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Clinicopathological significance of *c-KIT* mutation in gastrointestinal stromal tumors: a systematic review and meta-analysis

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Lin Yan^{1,*}, Lei Zou^{2,*}, Wenhua Zhao³, Yansen Wang¹, Bo Liu⁴, Hongliang Yao⁵ & Haihua Yu⁴

Many types of *KIT* mutations have been observed in gastrointestinal stromal tumors (GISTs), but their prognostic and predictive significance are still unclear. A meta-analysis and literature review were conducted to estimate the contribution of *KIT* mutations in prognostic parameters and clinicopathological significance of GISTs. A total of 18 relevant articles from PubMed, EMBASE and Web of Science databases were included in this study. The frequency of *KIT* mutation was significantly increased in the GIST patients with higher mitosis ($\geq 5/50$ high-power fields (HPFs) and larger size (≥ 5 cm) of tumors than in those with lower MI ($\leq 5/50$ HPFs) and smaller size (≤ 5 cm) of GISTs respectively. The rate of *KIT* mutation was not significantly changed between GISTs in stomachs and in small intestines. *KIT* mutational status has prognostic significance for patients' outcome. GIST patients with *KIT* exon 9 mutations have higher risk of progression than those with exon 11 mutations. 5 year relapse-free survival (RFS) rate was significantly higher in patients with *KIT* exon 11 deletion than in those with other type of *KIT* exon 11 mutations. The deletion involving *KIT* exon 11, particularly codons 557–558, is a valuable predictor of prognosis for patients with GISTs.

Gastrointestinal stromal tumors (GISTs) are the most common mesenchymal tumors in the gastrointestinal (GI) tract. The population-based studies showed that the annual incidence of GISTs ranges from 11 to 19.6 per million population^{1,2}. It has been a considerable debate regarding their cellular origin and diagnosis³. After gain-of-function mutations in the *c-KIT* protooncogene were discovered in 1998, GISTs were reliably distinguished from other histopathological subtypes of GI mesenchymal tumors⁴. GISTs occur primarily in older patients of either sex⁵, which are present anywhere along the GI tract from esophagus to the rectum, most commonly located in stomach (60%) and small intestine (25%)⁵. Local recurrence and metastasis are frequently observed in patients with GISTs after adequate resection and adjuvant therapy with tyrosine kinase inhibitor (TKIs)⁶. In addition, metastasis to the lung and bones occurs in patients with advanced disease⁶. Therefore, it is critical to identify risk criteria to predict their recurrence and metastasis.

c-KIT, the cellular homologue of the oncogene *v-KIT*, was isolated from feline fibrosarcoma, the Hardy-Zuckerman 4 feline sarcoma virus (HZ4-FeSV). The viral genome of HZ4-FeSV contains a new oncogene that was designated *v-KIT*, which encodes a transmembrane tyrosine kinase receptor called KIT⁷. Huizinga *et al.* revealed that mice with mutations in the *KIT* gene lacked the network of interstitial

¹Department of oncology, Shandong Jiaotong Hospital, Jinan, 250031, P.R.China. ²Department of gastrointestinal surgery, Shandong Cancer Hospital, Jinan, 250017, P.R.China. ³Department of oncology, Shandong Provincial Qianfoshan Hospital, Shandong University, Jinan, 250014, P.R.China. ⁴Department of gastrointestinal surgery, Shandong Provincial Qianfoshan Hospital, Shandong University, Jinan, 250014, P.R.China. ⁵Department of General Surgery, Second Xiangya Hospital of Central South University, Changsha 410011, China. ^{*}These authors contributed equally to this work. Correspondence and requests for materials should be addressed to H.Y. (email: haihuayu66@163.com)

***KIT* and *PDGFRA* Mutations in GIST**

'Wild-type' tumors: 15%

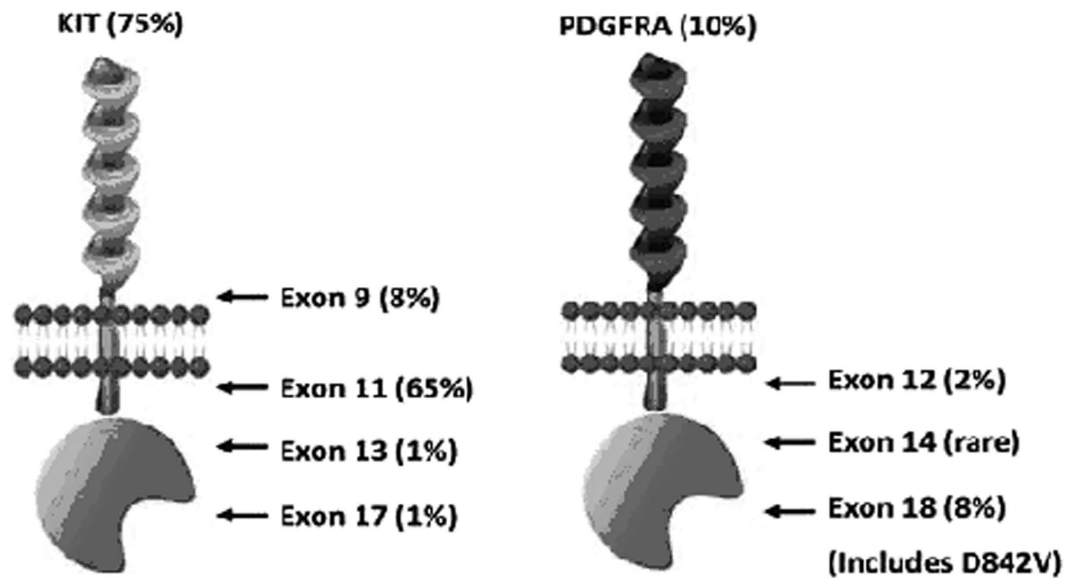


Figure 1. *KIT* and *PDGFRA* mutation in GIST.

cells of Cajal which was associated with Auerbach's nerve plexus and intestinal pacemaker activity, indicating that the interstitial cells of Cajal express the *KIT* receptor⁸. Mutations of the *KIT* gene in GISTs occur most frequently in *KIT* exon 11 (juxtamembrane domain), followed by *KIT* exon 9 (extracellular domain), less frequently, mutations occur in the adenosine triphosphate (ATP)-binding pocket (exon 13) or activation loop (exon 17) (Fig. 1)^{5,9}. Many types of *KIT* mutations have been observed in GISTs, but controversy still exists concerning their prognostic and predictive value¹⁰. Deletions in the *KIT* exon 11 most frequently involve the 5' portion between codons 550 and 560¹¹. A few studies have shown that tumors containing deletion in this area are clinically more aggressive than tumors with other type of mutations. However, several studies have reported inconsistent results^{6,12-14}. The aim of this study is to estimate the contribution of different types of *KIT* mutations in prognostic parameters and clinic-pathological significance of GISTs.

Methods

Search strategy and selection criteria. We conducted comprehensive literature searches in the PubMed, EMBASE and Web of Science databases in September 2014 with no low limit set for date of publication, using the following keywords: *c-KIT* or *KIT* and GIST or gastrointestinal stromal tumor. The language was limited to English and Chinese. A total of 1206 articles were identified with the initial search. Inclusion criteria for study selection were: 1) The articles in which the association between *c-KIT* mutation and the clinicopathological significance of GIST was evaluated; 2) The articles in that the association between *c-KIT* mutations and prognosis in patients with GIST was evaluated. Exclusion criteria were: 1) The studies which used the same population or overlapping database; 2) The studies of *in vitro* cell culture models; 3) The studies which showed insufficient data to calculate Odds Ratio or Hazard Ratio (Fig. 2). The search identified 18 articles of which were eligible for quantitative analysis in this meta-analysis. The detailed information of 18 relevant citations is listed in Table 1.

Data extraction and study assessment. Two investigators (LY and LZ) independently extracted data and reviewed the contents of the articles to determine whether or not they met the criteria for inclusion. Any discontent was discussed and resolved by a consensus including other two investigators (WZ and YW). A data extract form was developed accordingly. One review author (KL) extracted the following data from the included studies: first author's name, year of publication, number of patients, mitosis number per 50 HPFs in GISTs, size of GISTs, and *c-KIT* mutation status. The second author (LX) checked the extracted data, and disagreement was resolved by the discussion with other two authors (BL and HY) for all issues.

Statistics analysis. All analysis was performed with Review Manager 5.2. Heterogeneity between studies was assessed using the Q-test and I^2 index. Odds Ratio (OR) with 95% confidence intervals were calculated by using a fixed or random effect model depending on heterogeneity (a fixed effect model for

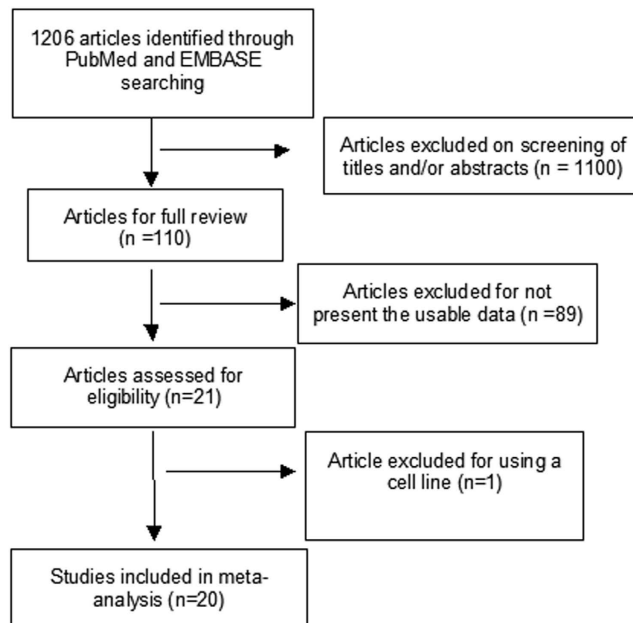


Figure 2. Schematic flow diagram for selection of included studies.

Author	Year	Country	Sample size	Follow-up (Median)	Treatment
Ma <i>et al.</i> ³¹	2014	China	68	91.3 mo	
Origone <i>et al.</i> ³²	2013	Italy	80		
Lv <i>et al.</i> ³³	2013	China	114	50 mo	Surgery
Kunstlinger <i>et al.</i> ³⁴	2013	Germany	1366		
Gao <i>et al.</i> ³⁵	2013	China	50	36 mo	Imatinib
Soreide <i>et al.</i> ³⁶	2012	Norway	38	8 year	Imatinib
Wozniak <i>et al.</i> ⁶	2012	Belgium	427	3.8 year	Surgery
Kang <i>et al.</i> ³⁷	2012	Korea	370	43.3	Imatinib
Daniels <i>et al.</i> ³⁸	2011	Germany	87		
Garces-Albir <i>et al.</i> ²⁸	2012	Spain	36	64.8 mo	Surgery
Kontogianni-Katsarou <i>et al.</i> ³⁹	2008	Greece	30		
Tzen <i>et al.</i> ⁴⁰	2008	China	134	47 mo	
DeMatteo <i>et al.</i> ¹⁴	2008	USA	127	5.2 year	Surgery
Imamura <i>et al.</i> ⁴¹	2007	Japan	95	160 mo	
Debiec-Rychter <i>et al.</i> ⁴²	2006	Belgium	476	25.3 mo	Imatinib
Yeh <i>et al.</i> ⁴³	2006	China	64	16.1 mo	Imatinib
Cho <i>et al.</i> ⁴⁴	2006	Japan	56	56.3 mo	Imatinib
Martin <i>et al.</i> ⁴⁵	2005	Spain	162	42 mo	

Table 1. Main characteristics of included studies. Abbreviations: mo, month.

$I^2 \leq 50\%$, a random effect model for $I^2 > 50\%$). Meta-analysis was performed to compare 5 year relapse free survival (RFS) in *c-KIT* exon 11 deletion and other type of *c-KIT* mutations in patients with GIST. *C-KIT* mutation frequency was compared in different size and different MI of tumors. The multivariate HRs were collected, and the log HRs and its standard errors were calculated for individual study. Pooled hazard ratio (HR) with a 95% confidence interval was calculated for the association between the risk of GISTs and *c-KIT* mutation status. All p values were two sided. Funnel plots were used for detection of publication bias. A sensitivity analysis, in which one study was removed at a time, was conducted to assess the result stability.

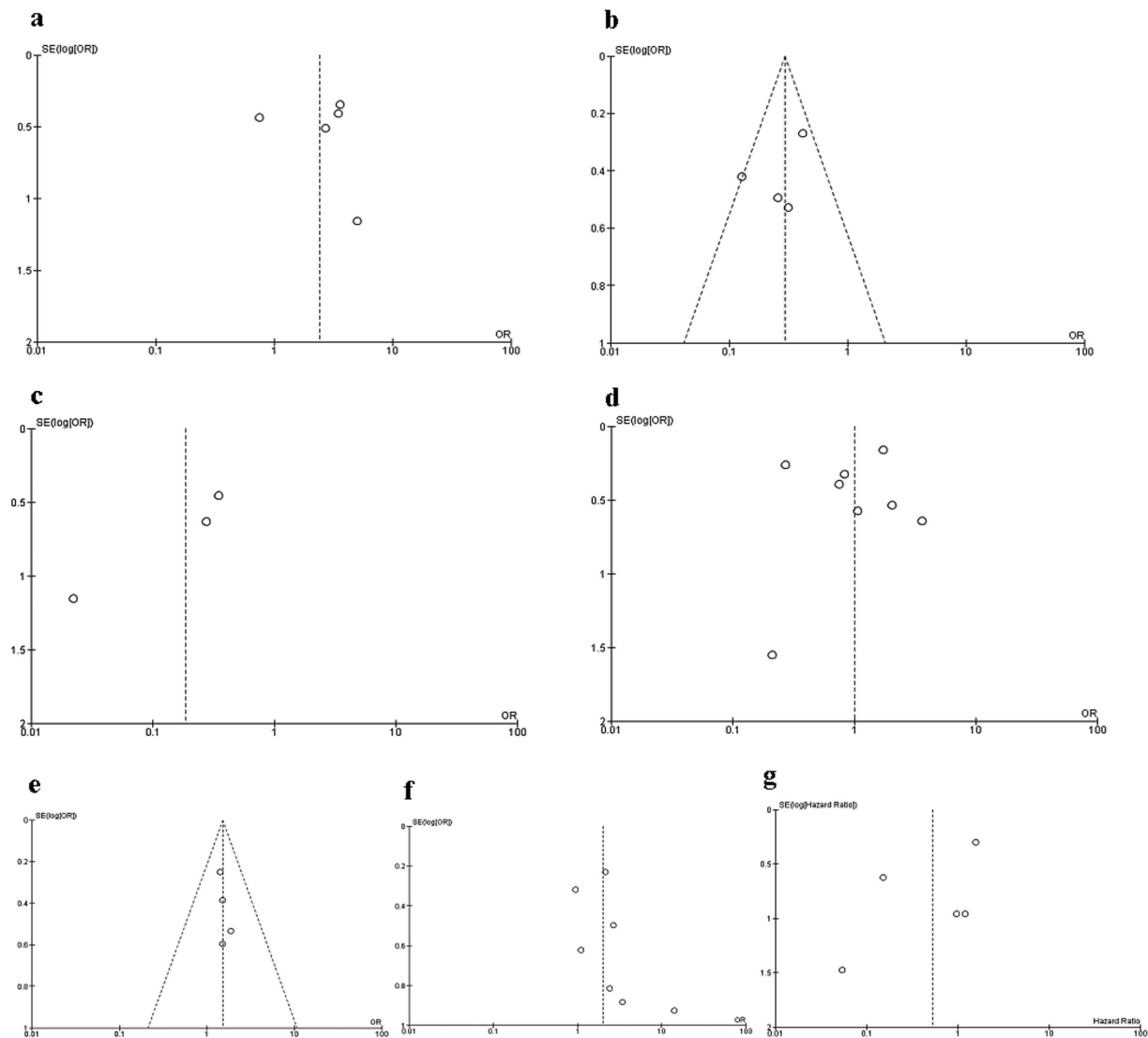


Figure 3. Funnel plot for publication bias. (a) Forest plot for PFS of GIST patients with *KIT* exon 11 mutation and *KIT* exon 9 mutation; (b) 5 year RFS of GIST patients with *KIT* 11 exon deletion and other *KIT* 11 exon mutation; (c) 5 year RFS of GIST patients with codons 557–558 of *KIT* 11 exon deletion and other *KIT* 11 deletion; (d) *KIT* mutation in different size of GIST; (e) *KIT* mutation in different of mitosis index of GIST; (f) *KIT* mutation in different of mitosis index of GIST; (g) the association of *c-KIT* mutation and the risk of GIST.

Results

Flow chart for study selection is reported in Fig. 2. There were 18 relevant articles available for meta-analysis, which included 3938 patients. The following items were collected from each study: first author's name, year of publication, number of patients, countries, the number of mitosis per 50 HPFs in GIST, tumor size, *c-KIT* mutation, treatment and the time of follow-up (Table 1).

The quality of each study was assessed with the Newcastle Ottawa Quality Assessment Scale (NOQAS). These scales were utilized to allocate a maximum of nine points for the quality of selection, comparability, exposure, and outcomes for study participants. Of the studies, one scored 8 points, ten scored 7 points, six scored 6 points, and one scored 5 points. Hence, the studies were of a relatively high quality (data not shown). The funnel plots were largely symmetric (Fig. 3) suggesting there were no publication biases in the meta-analysis of *c-KIT* mutation and clinicopathological features. We conducted a sensitivity analysis by removing a single study at one time. The pooled HR was not significantly changed, indicating the stability of our analyses.

Progression-free survival (PFS) of GIST patients was significantly worse in patients with *KIT* exon 9 mutations than in those with *KIT* exon 11 mutations, OR was 3.60, 95% CI 2.17–5.98, $z = 4.96$, $p < 0.00001$, heterogeneity $I^2 = 0\%$ (Fig. 4). 5-year RFS rate was significantly lower in patients with *KIT*

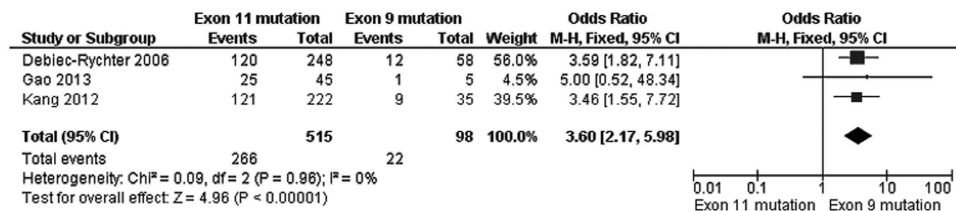


Figure 4. Forest plot for PFS of GIST patients with *KIT* exon 11 mutation and *KIT* exon 9 mutation.

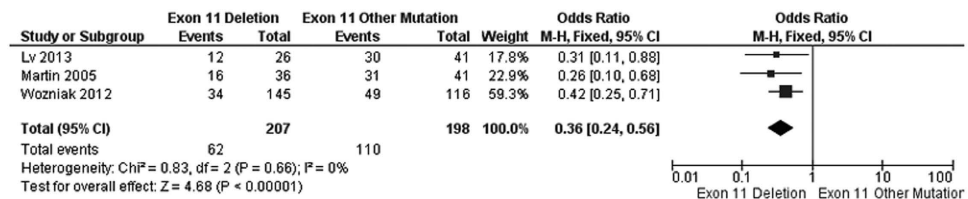


Figure 5. Forest plot for 5 year RFS of GIST patients with *KIT* 11 exon deletion and other *KIT* 11 exon mutations.

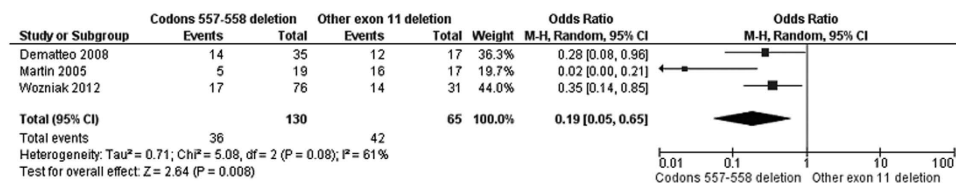


Figure 6. Forest plot for 5 year RFS of GIST patients with Codons 557–558 of *KIT* 11 exon deletion and other *KIT* 11 exon deletions.

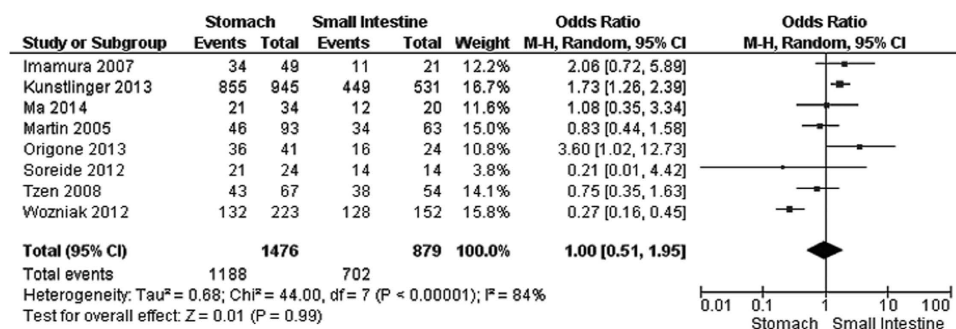
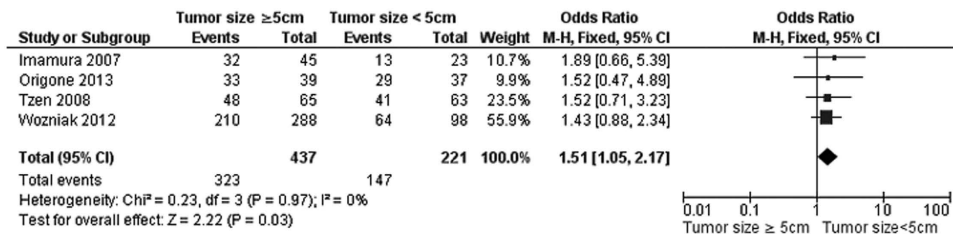
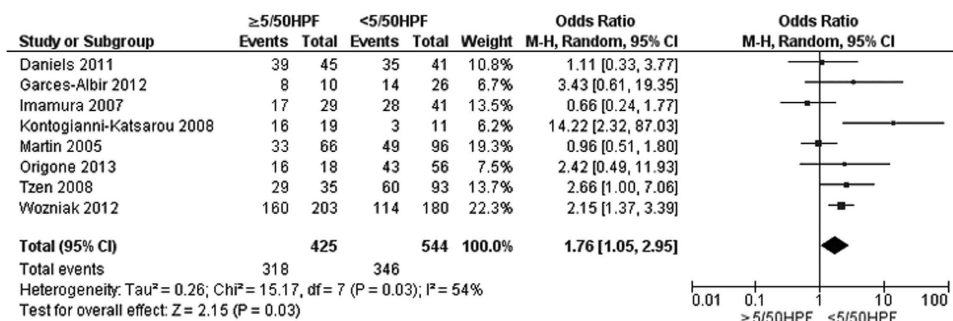
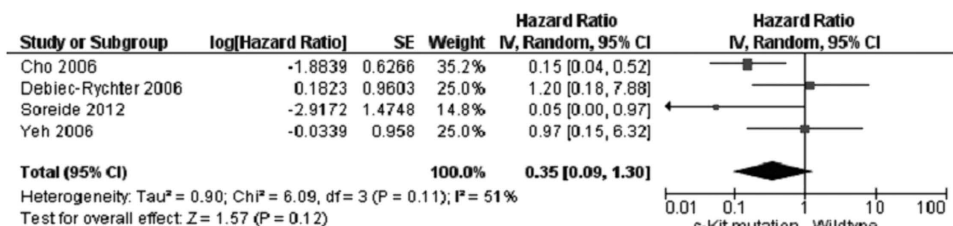


Figure 7. Forest plot for *KIT* mutation of patients with GIST in stomach and small Intestine.

exon 11 deletion than in those with other type of *KIT* exon 11 mutations, OR was 0.36, 95% CI 0.24–0.56, $z = 4.68$, $p < 0.00001$, heterogeneity $I^2 = 0\%$ (Fig. 5). Moreover, RFS for 5 year was significantly worse in patients with GISTs bearing deletions involving *KIT* codon 557–558 than in those bearing other deletions of *KIT* exon 11 (Fig. 6). The rate of *KIT* mutation was not significantly changed between GISTs in stomachs and those in small intestines, OR was 1.00, 95% CI 0.51–1.95, $z = 0.01$, $p = 0.99$, heterogeneity $I^2 = 84\%$, $p < 0.00001$ (Fig. 7). *KIT* mutations were significantly more frequently observed in the patients with larger size (≥ 5 cm) of GISTs than those with smaller size (≤ 5 cm) of GISTs, OR was 1.51, 95% CI 1.05–2.17, $z = 2.22$, $p = 0.03$, heterogeneity $I^2 = 0\%$, $p = 0.97$ (Fig. 8). *KIT* mutation was significantly increased in the patients with higher mitosis index (MI) ($\geq 5/50$ HPFs) of GISTs compared to the patients with lower MI ($\leq 5/50$ HPFs) of tumors. OR was 1.76, 95% CI 1.05–2.95, $z = 2.15$, $p = 0.03$, heterogeneity $I^2 = 57\%$, $p = 0.03$ (Fig. 9). *KIT* mutations were not significantly associated with the risk of mortality of patients with GIST. Hazard Ratio (HR) was 0.35 with a 95% confidence interval (CI) 0.09–1.30, $z = 1.57$, $p = 0.12$, heterogeneity $I^2 = 51\%$ (Fig. 10).

Figure 8. Forest plot for *KIT* mutation in different size of GIST.Figure 9. Forest plot for *KIT* mutation in different of mitosis index of GIST.Figure 10. Forest plot for the association of *KIT* mutation and the risk of GIST. Checklist S1. A PRIMA checklist.

Discussion

GISTs are the tumors with *KIT* expression, located in the gastrointestinal tract. Gain of function mutations in either *KIT* or platelet-derived growth factor receptor alpha (PDGFRA) were found in about 80%–85% of case^{4,15,16}. Many types of *KIT* mutations involved in exon 9, 11, 13 and 17 have been described in GISTs, including point mutation, insertion, deletion and duplication⁵. Treatment with tyrosine kinase inhibitor (TKIs) is effective in reducing disease recurrence after primary surgery and controlling unresectable disease¹⁷. Therefore, it is essential to identify mutation status to predict its response to TKIs and prognosis. Our analysis showed that *KIT* mutation was not associated with the risk of mortality of patients with GISTs. In the future, the stratified analysis by tumor size and mitosis index should be carried out to identify the prognosis power of *KIT* mutation, because tumor size and mitosis index are the most important confounding factors. In addition, the overall survival of patients with GISTs may depend on the specific type of *KIT* mutation. We performed a detailed subgroup analysis of relationship between different types of *KIT* mutations and prognosis of patients with GISTs. The result indicated that PFS of GIST patients was significantly worse in *KIT* exon 9 mutations than in *KIT* exon 11 mutations. Previous studies indicated the response to imatinib treatment was worse in patients whose tumors harbored *KIT* exon 9 mutations than in those with *KIT* exon 11 mutations^{18,19}. Patients with GIST treated with imatinib in all three studies were included in present meta-analysis (Fig. 2). There was no bias created from different treatments. Thus, GIST patients with *KIT* exon 9 mutations have higher risk of progression than those with exon 11 mutations.

Interestingly, deletions in the *KIT* exon 11 most frequently involve the 5' portion between codons 550 and 560, and less frequently involve codons 562–579^{12,13,20}. There is no significant difference in the response rate of imatinib or median progression-free survival among the patients with exon 11 deletion, point mutations and mixed-type mutations^{21,22}. Our result showed 5-year RFS was significantly worse in

patients with *KIT* exon 11 deletion than in those with other type of *KIT* exon 11 mutations. Moreover, RFS for 5 year was significantly worse in codon 557–558 deletion of *KIT* exon 11 than other deletion of *KIT* exon 11. Recently, a few studies reported controversial results of RFS for five year in patients of GIST with codon 557–558 deletion and other deletion of *KIT* exon 11 due to the small size of patient samples^{12–14,20}. For the first time, we pooled four studies in this meta-analysis with a total of 127 patients and more precisely assessed RFS for five year in patients of GIST with different parts of *KIT* exon 11 deletion.

KIT is a member of type III receptor tyrosine kinase family that contains platelet-derived growth factor receptors- α and - β (PDGFRA and PDGFRB), as well as the macrophage colony stimulating-factor receptor (CSF1R) and the Fl cytokine receptor (FLT3)²³. Mutations of the *KIT* gene in GISTs occur most frequently in *KIT* exon 11, the juxtamembrane domain that disrupts the normal juxtamembrane secondary structure and activate downstream signaling pathways, including the MAP kinase pathway (RAF, MEK, and ERK), the PI3 kinase/AKT pathway, and STAT3^{24–26}. The MAP and PI3 kinase pathway upregulate important transcriptional factors and lead to cell proliferation, and they downregulate the cell cycle inhibitor p27^{KIP} as well as anti-apoptotic signaling. Therefore, *KIT* mutation is a potential predictive factor for prognostic implication. We compared the frequency of *KIT* mutations in different size of tumors and different MIs. Our result indicated that *KIT* mutation was significantly more frequent in the patients with larger size (≥ 5 cm) and higher MI ($\geq 5/50$ HPFs) of GIST than in patients with smaller size (≤ 5 cm) and lower MI ($\leq 5/50$ HPFs) of GIST respectively. Taniguchi *et al.* have reported that there is a direct relationship between the presence of mutation in tumor size and mitotic count²⁷, which is in agreement with our result. Previous studies revealed that tumors larger than 5 cm and the presence of more than 5 mitoses/50 HPF were clearly associated with worse outcome²⁸. Tumor size and mitotic counts traditionally have been the two factors for estimation of prognosis²⁹. Zhao *et al.* conducted a meta-analysis and found that incidence of MI ($> 5/50$ HPFs) is not significantly higher in patients with mutated *KIT* than in the patients with wild type *KIT*³⁰. This discrepancy could be due to relatively small sample size (1751 patients). Present meta-analysis included 3980 patients and the result is more accurate. Taken together, our study indicated that *KIT* mutation status is another evaluable factor to estimate prognosis in GISTs in addition to tumor size and mitotic counts.

KIT exon 11 deletion may be associated with the risk of mortality of patients with GISTs. Additional research in the future especially larger prospective studies will be needed to evaluate this relationship. Finally, our study only selected the published articles, but it did not include some relevant unpublished papers which may result in certain publication bias. Thus the result should be interpreted carefully.

In conclusion, *KIT* mutational status has prognostic significance for patients with GISTs. GIST patients with *KIT* exon 9 mutations have higher risk of progression than those with exon 11 mutations. The deletion of *KIT* exon 11, particularly codon 557–558 deletion of *KIT* exon 11, was a valuable predictor of prognosis for patients with GISTs. The frequency of *KIT* mutation was significantly increased in the GIST patients with higher mitosis ($\geq 5/50$ HPFs) and larger size (≥ 5 cm) of tumors.

References

- Goettsch, W. G. *et al.* Incidence of gastrointestinal stromal tumours is underestimated: results of a nation-wide study. *Eur J Cancer* **41**, 2868–2872, doi: S0959-8049(05)00797-5 (2005).
- Nilsson, B. *et al.* Gastrointestinal stromal tumors: the incidence, prevalence, clinical course, and prognostication in the preimatinib mesylate era—a population-based study in western Sweden. *Cancer* **103**, 821–829, doi: 10.1002/cncr.20862 (2005).
- Joensuu, H. Gastrointestinal stromal tumor (GIST). *Ann Oncol* **17 Suppl 10**, x280–286, doi: 17/suppl_10/x280 (2006).
- Hirota, S. *et al.* Gain-of-function mutations of c-kit in human gastrointestinal stromal tumors. *Science* **279**, 577–580 (1998).
- Corless, C. L. Gastrointestinal stromal tumors: what do we know now? *Mod Pathol* **27 Suppl 1**, S1–16, doi: 10.1038/modpathol.2013.173 modpathol2013173 (2014).
- Wozniak, A. *et al.* Prognostic value of KIT/PDGFR mutations in gastrointestinal stromal tumours (GIST): Polish Clinical GIST Registry experience. *Ann Oncol* **23**, 353–360, doi: 10.1093/annonc/mdr127 mdr127 (2012).
- Chabot, B., Stephenson, D. A., Chapman, V. M., Besmer, P. & Bernstein, A. The proto-oncogene c-kit encoding a transmembrane tyrosine kinase receptor maps to the mouse W locus. *Nature* **335**, 88–89, doi: 10.1038/335088a0 (1988).
- Huizinga, J. D. *et al.* W/kit gene required for interstitial cells of Cajal and for intestinal pacemaker activity. *Nature* **373**, 347–349, doi: 10.1038/373347a0 (1995).
- Corless, C. L., Barnett, C. M. & Heinrich, M. C. Gastrointestinal stromal tumours: origin and molecular oncology. *Nat Rev Cancer* **11**, 865–878, doi: 10.1038/nrc3143 nrc3143 (2011).
- Lasota, J. & Miettinen, M. Clinical significance of oncogenic KIT and PDGFRA mutations in gastrointestinal stromal tumours. *Histopathology* **53**, 245–266, doi: 10.1111/j.1365-2559.2008.02977.x HIS2977 (2008).
- Miettinen, M. & Lasota, J. Gastrointestinal stromal tumors. *Gastroenterol Clin North Am* **42**, 399–415, doi: 10.1016/j.gtc.2013.01.001 S0889-8553(13)00019-8 (2013).
- Emile, J. F. *et al.* Clinicopathologic, phenotypic, and genotypic characteristics of gastrointestinal mesenchymal tumors. *Clin Gastroenterol Hepatol* **2**, 597–605, doi: S1542356504002435 (2004).
- Emile, J. F., Tabone-Eglinger, S., Theou-Anton, N. & Lemoine, A. Prognostic value of KIT exon 11 deletions in GISTs. *Gastroenterology* **131**, 976–977, doi: S0016-5085(06)01686-6 (2006).
- Dematteo, R. P. *et al.* Tumor mitotic rate, size, and location independently predict recurrence after resection of primary gastrointestinal stromal tumor (GIST). *Cancer* **112**, 608–615, doi: 10.1002/cncr.23199 (2008).
- Heinrich, M. C. *et al.* PDGFRA activating mutations in gastrointestinal stromal tumors. *Science* **299**, 708–710, doi: 10.1126/science.1079666 1079666 (2003).
- Miettinen, M. & Lasota, J. Gastrointestinal stromal tumors: review on morphology, molecular pathology, prognosis, and differential diagnosis. *Arch Pathol Lab Med* **130**, 1466–1478, doi: RA-5-1116 (2006).
- Ramamohan, A. *et al.* A gist of gastrointestinal stromal tumors: A review. *World J Gastrointest Oncol* **5**, 102–112, doi: 10.4251/wjgo.v5.i6.102 (2013).

18. Chen, P., Zong, L., Zhao, W. & Shi, L. Efficacy evaluation of imatinib treatment in patients with gastrointestinal stromal tumors: a meta-analysis. *World J Gastroenterol* **16**, 4227–4232 (2010).
19. Zhi, X., Zhou, X., Wang, W. & Xu, Z. Practical role of mutation analysis for imatinib treatment in patients with advanced gastrointestinal stromal tumors: a meta-analysis. *PLoS One* **8**, e79275, doi: 10.1371/journal.pone.0079275 PONE-D-13-25609 (2013).
20. Wardelmann, E. *et al.* Deletion of Trp-557 and Lys-558 in the juxtamembrane domain of the c-kit protooncogene is associated with metastatic behavior of gastrointestinal stromal tumors. *Int J Cancer* **106**, 887–895, doi: 10.1002/ijc.11323 (2003).
21. Gao, J., Dang, Y., Sun, N., Li, J. & Shen, L. C-KIT mutations were closely associated with the response to Imatinib in Chinese advanced gastrointestinal stromal tumor patients. *Med Oncol* **29**, 3039–3045, doi: 10.1007/s12032-012-0308-7 (2012).
22. Bachet, J. B. *et al.* Prognosis and predictive value of KIT exon 11 deletion in GISTs. *Br J Cancer* **101**, 7–11, doi: 10.1038/sj.bjc.6605117 6605117 (2009).
23. Hanks, S. K., Quinn, A. M. & Hunter, T. The protein kinase family: conserved features and deduced phylogeny of the catalytic domains. *Science* **241**, 42–52 (1988).
24. Duensing, A. *et al.* Mechanisms of oncogenic KIT signal transduction in primary gastrointestinal stromal tumors (GISTs). *Oncogene* **23**, 3999–4006, doi: 10.1038/sj.onc.1207525 1207525 (2004).
25. Rossi, F. *et al.* Oncogenic Kit signaling and therapeutic intervention in a mouse model of gastrointestinal stromal tumor. *Proc Natl Acad Sci U S A* **103**, 12843–12848, doi: 0511076103 (2006).
26. Rubin, B. P. *et al.* KIT activation is a ubiquitous feature of gastrointestinal stromal tumors. *Cancer Res* **61**, 8118–8121 (2001).
27. Taniguchi, M. *et al.* Effect of c-kit mutation on prognosis of gastrointestinal stromal tumors. *Cancer Res* **59**, 4297–4300 (1999).
28. Garcés-Albir, M. *et al.* Results on prognostic value of mutations in localized gastrointestinal stromal tumors (GIST) in one single center. *Rev Esp Enferm Dig* **104**, 405–410 (2012).
29. Casali, P. G., Jost, L., Reichardt, P., Schlemmer, M. & Blay, J. Y. Gastrointestinal stromal tumours: ESMO clinical recommendations for diagnosis, treatment and follow-up. *Ann Oncol* **20 Suppl 4**, 64–67, doi: 10.1093/annonc/mdp131 mdp131 (2009).
30. Zhao, W. Y., Cao, H., Zhang, Y., Shen, Z. Y. & Wu, Z. Y. [Effect of c-kit mutation on the prognosis of gastrointestinal stromal tumors: a meta-analysis]. *Zhonghua Wai Ke Za Zhi* **47**, 857–862 (2009).
31. Ma, Y. Y. *et al.* Involvement of c-KIT mutation in the development of gastrointestinal stromal tumors through proliferation promotion and apoptosis inhibition. *Onco Targets Ther* **7**, 637–643, doi: 10.2147/OTT.S60458 ott-7-637 (2014).
32. Origone, P. *et al.* Molecular characterization of an Italian series of sporadic GISTs. *Gastric Cancer* **16**, 596–601, doi: 10.1007/s10120-012-0213-y (2013).
33. Lv, A. *et al.* SKP2 high expression, KIT exon 11 deletions, and gastrointestinal bleeding as predictors of poor prognosis in primary gastrointestinal stromal tumors. *PLoS One* **8**, e62951, doi: 10.1371/journal.pone.0062951 PONE-D-13-03908 (2013).
34. Kunstlinger, H. *et al.* Gastrointestinal stromal tumors with KIT exon 9 mutations: Update on genotype-phenotype correlation and validation of a high-resolution melting assay for mutational testing. *Am J Surg Pathol* **37**, 1648–1659, doi: 10.1097/PAS.0b013e3182986b88 (2013).
35. Gao, J. *et al.* Secondary mutations of c-KIT contribute to acquired resistance to imatinib and decrease efficacy of sunitinib in Chinese patients with gastrointestinal stromal tumors. *Med Oncol* **30**, 522, doi: 10.1007/s12032-013-0522-y (2013).
36. Soreide, K. *et al.* Tyrosine-kinase mutations in c-KIT and PDGFR- α genes of imatinib naive adult patients with gastrointestinal stromal tumours (GISTs) of the stomach and small intestine: relation to tumour-biological risk-profile and long-term outcome. *Clin Transl Oncol* **14**, 619–629, doi: 10.1007/s12094-012-0851-x (2012).
37. Kang, H. J. *et al.* Imatinib efficacy by tumor genotype in Korean patients with advanced gastrointestinal stromal tumors (GIST): The Korean GIST Study Group (KGSG) study. *Acta Oncol* **51**, 528–536, doi: 10.3109/0284186X.2011.636753 (2012).
38. Daniels, M. *et al.* Spectrum of KIT/PDGFR α /BRAF mutations and Phosphatidylinositol-3-Kinase pathway gene alterations in gastrointestinal stromal tumors (GIST). *Cancer Lett* **312**, 43–54, doi: 10.1016/j.canlet.2011.07.029 S0304-3835(11)00454-X (2011).
39. Kontogianni-Katsarou, K. *et al.* KIT exon 11 codon 557/558 deletion/insertion mutations define a subset of gastrointestinal stromal tumors with malignant potential. *World J Gastroenterol* **14**, 1891–1897 (2008).
40. Tzen, C. Y., Wang, M. N. & Mau, B. L. Spectrum and prognostication of KIT and PDGFR α mutation in gastrointestinal stromal tumors. *Eur J Surg Oncol* **34**, 563–568, doi: S0748-7983(07)00180-1 (2008).
41. Imamura, M. *et al.* Prognostic significance of angiogenesis in gastrointestinal stromal tumor. *Mod Pathol* **20**, 529–537, doi: 3800767 (2007).
42. Debiec-Rychter, M. *et al.* KIT mutations and dose selection for imatinib in patients with advanced gastrointestinal stromal tumours. *Eur J Cancer* **42**, 1093–1103, doi: S0959-8049(06)00175-4 (2006).
43. Yeh, C. N. *et al.* Kinase mutations and imatinib mesylate response for 64 Taiwanese with advanced GIST: preliminary experience from Chang Gung Memorial Hospital. *Ann Surg Oncol* **14**, 1123–1128, doi: 10.1245/s10434-006-9288-1 (2007).
44. Cho, S. *et al.* Deletion of the KIT gene is associated with liver metastasis and poor prognosis in patients with gastrointestinal stromal tumor in the stomach. *Int J Oncol* **28**, 1361–1367 (2006).
45. Martin, J. *et al.* Deletions affecting codons 557-558 of the c-KIT gene indicate a poor prognosis in patients with completely resected gastrointestinal stromal tumors: a study by the Spanish Group for Sarcoma Research (GEIS). *J Clin Oncol* **23**, 6190–6198, doi: 23/25/6190 (2005).

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Author Contributions

L.Y., L.Z., W.Z. and H.Y. contributed substantially to the study and design, collection of data, and analysis and interpretation of data. L.Y., L.Z. and Y.W. contributed substantially to the acquisition, analysis, interpretation of data and performed the statistical analysis. Y.W., B.L. and H.I.Y. have been involved in the drafting and critical revision of the article for important intellectual content. The corresponding authors have full access to all data and the final responsibility for the decision to submit the article for publication. All authors read and approved the final manuscript.

Additional Information

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