

Lymphocutaneous spread of *Mycobacterium elephantis* in an immunocompetent individual: A case report

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Abstract

Mycobacterium elephantis was first described when isolated from an elephant that succumbed to lung abscess. However, despite this namesake, it is not associated with animals and has been described most often as a probable colonizer rather than pathogen in humans with chronic lung disease. In this report, we describe the first case of lymphocutaneous infection from *M. elephantis*, likely as a result of cutaneous inoculation with contaminated soil. This offers further evidence to its capabilities as a pathogen. We provide a review of the limited prior reports of *M. elephantis* and outline the available in vitro data on efficacy of various antimycobacterial agents.

Keywords

Lymphocutaneous, mycobacteria, atypical *Mycobacterium*, *Mycobacterium elephantis*

Introduction

Lymphocutaneous spread of infection may be caused by a broad variety of bacterial and fungal organisms, with nontuberculous mycobacteria (NTM) species representing particularly characteristic pathogens. While *Mycobacterium marinum* is classically described as causing this syndrome, it may be caused by a variety of atypical mycobacterial species and in particular, rapid-growing mycobacteria (RGM).

Mycobacterium elephantis is infrequently described in the literature and when referenced is often suspected as a colonizer of respiratory secretions in patients with pre-existing pulmonary disease. Its pathogenicity is debated, and it has never previously been described as a cutaneous pathogen. Here, we provide the first description of lymphocutaneous spread of *M. elephantis* and highlight the available data on pharmacological management. Similar to other atypical mycobacteria, it is likely best managed with a combination of antimycobacterial agents. Consent was provided from the patient to publish this report.

Case report

A 66-year-old immunocompetent female was seen in the dermatology clinic for ulcerations of her right hand, ascending

her arm in a lymphocutaneous distribution. She denied trauma preceding the lesions. There was no exposure to animals aside from a domesticated dog and cat. Her only travel was to Arizona 6 months before. She had exposure to floodwater in her home 5 months before. She was an avid gardener, working with soil on a daily basis. One month prior to symptom onset, she had received a potted plant (*Haworthia fasciata*) imported from the Eastern Cape in South Africa.

Clinically, she had punched-out ulcerations with yellow-white crust and minimal surrounding erythema (Figure 1).

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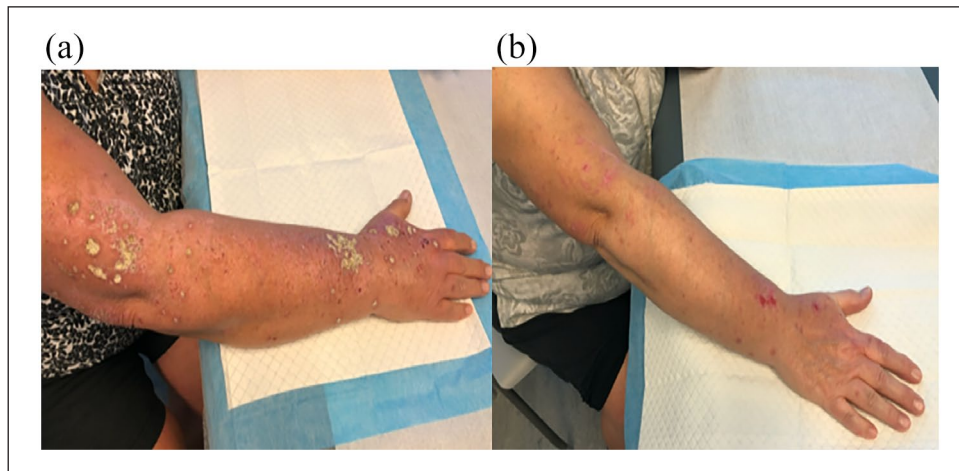


Figure 1. Lymphocutaneous infection from *Mycobacterium elephantis* (a) prior to treatment initiation and (b) 3 months into treatment with combination of azithromycin and moxifloxacin.

Histology of skin biopsy demonstrated a dense papillary dermal lymphohistiocytic infiltrate without well-formed granulomata. Gram, Fite, and Ziehl-Neelsen staining were negative. Aerobic culture was positive for *Candida parapsilosis*. She was treated with cephalexin and fluconazole without response. Mycobacterial culture was positive on BACTEC MGIT for *M. elephantis* at 23 days of incubation (acid-fast bacilli smear negative). Drug susceptibility testing results using microbroth dilution are outlined in Table 1. Additional scattered lesions on her legs were negative for acid-fast bacilli smear and mycobacterial culture. She was initiated on azithromycin and moxifloxacin with improvement after 3 months of treatment.

Discussion

M. elephantis was first described 20 years ago in an elephant that succumbed to chronic lung abscess.¹ The species is phenotypically similar to *Mycobacterium flavescens* though genotypically more closely related to *Mycobacterium pulveris*.^{1,2} *M. elephantis* is recognized as a rapid- or intermediate-growing *Mycobacterium* (growth detected at less than 7 days (rapid) or 10 days (intermediate)), although it appears to grow at a slower rate than other *rapid-growers* (RGM).^{2,3} RGM are frequently associated with cutaneous disease,⁴ but this is the first described case of *M. elephantis* infection in the skin.

Similar to other environmental mycobacteria, the pathogenicity of *M. elephantis* is debated. In the largest report, *M. elephantis* was described in 11 patients in Ontario, Canada (10 sputum, 1 lymph node excision).² Most were elderly patients with prior mycobacterial disease, and all sputum samples were smear negative with single-positive cultures, questioning the clinical significance.² Due to its relative scarcity of description, *M. elephantis* has not been described in international guidelines for diagnosis and management of

mycobacteria.^{4,5} Nonetheless, invasive disease has been confirmed in animals¹ and in humans.^{2,6}

Contact with animals is not a recognized risk factor. *M. elephantis* has environmental reservoirs including natural and man-made water sources and soil.^{7,8} Only one patient report described immunocompromise.⁶ Similarly, our case had no recognized predisposing factors but suggests that *M. elephantis* may extort breaks in cutaneous defenses. It is unknown if *M. elephantis* occupies a particular ecological niche to localize where infection was acquired. The most likely mechanism of acquisition was cryptogenic penetrating trauma to her extremity, contaminated by soil colonized with *M. elephantis*. Exposure to the plant/soil imported from South Africa is an interesting theory but could not be proven.

Lymphocutaneous spread of infection ascends the lymphatic pathway with intermittent subcutaneous nodules and ulcerations and is often termed a *sporotrichoid* pattern, named after *Sporothrix schenckii*. The infectious differential diagnosis includes *M. marinum* and RGM such as *Mycobacterium chelonae* and *Mycobacterium fortuitum*, but *M. elephantis* has not been reported.

Susceptibilities of *M. elephantis* are not established. Similar to other RGM, minimum inhibitory concentrations (MICs) to antituberculosis medications are usually elevated^{2,3,9,10} (Table 1). MICs to macrolides and fluoroquinolones are typically low.^{2,3,9} Due to the recognized potential of *Mycobacterium* to develop mutational macrolide resistance, we suggest a prudent approach is to utilize at least a two-drug therapy for several weeks beyond resolution of the lesions.

Herein, we report the first case of *M. elephantis* presenting as a cutaneous pathogen in a lymphocutaneous pattern. Although the pathogenicity of this organism in pulmonary samples is debated, our case adds further support to its potential pathogenicity in immunocompetent human hosts.

Table 1. Reported clinical isolation and drug susceptibility of *Mycobacterium elephantis* infection.

Study Country	Host	Type of infection	Clinically relevant infection	Treatment reported	Drug susceptibility reported (MIC if available)
Shojaei et al. ¹ Sri Lanka	Elephant	Lung abscess	Yes (fatal)	No	Isoniazid 1.4 µg/mL Rifampicin 16 µg/mL Pyrazinamide 66 µg/mL Ciprofloxacin >2.5 µg/mL Ethambutol >3.2 µg/mL
Turenne et al. ² Canada	11 Humans	Pulmonary (10) Lymph node (axillary), following recent tattoo (1)	Unclear Yes	No No	^a Amikacin ≤0.5 µg/mL Clarithromycin ≤1.25 µg/mL Ciprofloxacin ≤1.0 µg/mL Ethambutol ≤1.0 µg/mL Ethionamide 5.0 µg/mL Isoniazid 0.1 µg/mL Levofloxacin ≤0.5 µg/mL Rifabutin >2.0 µg/mL Rifampin >8.0 µg/mL Streptomycin ≤0.5 µg/mL
Tortoli et al. ³ (from strains isolated between 1995 and 2000) Italy	3 Humans	Sputum (2) BAL (1)	Possible, though did not meet American Thoracic Society/ Infectious Diseases Society of America (ATS/IDSA) criteria for clinical disease	No	Amikacin ≤2 µg/mL Ciprofloxacin ≤1 µg/mL Clarithromycin ≥32 µg/mL Ethambutol ≤2(2), 8(1) µg/mL Rifampin >8 µg/mL Streptomycin ≤2 µg/mL
Potters et al. ¹⁰ (from strain isolated in 1999) Belgium	1 Human	Sputum	Possible; presented with enteritis and respiratory tract infection	No	Reported “susceptible” to: Isoniazid 1 µg/mL Streptomycin 2 µg/mL Kanamycin 5 µg/mL Capreomycin 10 µg/mL Cycloserine 30 µg/mL Clarithromycin 4 µg/mL

(Continued)

Table 1. (Continued)

Study Country	Host	Type of infection	Clinically relevant infection	Treatment reported	Drug susceptibility reported (MIC if available)
Heidarieh et al. ⁹ Iran	1 Human	BAL (1)	Yes; febrile chronic respiratory illness with pulmonary infiltrate	Yes; amikacin + ciprofloxacin × 2 months	Reported “susceptible” to: Amikacin, clarithromycin, ciprofloxacin, ethambutol, isoniazid, doxycycline, streptomycin, ceftazidime, sulfamethoxazole, imipenem Reported “resistant” to: Rifampicin
Chin'ombe et al. ⁸ Zimbabwe	2 Humans 1 Cow	Sputum (2) Stool (1)	Unclear	No	
Ulusoy et al. ⁶ Turkey	1 Human (with interferon gamma receptor defect)	Lymph node	Yes; immunocompromised	No	
Present Case Canada	1 Human	Lymphocutaneous	Yes	Yes; azithromycin + moxifloxacin	“Susceptible” Amikacin < 1 µg/mL Ciprofloxacin 1 µg/mL Moxifloxacin ≤ 0.25 µg/mL Imipenem 4 µg/mL Clarithromycin 1 µg/mL Linezolid 4 µg/mL “Intermediate” Cefoxitin 32 µg/mL Doxycycline 2 µg/mL “Resistant” Trimethoprim-sulfamethoxazole 8 µg/mL

MIC: minimum inhibitory concentration; BAL: bronchoalveolar lavage.

^aMICs from lymph node sample.

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Informed consent

Informed patient consent was provided for the publication of this report.

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References

1. Shojaei H, Magee JG, Freeman R, et al. *Mycobacterium elephantis* sp. nov., a rapidly growing non-chromogenic *Mycobacterium* isolated from an elephant. *Int J Syst Evol Microbiol* 2000; 50(Pt 5): 1817–1820.
2. Turenne C, Chedore P, Wolfe J, et al. Phenotypic and molecular characterization of clinical isolates of *Mycobacterium elephantis* from human specimens. *J Clin Microbiol* 2002; 40(4): 1230–1236.
3. Tortoli E, Rindi L, Bartoloni A, et al. *Mycobacterium elephantis*: not an exceptional finding in clinical specimens. *Eur J Clin Microbiol Infect Dis* 2003; 22(7): 427–430.
4. Griffith DE, Aksamit T, Brown-Elliott BA, et al. An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am J Respir Crit Care Med* 2007; 175(4): 367–416.
5. Daley CL, Iaccarino JM, Lange C, et al. Treatment of nontuberculous mycobacterial pulmonary disease: an official ATS/ERS/ESCMID/IDSA clinical practice guideline: executive summary. *Clin Infect Dis* 2020; 71(4): e1–e36.
6. Ulusoy E, Edeer Karaca N, Aksu G, et al. Frequency of *Mycobacterium bovis* and mycobacteria in primary immunodeficiencies. *Turk Pediatri Ars* 2017; 52(3): 138–144.
7. Lande L. Environmental niches for NTM and their impact on NTM disease. In: Griffith DE (ed.) *Nontuberculous mycobacterial disease: a comprehensive approach to diagnosis and management*. Cham: Springer Nature; 2019, pp. 131–144.
8. Chin'ombe N, Munemo E, Magwenzi M, et al. First cases of *Mycobacterium elephantis* in Zimbabwe revealed by 16s ribosequencing. *Arch Clin Microbiol* 2015; 6(4): 10–13.
9. Heidarieh P, Shojaei H, Hashemi A, et al. First report of isolation of *Mycobacterium elephantis* from bronchial lavage of a patient in Asia. *JRSM Short Rep* 2011; 2(4): 1–3.
10. Potters D, Seghers M, Muyltermans G, et al. Recovery of *Mycobacterium elephantis* from sputum of a patient in Belgium. *J Clin Microbiol* 2003; 41(3): 1344.