Clinical manifestations and genetic analysis of 4 children with chronic granulomatous disease

Chunyan Guo, MD^{* (D}, Xing Chen, MD, Jinrong Wang, MD, Fengqin Liu, MD, Yan Liang, MD, Juan Yang, MD, Fangfang Dai, MS, Ning Ding, MS

Abstract

Pediatricians are unfamiliar with chronic granulomatous disease (CGD) because of its rarity and paucity of available data, potentially leading to misdiagnosis, late treatments, and mortality. The main purpose of this study was to summarize the clinical manifestations and auxiliary examination findings of four children with CGD confirmed by genetic testing.

This was a case series study of children hospitalized at the Pediatric Respiratory Department of Shandong Provincial Hospital. The clinical, laboratory, treatment, and prognosis data were analyzed.

All 4 children were boys. Two were brothers. The children's age was from 34 days to 3 years and 2 months at disease onset. The manifestations were repeated pulmonary infection, lymphadenitis, skin infection, and granuloma formation. Pulmonary infections were common. Abnormal responses were common after BCG vaccination. Thoracic computed tomography (CT) mainly showed nodules and masses, while the consolidation area in CT images reduced slowly. No abnormalities in cellular immune functions and immunoglobulin were found. The disease in all four children was confirmed by genetic testing. Long-term antibiotics and anti-fungal drugs were needed to prevent bacterial and fungal infections.

CGD should be considered in children with repeated severe bacterial and fungal infections. Abnormal responses after BCG vaccination and nodular or mass-shaped consolidation in thoracic CT images should hint toward CGD. Gene sequencing could provide molecular evidence for diagnosis. The treatments of CGD include the prevention and treatment of infections and complications. Immunologic reconstitution treatment is currently the only curative treatment for CGD.

Abbreviations: BALF = bronchoalveolar lavage fluid, CGD = chronic granulomatous disease, CRP = C-reactive protein, CT = computed tomography, NADPH = nicotinamide adenine dinucleotide phosphate.

Keywords: children, chronic granulomatous disease, diagnosis, imaging, treatment

1. Introduction

Chronic granulomatous disease (CGD) is a rare hereditary disease characterized by the congenital dysfunction of phagocytes (including neutrophils, monocytes, macrophages, and eosino-phils).^[1-3] The mean age at diagnosis is 3.2 years for the X-linked recessive form and 11 years in patients with the autosomal recessive form.^[4,5] Management of CGD typically consists of prophylactic

Editor: Bing Rong Zhou.

The study was supported by the Natural Science Foundation of Shandong Province (No. ZR2012HM073).

The authors have no conflicts of interest to disclose.

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Department of Pediatric Respiratory, Provincial Hospital Affiliated to Shandong University, Jinan, China.

* Correspondence: Xing Chen, Provincial Hospital Affiliated to Shandong University, Jinan, Shandong, China (e-mail: chen_xing9999@163.com).

Copyright © 2020 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Guo C, Chen X, Wang J, Liu F, Liang Y, Yang J, Dai F, Ding N. Clinical manifestations and genetic analysis of four children with chronic granulomatous disease. Medicine 2020;99:23(e20599).

Received: 14 March 2020 / Received in final form: 30 April 2020 / Accepted: 5 May 2020

http://dx.doi.org/10.1097/MD.000000000020599

therapy and aggressive treatment of acute infections.^[1–3] The only curative options are hematopoietic cell transplantation and gene therapy.^[6]

Medicine

The nicotinamide adenine dinucleotide phosphate (NADPH) oxidase complex is composed of five subunits (gp91phox, p22phox, p47phox, p67phox, and p40phox) encoded by the *CYBB*, *CYBA*, *NCF1*, *NCF2*, and *NCF4* genes, respectively. CGD caused by *CYBB* gene mutation is an X-linked CGD, while CGD caused by mutations in the *CYBA*, *NCF1*, *NCF2*, and *NCF4* genes is called autosomal recessive inherited CGD. Any of those mutations will lead to a deficiency in the reduced NADPH oxidase complex in macrophages.^[1-3] This deficiency leads to respiratory burst dysfunction and inhibits the generation of superoxide in macrophages, consequently abrogating their capability of killing the peroxidase-positive bacteria and fungi.^[1,2]

The major characteristic of CGD is granuloma formation induced by repeated severe bacterial and fungal infection, as well as by excessive inflammatory responses.^[1–3] The typical manifestations of CGD include repeated fever and local purulent inflammation such as repeated pulmonary infection, lymphadenitis, hepatapostema, osteomyelitis, dermapostasis, and cellulitis; pulmonary lesions are the most common manifestations of this disease.^[4]

Despite its severity, pediatricians are unfamiliar with CGD because of the rarity of the disease and the paucity of available data, potentially leading to misdiagnosis, late treatments, and mortality.^[7,8] Therefore, the aim of the present study was to summarize the clinical manifestations and auxiliary examination

findings of four children with CGD confirmed by genetic testing between 2012 and 2016. Improving pediatricians' knowledge and awareness of the disease should improve its management.

2. Methods

2.1. Study design and patients

This was a case series study of children hospitalized at the Pediatric Respiratory Department of Shandong Provincial Hospital between October 2012 and August 2016. This study was approved by the Ethics Committee of the Shandong Provincial Hospital. All patients' parents provided written informed consent for the publication of this case series report.

2.2. Data collection

The laboratory examination results, imaging findings, and genetic testing results were collected. The laboratory examinations included blood routine examinations, C-reactive protein (CRP), cellular immune function, immunoglobulin test, and pathogenic examinations. The pathogenic examinations included blood culture of bacteria and fungi, sputum culture, bronchoal-veolar lavage fluid (BALF) culture of bacteria and fungi, contamination-proof fiberoptic bronchoscopic brushing assay for bacteria and fungi culture, culture of perianal abscess discharge, quantification of mycobacterial DNA in BALF, acid-fast bacillus smear, GM test of BALF, G test of serum, GM test, PPD test, and T-sport test. The imaging examinations included thoracic X-ray and CT scanning.

For the routine genetic testing, 1 to 2 ml of EDTAanticoagulated peripheral blood were collected from the children and their parents. Genomic DNA were extracted from peripheral blood samples using standard procedures. Five candidate genes of primary immunodeficiency diseases, including *CYBB*, *CYBA*, *NCF1*, *NCF2*, and *NCF4*, were sequenced using the Illumina Nextseq500 platform (Illumina Inc., San Diego, CA). Sanger sequencing was performed to confirm the candidate pathogenic variants.

2.3. Follow-up

The children were routinely followed by an outpatient visit, medical record during hospitalization, and telephone. The last follow-up was on September 20th, 2019.

Medicine

2.4. Statistical analysis

All statistical analyses were performed using SPSS 20.0 (IBM, Armonk, NY). Only descriptive statistics were used.

3. Results

3.1. Clinical characteristics

Four children, all boys, were included in this study. Their ages ranged from 34 days to 2 years and 2 months. These children were from 3 families, and patient #4 was the elder brother of patient #3. Family history was found for patient #1 (his grandfather had been diagnosed with mycotic pneumonia).

All 4 children were admitted to the hospital for symptoms of pulmonary infection (fever and/or cough). Patients #3 and #4 had a history of perianal abscesses. Patients #2 and #3 had a history of suppuration at the site of BCG vaccination. Patient #4 had a history of sepsis, purulent meningitis, and infection-related hemophagocytic syndrome (Table 1).

Physical examinations showed multiple enlarged lymph nodes in the left axilla of patients #2 and #3. The size of the BCG vaccination scar was about $5 \times 2 \text{ cm}$ in patient #3.

3.2. Laboratory examinations

For all 4 children, the blood routine examinations in the acute infection phase showed evidently increased white blood cell count, as high as $20 \text{ to } 40 \times 10^9$ /L, mainly neutrophils. The CRP levels were 40 to 150 mg/L. No abnormalities were found in the cellular immune functions and immunoglobulin tests (Table 1).

For patient #1, BALF culture (Fig. 1A-B) and contaminationproof brushing assay showed infection by *Aspergillus fumigatus*. For patient #2, serum GM test results were high, the PPD test was strongly positive, the T-spot test showed positive results, and the BALF culture showed infection by *Pseudomonas aeruginosa*. For patient #3, serum G test results were high, and the PPD test was strongly positive. For patient #4, the PPD test was strongly positive, and the T-spot test showed positive results.

3.3. Imaging examinations

For patients #1 (Fig. 1C-D) and #2 (Fig. 2A-B), thoracic computed tomography (CT) at disease onset showed nodular high-density shadows or mass-like shadows at the site of infection. For patient #2, the thoracic CT at disease onset

Table 1

Demographic and clinical characteristics of the 4 children.

2 chiegi aprile and chinear chiare		••••		
	#1	#2	#3	#4
Age	34 days	4 months and 28 days	2 years and 2 months	2 years and 2 months
Sex	Male	Male	Male	Male
Family history	Yes	Yes	No	No
Symptom	Cough and fever	Cough	Repeated fever	Cough and fever
History of perianal abscess	No	No	Yes	Yes
Enlargement of left axillary lymph nodes	No	No	Yes	No
PPD test	Negative	Strongly positive	Negative	Strongly positive
T-spot	Negative	Positive	Negative	Positive
G test	Elevation	Negative	Positive	Negative
GM test	Normal	Positive	Negative	Negative
Bacterial and Fungal culture	Aspergillus fumigatus positive	Pseudomonas aeruginosa	Normal upper respiratory tract flora	Normal upper respiratory tract flora
Genetic testing	NCF2 mutation	CYBB mutation	CYBB mutation	CYBB mutation



Figure 1. Patient #1, a 34-day-old boy admitted to the hospital for cough for 1 week, and fever for 1 day. (A) BALF culture for bacteria showed the growth of *Aspergillus fumigatus*. (B) BALF culture for fungi showed the growth of *A fumigatus*. Thoracic computed tomography at admission (C-D) showed multiple nodular high-density shadows or mass-like shadows.

showed multiple enlarged lymph nodes in the left axilla, while the chest X-ray 2 years later showed calcification of the lymph nodes in the left axilla (Fig. 2C). For patient #3, thoracic imaging at initial admission showed the manifestations of bronchopneumonia. For patient #4, thoracic CT at disease onset showed interstitial pneumonia. Thoracic CT after disease progression in patients #3 and #4 showed multiple nodular high-density shadows or mass-like shadows (Figs. 3 and 4).

3.4. Genetic testing

Patient #1 was with autosomal recessive inherited CGD with an *NCF2* gene mutation. Patients #2, #3, and #4 were with X-linked CGD with a *CYBB* gene mutation. For Case 1, both parents were carriers of the pathogenic gene. For Case 2, 3 and 4, their mothers were gene carriers (Figs. 5–8).

3.5. Treatments and prognosis

All 4 children received treatments with voriconazole, imipenem, and cilastatin. In addition, patient #1 also received vancomycin, while patient #2 received linezolid. After diagnosis, patient #2 received long-term treatment with cotrimoxazole and voriconazole. The last follow-up of patient #2 was on September 20, 2019, at which time his general condition was good, and the patient was



Figure 2. Patient #2, a boy of 4 months and 28 days, admitted to the hospital for cough for over 2 months and aggravated for 4 days. (A-B) Thoracic computed tomography showing multiple nodular high-density shadows or mass-like shadows. (C) Thoracic X-ray image 2 years later, showing calcification of the left axillary lymph nodes.



Figure 3. Patient #3, a boy of 2 years and 2 months, admitted to the hospital for repeated fever for over half a month. Thoracic computed tomography 7 years after disease onset, showing multiple nodular high-density shadows or mass-like shadows.



Figure 4. Patient #4, a boy of 2 years and 2 months who was the elder brother of patient #3, admitted to the hospital for "intermittent fever for over half a month, and cough for over 10 days." Thoracic computed tomography 7 years after disease onset, showing multiple nodular high-density shadows or mass-like shadows.

on the waiting list for hematopoietic stem cell transplantation. The parents of the patients #1, #3, and #4 did not abide by the doctors' orders to continue the long-term preventive treatment. Although they are still alive, repeated infections and progressive pulmonary lesions were found.

4. Discussion

Most pediatricians are unfamiliar with CGD because of its rarity and paucity of available data, potentially leading to misdiagnosis, late treatments, and mortality.^[7,8] The results of this study suggest that CGD should be considered in children with repeated severe bacterial and fungal infections. Abnormal responses after BCG vaccination and nodular or mass-shaped consolidation in thoracic CT images should hint toward CGD.

Abnormalities in any of the five subunits of the NADPH oxidase complex or in the G-proteins lead to dysfunctions of the oxidase complex.^[9–11] Most cases of CGD are X-linked recessive inherited CGD caused by mutation in the *CYBB* gene, while some cases are autosomal recessive inherited CGD caused by mutations in the *CYBA*, *NCFI*, *NCF2*, and *NCF4* genes.^[9,11] Over 700 mutations in the *CYBB* gene have been reported.^[12] In the present study, all 4 boys were confirmed with CGD by genetic testing: 3 were with gene mutations in CYBB, and one was with a mutation in *NCF2*. Their mothers were all carriers of their offspring's mutation.

GeneticTesting

Sample ID :	C160829C06001	Name :		Gender :	Male
Age :	1	Sample Type : Blood Specimen Received		Specimen Received :	2016/8/27
Hospital :	Shandong Provinc		Medical Record Number :	G (1	
Test Name :	Immune Diseases	Panel			
Clinical Inform	ation : Congenital g	ranuloma			

Test Result : Pathogenic variant is identified

Variant Information :

Gene	Position	Transcr ipt ID	Exon	Nucleoti de Chan ges	Amino A cid Chan ges	Hom/ Het	MAF	Variant classifi cation	Inheri tance	Disease / phenotyp e	Variant Origin
NCF 2	chr1-183 532440	NM_00 0433	exon 13	c.1180T> G	p.Y394D	het	-	pathog enic	AR	Chronic granuloma tous disease due to deficiency of NCF-2	Father
NCF 2	chr1-183 532442	NM_00 0433	exon 13	c.1179-1 G>C	splicing	het	5	pathog enic	AR	Chronic granuloma tous disease due to deficiency of NCF-2	Mother

Figure 5. Genetic test report of Case 1 showed that the child was an autosomal recessive CGD caused by NCF2 gene mutation and both parents were carriers of the pathogenic gene.

Name	Serial Nu	umber	Sample Type		Blo	Blood			Sequencing	
	1012586	012586NIt01 Speci		nen April 30		0,2015	Research		of Site 301 in	
			Received				Project		The Coding	
Father	101400)7Gs Bonort		Date June 1		5 2015		oject	Region of	
Mother	101400	08Gs	Gs		Date June 15,				CYBB Gene	
Result:										
Name	Name Gene Mutat			Nucleotide		Amino Acid		Mutation Affecting		
Ivanie			M		utation Mutation		n			
								Protein structure		
	CYBB	home	zygote c.30		1C>G(E4)	p.H101D) pre		edicted to be	
									harmful.	
Father	CYBB	/	Wil		Type 301C					
							Pro	tein structure		
Mother	CYBB	CYBB hetero		c.30	1C>G(E4)	p.H101D) pr		edicted to be	
									harmful.	

Genetic Testing

Figure 6. Genetic test report of Case 2 showed that the child was X-linked CGD caused by CYBB gene mutation, and the mother was the carrier of the pathogenic gene.

The age at onset is associated with the residual activity of NADPH oxidase in macrophages.^[11,13] The clinical manifestations of CGD are very complex. The symptoms of CGD can appear at a very young age in serious cases or in adolescence or adulthood in relatively mild cases^[14] and include repeated fever and repeated pulmonary infection, lymphadenitis, hepatapostema, osteomyelitis, dermapostasis, and cellulitis.^[1-3,15] Pulmonary lesions are the major cause of death of this disease.^[1-4,15,16]

All 4 children presented here were hospitalized for pneumonia, among whom 2 children had *Aspergillus* pneumonia (patients #1 and #2), while the other 2 were with *Mycobacterium tuberculosis* infection. In addition, one of the 2 children with *Mycobacterium tuberculosis* developed *Aspergillus* pneumonia during treatment for pulmonary tuberculosis. Skin or lymph node infection is relatively common in patients with this CGD.^[17] Two children had a history of perianal abscess, and



Figure 7. Genetic test report of Case 3 showed that the child was X-linked CGD caused by CYBB gene mutation, and the mother was the carrier of the pathogenic gene.

Name	Serial Nur	nber	Sample 1	Гуре	Bloc	d	Research		Sequencing
	105433	Gs	Specim Receive	en ed	September	25,2013			of Site 1234 in The Coding
Mother	105434	Gs	Report D	Date	November	5,2013		Jeci	Region of CYBB Gene
Result	:	50		01		6			
Name	Gene	Muta	ation Type	N T	Nucleotide Amino Acid Mutation Mutation		Mut	ation Affecting	
	СҮВВ	hon	nozygote		1234G>A	p.G412R		It has the potential to cause disease	
Mother	СҮВВ	hete	erozygote	C.	1234G>A	p.G412	R.	It has ca	s the potential to ause disease

Genetic Testing

Figure 8. Genetic test report of Case 4 showed that the child was X-linked CGD caused by CYBB gene mutation, and the mother was the carrier of the pathogenic gene.

2 children were with skin or lymph node infection with local scar formation.

Abnormal reactions after BCG vaccination occur in children with CGD.^[17,18] In addition, positive results of blood culture for *Mycobacterium tuberculosis* can be found in some cases. Abnormal responses after BCG vaccination should be a cue for the diagnosis of CGD. Findings in two children included here strongly suggest that abnormal responses after BCG vaccination have a high value for the diagnosis of CGD.

In CGD, the typical granuloma manifests as non-caseous necrosis and may occur in the brain, lung, liver, spleen, and gastrointestinal tract.^[13,19] Generally, there is no pathogen in the granuloma, and response to glucocorticoid therapy is generally good, but complications may occur.^[20] Gastrointestinal tract involvement, especially inflammatory bowel disease, can be the initial manifestation of CGD.^[17]

As pulmonary infection is the most frequent symptom of CGD, imaging of the lungs mainly shows multiple nodular high-density shadows, masses, or lobe consolidation, accompanied by nodular consolidation.^[17] In the present series, the initial thoracic CT images in 2 of the 4 children showed multiple nodular consolidate shadows or mass-like shadows. Therefore, imaging findings of multiple nodular consolidations have important indicative value for the diagnosis of CGD.^[1–3] Because of the over-activated inflammatory responses in CGD, the decrease of the consolidation area with treatment could be slower than in patients with pulmonary infections.^[21]

CGD is a primary immunodeficiency disease due to decreased function of macrophages, but the routine cellular immune examinations and immunoglobulin examinations mainly show no evident abnormalities, and therefore, regular immunological function examinations have no indicative value for the diagnosis of this disease.^[1–3] Thus, the regular immunological function examinations in all four children were normal. The principle of the DHR test is that the non-fluorescent DHR can freely enter the neutrophils; when the NADPH oxidase is activated by phorbol ester, reactive oxygen species (ROS) are generated, and oxidized DHR has a green fluorescence. Therefore, children with CGD should have low or absent DHR signals.^[14] Of course, gene sequencing allows the characterization of the exact mutation that

could cause CGD.^[14] In the present study, all 4 children with CGD underwent gene sequencing, which confirmed *CYBB* gene mutations in three children, and an *NCF2* gene mutation in one child.

The management of CGD mainly includes controlling the infections in their acute phase, radical treatment of the underlying diseases, and prevention of infection. For each child with CGD, aggressive pathogenic examinations and drug sensitivity tests should be conducted, the presence of *Mycobacterium tuberculosis* should be controlled for, and attention should be paid to fungal infections.^[1-3,22] Antibiotics should be selected according to the results of the sensitivity tests. For patients with unavailable results, empirical treatment with antibiotics should cover the oxidase-positive bacteria. For patients with empyema or abscess, surgical drainage is needed.^[1-3,17] For CGD children with pulmonary symptoms and/or fever from unknown causes, empirical antifungal therapy should be conducted.^[1-3] For patients with *Mycobacterium tuberculosis*, anti-tuberculosis drugs should be given according to the patient's condition.

Immunologic reconstitution treatment, including stem cell transplantation and gene therapy, is the only curative treatment for CGD.^[4,6,23-26] Nevertheless, gene therapy is still in its preliminary stage, and stem cell transplantation is still the most effective treatment method for most CGD patients. After diagnosis, transplantation should be conducted in the early life stage to protect the organ functions as much as possible.^[6]

Long-term or even life-long trimethoprim-sulfamethoxazole should be provided for CGD patients to prevent bacterial infection before effective immunologic reconstitution treatment.^[8,27] The patients should avoid exposure to fungi-breeding environments or materials. Long-term or even life-long itraconazole treatment should be provided for CGD children to prevent fungal infection.^[8,27] The liver functions should be monitored regularly during itraconazole treatment. The use of INF- γ in treating CGD is still controversial.^[17]

In conclusion, the understanding of CGD in pediatricians should be improved to avoid misdiagnosis and complications.^[28,29] CGD should be considered in children with severe bacterial or fungal infections in the lung, lymph nodes, liver, spleen, bone, or skin with an onset at a very young age, in the presence of abnormal responses after BCG vaccination, and with multiple nodular high-density shadows or mass-like shadows in thoracic CT images that disappear very slowly or not all after treatment. For CGD patients or carriers, genetic counseling and prenatal examinations should be considered. BCG vaccination should be avoided for children with abnormal results. Currently, stem cell transplantation is the most effective method for the treatment of CGD in most patients.

Acknowledgments

We gratefully acknowledge the Shandong Medical Imaging Research Institute and the Microbiology Laboratory of Shandong Provincial Hospital for their efforts.

Author contributions

Chunyan Guo: The main finisher of the article. Xing Chen: Responsible for the design of the article. Jingrong wang: Responsible for collecting imaging data. Fengqin Liu: Responsible for collecting imaging data. Yan Liang: Responsible for the collection of case data. Juan Yang: Responsible for the collection of case data. Fangfang Dai: Responsible for the follow-up of cases. Ning Ding: Responsible for the follow-up of cases.

References

- Thomsen IP, Smith MA, Holland SM, et al. A comprehensive approach to the management of children and adults with chronic granulomatous disease. J Allergy Clin Immunol Pract 2016;4:1082–8.
- [2] Chiriaco M, Salfa I, Di Matteo G, et al. Chronic granulomatous disease: clinical, molecular, and therapeutic aspects. Pediatr Allergy Immunol 2016;27:242–53.
- [3] Bonilla FA, Khan DA, Ballas ZK, et al. Practice parameter for the diagnosis and management of primary immunodeficiency. J Allergy Clin Immunol 2015;136:1186–205. e1181-1178.
- [4] Marciano BE, Spalding C, Fitzgerald A, et al. Common severe infections in chronic granulomatous disease. Clin Infect Dis 2015;60:1176–83.
- [5] Gennery AR. Editorial commentary: severe infections in patients with chronic granulomatous disease. Clin Infect Dis 2015;60:1184–5.
- [6] Lacerda-Pontes R, Gomes LN, Albuquerque RS, et al. The extended understanding of chronic granulomatous disease. Curr Opin Pediatr 2019;31:869–73.
- [7] Rawat A, Vignesh P, Sharma A, et al. Infection profile in chronic granulomatous disease: a 23-year experience from a tertiary care center in North India. J Clin Immunol 2017;37:319–28.
- [8] Arnold DE, Heimall JR. A Review of chronic granulomatous disease. Adv Ther 2017;34:2543–57.

- [9] Johnston RBJr. Clinical aspects of chronic granulomatous disease. Curr Opin Hematol 2001;8:17–22.
- [10] Leusen JH, de Klein A, Hilarius PM, et al. Disturbed interaction of p21rac with mutated p67-phox causes chronic granulomatous disease. J Exp Med 1996;184:1243–9.
- [11] Rider NL, Jameson MB, Creech CB. Chronic granulomatous disease: epidemiology, pathophysiology, and genetic basis of disease. J Pediatric Infect Dis Soc 2018;7(suppl_1):S2–5.
- [12] Boonyawat B, Suksawat Y, Pacharn P, et al. X-linked chronic granulomatous disease: initial presentation with intracranial hemorrhage from vitamin K deficiency in infant. Case Rep Pediatr 2018;2018:7041204.
- [13] Song E, Jaishankar GB, Saleh H, et al. Chronic granulomatous disease: a review of the infectious and inflammatory complications. Clin Mol Allerg 2011;9:10.
- [14] Yu JE, Azar AE, Chong HJ, et al. Considerations in the diagnosis of chronic granulomatous disease. J Pediatric Infect Dis Soc 2018;7 (suppl_1):S6-11.
- [15] Mahdaviani SA, Mohajerani SA, Rezaei N, et al. Pulmonary manifestations of chronic granulomatous disease. Expert Rev Clin Immunol 2013;9:153–60.
- [16] Salvator H, Mahlaoui N, Catherinot E, et al. Pulmonary manifestations in adult patients with chronic granulomatous disease. Eur Respir J 2015;45:1613–23.
- [17] Zhao SY, Zhao MJ, Liu H. Chronic granulomatous disease in children: diagnosis and treatment. Chin J Pediatr 2016;54:303–6.
- [18] Ying WJ, Wang XC, Sun JQ, et al. Clinical features of chronic granulomatous disease. Chin J Pediatr 2012;50:380–5.
- [19] Labrosse R, Abou-Diab J, Blincoe A, et al. Very early-onset inflammatory manifestations of X-linked chronic granulomatous disease. Front Immunol 2017;8:1167.
- [20] Siddiqui MR, Sanford T, Nair A, et al. Chronic colovesical fistula leading to chronic urinary tract infection resulting in end-stage renal disease in a chronic granulomatous disease patient. Urol Case Rep 2017;11:37–8.
- [21] Khanna G, Kao SC, Kirby P, et al. Imaging of chronic granulomatous disease in children. Radiographics 2005;25:1183–95.
- [22] King J, Henriet SSV, Warris A. Aspergillosis in chronic granulomatous disease. J Fungi 2016;2:15.
- [23] Shin KS, Lee MS. Concomitant use of corticosteroid and antimicrobials for liver abscesses in patients with chronic granulomatous disease. Korean J Pediatr 2016;59:196–201.
- [24] Candotti F. Gene transfer into hematopoietic stem cells as treatment for primary immunodeficiency diseases. Int J Hematol 2014;99:383–92.
- [25] Battersby AC, Cale AM, Goldblatt D, et al. Clinical manifestations of disease in X-linked carriers of chronic granulomatous disease. J Clin Immunol 2013;33:1276–84.
- [26] Kang EM, Malech HL. Gene therapy for chronic granulomatous disease. Method Enzymol 2012;507:125–54.
- [27] Slack MA, Thomsen IP. Prevention of Infectious complications in patients with chronic granulomatous disease. J Pediatric Infect Dis Soc 2018;7(suppl_1):S25–30.
- [28] Henrickson SE, Jongco AM, Thomsen KF, et al. Noninfectious manifestations and complications of chronic granulomatous disease. J Pediatric Infect Dis Soc 2018;7(suppl_1):S18–24.
- [29] Marciano BE, Rosenzweig SD, Kleiner DE, et al. Gastrointestinal involvement in chronic granulomatous disease. Pediatrics 2004;114:462–8.