MBRS-57. IDENTIFICATION OF MYC-DEPENDENT THERAPEUTIC VULNERABILITIES FOR TARGETING GROUP 3 MEDULLOBLASTOMA

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Group 3 medulloblastoma (MB_{Group3}) is a highly aggressive tumour char-acterised by MYC amplification and elevated expression (17% of MB_{Group3}). MYC amplification in MB_{Group3} confers a dismal prognosis using standard therapies and there is an urgent upper to nod for accord the second theory of the second standard theory of theory of the second stan therapies, and there is an urgent unmet need for novel therapeutic approaches. The identification and targeting of MYC's biological dependencies thus represents a promising strategy to treat MYC-MB_{Group3} tumours. Three independent isogenic MYC-regulable MB_{Group3} human cell-based models, in which elevated MYC expression can be directly down-regulated by doxycycline-inducible shRNAs, were developed and used initially to establish MYC-dependent growth of each model. Our novel models were then used to investigate MYCdependent drug sensitivity, by characterising responses to a panel of candidate cancer therapeutics and small molecule inhibitors, including a high-throughput compound screen of >500 established/clinically-relevant small molecule inhibitors. This approach identified several specific, consistently observed, druggable MYC-dependencies (e.g. cell cycle regulators, DNA-damage response controllers, mitotic control machinery) with potential for the development of treatments against MYC-MB_{Group3} tumours. *PLK1*, *CHK1* and *AURK* were identified as prime candidate targets with consistent MYC-dependent response profiles. Subsequent validation of each candidate, by genetic and pharmacological target inhibition, confirmed their MYC-dependent effects, associated with downregulation of MYC and established target-dependent pharmacodynamic biomarkers/pathways. Results were consistent across all of our MB_{Group3} models. In summary, our novel models reveal druggable *MYC*-associated dependencies as a feature of MB_{Group3}. Our findings support the de-velopment of *PLK1*, *CHK1* and *AURK* inhibition as therapeutic approaches against MYC-dependent MB_{Group3} . Future work is now essential to validate our findings *in vivo*, to support the design of future clinical trials.

MBRS-59. SINGLE-CELL WHOLE-GENOME SEQUENCING DISSECTS INTRA-TUMOURAL GENOMIC HETEROGENEITY AND CLONAL EVOLUTION IN CHILDHOOD MEDULLOBLASTOMA Marina Danilenko¹, Masood Zaka², Claire Keeling¹, Stephen Crosier¹, Rafiqul Hussain³, Edward Schwalbe⁴, Dan Williamson¹, Jonathan Coxhead³, Vikki Rand², Simon Bailey¹, and Steven Clifford¹; ¹Wolfson Childhood Cancer Research Centre, Translational & Clinical Research Institute, Newcastle University Centre for Cancer, Newcastle upon Tyne, United Kingdom, ²National Horizons Centre, Darlington, United Kingdom, ³Genomics Core Facility, Biosciences Institute, Newcastle University, Newcastle upon Tyne, United Kingdom, ⁴Department of Applied Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom

Medulloblastomas harbor clinically-significant intra-tumoral heterogeneity for key biomarkers (e.g. MYC/MYCN, β-catenin). Recent studies have characterized transcriptional heterogeneity at the single-cell level, however the underlying genomic copy number and mutational architecture remains to be resolved. We therefore sought to establish the intra-tumoural genomic heterogeneity of medulloblastoma at single-cell resolution. Copy number patterns were dissected by whole-genome sequencing in 1024 single cells isolated from multiple distinct tumour regions within 16 snap-frozen medulloblastomas, representing the major molecular subgroups (WNT, SHH, Group3, Group4) and genotypes (i.e. MYC amplification, TP53 mutation). Common copy number driver and subclonal events were identified, providing clear evidence of copy number evolution in medulloblastoma development. Moreover, subclonal whole-arm and focal copy number alterations covering important genomic loci (e.g. on chr10 of SHH patients) were detected in single tumour cells, yet undetectable at the bulk-tumor level. Spatial copy number heterogeneity was also common, with differences between clonal and subclonal events detected in distinct regions of individual tumours. Mutational analysis of the cells allowed dissection of spatial and clonal heterogeneity patterns for key medulloblastoma mutations (e.g. CTNNB1, TP53, SMARCA4, PTCH1) within our cohort. Integrated copy number and mutational analysis is underway to establish their inter-relationships and relative contributions to clonal evolution during tumourigenesis. In summary, single-cell analysis has enabled the resolution of common mutational and copy number drivers, alongside sub-clonal events and distinct patterns of clonal and spatial evolution, in medulloblastoma development. We anticipate these findings will provide a critical foundation for future improved biomarker selection, and the development of targeted therapies.

MBRS-60. THE ACTIONABLE GENOMIC LANDSCAPE OF RELAPSED MEDULLOBLASTOMA IS DEFINED BY MAINTENANCE AND ACQUISITION OF DRIVER EVENTS

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Medulloblastoma relapse (rMB) occurs in 30-40% of patients and is almost universally fatal. Understanding the genomic landscape of rMB, and its relationship to disease characteristics at diagnosis, will be essential to underpin the development of improved therapeutic strategies, delivered at both diagnosis and relapse. Utilising NGS and Illumina DNA methylation arrays, we interrogated the molecular landscape of >100 rMBs, alongside matched diagnostic samples (n>80), encompassing molecular subgroup, novel subtypes, copy number (CNV) and mutational variants. Molecular subgroup and novel subtypes were stable over disease-course. The majority of genomic aberrations were also maintained (total arm-level CNVs at relapse, 60% maintained/40% acquired; deleterious/driver mutations, 75% maintained/25% acquired). Importantly, however, the landscape of alterations differed markedly at relapse, through both selective maintenance and acquisition of specific gene and pathway aberrations. For instance, we observed significant enrichment of subgroup-specific events at relapse, including focal CDK6/CDK14 amplifications (4/26 (15%) of MB_{Group4}) and CDKN2A/CDKN2B deletions (3/48 (6%) of MB_{SHH}). In contrast, mutations in DNA damage response pathways were commonly enriched across all molecular subgroups, most significantly in MB_{SHH} (~40% of rMB_{SHH}; TP53, 9/36 (25%); ATM, 5/36 (14%)). Driver events in rMB are characterised by both selective maintenance and acquisition across disease-course, and together combine to define its actionable genetic landscape. Evaluation of their clinical and biological significance will be essential to establish their potential (i) as biomarkers to direct disease management and (ii) as a basis for therapeutic strategies targeted against medulloblastoma relapse.

MBRS-61. MOLECULAR SUB-GROUPING OF PEDIATRIC MEDULLOBLASTOMA: CORRELATION WITH CLINICAL AND HISTOLOGICAL FEATURES, A SINGLE INSTITUTIONAL STUDY Gauri Deshpande, Mamta Gurav, Omshree Shetty, Vinayak Kadam, Vishal Chaubey, Tejpal Gupta, Aliasgar Moiyadi, Girish Chinnaswamy, and Sridhar Epari; Tata Memorial Centre, Mumbai, Maharashtra, India

INTRODUCTION: Molecular subgroups of pediatric medulloblastomas are distinctive in infantile and non-infantile age-groups. METHODS: Realtime quantitative PCR based GEP of customized 12 protein-coding genes was performed on 206 FFPE childhood medulloblastoma samples. FISH for MYC amplification, monosomy 6 and sequencing for CTNNB1 exon 3 mutation were done in relevant cases. H&E and reticulin-stained slides were used for histological subtyping. p53-protein immunoreactivity pattern was noted. RESULTS: Infantile (n=33) comprised 57.6% SHH-activated (desmoplastic: 73.7%; MBEN: 15.8% and classic: 10.5%), 21.2% group 3 (large cell/anaplastic [LCA]: 28.6% and none were desmoplastic) and 12% group 4. 40% of group 3 patients died of disease and 21% of the SHHactivated (all desmoplastic) had subsequent local recurrence. Non-infantile (n=173) comprised 19.4% WNT-activated, 12.9% SHH-activated (15% classic, 30% desmoplastic, 10% paucinodular), 19.4% group 3 (63.3% classic & 26.7% LCA), 48.4% group 4 (73.3% classic, 5.3% desmoplastic, 10.7% paucinodular & 1.4% LCA), and non-WNT/non-SHH (NWNS), NOS (n=14,9%) and unclassified (n=4,2.6%). None of WNT-activated were desmoplastic/LCA histology. Non-infantile WNT-activated and group 3 MBs showed 90% monosomy 6 & CTNNB1 mutation, and 16.7% MYCamplification respectively. 17.4% (13% spinal, 4.4% local) WNT-activated, 31% (12.5% local, 18.5% distant [spinal: 12.5%, intracranial:6%]) SHHactivated, 27% (18% both spinal and local, 9% spinal) group 3 and 31.5% (7.4% local, 5.5% intracranial, 11.2% spinal, 7.4% both spinal and local) group 4 showed metastases during follow up. CONCLUSIONS: SHH-activated and group 3 are the common infantile subgroups but group 4 is not non-existent in infantile age. No desmoplastic (including paucinodular) histological subtype is of WNT- activated and group 3.