNA for NGS was processed as described previously ([11\)](#page-3-10), including DNase treatment and RNA purification prior to first-strand cDNA synthesis. Illumina Nextera XT sequencing libraries were prepared according to the manufacturer's instructions and sequenced on a MiSeq instrument using a V3 cycling kit (Illumina). Raw sequencing data were processed with the nf-villumina Nextflow ([12](#page-3-11)) workflow ([https://github](https://github.com/CFIA-NCFAD/nf-villumina) [.com/CFIA-NCFAD/nf-villumina](https://github.com/CFIA-NCFAD/nf-villumina)) for quality control, taxonomic classification, and de novo assembly ([13](#page-3-12)). De novo assembly was performed with Unicycler v0.4.7 [\(14](#page-3-13)) in "conservative" mode to find an optimal SPAdes assembly ([15](#page-3-14)) for each set of paired-end Illumina reads. Contigs were identified as FMDV using nucleotide BLAST ([16](#page-3-15), [17](#page-3-16)) search against the NCBI nucleotide database. The gene segments, including VP1, were identified with Annotate in Geneious v9.1.8 using a publicly available annotated FMDV genome. Phylogenetic analysis of obtained and previously published VP1 sequences in GenBank was performed using the Molecular Evolutionary Genetics Analysis (MEGA X) software ([18](#page-3-17)), with the evolutionary history inferred using the maximum likelihood method and Hasegawa-Kishino-Yano model. A discrete gamma distribution was used to model evolutionary rate differences among sites (5 categories [+G, parameter = 0.3453]). A bootstrap value of  $\geq$ 70% was considered significant.

The VP1 sequences in this study clustered within topotype VII of SAT2 and were closely related to 2018 isolates from Ghana. The 2017 PL isolates clustered together ([Fig. 1\)](#page-1-0). The 2018 BAU isolate was separated from the PL isolates and was closely related to isolates from Nigeria in 2014 and Cameroon in 2015.

SAT2, previously restricted to southern Africa, has become established in other sub-Saharan African countries, with sporadic spread into North Africa and the Middle East. Topotype VII is the predominant SAT2 outside southern Africa, and our data confirm its presence in Nigeria in 2017 to 2018, in agreement with recent reports of SAT2 in Nigeria, Cameroon, and other West African countries from 2013 to 2018 [\(5](#page-3-4), [19](#page-3-18)). Animal movement, especially for trade in Nigeria from neighboring Cameroon, Chad, and Sudan [\(6,](#page-3-5) [20](#page-3-19)), facilitates FMDV spread ([21\)](#page-3-20). Therefore, the FMD situation in Nigeria and neighboring countries is dynamic, and a regional control strategy remains vital.

**Data availability.** The sequences in this report and the associated raw data have been deposited in GenBank and the NCBI SRA under the accession numbers and identifiers shown in [Table 1](#page-0-0).

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## **FIG 1** Legend (Continued)

to the branches. Initial trees for the heuristic search were obtained automatically by applying the neighbor-join and BioNJ algorithms to a matrix of pairwise distances estimated using the maximum composite likelihood (MCL) approach and then selecting the topology with the superior log likelihood value. A discrete gamma distribution was used to model evolutionary rate differences among sites (5 categories [+G, parameter = 0.3453]). This analysis involved 94 nucleotide sequences. All positions containing gaps and missing data were eliminated (complete deletion option). There was a total of 635 positions in the final data set. Evolutionary analyses were conducted in MEGA X. The Nigerian isolates in this study are represented by dark stars. The original tree was exported from MEGA X as a Newick tree into ITOL v5 [\(https://itol.embl.de\)](https://itol.embl.de) for tree display and annotation. A bootstrap value of  $\geq$ 70% was considered significant. The Roman numerals i to xiv represent the 14 known topotypes of FMDV SAT2.

## **REFERENCES**

- <span id="page-3-0"></span>1. Grubman MJ, Baxt B. 2004. Foot-and-mouth disease. Clin Microbiol Rev 17:465–493. [https://doi.org/10.1128/CMR.17.2.465-493.2004.](https://doi.org/10.1128/CMR.17.2.465-493.2004)
- <span id="page-3-1"></span>2. Knight-Jones TJ, Rushton J. 2013. The economic impacts of foot and mouth disease: what are they, how big are they and where do they occur? Prev Vet Med 112:161–173. [https://doi.org/10.1016/j.prevetmed.2013.07.013.](https://doi.org/10.1016/j.prevetmed.2013.07.013)
- <span id="page-3-2"></span>3. Knowles NJ, Samuel AR. 2003. Molecular epidemiology of foot-and-mouth disease virus. Virus Res 91:65–80. [https://doi.org/10.1016/S0168-1702\(02\)00260-5.](https://doi.org/10.1016/S0168-1702(02)00260-5)
- <span id="page-3-3"></span>4. Wungak YS, Ishola OO, Olugasa BO, Lazarus DD, Ehizibolo DO, Ularamu HG. 2017. Spatial pattern of foot-and-mouth disease virus serotypes in North Central Nigeria. Vet World 10:450–456. [https://doi.org/10.14202/vetworld.2017](https://doi.org/10.14202/vetworld.2017.450-456) [.450-456.](https://doi.org/10.14202/vetworld.2017.450-456)
- <span id="page-3-4"></span>5. Ularamu HG, Lefebvre DJ, Haegeman A, Wungak YS, Ehizibolo DO, Lazarus DD, De Vleeschauwer AR, De Clercq K. 2020. Complex circulation of foot-and-mouth disease virus in cattle in Nigeria. Front Vet Sci 7:466. <https://doi.org/10.3389/fvets.2020.00466>.
- <span id="page-3-5"></span>6. Ehizibolo DO, Fish IH, Brito B, Bertram MR, Ardo A, Ularamu HG, Lazarus DD, Wungak YS, Nwosuh CI, Smoliga GR, Hartwig EJ, Pauszek SJ, Dickmu S, Abdoulkadiri S, Arzt J. 2020. Characterization of transboundary footand-mouth disease viruses in Nigeria and Cameroon during 2016. Transbound Emerg Dis 67:1257–1270. [https://doi.org/10.1111/tbed.13461.](https://doi.org/10.1111/tbed.13461)
- <span id="page-3-6"></span>7. Kittelberger R, Nfon C, Swekla K, Zhang Z, Hole K, Bittner H, Salo T, Goolia M, Embury-Hyatt C, Bueno R, Hannah M, Swainsbury R, O'Sullivan C, Spence R, Clough R, McFadden A, Rawdon T, Alexandersen S. 2017. Foot-and-mouth disease in red deer: experimental infection and test methods performance. Transbound Emerg Dis 64:213–225. [https://doi.org/10.1111/tbed.12363.](https://doi.org/10.1111/tbed.12363)
- <span id="page-3-7"></span>8. Moniwa M, Embury-Hyatt C, Zhang Z, Hole K, Clavijo A, Copps J, Alexandersen S. 2012. Experimental foot-and-mouth disease virus infection in white tailed deer. J Comp Pathol 147:330–342. <https://doi.org/10.1016/j.jcpa.2012.01.010>.
- <span id="page-3-8"></span>9. Senthilkumaran C, Yang M, Bittner H, Ambagala A, Lung O, Zimmerman J, Giménez-Lirola LG, Nfon C. 2017. Detection of genome, antigen, and antibodies in oral fluids from pigs infected with foot-and-mouth disease virus. Can J Vet Res 81:82–90.
- <span id="page-3-9"></span>10. Hicks HM, Wadsworth J, Azhar M, Afzal M, Manzoor S, Abubakar M, Khan E-u-H, King DP, Knowles NJ. 2020. Genome sequences of foot-and-mouth disease virus O/ME-SA/Ind-2001e strains isolated in Pakistan. Microbiol Resour Announc 9:e00165-20. <https://doi.org/10.1128/MRA.00165-20>.
- <span id="page-3-10"></span>11. Logan G, Freimanis GL, King DJ, Valdazo-González B, Bachanek-Bankowska K, Sanderson ND, Knowles NJ, King DP, Cottam EM. 2014. A universal protocol to generate consensus level genome sequences for foot-and-mouth disease virus and other positive-sense polyadenylated RNA viruses using the Illumina MiSeq. BMC Genomics 15:828. [https://doi.org/10.1186/1471-2164-15-828.](https://doi.org/10.1186/1471-2164-15-828)
- <span id="page-3-11"></span>12. Di Tommaso P, Chatzou M, Floden EW, Barja PP, Palumbo E, Notredame C. 2017. Nextflow enables reproducible computational workflows. Nat Biotechnol 35:316–319. <https://doi.org/10.1038/nbt.3820>.
- <span id="page-3-12"></span>13. Fisher M, Harrison TMR, Nebroski M, Kruczkiewicz P, Rothenburger JL, Ambagala A, Macbeth B, Lung O. 2020. Discovery and comparative genomic analysis of elk circovirus (ElkCV), a novel circovirus species and the first reported from a cervid host. Sci Rep 10:19548. [https://doi.org/10.1038/s41598](https://doi.org/10.1038/s41598-020-75577-6) [-020-75577-6](https://doi.org/10.1038/s41598-020-75577-6).
- <span id="page-3-13"></span>14. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. [https://doi.org/10.1371/journal.pcbi.1005595.](https://doi.org/10.1371/journal.pcbi.1005595)
- <span id="page-3-14"></span>15. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- <span id="page-3-15"></span>16. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. [https://doi.org/10.1186/1471-2105-10-421.](https://doi.org/10.1186/1471-2105-10-421)
- <span id="page-3-16"></span>17. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. [https://doi.org/10.1016/](https://doi.org/10.1016/S0022-2836(05)80360-2) [S0022-2836\(05\)80360-2.](https://doi.org/10.1016/S0022-2836(05)80360-2)
- <span id="page-3-17"></span>18. Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. Mol Biol Evol 35:1547–1549. <https://doi.org/10.1093/molbev/msy096>.
- <span id="page-3-18"></span>19. Bertram MR, Bravo de Rueda C, Garabed R, Dickmu Jumbo S, Moritz M, Pauszek S, Abdoulkadiri S, Rodriguez LL, Arzt J. 2018. Molecular epidemiology of foot-and-mouth disease virus in the context of transboundary animal movement in the far north region of Cameroon. Front Vet Sci 5:320. [https://doi.org/10.3389/fvets.2018.00320.](https://doi.org/10.3389/fvets.2018.00320)
- <span id="page-3-19"></span>20. Ehizibolo DO, De Vleeschauwer AR, Haegeman A, Lefebvre D, Nwosuh CI, Umoh JU, Okolocha EC, Kazeem HM, Van Borm S, De Clercq K. 2019. Serological and molecular epidemiology of foot-and-mouth disease viruses in agropastoralist livestock herds in the Kachia Grazing Reserve, Nigeria. Transbound Emerg Dis 66:1575–1586. [https://doi.org/10.1111/tbed.13182.](https://doi.org/10.1111/tbed.13182)
- <span id="page-3-20"></span>21. Habiela M, Ferris NP, Hutchings GH, Wadsworth J, Reid SM, Madi M, Ebert K, Sumption KJ, Knowles NJ, King DP, Paton DJ. 2010. Molecular characterization of foot-and-mouth disease viruses collected from Sudan. Transbound Emerg Dis 57:305–314. <https://doi.org/10.1111/j.1865-1682.2010.01151.x>.