

PCSK9 Manipulates Lipid Metabolism and the Immune Microenvironment in Cancer

Chaochu Cui^{1,*}, Aiwei Yan^{1,*}, Shengming Huang^{1,2}, Yifan Chen¹, Jinyu Zhao¹, Cixia Li^{1,3}, Xianwei Wang¹, Jianbo Yang^{1,4}

¹Henan Key Laboratory of Medical Tissue Regeneration, Xinxiang Medical University, Xinxiang, Henan, People's Republic of China; ²Rudong County Hospital of Traditional Chinese Medicine, Nantong, Jiangsu, People's Republic of China; ³School of Life Sciences and Technology, Xinxiang Medical University, Xinxiang, Henan, People's Republic of China; ⁴School of Medical Technology, Xinxiang Medical University, Xinxiang, Henan, 453003, People's Republic of China

*These authors contributed equally to this work

Correspondence: Xianwei Wang; Jianbo Yang, Email Wangxianwei1116@126.com; Yangjianbo12@126.com

Abstract: Cancer remains the foremost cause of mortality on a global scale. Immunotherapy has yielded remarkable outcomes in the fight against cancer and is regarded as one of the most crucial and promising therapeutic modalities. PCSK9, a critical target for plasma lipids control, has been extensively and deeply studied in multiple diseases. Currently, the functions of PCSK9 in cancer, particularly its immunomodulatory role, have been progressively revealed. PCSK9 is capable of modulating a variety of immune response throughout tumor progression by orchestrating lipid metabolism. Moreover, PCSK9 governs the cell fate of diverse immune cells, such as inflammatory factor signals, MHC signals, and TCR signals. This review comprehensively summarizes the current state of knowledge regarding the role and underlying mechanisms of PCSK9 in tumorigenesis, progression, immune escape, and drug resistance.

Keywords: PCSK9, lipid metabolism, immunotherapy, cancer

Introduction

Cancer is still a leading cause of mortality worldwide, and immunotherapy has achieved notable success in cancer treatment by modulating the function of the human immune system.¹ Abnormal lipid metabolism plays critical roles in cancer cells and the immune response in the progression of malignant tumors.² Dysregulation of lipid metabolism enables tumor cells to obtain the energy necessary for proliferation, invasion and the formation of favorable tumor microenvironment (TME). Furthermore, lipid metabolism disorders may impede immune cell function, thereby impacting the efficacy of immunotherapy.^{3–5}

Dysregulation of PCSK9 has been observed in a multitude of different cancers and affects the proliferation, invasion and drug resistance of tumor cells.⁶ PCSK9 is essential for regulating lipid metabolism, apoptosis, immune cell signaling, etc.^{7–10} As a soluble secreted serine protease that is synthesized primarily in the liver, PCSK9 functions through interactions with the low-density lipoprotein (LDL) receptor and CD36, thereby affecting cholesterol homeostasis, which may in turn be implicated in oncogenesis.¹¹

In addition to affecting lipid metabolism, PCSK9 also regulates cuproptosis, which is essential in TME. Cuproptosis correlates with immunosuppressive tumor microenvironment based on pan-cancer multiomics and single-cell sequencing analysis, Evolocumab attenuates myocardial ischemia/reperfusion injury by blocking PCSK9/LIAS-mediated cuproptosis of cardiomyocyte.¹² Besides, PCSK9 manipulates TME immune cells through a range of pathways, including MHC and PD-1 signaling. PCSK9 binds to MHC-I and then recruits adaptor proteins, which facilitate the internalization of the MHC-I, impeding the ability of cytotoxic T cells to recognize and eliminate tumor cells. PCSK9 may suppress the production of cytokines by T cells, which are essential for the anti-tumor immune response.¹³ This can influence

processes such as monocyte differentiation, DC maturation, and T-cell and natural killer cell functions.¹⁴ In the context of cancer, PCSK9 inhibitors can reverse the immunosuppressive effects of PCSK9 or displayed synergistic anti-tumor effects with immune checkpoint inhibitors, or even overcome therapy resistance in some cancers.^{15–17} This review aims to comprehensively summarize the current research progress of PCSK9 in tumor immunotherapy, covering its molecular mechanisms in immune pathways and specific impacts on different cancer types.

Lipid Metabolism Regulates Immunity

Lipid metabolism is a primary energy supply of immune cells. It has been demonstrated that lipids regulate the proliferation and differentiation of immune cells. For example, following the activation of lymphocytes, there is a rapid increase in cholesterol and fatty acid synthesis to accommodate the demands of rapid proliferation and differentiation.^{18,19} An increase in membrane cholesterol specifically promotes the differentiation of CD4⁺ T cells into the Th1 phenotype.³ The inhibition of HMG-CoA reductase, an enzyme crucial for cholesterol synthesis, can suppress the proliferation of immune cells through mitogen activation.^{20,21}

Moreover, lipids play vital roles in the structure and function of immune cell membranes, such as establishing immune synapses, which exerts a direct influence on TCR signal transduction that then affects immune cell activity.^{22,23} In T cells, activation and effector functions are facilitated by specialized signal centers located in the cytoplasmic membrane, which are composed of cholesterol and sphingolipids. CD8⁺ T cells constitute one primary mechanism by which the immune system combats cancer cells, with their proliferation and effector functions closely associated with cholesterol levels.^{23–26} Inhibition of ACAT1, the main enzyme involved in cholesterol esterification in CD8⁺ T cells, leads to an increase in the cholesterol content of CD8⁺ T-cell membranes and increased TCR clustering and signal transduction. This ultimately results in increased production of cytotoxic granules and cytokines by CD8⁺ T cells, along with increased cytotoxic capacity. Such enhancements facilitate the establishment of mature immune synapses and bolster the antitumor activity of CD8⁺ T cells.^{27–29}

Macrophages combat pathogens primarily through phagocytosis, a process involving the dynamic fusion and division of cellular membranes. Upon exposure to inflammatory stimuli, macrophages increase lipid synthesis through the activation of the mTOR pathway and increase sterol regulatory element-binding protein (SREBP) 1a activity, thereby increasing phagocytic ability.^{30,31} Moreover, during atherogenesis, monocytes differentiate into macrophages in the subendothelial space and internalize oxidized LDL (oxLDL) through scavenger receptors such as SR-A and CD36.³² Inhibiting cholesterol synthesis has been demonstrated to diminish macrophage-driven inflammation.^{33–35} NFATc3 deficiency in macrophages potentiates SR-A- and CD36-mediated lipid uptake, both of which are targeted by PCSK9, thereby reducing the development of atherosclerotic plaques.³⁶

Additionally, elevated cholesterol levels can activate NK cells, which can effectively eliminate liver cancer cells.³⁷ Following NK cell activation, there is an upregulation of intracellular SREBP and mTORC1, which enhances aerobic glycolysis and OXPHOS metabolic rates. However, excessive fatty acid uptake by cells can activate the PPAR- γ /PPAR δ signaling pathway, which has been demonstrated to not only inhibit NK cell functions but also induce cellular toxicity.³⁸ Therefore, an appropriate lipid level is conducive to the killing function of NK cells.

PCSK9 Manipulates Lipid Metabolism

It is widely accepted that PCSK9 is critical for maintaining lipid metabolism homeostasis.³⁹ PCSK9 is synthesized primarily in hepatocytes, which are the major source of PCSK9 in circulating blood. While some PCSK9 can be synthesized in the kidney, intestine, macrophages, and endothelial cells, this type of PCSK9 is considered to be used primarily for autocrine or intracellular functions.⁴⁰ The structure of PCSK9 consists of a signal peptide, a precursor domain, a catalytic domain and a carboxyl terminal domain. The zymogen PCSK9 (approximately 74 kDa) undergoes self-catalyzed cleavage, resulting in the maturation of PCSK9 (approximately 60 kDa), which is then secreted into the extracellular space. PCSK9 has a variety of biological functions, such as regulating plasma lipid homeostasis, liver regeneration, insulin production, neurodevelopment, cardiovascular diseases and the tumor immune response.^{41–46}

PCSK9 influences lipid metabolism primarily in diverse manners (Figure 1). PCSK9 can promote the endocytosis and degradation of LDL receptors (LDLRs), thereby inhibiting the clearance of LDLC from the blood.⁴⁷ PCSK9 also

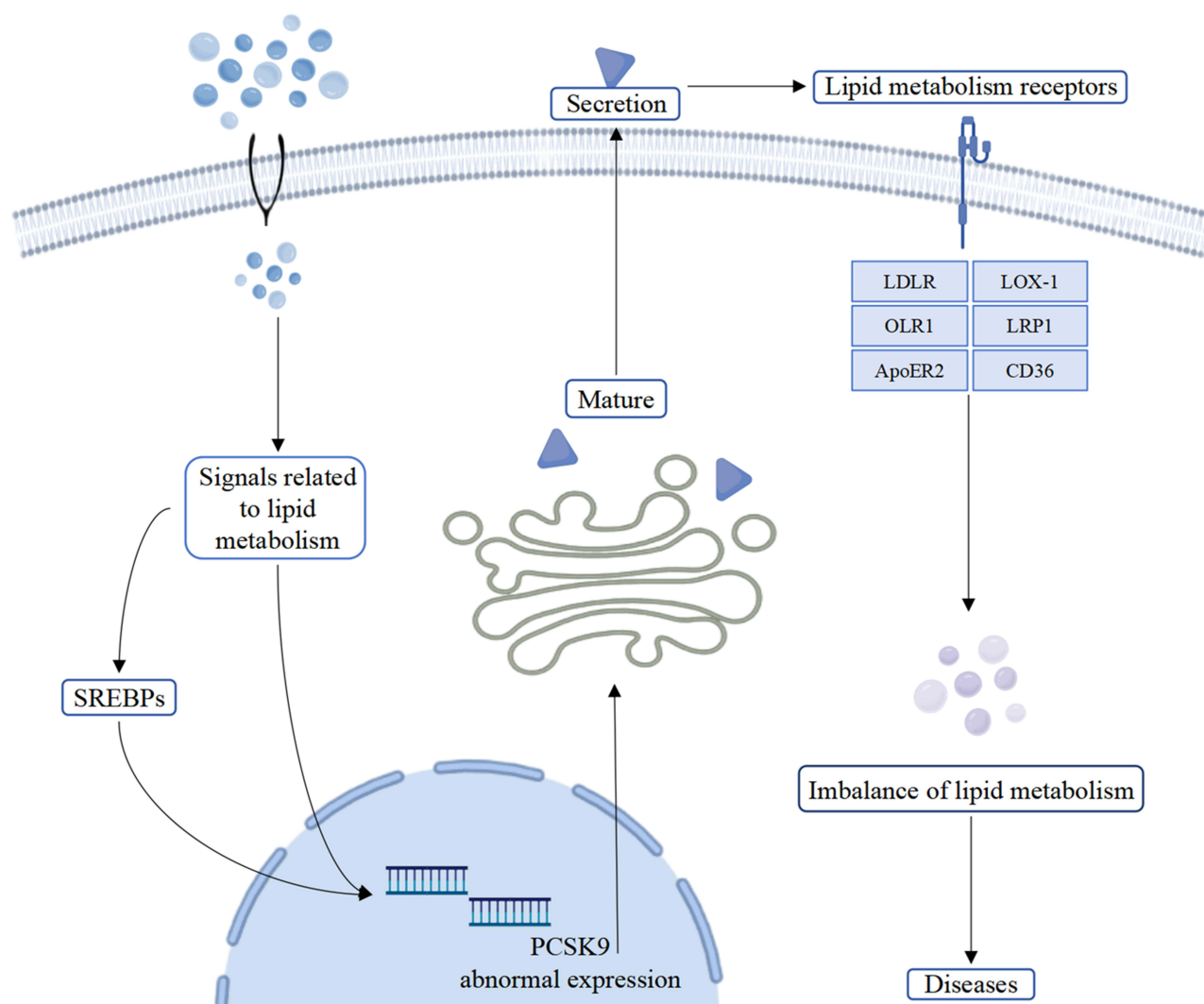


Figure 1 The mechanism by which PCSK9 regulates lipid metabolism. The level of PCSK9 on the cell surface is influenced by blood cholesterol levels. After undergoing maturation through Golgi modifications, PCSK9 becomes active on the cell membrane surface. It then binds to the low-density lipoprotein receptor (LDLR) and facilitates its degradation via lysosomal pathways. Additionally, PCSK9 plays a role in modulating the expression of SREBP-2, ApoER2, and HMG-CoA reductase, thereby impacting the accumulation of lipids within cells.

upregulates the expression of lectin-like oxidized low-density lipoprotein receptor 1 (LOX-1), promoting the uptake of oxidized low-density lipoprotein cholesterol (ox-LDL) by macrophages.⁴⁸ Furthermore, SREBPs serve as crucial transcription factors that govern lipid metabolism. These proteins play pivotal roles by modulating PCSK9, which in turn impacts the synthesis of cholesterol and fatty acids. Consequently, the regulation of PCSK9 by SREBPs can lead to an increase in lipid levels.⁴⁹ Moreover, PCSK9 regulates the expression of apolipoprotein E receptor 2 (ApoER2), low-density lipoprotein receptor-associated protein 1 (LRP1), and oxidized low-density lipoprotein (lectin-like) receptor 1 (OLR1), which are proteins with central roles in lipid metabolism, thus mediating neuronal cell death. Therefore, PCSK9 is an important target for the treatment of LDL-mediated cell dysregulation.^{50,51} Since the discovery that PCSK9 is related to lipid metabolism, PCSK9 has quickly become a research target for the treatment of diseases.^{52–54}

PCSK9 Mediated-Immune Response

Interestingly, it was recently demonstrated that PCSK9 is also a promising target for cancer therapy.^{13,55} Investigations have revealed diverse immunomodulatory capabilities of PCSK9 through modulation of the expression of cytokines or immune signal transduction proteins (Figure 2 and Table 1).

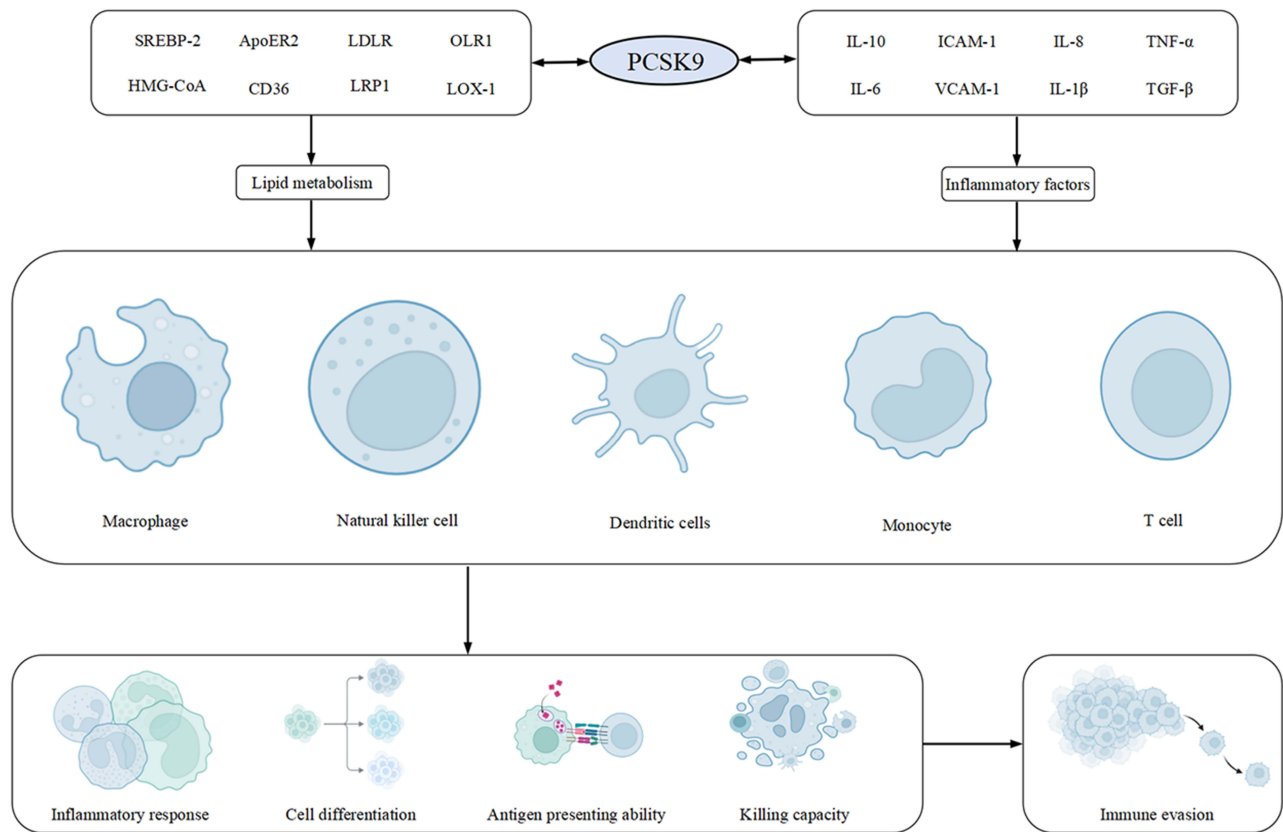


Figure 2 The role of PCSK9 in the immune response. The role of PCSK9 in the immune response influences the immune response in the tumor microenvironment and the differentiation, antigen presentation and killing ability of a variety of immune cells by interacting with many lipid metabolism-related molecules and inflammatory factors.

PCSK9 Regulates Inflammation

PCSK9 has been found to regulate nonspecific immune responses. During viral infections, PCSK9 levels increase, resulting in the induction of localized inflammatory reactions by modulating various inflammatory mediators.⁸ In certain cell types and under specific physiological or pathological conditions, PCSK9 can promote inflammation. Elevated PCSK9 levels can increase the incidence of liver cancer in hepatitis patients.^{9,56} Moreover, monoclonal antibodies (mAbs) targeting PCSK9 significantly alter the expression of the chemokine receptors CX3CR1, CXCR6, and CCR2 on certain leukocyte subsets. Concurrently, the plasma levels of the proinflammatory cytokine TNF-α are reduced, whereas those of the anti-inflammatory cytokine IL-10 are increased.⁵⁷

Table I The Regulation of Immune Cells by PCSK9

Cell type	Disease Type	Signaling Pathway	Reference
Monocytes	Coronary Artery Disease	Inflammatory Cytokines	[61]
	Atherosclerosis	Cholesterol Metabolism	[62]
	Hepatocellular Carcinoma		[63]
	Familial Hypercholesterolaemia		[64]
Macrophages	Hepatocellular Carcinoma	Cholesterol Metabolism	[71]
	Breast Cancer	CXCL12	[77]
	Skin Damage	IRF3	[78]

(Continued)

Table 1 (Continued).

Cell type	Disease Type	Signaling Pathway	Reference
Dendritic Cells	Hepatocellular Carcinoma	MHC-I	[79]
	Melanoma	TLR	[80]
	Atherosclerotic	Inflammatory Cytokines	[81]
	Subarachnoid Hemorrhage	SIRT6	[82]
	Systemic Lupus Erythematosus	CD86/HLA-DR	[83]
Natural Killer Cells	Neuroblastoma	Cholesterol Metabolism	[41]

Conversely, in other situations, PCSK9 can play a role in suppressing inflammation. In endothelial cells, PCSK9 may interact with specific membrane proteins and modulate the production of anti-inflammatory mediators. By inhibiting certain proinflammatory signaling cascades, such as the MAPK pathway under certain conditions, PCSK9 can reduce the expression of adhesion molecules and chemokines. This results in a decreased recruitment of immune cells to the site of potential inflammation, thereby dampening the overall inflammatory response.^{58,59} PCSK9 intervention has also been demonstrated to be effective in mitigating the circulating levels of chemokines such as IL-8/CXCL8 and eosinophilic chemokine-2/CCL24. This has been shown to diminish the activation of neutrophils and eosinophils. Moreover, the inhibition of PCSK9 has been shown to reduce the expression of adhesion molecules such as ICAM-1 and VCAM-1, along with the chemokines CX3CL1 and CXCL16, thereby inhibiting interactions between leukocytes and endothelial cells. This results in a reduction in systemic inflammation and endothelial dysfunction.^{57,60}

Inflammation exhibits a dual-edged sword effect in tumors. On one hand, chronic inflammation can create a microenvironment conducive to tumor initiation, growth, and metastasis by promoting cell proliferation, angiogenesis, and immune evasion. On the other hand, in some cases, an appropriate inflammatory response can also trigger anti-tumor immune reactions. This complex role of inflammation further complicates the function of PCSK9 in tumors.

PCSK9 Regulates Monocytes

PCSK9 plays a pivotal role in the differentiation of monocytes into macrophages by modulating their surface receptors and signaling cascades. For example, the regulatory impact of PCSK9 on LDLRs influences the cholesterol metabolism of monocytes, thereby shaping their differentiation trajectories.⁶¹ The overexpression of PCSK9 has been demonstrated to amplify the inflammatory response of monocytes, increasing the secretion of proinflammatory cytokines such as IL-1 β , IL-6, and TNF- α , which can promote tumor progression and metastasis.⁶²

PCSK9 has also been shown to enhance the immunosuppressive capabilities of monocytes, steering their differentiation toward immunosuppressive M2-type macrophages. These M2 macrophages secrete anti-inflammatory cytokines and facilitate cancer cell immune escape.⁶³ In the context of cardiovascular disease, circulating PCSK9 levels reflect the distribution of monocyte subsets in individuals with stable coronary artery disease. Moreover, monoclonal antibodies targeting PCSK9 have been demonstrated to mitigate the proinflammatory characteristics of monocytes, consequently reducing the expression of inflammatory markers.^{61,64}

PCSK9 Regulates Macrophages

Elevated levels of PCSK9 are correlated with unfavorable outcomes in diverse cancers, potentially by modulating the function of immune cells, including tumor-associated macrophages (TAMs).⁴¹ For example, in macrophages, PCSK9 can bind to cell-surface receptors, such as LRP1. This activates intracellular signaling pathways, like the nuclear factor- κ B (NF- κ B) mediated upregulation of pro-inflammatory cytokines, such as TNF- α , IL-1 β , and IL-6. These cytokines then initiate and amplify the inflammatory response, contributing to the development and progression of various inflammatory-related diseases, including those associated with tumor microenvironments.^{73–75} Meanwhile, PCSK9 upregulates

the expression of genes associated with M1 polarization, such as iNOS. iNOS produces nitric oxide (NO), a potent antimicrobial and pro-inflammatory molecule. In contrast, it may downregulate genes associated with M2 polarization.^{76–78}

Supernatant from PCSK9-overexpressing MHCC97H cells inhibited THP-1 macrophage migration and M2-like TAMs polarization. In contrast, supernatant from PCSK9-silenced Huh7 cells enhanced these functions. In tumor bearing mice, PCSK9 overexpression promoted OX40L, suppressing M2-like TAM polarization.⁶³ Arenobufagin has been demonstrated to impede the progression of hepatocellular carcinoma by modulating PCSK9-mediated cholesterol metabolism, which in turn influences the polarization of TAMs. This process involves the suppression of PCSK9 expression, the regulation of cholesterol metabolism, the promotion of M1-type macrophage polarization, and the amplification of the antitumor immune response.⁷¹

Efferocytosis, mediated by macrophages, is a process to eliminate apoptotic cells. This is fundamental for maintaining homeostasis, preventing autoimmune diseases, and facilitating inflammation resolution. In cancer, efferocytosis often becomes dysregulated, while defects in efferocytosis can lead to the accumulation of apoptotic cells in TME. Cancer cells can actively interfere with efferocytosis to evade immune surveillance.^{79–81} In vascular endothelial cells, recombinant PCSK9 protein has been shown to induce efferocytic defects and downregulate the expression of the efferocytosis receptor MerTK. During the aging process of endothelial cells (ECs), PCSK9 levels increase while MerTK levels decline, and knockout of the PCSK9 gene can partially restore the associated functions. It likely involves the induction of ROS production, which inhibits MerTK expression and causes its cleavage. Additionally, PCSK9 may activate the MAPK pathway and NF- κ B signaling, thereby indirectly suppressing efferocytosis through an inflammatory response.^{75,76}

High expression of CXCL12 in breast cancer tissues is linked to poor prognosis and is associated with the extent of infiltration by various immune cells, especially in high-risk patients, who have increased infiltration of M2-like macrophages. Through targeted gene screening, PCSK9 was identified as a key factor correlated with the infiltration of TAMs and regulatory T cells (Tregs), both of which typically exhibit immunosuppressive functions and can facilitate tumor immune evasion and growth.⁷⁷ PCSK9 also promotes the transport of double-stranded DNA fragments into macrophages, resulting in interferon regulatory factor 3 (IRF3) and STING activation.⁷⁸ Understanding the complex interactions between PCSK9 and macrophage is essential for developing targeted therapeutic strategies.

PCSK9 Regulates DCs

The regulatory effects of PCSK9 on DCs are manifested primarily through its ability to modulate their maturation process, as well as its influence on cytokine and miRNA expression. The combination of chemotherapy drugs with PCSK9 inhibitors has been shown to enhance the maturation of DCs within tumor sites. Furthermore, the expression level of PCSK9 is closely associated with the activation of DCs.^{79,80}

In the context of other systemic diseases, the exposure of DCs to oxLDL has been shown to prompt the maturation process, leading to T-cell activation and potentially fostering a proinflammatory state.⁸¹ By inhibiting DC maturation, PCSK9 inhibitors curtail the production of proinflammatory cytokines such as TNF- α , IL-1 β , and IL-6 while simultaneously promoting the secretion of anti-inflammatory cytokines, including TGF- β and IL-10. This action helps prevent the polarization of Th1 and/or Th17 cells and instead encourages the emergence of T regulatory cells.^{82,83}

PCSK9 Regulates Natural Killer (NK) Cells

Following the genetic ablation of PCSK9, there is a notable increase in the number of NK cells in close proximity to the tumor. Furthermore, PCSK9 controls the antitumor efficacy of NK cells by affecting their cholesterol metabolism. Through the attenuation of PCSK9 activity, PCSK9 inhibitors increase the cholesterol content within NK cells, thereby reinforcing their antitumor capabilities.^{6,41} Analyses of pancreatic adenocarcinoma (PAAD) patients and normal controls have revealed marked disparities in the infiltration of NK cells within tumor tissues compared to normal tissues. Notably, PCSK9 expression exhibits a correlation with the infiltration level of NK cells, with elevated PCSK9 expression inversely associated with the infiltration of activated NK cells. This inverse relationship suggests that PCSK9 may

impede the infiltration of NK cells, thereby potentially compromising the immune surveillance and anti-tumor cytotoxicity of NK cells against tumor cells. Subsequent experimental investigations further validated that modulating PCSK9 levels can effectively regulate NK cell function, ultimately influencing tumorigenesis and tumor progression. These findings highlight the crucial role of PCSK9 - NK cell interactions in the pathogenesis of PAAD and underscore the potential of targeting this axis for therapeutic intervention.^{84,85}

Currently, the direct relationship between PCSK9 and NK cells across diverse cancers remains unclarified. However, within the framework of immune regulation, both PCSK9 and NK cells exert influence on disease development and treatment strategies.⁸⁵ They thus hold promise as potential therapeutic targets, warranting further investigation to fully understand their roles and exploit their therapeutic potential.

PCSK9 Regulates T Cells

In the complex microenvironments of the immune system and tumors, PCSK9 has emerged as a crucial regulator with far-reaching implications. Unraveling the precise mechanisms by which it exerts its differential effects on T cells and tumor cells has become a focal point of recent research endeavors (Figure 3).

When considering the interaction between PCSK9 and LDLR, it becomes evident that this binding event has profound consequences. LDLR, a key player in lipid uptake and homeostasis, normally undergoes a recycling process to return to the plasma membrane for continued function. However, when PCSK9 latches onto it, this recycling pathway is effectively blocked. This not only disrupts normal lipid metabolism but also directly affects the function of cytotoxic T cells. The TCR, which is integral for T-cell activation and antigen recognition, is also affected by this perturbed recycling, leading to impaired effector function. As a result, the ability of cytotoxic T cells to mount an effective immune response against tumor cells or pathogens is compromised.^{55,83,86}

In the context of tumor cells, the physical association of PCSK9 with major histocompatibility complex class I (MHC I) initiates a series of intracellular events. MHC-I molecules are responsible for presenting antigenic peptides to cytotoxic T cells, thereby alerting the immune system to the presence of abnormal or tumorigenic cells. PCSK9, however, promotes the trafficking of MHC-I to lysosomes within tumor cells. Once inside lysosomes, MHC-I undergoes degradation, leading to a significant reduction in its surface expression. This devious tactic employed by tumor cells, facilitated by PCSK9, allows them to evade detection and destruction by the immune system, providing a survival advantage and promoting tumor progression.^{13,87,88}

The impact of PCSK9 on intracellular cholesterol levels is yet another aspect that demands attention. By interfering with lipid metabolism pathways, PCSK9 causes a buildup of cholesterol within cells. Macrophages, which are highly sensitive to changes in their lipid environment, are particularly affected. Elevated cholesterol levels lead to lipid accumulation, altering the macrophage phenotype and function. These lipid-laden macrophages may exhibit impaired phagocytic activity or secrete factors that contribute to the tumor microenvironment, either directly or indirectly promoting tumor growth.^{42,89}

The role of PCSK9 in modulating T-cell activation is multifaceted. Through the activation of MAPK, which subsequently phosphorylates and activates NFATc1, PCSK9 fine-tunes the activation state of cytotoxic T cells. This signaling cascade is a critical determinant of whether T cells become fully activated and carry out their effector functions.^{71,90,91} On the other hand, the PI3K/Akt pathway represents a double-edged sword. Some therapeutic interventions have harnessed the power of this pathway to protect T cells from apoptosis, ensuring their survival and enhancing the immune response. In contrast, PCSK9 can act as an antagonist, inhibiting the PI3K/Akt pathway and tipping the balance toward apoptosis, potentially dampening immune defense mechanisms.^{92,93}

With respect to Th cell differentiation, PCSK9 influences Th cell differentiation through activation of the NF- κ B signaling pathway. This pathway is a master regulator of gene expression, dictating the fate of Th cells. By activating NF- κ B, PCSK9 nudges Th cells toward a particular differentiation pathway, which can have implications for the type of immune response. For example, it may skew the response toward a more proinflammatory or anti-inflammatory phenotype, depending on the context.⁹⁴ Furthermore, the upregulation or activation of the TLR-4/NF- κ B pathway by PCSK9 has far-reaching consequences. This not only fuels vascular inflammation, which can create a conducive environment for tumor growth and metastasis but also further drives Th cell differentiation. When engaged, the TLR-

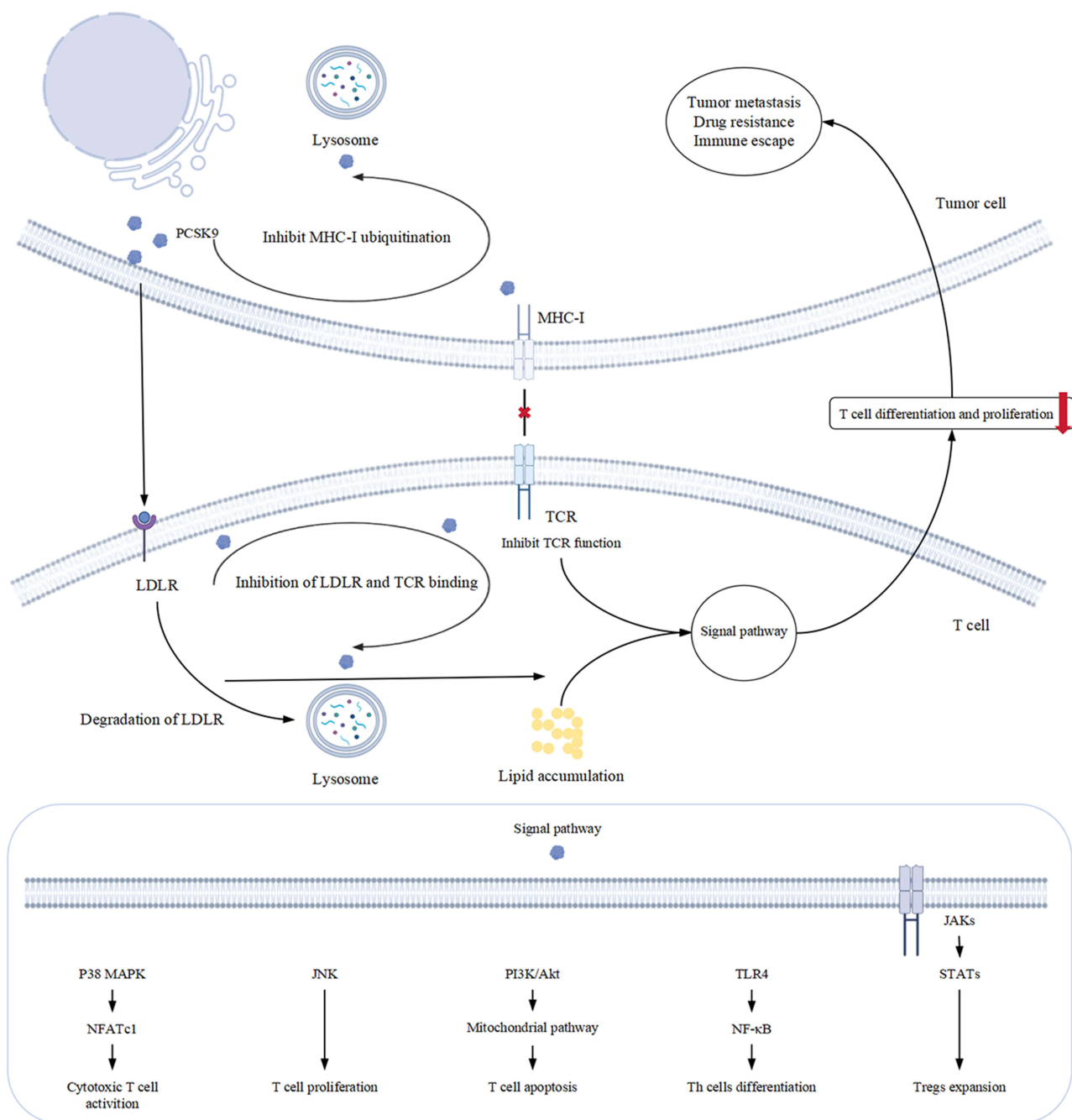


Figure 3 Mechanisms of the regulatory effects of PCSK9 in T cells and tumor cells. The binding of PCSK9 to LDLR impedes the recycling of LDLR and TCR back to the plasma membrane, consequently suppressing the effector function of cytotoxic T cells. By physically associating with MHC-I, PCSK9 instigates lysosomal degradation within tumor cells, which in turn diminishes the surface presentation of MHC-I, thereby facilitating tumor immune evasion. Moreover, PCSK9 triggers an increase in intracellular cholesterol levels, culminating in lipid accumulation within macrophages. PCSK9 modulates the activation of cytotoxic T cells via MAPK-mediated activation of NFATc1. Certain treatments can prevent T-cell apoptosis and promote T-cell survival through the PI3K/Akt pathway. Conversely, PCSK9 can promote apoptosis by inhibiting the PI3K/Akt pathway. PCSK9 drives Th cell differentiation by activating the NF- κ B signaling pathway. Additionally, PCSK9 augments vascular inflammation and promotes Th cell differentiation by upregulating or activating the TLR-4/NF- κ B pathway. Finally, PCSK9 facilitates the expansion of Tregs through the JAK/STAT pathway.

4 receptor initiates a signaling cascade that converges with the NF- κ B pathway, leading to a complex interplay of gene activation and cell fate determination.^{95,96}

PCSK9 promotes the amplification of Tregs through the JAK/STAT pathway, thus affecting immune balance. Tregs play a crucial role in maintaining immune homeostasis, preventing overactive immune responses that could lead to autoimmunity.^{15,97–99} However, an overabundance of Tregs, potentially induced by PCSK9, could also suppress the

immune response against tumors, allowing them to thrive. Reducing the expression of PCSK9 in cancer cells leads to increased clonal expansion of CD8⁺ T cells, while the number of Tregs remains unchanged and the expression of PD-L1 does not change significantly, thus enhancing the immunotherapy response. Single-cell sequencing analysis revealed that PCSK9 may influence the functionality of circulating T cells via the PD-1/CTLA-4 pathway.¹⁰⁰ Overall, understanding these intricate mechanisms of PCSK9 is essential for devising novel therapeutic strategies that can either target PCSK9 directly or manipulate its downstream pathways to combat diseases such as cancer and autoimmune disorders.

In addition to its direct regulatory relationship, PCSK9 can also indirectly regulate T cells. Inhibition of PCSK9 can promote DCs infiltration and the expression of MHC-II, improving the activation of CD8⁺ T cells within the TME.⁴³ In the TME, PCSK9 downregulates the expression of LDLR and TCR signaling in CD8⁺ T cells, which ultimately blunts the functions of cytotoxic T lymphocyte effectors. The elimination of PCSK9 or administration of PCSK9 inhibitors has the potential to alleviate this inhibitory effect on CD8⁺ T cells, thereby increasing their antitumor activity and curtailing tumor progression. Combining PCSK9 inhibitors with OVA-II tumor vaccines can enhance the therapeutic efficacy of monotherapy.^{43,101}

The intracellular cholesterol content of $\gamma\delta$ T lymphocytes is positively associated with the activation of the TCR signaling pathway, cell activation, and proliferation. PCSK9 influences the antitumor activity of CD8⁺ T cells through the LDLR/TCR axis, thereby affecting tumor progression. The combination of PCSK9 inhibitors and PD-1-related drugs has synergistic antitumor effects.^{44,55}

Elevated levels of PCSK9 can attenuate the efficacy of anti-PD-1 immunotherapy in patients diagnosed with advanced non-small cell lung cancer (NSCLC). Moreover, the concurrent administration of PCSK9 inhibitors and anti-CD137 agonists has been shown to increase the recruitment of CD8⁺ and GzmB⁺ CD8⁺ T cells, thereby extending the lifespan of lung cancer model mice.¹⁰² In the context of colorectal cancer, the inhibition of PCSK9 expression in conjunction with the PD-1/PD-L1 pathway can restrain tumor growth by encouraging the infiltration of cytotoxic T cells and the expression of IFN- γ .¹⁶ Furthermore, the combination of evolocumab and DOX increased the proportions of mature DCs, tumor-infiltrating CD8⁺ T cells, and NK cells, thereby enhancing the efficacy of immunotherapy in hepatocellular carcinoma.⁷⁹

Furthermore, cholesterol metabolism regulates the activation, clonal expansion, and effector functions of CD8⁺ T cells, particularly with respect to their antitumor efficacy.^{27,103} Within the TME, PCSK9 has been identified as an inhibitory factor of cholesterol metabolism in CD8⁺ T cells. This results in a reduction in the antitumor capabilities of these T cells due to a decrease in their cholesterol levels. Notably, two monoclonal antibodies targeting PCSK9, namely, evolocumab and alirocumab, which have been granted approval for clinical use in lowering cholesterol levels, enhances the response to tumor immunotherapy.^{104,105} PCSK9 knockout tumors exhibit a marked increase in the number of CD8⁺ cytotoxic T cells (CTLs), CD4⁺ T-helper (TH) cells, and $\gamma\delta$ T cells within the tumor.^{98,106} These findings suggest that PCSK9 deficiency may hinder tumor growth by enhancing the antitumor activity of T cells.

Mechanisms of PCSK9 and the Application of PCSK9 Inhibitors in Tumor Therapy

Breast Cancer

The levels of PCSK9 exhibit progressively increase as breast cancer advances. Remarkably, following tumor regression, PCSK9 levels decrease to levels similar to those of the control group.¹⁰⁷ The overexpression of PCSK9 is correlated with increased breast cancer cell proliferation and invasion and avoidance of apoptosis, and a reduction in PCSK9 is correlated with a decreased risk of breast cancer.^{108,109}

Pseurotin A has been demonstrated to effectively inhibit the secretion and interaction of PCSK9 with LDLR. This action inhibits the uptake of LDL-C, thereby limiting the growth and metastatic potential of breast cancer.¹¹⁰ These findings strongly imply that PCSK9 has considerable potential as a diagnostic and prognostic biomarker in the context of breast cancer, heralding new avenues for enhanced disease management and patient outcomes.

Melanoma

Either genetic ablation of the PCSK9 gene or the administration of a PCSK9 inhibitor will delay melanoma progression.¹¹¹ This compelling evidence implies that curbing PCSK9 activity could exert a direct inhibitory effect on melanoma growth. PCSK9 plays a pivotal role in modulating cholesterol homeostasis within the TME. Cholesterol accumulation has been implicated in furnishing the requisite lipid substrates that fuel rapid tumor cell proliferation. By impeding PCSK9 function, alterations in the lipid metabolic circuitry impede the growth and expansion of melanoma cells.^{6,112} In parallel, PCSK9 knockout mice engrafted with B16F1 mouse melanoma cells has also been examined. Intriguingly, in this model, cancer cells were found to have metastasized to the liver, accompanied by a pronounced increase in hepatic interstitial and tumor cell apoptosis. This phenomenon is plausibly attributed to the activation of the TNF- α pathway.^{113,114}

Prostate Cancer

Elevated PCSK9 expression in prostate cancer (PC) is correlated with a reduced disease-specific survival time. This finding implies that the expression level of PCSK9 could serve as a predictive biomarker for the prognosis of PC patients.¹¹⁵ Suppressing PCSK9 in prostate cancer cells can confer protection against ionizing radiation (IR)-induced damage. This is achieved through promoting cell viability and concomitantly inhibiting apoptosis and the activity of matrix metalloproteinases (MMPs). Upon IR exposure, characteristic changes, such as increases in CytC, caspase-3, and Bax levels, along with decreases in Bcl-2 levels, are observed. Notably, siRNAs targeting PCSK9 lead to increased radioresistance. These results suggest that PCSK9 might modulate the radiosensitivity of prostate cancer cells by orchestrating mitochondrial signaling pathways.¹¹⁶

Furthermore, treatment with Pseurotin A elicited dose-dependent suppression of migration, colony formation, and PCSK9 expression in PC cell lines. Moreover, Pseurotin A effectively curbed the *in vivo* progression of PC-3 cells orthotopically xenografted in nude mice and thwarted both locoregional and distant tumor recurrences following primary tumor surgical excision.¹¹⁷

Colorectal Cancer

By modulating the epithelial–mesenchymal transition (EMT) and PI3K/AKT signaling pathways within tumor cells, PCSK9 plays pivotal roles in both the progression and metastasis of colon cancer. Concurrently, by controlling inhibitory factors and lactate levels, it significantly impacts the phenotype polarization of macrophages.¹¹⁸ Moreover, PCSK9 expedites the development of colon cancer via the activation of the JAK2/STAT3/SOCS3 signaling cascades.¹¹⁹ In the context of APC/KRAS-mutant colorectal cancer, PCSK9 promotes tumor progression by facilitating cholesterol uptake and triggering associated signaling cascades.¹²⁰ Berberine, a natural inhibitor of PCSK9, has the capacity to influence tumorigenesis and progression by regulating the expression of LDLR, HNF-1 α , EGF-A, and SREBP-2.¹²¹

Liver Cancer

HCV-positive patients typically exhibit elevated plasma PCSK9 protein levels compared with those of HCV-negative individuals.⁸ Interestingly, tumor tissue from HCC patients presents significantly lower PCSK9 mRNA and protein levels and significantly higher LDLR levels than adjacent cirrhotic liver tissue.¹²² Moreover, serum PCSK9 protein levels are significantly higher in patients with HCC than in patients with chronic liver disease without HCC, whereas high expression of PCSK9 is associated with poorer overall survival (OS).¹²³ In HepG2 cells, PCSK9 expression is correlated with the level of TNF- α .¹²⁴ An increase in PCSK9 expression is associated with drug resistance in certain cancers, contributing to their resistance to chemotherapeutic drugs.^{93,125} A reduction in PCSK9 results in cholesterol accumulation within liver cancer cells, augmenting the proinflammatory actions of NF- κ B and thereby limiting tumor growth.¹²⁶ Moreover, the loss of PCSK9 has been shown to result in elevated cholesterol levels in the liver, which can promote inflammation and carcinogenesis. This illustrates the intricate role of PCSK9 in liver disease and liver cancer, where it may confer some protective effects while potentially contributing to disease progression in certain instances.¹²⁷ PCSK9 interacts with GSTP1, thereby inhibiting the JNK signaling pathway and regulating apoptosis in HCC.¹²⁸

Nanoparticles containing berberine can enhance the regulatory effects of berberine on PCSK9 expression in HepG2 cells.¹²⁹ Curcumin can downregulate the expression of PCSK9 by affecting the nuclear translocation of HNF-1 α , resulting in increased uptake of LDL in HepG2 cells.¹³⁰ Increasingly, extracts, such as *Sparassis crispa*, *Protium heptaphyllum* gum resin extract, *Scutellaria baicalensis*, olive oil phenol extracts, and Welsh onion extract, are being shown to affect the occurrence and development of tumors by regulating the expression level of PCSK9.^{131–135}

Ovarian Cancer

PCSK9 might also play a part in governing ovarian cancer cell fate. The inhibition of annexin A11, a protein implicated in conferring chemoresistance to ovarian cancer cells, could be under the control of PCSK9. These findings suggest that the latent capacity of PCSK9 modulates both apoptosis and drug resistance in ovarian cancer.¹³⁶ Moreover, synergistic interactions have been revealed between PCSK9 and its inhibitors and between PCSK9 and mTOR inhibitors. By perturbing ovarian cancer cell metabolism and signaling cascades, this combination amplifies therapeutic efficacy, opening new avenues for treatment optimization.¹³⁷

Gastric Cancer

Both PCSK9 and transcriptional enhancer factor domain-containing protein 4 (TEAD4) are markedly overexpressed in stomach cancer (STAD). These proteins collaborate to bolster cancer stem cell (CSC) traits, with fatty acid metabolism (FAM) serving as a crucial mediator. Consequently, targeting the TEAD4/PCSK9/lipid axis could herald a novel therapeutic paradigm for quelling CSCs in STAD.¹³⁸ Additionally, PCSK9 has been shown to fuel gastric cancer metastasis and impede apoptosis by activating the MAPK signaling pathway and upregulating HSP70 expression.¹³⁹ Intriguingly, in esophageal cancer patients, those harboring higher levels of anti-PCSK9 antibodies appear to be better in the postoperative period.¹⁴⁰

Other Cancers

In addition to its well-established role in modulating lipid metabolism, PCSK9 has emerged as a key regulator of cell death in lung adenocarcinoma, operating through diverse molecular pathways. For example, in A549 cells, PCSK9 orchestrates apoptotic processes by affecting mitochondrial and endoplasmic reticulum stress responses.^{141,142} PCSK9 could also potentially hold a pivotal position in the genesis, progression, and therapeutic management of cholangiocarcinoma.^{143–145} However, the precise mechanistic underpinnings and the exact nature of this correlation await further elucidation.^{146,147} Nevertheless, certain malignancies, such as sebaceous carcinoma and pancreatic cancer, have established links to lipid metabolism, yet their associations with PCSK9 remain largely unexplored.^{148,149} These uncharted territories require further investigation to fully elucidate the potential roles of PCSK9 in the oncogenic processes of these cancers.

Summary

PCSK9 plays complex and essential roles in cancer. PCSK9 not only indirectly influences the TME through regulating lipid metabolism but also directly affects immune cells, thereby affecting their occurrence, development, and therapeutic effects. The facts that PCSK9 has been extensively studied, and that numerous pharmacological agents have been extensively used and have achieved remarkable outcomes in the treatment of multiple diseases in the clinic, will provide many useful clues for its study in cancer.

Although PCSK9 has demonstrated significant potential across multiple cancers and promising progress has been made in the research of PCSK9, the role of PCSK9 in cancer has not been fully elucidated and further research is needed. For example, the specific mechanism of action of PCSK9 may differ among various tumor types. Furthermore, drug delivery of PCSK9 inhibitors to reach the tumor microenvironment effectively is also one potential challenge for cancer treatment. This is due to the unique and complex architecture of tumors and the presence of physiological barriers in TME that is completely different to liver.

Efforts should focus on delving deeper into the mechanisms by which PCSK9 regulates tumor immunity. This includes exploring the interactions between PCSK9 and other molecules to uncover its comprehensive role in cancer. In

addition, resistance mechanisms may develop, which may due to alternative ways are activated to evade the immune system or maintain their growth despite PCSK9 inhibition. Another important aspect is the clinical feasibility of combining PCSK9 inhibitors with existing cancer treatments, such as potential drug-drug interactions. Development of next-generation inhibitors with enhanced specificity and reduced off-target effects represents a key frontier. In the future, artificial intelligence may be used to make new breakthroughs in PCSK9-related drug development, tumor resistance, drug delivery, and drug combination. Such endeavors will definitely lay a solid theoretical foundation for the formulation of novel treatment strategies that target PCSK9 in cancer immunotherapy.

Funding

This work was sponsored by the Natural Science Foundation of Henan (No.202300410309), the Key Science and Technology Program of Henan Province (No.242102111037), and the Xinxiang Medical University Grant for Talents (No.XYBSKYZZ201828).

Disclosure

The authors declare that they have no conflicts of interest in this work.

References

1. Zhang Y, Zhang Z. The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications [J]. *Cell mol Immunol*. 2020;17(8):807–821. doi:10.1038/s41423-020-0488-6
2. Bleve A, Durante B, Sica A, et al. Lipid metabolism and cancer immunotherapy: immunosuppressive myeloid cells at the crossroad [J]. *Int J mol Sci*. 2020;21(16). doi:10.3390/ijms21165845.
3. Perucha E, Melchiotti R, Bibby JA, et al. The cholesterol biosynthesis pathway regulates IL-10 expression in human Th1 cells [J]. *Nat Commun*. 2019;10(1):498. doi:10.1038/s41467-019-08332-9
4. Li K, Deng Y, Deng G, et al. High cholesterol induces apoptosis and autophagy through the ROS-activated AKT/FOXO1 pathway in tendon-derived stem cells [J]. *Stem Cell Res Ther*. 2020;11(1):131. doi:10.1186/s13287-020-01643-5
5. Huang B, Song BL, Xu C. Cholesterol metabolism in cancer: mechanisms and therapeutic opportunities [J]. *Nat Metab*. 2020;2(2):132–141. doi:10.1038/s42255-020-0174-0
6. Gu Y, Lin X, Dong Y, et al. PCSK9 facilitates melanoma pathogenesis via a network regulating tumor immunity [J]. *J Exp Clin Cancer Res*. 2023;42(1):2. doi:10.1186/s13046-022-02584-y
7. Wang F, Li M, Zhang A, et al. PCSK9 modulates macrophage polarization-mediated ventricular remodeling after myocardial infarction [J]. *J Immunol Res*. 2022;2022:7685796. doi:10.1155/2022/7685796
8. Fasolato S, Pigozzo S, Pontisso P, et al. PCSK9 levels are raised in chronic HCV patients with hepatocellular carcinoma [J]. *J Clin Med*. 2020;9(10):3134. doi:10.3390/jcm9103134
9. Quagliariello V, Bisceglia I, Berretta M, et al. PCSK9 inhibitors in cancer patients treated with immune-checkpoint inhibitors to reduce cardiovascular events: new frontiers in cardioncology [J]. *Cancers (Basel)*. 2023;15(5):1397. doi:10.3390/cancers15051397
10. Demers A, Samami S, Lauzier B, et al. PCSK9 Induces CD36 degradation and affects long-chain fatty acid uptake and triglyceride metabolism in adipocytes and in mouse liver [J]. *Arterioscler Thromb Vasc Biol*. 2015;35(12):2517–2525. doi:10.1161/ATVBAHA.115.306032
11. Liu C, Chen J, Chen H, et al. PCSK9 inhibition: from current advances to evolving future [J]. *Cells*. 2022;11(19):2972. doi:10.3390/cells11192972
12. Li ZZ, Guo L, Yi A, et al. Evolocumab attenuates myocardial ischemia/reperfusion injury by blocking PCSK9/LIAS-mediated cuproptosis of cardiomyocytes [J]. *Basic Res Cardiol*. 2025; 1–20.
13. Liu X, Bao X, Hu M, et al. Inhibition of PCSK9 potentiates immune checkpoint therapy for cancer [J]. *Nature*. 2020;588(7839):693–698. doi:10.1038/s41586-020-2911-7
14. Paciullo F, Fallarino F, Bianconi V, et al. PCSK9 at the crossroad of cholesterol metabolism and immune function during infections [J]. *J Cell Physiol*. 2017;232(9):2330–2338. doi:10.1002/jcp.25767
15. Wiciński M, Żak J, Malinowski B, et al. PCSK9 signaling pathways and their potential importance in clinical practice [J]. *Epma j*. 2017;8(4):391–402. doi:10.1007/s13167-017-0106-6
16. Guo W, Gao H, Li H, et al. Self-assembly of a multifunction DNA tetrahedron for effective delivery of aptamer PL1 and Pcsk9 siRNA potentiate immune checkpoint therapy for colorectal cancer [J]. *ACS Appl Mater Interfaces*. 2022;14(28):31634–31644. doi:10.1021/acsami.2c06001
17. Yang Z, Li H, Dong T, et al. Comprehensive analysis of resistance mechanisms to EGFR-TKIs and establishment and validation of prognostic model [J]. *J Cancer Res Clin Oncol*. 2023;149(15):13773–13792. doi:10.1007/s00432-023-05129-8
18. Lochner M, Berod L, Sparwasser T. Fatty acid metabolism in the regulation of T cell function [J]. *Trends Immunol*. 2015;36(2):81–91. doi:10.1016/j.it.2014.12.005
19. Poggi P, Mirabella R, Neri S, et al. Membrane fatty acid heterogeneity of leukocyte classes is altered during in vitro cultivation but can be restored with ad-hoc lipid supplementation [J]. *Lipids Health Dis*. 2015;14:165. doi:10.1186/s12944-015-0166-3
20. Kansal V, Burnham AJ, Kinney BLC, et al. Statin drugs enhance responses to immune checkpoint blockade in head and neck cancer models [J]. *J Immunother Cancer*. 2023;11(1). doi:10.1136/jitc-2022-005940.

21. Ni W, Mo H, Liu Y, et al. Targeting cholesterol biosynthesis promotes anti-tumor immunity by inhibiting long noncoding RNA SNHG29-mediated YAP activation [J]. *Mol Ther*. 2021;29(10):2995–3010. doi:10.1016/j.ymthe.2021.05.012
22. Nicoli F, Cabral-Piccin MP, Papagno L, et al. Altered basal lipid metabolism underlies the functional impairment of naive CD8(+) T cells in elderly humans [J]. *J Immunol*. 2022;208(3):562–570. doi:10.4049/jimmunol.2100194
23. Saveanu L, Zucchetti AE, Evnouchidou I, et al. Is there a place and role for endocytic TCR signaling? *Immunol Rev*. 2019;291(1):57–74. doi:10.1111/imr.12764
24. Nava Lauson CB, Tiberti S, Corsetto PA, et al. Linoleic acid potentiates CD8(+) T cell metabolic fitness and antitumor immunity [J]. *Cell Metab*. 2023;35(4):633–50.e9. doi:10.1016/j.cmet.2023.02.013
25. Pan Y, Tian T, Park CO, et al. Survival of tissue-resident memory T cells requires exogenous lipid uptake and metabolism [J]. *Nature*. 2017;543(7644):252–256. doi:10.1038/nature21379
26. Yang W, Bai Y, Xiong Y, et al. Potentiating the antitumor response of CD8(+) T cells by modulating cholesterol metabolism [J]. *Nature*. 2016;531(7596):651–655. doi:10.1038/nature17412
27. Ma X, Bi E, Lu X, et al. Cholesterol Induces CD8(+) T cell exhaustion in the tumor microenvironment [J]. *Cell Metab*. 2019;30(1):143–56.e5. doi:10.1016/j.cmet.2019.04.002
28. Sugi T, Katoh Y, Ikeda T, et al. SCD1 inhibition enhances the effector functions of CD8(+) T cells via ACAT1-dependent reduction of esterified cholesterol [J]. *Cancer Sci*. 2024;115(1):48–58. doi:10.1111/cas.15999
29. Li M, Yang Y, Wei J, et al. Enhanced chemo-immunotherapy against melanoma by inhibition of cholesterol esterification in CD8(+) T cells [J]. *Nanomedicine*. 2018;14(8):2541–2550. doi:10.1016/j.nano.2018.08.008
30. Mesquita I, Ferreira C, Moreira D, et al. The absence of HIF-1 α increases susceptibility to leishmania donovani infection via activation of BNIP3/mTOR/SREBP-1c Axis [J]. *Cell Rep*. 2020;30(12):4052–64.e7. doi:10.1016/j.celrep.2020.02.098
31. O'rourke SA, Neto NGB, Devilly E, et al. Cholesterol crystals drive metabolic reprogramming and M1 macrophage polarisation in primary human macrophages [J]. *Atherosclerosis*. 2022;352:35–45. doi:10.1016/j.atherosclerosis.2022.05.015
32. Naito C, Hashimoto M, Watanabe K, et al. Facilitatory effects of fetuin-A on atherosclerosis [J]. *Atherosclerosis*. 2016;246:344–351. doi:10.1016/j.atherosclerosis.2016.01.037
33. Xu F, Yu Z, Liu Y, et al. A high-fat, high-cholesterol diet promotes intestinal inflammation by exacerbating gut microbiome dysbiosis and bile acid disorders in cholecystectomy [J]. *Nutrients*. 2023;15(17):3829. doi:10.3390/nu15173829
34. Rivas-Urbina A, Rotllan N, Santos D, et al. Monitoring atheroprotective macrophage cholesterol efflux in vivo [J]. *Methods mol Biol*. 2022;2419:569–581. doi:10.1007/978-1-0716-1924-7_35
35. Yan J, Horng T. Lipid metabolism in regulation of macrophage functions [J]. *Trends Cell Biol*. 2020;30(12):979–989. doi:10.1016/j.tcb.2020.09.006
36. Liu X, Guo JW, Lin XC, et al. Macrophage NFATc3 prevents foam cell formation and atherosclerosis: evidence and mechanisms [J]. *Eur Heart J*. 2021;42(47):4847–4861. doi:10.1093/eurheartj/ehab660
37. Qin WH, Yang ZS, Li M, et al. High serum levels of cholesterol increase antitumor functions of nature killer cells and reduce growth of liver tumors in mice [J]. *Gastroenterology*. 2020;158(6):1713–1727. doi:10.1053/j.gastro.2020.01.028
38. Choi C, Finlay DK. Optimising NK cell metabolism to increase the efficacy of cancer immunotherapy [J]. *Stem Cell Res Ther*. 2021;12(1):320. doi:10.1186/s13287-021-02377-8
39. Alannan M, Trézéguet V, Amoêdo ND, et al. Rewiring lipid metabolism by targeting PCSK9 and HMGCR to treat liver cancer [J]. *Cancers (Basel)*. 2022;15(1):3. doi:10.3390/cancers15010003
40. Abifadel M, Guerin M, Benjannet S, et al. Identification and characterization of new gain-of-function mutations in the PCSK9 gene responsible for autosomal dominant hypercholesterolemia [J]. *Atherosclerosis*. 2012;223(2):394–400. doi:10.1016/j.atherosclerosis.2012.04.006
41. Sun C, Zhu G, Shen C, et al. Identification and validation of PCSK9 as a prognostic and immune-related influencing factor in tumorigenesis: a pan-cancer analysis [J]. *Front Oncol*. 2023;13:1134063. doi:10.3389/fonc.2023.1134063
42. Alannan M, Seidah NG, Merched AJ. PCSK9 in liver cancers at the crossroads between lipid metabolism and immunity [J]. *Cells*. 2022;11(24):4132. doi:10.3390/cells11244132
43. Wang H, Zhang X, Zhang Y, et al. Targeting PCSK9 to upregulate MHC-II on the surface of tumor cells in tumor immunotherapy [J]. *BMC Cancer*. 2024;24(1):445. doi:10.1186/s12885-024-12148-2
44. Wang R, Liu H, He P, et al. Inhibition of PCSK9 enhances the antitumor effect of PD-1 inhibitor in colorectal cancer by promoting the infiltration of CD8(+) T cells and the exclusion of Treg cells [J]. *Front Immunol*. 2022;13:947756. doi:10.3389/fimmu.2022.947756
45. Da Dalt L, Ruscica M, Bonacina F, et al. PCSK9 deficiency reduces insulin secretion and promotes glucose intolerance: the role of the low-density lipoprotein receptor [J]. *Eur Heart J*. 2019;40(4):357–368. doi:10.1093/eurheartj/ehy357
46. Wagner J, Park LM, Mukhopadhyay P, et al. PCSK9 inhibition attenuates alcohol-associated neuronal oxidative stress and cellular injury [J]. *Brain Behav Immun*. 2024;119:494–506. doi:10.1016/j.bbi.2024.04.022
47. Ma N, Fan L, Dong Y, et al. New PCSK9 inhibitor miR-552-3p reduces LDL-C via enhancing LDLR in high fat diet-fed mice [J]. *Pharmacol Res*. 2021;167:105562. doi:10.1016/j.phrs.2021.105562
48. Yan L, Jia Q, Cao H, et al. Fisetin ameliorates atherosclerosis by regulating PCSK9 and LOX-1 in apoE(-/-) mice [J]. *Exp Ther Med*. 2021;21(1):25. doi:10.3892/etm.2020.9457
49. Jia YJ, Xu RX, Sun J, et al. Enhanced circulating PCSK9 concentration by berberine through SREBP-2 pathway in high fat diet-fed rats [J]. *J Transl Med*. 2014;12:103. doi:10.1186/1479-5876-12-103
50. Singh A, Kumar P, Sonkar AB, et al. A comprehensive review on PCSK9 as mechanistic target approach in cancer therapy [J]. *Mini Rev Med Chem*. 2023;23(1):24–32. doi:10.2174/1389557521666211202115823
51. Wang L, Wang Z, Shi J, et al. Inhibition of proprotein convertase subtilisin/kexin type 9 attenuates neuronal apoptosis following focal cerebral ischemia via apolipoprotein E receptor 2 downregulation in hyperlipidemic mice [J]. *Int J Mol Med*. 2018;42(4):2098–2106. doi:10.3892/ijmm.2018.3797
52. Sun H, Meng W, Zhu J, et al. Antitumor activity and molecular mechanism of proprotein convertase subtilisin/kexin type 9 (PCSK9) inhibition [J]. *Naunyn Schmiedeberg Arch Pharmacol*. 2022;395(6):643–658. doi:10.1007/s00210-022-02200-y

53. Rabow Z, Laubach K, Kong X, et al. p73 α 1, an Isoform of the p73 tumor suppressor, modulates lipid metabolism and cancer cell growth via stearoyl-CoA desaturase-1 [J]. *Cells*. 2022;11(16):2516. doi:10.3390/cells11162516
54. Lebeau PF, Byun JH, Platko K, et al. Caffeine blocks SREBP2-induced hepatic PCSK9 expression to enhance LDLR-mediated cholesterol clearance [J]. *Nat Commun*. 2022;13(1):770. doi:10.1038/s41467-022-28240-9
55. Yuan J, T CAI, Zheng X, et al. Potentiating CD8(+) T cell antitumor activity by inhibiting PCSK9 to promote LDLR-mediated TCR recycling and signaling [J]. *Protein and Cell*. 2021;12(4):240–260.
56. Seidah NG. The PCSK9 discovery, an inactive protease with varied functions in hypercholesterolemia, viral infections, and cancer [J]. *J Lipid Res*. 2021;62:100130. doi:10.1016/j.jlr.2021.100130
57. Leung AKK, Xue YC, De Guzman A, et al. Modulation of vascular endothelial inflammatory response by proprotein convertase subtilisin-kexin type 9 [J]. *Atherosclerosis*. 2022;362:29–37. doi:10.1016/j.atherosclerosis.2022.09.008
58. Zhang XY, Lu QQ, Li YJ, et al. Conditional knockdown of hepatic PCSK9 ameliorates high-fat diet-induced liver inflammation in mice [J]. *Front Pharmacol*. 2025;16:1528250. doi:10.3389/fphar.2025.1528250
59. Miao M, Zhang XY, Yu HX, et al. Mechanisms underlying the effects of the conditional knockdown of hepatic PCSK9 in attenuating lipopolysaccharide-induced acute liver inflammation [J]. *Int J Biol Macromol*. 2025;291:139066. doi:10.1016/j.ijbiomac.2024.139066
60. Marques P, Domingo E, Rubio A, et al. Beneficial effects of PCSK9 inhibition with alirocumab in familial hypercholesterolemia involve modulation of new immune players [J]. *Biomed Pharmacother*. 2022;145:112460. doi:10.1016/j.biopha.2021.112460
61. Krychtiuk KA, Lenz M, Hohensinner P, et al. Circulating levels of proprotein convertase subtilisin/kexin type 9 (PCSK9) are associated with monocyte subsets in patients with stable coronary artery disease [J]. *J Clin Lipidol*. 2021;15(3):512–521. doi:10.1016/j.jacl.2021.02.005
62. Greco MF, Rizzuto AS, Zarà M, et al. PCSK9 confers inflammatory properties to extracellular vesicles released by vascular smooth muscle cells [J]. *Int J mol Sci*. 2022;23(21):13065. doi:10.3390/ijms232113065
63. Hu J, Zhang M, Gui L, et al. PCSK9 suppresses M2-like tumor-associated macrophage polarization by regulating the secretion of OX40L from hepatocellular carcinoma cells [J]. *Immunol Invest*. 2022;51(6):1678–1693. doi:10.1080/08820139.2022.2027439
64. Bernelot Moens SJ, Neele AE, Kroon J, et al. PCSK9 monoclonal antibodies reverse the pro-inflammatory profile of monocytes in familial hypercholesterolaemia [J]. *Eur Heart J*. 2017;38(20):1584–1593. doi:10.1093/eurheartj/ehx002
65. Seidah NG, Garçon D. Expanding Biology of PCSK9: roles in Atherosclerosis and Beyond [J]. *Curr Atheroscler Rep*. 2022;24(10):821–830. doi:10.1007/s11883-022-01057-z
66. Huang L, Li Y, Cheng Z, et al. PCSK9 promotes endothelial dysfunction during sepsis via the TLR4/MyD88/NF- κ B and NLRP3 Pathways [J]. *Inflammation*. 2023;46(1):115–128. doi:10.1007/s10753-022-01715-z
67. Waiz M, Alvi SS, Khan M. Association of circulatory PCSK-9 with biomarkers of redox imbalance and inflammatory cascades in the prognosis of diabetes and associated complications: a pilot study in the Indian population [J]. *Free Radic Res*. 2023;57(4):294–307. doi:10.1080/10715762.2023.2237180
68. Zheng Y, Zhu T, Li G, et al. PCSK9 inhibitor protects against ischemic cerebral injury by attenuating inflammation via the GPNMB/CD44 pathway [J]. *Int Immunopharmacol*. 2024;126:111195. doi:10.1016/j.intimp.2023.111195
69. Ly K, Saavedra YG, Canuel M, et al. Annexin A2 reduces PCSK9 protein levels via a translational mechanism and interacts with the M1 and M2 domains of PCSK9 [J]. *J Biol Chem*. 2014;289(25):17732–17746. doi:10.1074/jbc.M113.541094
70. Song F, Z LJ, Wu Y, et al. Ubiquitinated ligation protein NEDD4L participates in MiR-30a-5p attenuated atherosclerosis by regulating macrophage polarization and lipid metabolism [J]. *mol Ther Nucleic Acids*. 2021;26:1303–1317. doi:10.1016/j.omtn.2021.10.030
71. Li Y, Chen Y, Zhao C, et al. Arenobufagin modulation of PCSK9-mediated cholesterol metabolism induces tumor-associated macrophages polarisation to inhibit hepatocellular carcinoma progression [J]. *Phytomedicine*. 2024;128:155532. doi:10.1016/j.phymed.2024.155532
72. Qiu H, Shao Z, Wen X, et al. Efferocytosis: an accomplice of cancer immune escape [J]. *Biomed Pharmacother*. 2023;167:115540. doi:10.1016/j.biopha.2023.115540
73. Tajbakhsh A, Gheibi Hayat SM, Movahedpour A, et al. The complex roles of efferocytosis in cancer development, metastasis, and treatment [J]. *Biomed Pharmacother*. 2021;140:111776. doi:10.1016/j.biopha.2021.111776
74. Cheng M, Chen S, Li K, et al. CD276-dependent efferocytosis by tumor-associated macrophages promotes immune evasion in bladder cancer [J]. *Nat Commun*. 2024;15(1):2818. doi:10.1038/s41467-024-46735-5
75. Liu S, Wu J, Stolarz A, et al. PCSK9 attenuates efferocytosis in endothelial cells and promotes vascular aging [J]. *Theranostics*. 2023;13(9):2914–2929. doi:10.7150/thno.83914
76. Wu J, Liu S, Banerjee O, et al. Disturbed flow impairs MerTK-mediated efferocytosis in aortic endothelial cells during atherosclerosis [J]. *Theranostics*. 2024;14(6):2427–2441. doi:10.7150/thno.93036
77. Gao ZJ, Fang Z, Yuan JP, et al. Integrative multi-omics analyses unravel the immunological implication and prognostic significance of CXCL12 in breast cancer [J]. *Front Immunol*. 2023;14:1188351. doi:10.3389/fimmu.2023.1188351
78. Luan C, He Y, Liu W, et al. PCSK9 inhibition interrupts the cross-talk between keratinocytes and macrophages and prevents UVB-induced skin damage [J]. *J Biol Chem*. 2023;299(7):104895. doi:10.1016/j.jbc.2023.104895
79. Fang S, Zheng L, Shu GF, et al. Multiple immunomodulatory strategies based on targeted regulation of proprotein convertase subtilisin/kexin type 9 and immune homeostasis against hepatocellular carcinoma [J]. *ACS Nano*. 2024;18(12):8811–8826. doi:10.1021/acsnano.3c11775
80. Kuai R, Sun X, Yuan W, et al. Dual TLR agonist nanodiscs as a strong adjuvant system for vaccines and immunotherapy [J]. *J Control Release*. 2018;282:131–139. doi:10.1016/j.jconrel.2018.04.041
81. Liu A, Frosteigård J. PCSK 9 plays a novel immunological role in oxidized LDL-induced dendritic cell maturation and activation of T cells from human blood and atherosclerotic plaque. *J Internal Med*. 2018;284(2):193–210. doi:10.1111/joim.12758
82. Wang Y, Zhang L, Lv L, et al. Dendritic cell-derived exosomal miR-3064-5p inhibits SIRT6/PCSK9 to protect the blood-brain barrier after subarachnoid hemorrhage [J]. *J Biochem mol Toxicol*. 2023;37(6):e23346. doi:10.1002/jbt.23346
83. Liu A, Rahman M, Hafström I, et al. Proprotein convertase subtilisin kexin 9 is associated with disease activity and is implicated in immune activation in systemic lupus erythematosus [J]. *Lupus*. 2020;29(8):825–835. doi:10.1177/0961203320926253
84. Zhou S, Guo Q, Chen A, et al. Integrated bioinformatics analysis identifies PCSK9 as a prognosticator correlated with lipid metabolism in pancreatic adenocarcinoma [J]. *World J Surg Oncol*. 2024;22(1):256. doi:10.1186/s12957-024-03532-0

85. Xiao Q, Li X, Li Y, et al. Biological drug and drug delivery-mediated immunotherapy [J]. *Acta Pharm Sin B*. 2021;11(4):941–960. doi:10.1016/j.apsb.2020.12.018
86. Elemans M, Florins A, Willems L, et al. Rates of CTL killing in persistent viral infection in vivo [J]. *PLoS Comput Biol*. 2014;10(4):e1003534. doi:10.1371/journal.pcbi.1003534
87. Kidani Y, Elsaesser H, Hock MB, et al. Sterol regulatory element-binding proteins are essential for the metabolic programming of effector T cells and adaptive immunity [J]. *Nat Immunol*. 2013;14(5):489–499. doi:10.1038/ni.2570
88. Naslavsky N, Weigert R, Donaldson JG. Characterization of a nonclathrin endocytic pathway: membrane cargo and lipid requirements [J]. *mol Biol Cell*. 2004;15(8):3542–3552. doi:10.1091/mbc.e04-02-0151
89. Badimon L, Luquero A, Crespo J, et al. PCSK9 and LRP5 in macrophage lipid internalization and inflammation [J]. *Cardiovasc Res*. 2021;117(9):2054–2068. doi:10.1093/cvr/cvaa254
90. Klein-Hessling S, Muhammad K, Klein M, et al. NFATc1 controls the cytotoxicity of CD8(+) T cells [J]. *Nat Commun*. 2017;8(1):511. doi:10.1038/s41467-017-00612-6
91. Gabriel CH, Gross F, Karl M, et al. Identification of novel nuclear factor of activated T cell (NFAT)-associated Proteins in T Cells [J]. *J Biol Chem*. 2016;291(46):24172–24187. doi:10.1074/jbc.M116.739326
92. Hsu CY, Abdulrahim MN, Mustafa MA, et al. The multifaceted role of PCSK9 in cancer pathogenesis, tumor immunity, and immunotherapy [J]. *Med Oncol*. 2024;41(8):202. doi:10.1007/s12032-024-02435-0
93. Sun Y, Zhang H, Meng J, et al. S-palmitoylation of PCSK9 induces sorafenib resistance in liver cancer by activating the PI3K/AKT pathway [J]. *Cell Rep*. 2022;40(7):111194. doi:10.1016/j.celrep.2022.111194
94. Mondal K, Chakraborty P, Kabir SN. Hyperhomocysteinemia and hyperandrogenemia share PCSK9-LDLR pathway to disrupt lipid homeostasis in PCOS [J]. *Biochem Biophys Res Commun*. 2018;503(1):8–13. doi:10.1016/j.bbrc.2018.04.078
95. Cai J, Jiang Y, Chen F, et al. PCSK9 promotes T helper 1 and T helper 17 cell differentiation by activating the nuclear factor- κ B pathway in ankylosing spondylitis [J]. *Immun Inflamm Dis*. 2023;11(5):e870. doi:10.1002/iid3.870
96. Rose M, Duhamel M, Rodet F, et al. The role of proprotein convertases in the regulation of the function of immune cells in the oncoimmune response [J]. *Front Immunol*. 2021;12:667850. doi:10.3389/fimmu.2021.667850
97. Saud AH, Ali NAJ, Gali FY, et al. The effect of evolocumab alone and in combination with atorvastatin on lipid profile [J]. *Wiad Lek*. 2021;74(12):3184–3187. doi:10.36740/WLek202112111
98. Yu M, Tang W, Liang W, et al. PCSK9 inhibition ameliorates experimental autoimmune myocarditis by reducing Th17 cell differentiation through LDLR/STAT-3/ROR- γ t pathway [J]. *Int Immunopharmacol*. 2023;124(Pt B):110962. doi:10.1016/j.intimp.2023.110962
99. Goswami R, Kaplan MH. STAT transcription factors in T cell control of health and disease [J]. *Int Rev Cell mol Biol*. 2017;331:123–180. doi:10.1016/bs.ircmb.2016.09.012
100. Qin L, Shi L, Wang Y, et al. Fumarate hydratase enhances the therapeutic effect of PD-1 antibody in colorectal cancer by regulating PCSK9 [J]. *Cancers (Basel)*. 2024;16(4). doi:10.3390/cancers16040713.
101. Xu W, Hu M, Lu X, et al. Inhibition of PCSK9 enhances the anti-hepatocellular carcinoma effects of TCR-T cells and anti-PD-1 immunotherapy [J]. *Int J Biol Sci*. 2024;20(10):3942–3955. doi:10.7150/ijbs.93668
102. X GAO, Yi L, Jiang C, et al. PCSK9 regulates the efficacy of immune checkpoint therapy in lung cancer [J]. *Front Immunol*. 2023;14:1142428. doi:10.3389/fimmu.2023.1142428
103. Baziotti V, Halmos B, Westertorp M. T-cell cholesterol accumulation, aging, and atherosclerosis [J]. *Curr Atheroscler Rep*. 2023;25(9):527–534. doi:10.1007/s11883-023-01125-y
104. Kanda N, Okajima F. Atopic dermatitis-like rash during evolocumab treatment of familial hypercholesterolemia [J]. *J Nippon Med Sch*. 2019;86(3):187–190. doi:10.1272/jnms.JNMS.2019_86-309
105. Ji E, Lee S. Antibody-based therapeutics for atherosclerosis and cardiovascular diseases [J]. *Int J mol Sci*. 2021;22(11):5770. doi:10.3390/ijms22115770
106. Yang QC, Wang S, Liu YT, et al. Targeting PCSK9 reduces cancer cell stemness and enhances antitumor immunity in head and neck cancer [J]. *iScience*. 2023;26(6):106916. doi:10.1016/j.isci.2023.106916
107. Pitteri SJ, Kelly-Spratt KS, Gurley KE, et al. Tumor microenvironment-derived proteins dominate the plasma proteome response during breast cancer induction and progression [J]. *Cancer Res*. 2011;71(15):5090–5100. doi:10.1158/0008-5472.CAN-11-0568
108. Momtazi-Borojeni AA, Nik ME, Jaafari MR, et al. Effects of immunization against PCSK9 in an experimental model of breast cancer [J]. *Arch Med Sci*. 2019;15(3):570–579. doi:10.5114/aoms.2019.84734
109. Nowak C, Ärnlov J. A Mendelian randomization study of the effects of blood lipids on breast cancer risk [J]. *Nat Commun*. 2018;9(1):3957. doi:10.1038/s41467-018-06467-9
110. Abdelwahed KS, Siddique AB, Mohyeldin MM, et al. Pseurotin A as a novel suppressor of hormone dependent breast cancer progression and recurrence by inhibiting PCSK9 secretion and interaction with LDL receptor [J]. *Pharmacol Res*. 2020;158:104847. doi:10.1016/j.phrs.2020.104847
111. Suh JM, Son Y, Yoo JY, et al. Proprotein convertase subtilisin/kexin Type 9 is required for Ahnak-mediated metastasis of melanoma into lung epithelial cells [J]. *Neoplasia*. 2021;23(9):993–1001. doi:10.1016/j.neo.2021.07.007
112. Yang B, Wang H, Song W, et al. Lipid-lowering medications and risk of malignant melanoma: a Mendelian randomization study [J]. *Front Oncol*. 2024;14:1408972. doi:10.3389/fonc.2024.1408972
113. Sun X, Essalmani R, Day R, et al. Proprotein convertase subtilisin/kexin type 9 deficiency reduces melanoma metastasis in liver [J]. *Neoplasia*. 2012;14(12):1122–1131. doi:10.1593/neo.121252
114. Momtazi-Borojeni AA, Nik ME, Jaafari MR, et al. Effects of immunisation against PCSK9 in mice bearing melanoma [J]. *Arch Med Sci*. 2020;16(1):189–199. doi:10.5114/aoms.2020.91291
115. Fang S, Yarmolinsky J, Gill D, et al. Association between genetically proxied PCSK9 inhibition and prostate cancer risk: a Mendelian randomisation study [J]. *PLoS Med*. 2023;20(1):e1003988. doi:10.1371/journal.pmed.1003988
116. Gan SS, Ye JQ, Wang L, et al. Inhibition of PCSK9 protects against radiation-induced damage of prostate cancer cells [J]. *Onco Targets Ther*. 2017;10:2139–2146. doi:10.2147/OTT.S129413

117. Abdelwahed KS, Siddique AB, Qusa MH, et al. PCSK9 axis-targeting pseurotin a as a novel prostate cancer recurrence suppressor lead [J]. *ACS Pharmacol Transl Sci*. 2021;4(6):1771–1781. doi:10.1021/acspstsci.1c00145
118. Wang L, Li S, Luo H, et al. PCSK9 promotes the progression and metastasis of colon cancer cells through regulation of EMT and PI3K/AKT signaling in tumor cells and phenotypic polarization of macrophages [J]. *J Exp Clin Cancer Res*. 2022;41(1):303. doi:10.1186/s13046-022-02477-0
119. Yang K, Zhu J, Luo HH, et al. Pro-protein convertase subtilisin/kexin type 9 promotes intestinal tumor development by activating Janus kinase 2/signal transducer and activator of transcription 3/SOCS3 signaling in Apc(Min/+) mice [J]. *Int J Immunopathol Pharmacol*. 2021;35:20587384211038345. doi:10.1177/20587384211038345
120. Wong CC, Wu JL, Ji F, et al. The cholesterol uptake regulator PCSK9 promotes and is a therapeutic target in APC/KRAS-mutant colorectal cancer [J]. *Nat Commun*. 2022;13(1):3971. doi:10.1038/s41467-022-31663-z
121. Coppinger C, Pomales B, Movahed MR, et al. Berberine: a multi-target natural PCSK9 inhibitor with the potential to treat diabetes, Alzheimer's, cancer and cardiovascular disease. *Curr Rev Clin Exp Pharmacol*. 2024;19(4):312–326. doi:10.2174/0127724328250471231222094648
122. Bhat M, Skill N, Marcus V, et al. Decreased PCSK9 expression in human hepatocellular carcinoma [J]. *BMC Gastroenterol*. 2015;15(1):176. doi:10.1186/s12876-015-0371-6
123. Zhang SZ, Zhu XD, Feng LH, et al. PCSK9 promotes tumor growth by inhibiting tumor cell apoptosis in hepatocellular carcinoma [J]. *Exp Hematol Oncol*. 2021;10(1):25. doi:10.1186/s40164-021-00218-1
124. Ruscica M, Ricci C, Macchi C, et al. Suppressor of Cytokine Signaling-3 (SOCS-3) induces proprotein convertase subtilisin kexin type 9 (PCSK9) expression in hepatic HepG2 cell line [J]. *J Biol Chem*. 2016;291(7):3508–3519. doi:10.1074/jbc.M115.664706
125. Kim HJ, Lee J, Chung MY, et al. Piceatannol reduces resistance to statins in hypercholesterolemia by reducing PCSK9 expression through p300 acetyltransferase inhibition [J]. *Pharmacol Res*. 2020;161:105205. doi:10.1016/j.phrs.2020.105205
126. He M, Zhang W, Dong Y, et al. Pro-inflammation NF-κB signaling triggers a positive feedback via enhancing cholesterol accumulation in liver cancer cells [J]. *J Exp Clin Cancer Res*. 2017;36(1):15. doi:10.1186/s13046-017-0490-8
127. Ioannou GN, Lee SP, Linsley PS, et al. Pcsk9 deletion promotes murine nonalcoholic steatohepatitis and hepatic carcinogenesis: role of cholesterol [J]. *Hepatol Commun*. 2022;6(4):780–794. doi:10.1002/hep4.1858
128. He M, Hu J, Fang T, et al. Protein convertase subtilisin/Kexin type 9 inhibits hepatocellular carcinoma growth by interacting with GSTP1 and suppressing the JNK signaling pathway [J]. *Cancer Biol Med*. 2021;19(1):90–103. doi:10.20892/j.issn.2095-3941.2020.0313
129. Ochinn CC, Garelnabi M. Berberine Encapsulated PLGA-PEG Nanoparticles Modulate PCSK-9 in HepG2 Cells [J]. *Cardiovasc Hematol Disord Drug Targets*. 2018;18(1):61–70. doi:10.2174/1871529X18666180201130340
130. Tai MH, Chen PK, Chen PY, et al. Curcumin enhances cell-surface LDLR level and promotes LDL uptake through downregulation of PCSK9 gene expression in HepG2 cells [J]. *mol Nutr Food Res*. 2014;58(11):2133–2145. doi:10.1002/mnfr.201400366
131. Choi HK, Hwang JT, Nam TG, et al. Welsh onion extract inhibits PCSK9 expression contributing to the maintenance of the LDLR level under lipid depletion conditions of HepG2 cells [J]. *Food Funct*. 2017;8(12):4582–4591. doi:10.1039/C7FO00562H
132. Lammi C, Mulinacci N, Cecchi L, et al. Virgin olive oil extracts reduce oxidative stress and modulate cholesterol metabolism: comparison between oils obtained with traditional and innovative processes [J]. *Antioxidants (Basel)*. 2020;9(9). doi:10.3390/antiox9090798
133. Bang S, Chae HS, Lee C, et al. New aromatic compounds from the fruiting body of *sparassis crispa* (Wulf.) and their inhibitory activities on proprotein convertase subtilisin/kexin type 9 mRNA expression [J]. *J Agric Food Chem*. 2017;65(30):6152–6157. doi:10.1021/acs.jafc.7b02657
134. Nhoe KP, Chae HS, Masagalli JN, et al. Discovery of flavonoids from *Scutellaria baicalensis* with inhibitory activity against PCSK 9 expression: isolation, synthesis and their biological evaluation [J]. *Molecules*. 2018;23(2): 504.
135. Mannino G, Iovino P, Lauria A, et al. Bioactive triterpenes of protium heptaphyllum gum resin extract display cholesterol-lowering potential [J]. *Int J mol Sci*. 2021;22(5): 2664.
136. Song J, Shih IEM, Chan DW, et al. Suppression of annexin A11 in ovarian cancer: implications in chemoresistance [J]. *Neoplasia*. 2009;11(6):605–14,1pfollowing14. doi:10.1593/neo.09286
137. Jacome Sanz D, Raivola J, Karvonen H, et al. Evaluating targeted therapies in ovarian cancer metabolism: novel role for PCSK9 and second generation mTOR inhibitors [J]. *Cancers (Basel)*. 2021;13(15):3727. doi:10.3390/cancers13153727
138. Xu D, Han G, Zhou X, et al. TEAD4 activates PCSK9 to promote stomach adenocarcinoma cell stemness through fatty acid metabolism [J]. *Digestion*. 2024;105(4):243–256. doi:10.1159/000538329
139. Xu B, Li S, Fang Y, et al. Proprotein convertase subtilisin/kexin type 9 promotes gastric cancer metastasis and suppresses apoptosis by facilitating MAPK signaling pathway through HSP70 up-regulation [J]. *Front Oncol*. 2020;10:609663. doi:10.3389/fonc.2020.609663
140. Ito M, Hiwasa T, Oshima Y, et al. Association of Serum Anti-PCSK9 antibody levels with favorable postoperative prognosis in esophageal cancer [J]. *Front Oncol*. 2021;11:708039. doi:10.3389/fonc.2021.708039
141. Piao MX, Bai JW, Zhang PF, et al. PCSK9 regulates apoptosis in human neuroglioma u251 cells via mitochondrial signaling pathways [J]. *Int J Clin Exp Pathol*. 2015;8(3):2787–2794.
142. Xu X, Cui Y, Cao L, et al. PCSK9 regulates apoptosis in human lung adenocarcinoma A549 cells via endoplasmic reticulum stress and mitochondrial signaling pathways. *Exp Ther Med*. 2017;13(5):1993–1999. doi:10.3892/etm.2017.4218
143. Rizzo A, Brandi G. Neoadjuvant therapy for cholangiocarcinoma: a comprehensive literature review. *Cancer Treat Res Commun*. 2021;27:100354. doi:10.1016/j.ctarc.2021.100354
144. Rizzo A, Ricci AD, Tober N, et al. Second-line Treatment in Advanced Biliary Tract Cancer: today and Tomorrow [J]. *Anticancer Res*. 2020;40(6):3013–3030. doi:10.21873/anticancer.14282
145. Rizzo A, Mollica V, Tateo V, et al. Hypertransaminasemia in cancer patients receiving immunotherapy and immune-based combinations: the MOUSEION-05 study. *Cancer Immunol Immunother*. 2023;72(6):1381–1394. doi:10.1007/s00262-023-03366-x
146. Guven DC, Sahin TK, Erul E, et al. The association between albumin levels and survival in patients treated with immune checkpoint inhibitors: a systematic review and meta-analysis. *Front Mol Biosci*. 2022;9:1039121. doi:10.3389/fmolb.2022.1039121
147. Huang HC, Hsu SJ, Chang CC, et al. Effects of PCSK-9 inhibition by alirocumab treatments on biliary cirrhotic rats [J]. *Int J mol Sci*. 2022;23(13): 7292.

148. Hossini AM, Hou X, Exner T, et al. Free Fatty Acids Induce Lipid Accumulation, Autophagy, and Apoptosis in Human Sebocytes. *Skin Pharmacol Physiol*. 2023;36(1):1–15. doi:10.1159/000527471
149. Gao L, Xu Z, Huang Z, et al. CPI-613 rewires lipid metabolism to enhance pancreatic cancer apoptosis via the AMPK-ACC signaling. *J Exp Clin Cancer Res*. 2020;39(1):73. doi:10.1186/s13046-020-01579-x

OncoTargets and Therapy

Publish your work in this journal

OncoTargets and Therapy is an international, peer-reviewed, open access journal focusing on the pathological basis of all cancers, potential targets for therapy and treatment protocols employed to improve the management of cancer patients. The journal also focuses on the impact of management programs and new therapeutic agents and protocols on patient perspectives such as quality of life, adherence and satisfaction. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/oncotargets-and-therapy-journal>

Dovepress
Taylor & Francis Group