

MVC administration for 3 days blocked *ex vivo* virus infection of LCs as well as subsequent virus transmission when different R5 HIV strains, HIV<sub>AD8</sub> and HIV<sub>JR-FL</sub>, were utilized for an additional six subjects ( $n=3$  for each strain, Figures 1 and 2). These data demonstrate that oral administration of MVC for at least 3 days is capable of fully protecting HIV infection of LCs within epithelial tissue.

These experiments provide perhaps the best proof-of-concept test for MVC as a potential PrEP drug, as it would be unethical to expose MVC-treated volunteers to HIV *in vivo*. As proven here, orally delivered MVC rapidly distributes to skin and functionally acts to block infection of relevant target cells, LCs, supporting randomized controlled trials of MVC as a PrEP therapy for individuals at high risk of becoming infected with HIV through sexual exposure.

#### CONFLICT OF INTEREST

The authors state no conflict of interest.

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#### SUPPLEMENTARY MATERIAL

Supplementary material is linked to the online version of the paper at <http://www.nature.com/jid>

#### REFERENCES

- Cohen MS, Chen YQ, McCauley M *et al.* (2011) Prevention of HIV-1 infection with early antiretroviral therapy. *N Engl J Med* 365: 493–505
- Dumond JB, Patterson KB, Pecha AL *et al.* (2009) Maraviroc concentrates in the cervicovaginal fluid and vaginal tissue of HIV-negative women. *J Acquir Immune Defic Syndr* 51:546–53
- Ganor Y, Zhou Z, Tudor D *et al.* (2010) Within 1 h, HIV-1 uses viral synapses to enter efficiently the inner, but not outer, foreskin mucosa and engages Langerhans-T cell conjugates. *Mucosal Immunol* 3:506–22
- Grant RM, Lama JR, Anderson PL *et al.* (2010) Preexposure chemoprophylaxis for HIV prevention in men who have sex with men. *N Engl J Med* 363:2587–99
- Hu J, Gardner MB, Miller CJ (2000) Simian immunodeficiency virus rapidly penetrates the cervicovaginal mucosa after intravaginal inoculation and infects intra-

epithelial dendritic cells. *J Virol* 74: 6087–95

- Kawamura T, Bruse SE, Abraha A *et al.* (2004) PSC-RANTES blocks R5 human immunodeficiency virus infection of Langerhans cells isolated from individuals with a variety of CCR5 diplotypes. *J Virol* 78:7602–9
- Kawamura T, Cohen SS, Borris DL *et al.* (2000) Candidate microbicides block HIV-1 infection of human immature Langerhans cells within epithelial tissue explants. *J Exp Med* 192:1491–500
- Lederman MM, Offord RE, Hartley O (2006) Microbicides and other topical strategies to prevent vaginal transmission of HIV. *Nat Rev Immunol* 6:371–82
- Ogawa Y, Kawamura T, Matsuzawa T *et al.* (2013) Antimicrobial Peptide LL-37 Produced by HSV-2-Infected Keratinocytes Enhances HIV Infection of Langerhans Cells. *Cell Host Microbe* 13:77–86
- Patterson KB, Prince HA, Kraft E *et al.* (2011) Penetration of tenofovir and emtricitabine in mucosal tissues: implications for prevention of HIV-1 transmission. *Sci Transl Med* 3:112re4
- Takahashi M, Hirano A, Okubo N *et al.* (2010) Development and application of a simple LC-MS method for the determination of plasma maraviroc concentrations. *J Med Invest* 57:245–50
- van der Straten A, Van Damme L, Haberer JE *et al.* (2012) Unraveling the divergent results of pre-exposure prophylaxis trials for HIV prevention. *AIDS* 26:F13–9
- Veazey RS, Ketas TJ, Dufour J *et al.* (2010) Protection of rhesus macaques from vaginal infection by vaginally delivered maraviroc, an inhibitor of HIV-1 entry via the CCR5 co-receptor. *J Infect Dis* 202:739–44
- Veazey RS, Springer MS, Marx PA *et al.* (2005) Protection of macaques from vaginal SHIV challenge by an orally delivered CCR5 inhibitor. *Nat Med* 11:1293–4

**JID Open**

## Heterozygous Mutations in AAGAB Cause Type 1 Punctate Palmoplantar Keratoderma with Evidence for Increased Growth Factor Signaling

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#### TO THE EDITOR

Punctate palmoplantar keratoderma (punctate PPK or PPKP) is a rare

autosomal dominant disorder of keratinization. Three variants of this disease have been described; PPKP1 (OMIM

#148600, Buschke–Fischer–Brauer type) is characterized by the progressive development of discrete areas of hyperkeratosis on the palms and soles, followed by more extensive diffuse hyperkeratosis on the pressure-bearing areas of plantar skin.

Linkage analyses of families affected by PPKP1 have previously identified a

Abbreviations: AAGAB, gene encoding alpha- and gamma-adaptin-binding protein p34; bp, base pair; OMIM, Online Mendelian Inheritance in Man; PPK(P), (punctate) palmoplantar keratoderma; RTK, receptor tyrosine kinase

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locus within 15q22–q24 (Martinez-Mir *et al.*, 2003; Gao *et al.*, 2005), but two Chinese pedigrees with a PPKP1 phenotype demonstrated linkage on chromosome 8q24.13–q24.21 (Zhang *et al.*, 2004). Recently, nonsense mutations in *AAGAB*, the gene encoding alpha- and gamma-adaptin-binding protein p34, were reported in three PPKP1 families (Giehl *et al.*, 2012). Simultaneously, we applied whole-exome sequencing and reported heterozygous loss-of-function mutations in *AAGAB* in 18 PPKP1 kindreds (Pöhler *et al.*, 2012). *AAGAB* is located on chromosome 15q22, within one of the previously reported linkage regions (Martinez-Mir *et al.*, 2003; Pöhler *et al.*, 2012). We now report the *AAGAB* genotype in a further 12 PPKP1 patients from 6 independent kindreds of Scottish, English, and Mexican ancestry. This study was carried out in accordance with the Declaration of Helsinki Principles, and all subjects gave written informed consent.

### KINDRED 1

A 56-year-old Scottish woman reported a 15-year history of the progressive development of tender callosities on her soles and palms. Examination revealed multiple hyperkeratotic foci on the palmar skin of hands and fingers (Figure 1a) and diffuse hyperkeratosis on weight-bearing areas of plantar skin (Figure 1b). The proband's parents were unaffected, but one of her three offspring demonstrated a PPKP1 phenotype. Sequencing of *AAGAB* using reported methodology (Pöhler *et al.*, 2012) identified a heterozygous 1-bp deletion, mutation c.344del resulting in a premature termination codon (p.Asp115Valfs\*7; Supplementary Figure S1 online). This mutation has previously been observed in two apparently unrelated Scottish families (Pöhler *et al.*, 2012).

### KINDRED 2

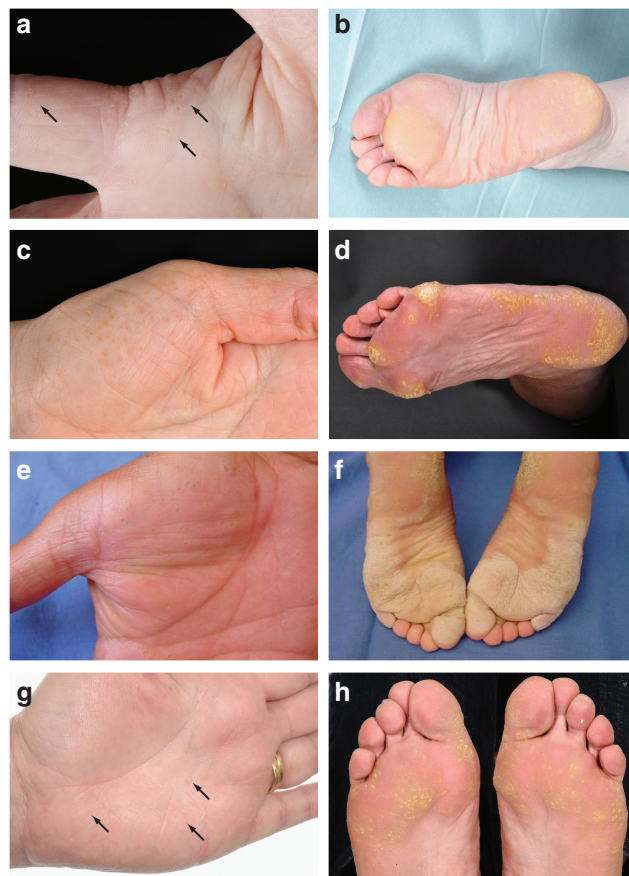
A 74-year-old Scottish woman presented with a 25-year history of progressive hyperkeratosis of her plantar skin (Figure 1d), with lesser involvement of the palmar skin (Figure 1c). The proband's parents were unaffected, but one of her three adult children had keratoderma. DNA sequencing identified a previously unreported heterozygous

nonsense mutation, c.390G>A, predicting the protein change p.Trp130\*0, within *AAGAB* (Supplementary Figures S1 and S2b online). This mutation was not detected in ~11,000 European and African American individuals' exome sequencing data (NHLBI Exome Sequencing Project <http://evs.gs.washington.edu/EVS/> accessed 27 March 2013), nor is it reported in the 1000 Genomes Catalogue of Human Genetic Variation (<http://www.1000genomes.org/ensembl-browser> accessed 27 March 2013).

### KINDRED 3

A 79-year-old Scottish man presented with a 30-year history of hyperkeratosis

of the soles and palms (Supplementary Figure S3a and b online). His medical history included a cutaneous squamous cell carcinoma on the hand and a basal cell carcinoma on the nose, but these were not at sites affected by keratoderma. The patient's parents, eight siblings, and two children were unaffected. DNA sequencing revealed a heterozygous c.870+1G>A splice-site mutation within *AAGAB* (Supplementary Figure S1 online), previously reported in an unrelated Scottish family (Pöhler *et al.*, 2012). The protein consequences of this mutation are unknown but is likely to represent a frameshift mutation.



**Figure 1. Clinical images of punctate palmoplantar keratoderma cases.** (a) Proband from kindred 1 illustrating focal punctate callosity on palmar skin and diffuse hyperkeratosis on plantar skin (b). Palm (c) and sole (d) of a 74-year-old woman from kindred 2 showing punctate lesions coalescing to form focal areas of marked hyperkeratosis. Affected individuals from kindred 5 showing typical focal calluses on palmar skin (e) and coalescence of the lesions on plantar pressure points (f). The proband from kindred 6 is shown in g and h, demonstrating subtle discrete hyperkeratotic papules on the palmar skin with more marked hyperkeratotic foci, which coalesce in weight-bearing areas on the plantar skin. Arrows indicate punctate hyperkeratotic lesions. Clinical images from the cases in kindreds 3 and 4 are shown in Supplementary Figures 3a, b, and 4a, b online.

#### KINDRED 4

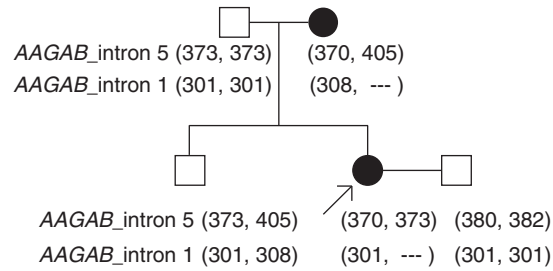
A 55-year-old female patient of English ancestry presented at the age of 30 years with typical features of PPK1. She had multiple keratotic papules on palmar and plantar surfaces, which had gradually increased in number, becoming confluent and tender (Supplementary Figure S4a and b online). The patient has one affected and four unaffected siblings; her two daughters are similarly affected and her one grandson shows early signs of punctate keratoderma. DNA sequencing revealed a heterozygous frameshift mutation c.472del, leading to a premature termination codon (p.Gly158Glufs\*0; Supplementary Figure S1 online), previously reported in five apparently unrelated Scottish families (Pöhler *et al.*, 2012).

#### KINDRED 5

Thirty-one members of this Mexican family over four generations demonstrate a PPK1P phenotype with an autosomal dominant pattern of inheritance. The affected living relatives showed multiple hyperkeratotic papules on the palms and soles with no associated nail disease (representative clinical images are shown in Figure 1e and f). Linkage analysis in this family has identified the *AAGAB* locus at chromosome 15q22 (Martinez-Mir *et al.*, 2003). *AAGAB* sequencing of DNA from five affected family members has now identified a previously unreported heterozygous 1-bp deletion in exon 3, c.275del predicting the protein change p.Leu92Leufs\*18, in each affected individual (Supplementary Figures S1 and S2d online). This mutation was not detected in ~11,000 European and African American individuals' exome sequencing data (NHLBI Exome Sequencing Project <http://evs.gs.washington.edu/EVS/> accessed 27 March 2013), nor is it reported in the 1000 Genomes Catalogue of Human Genetic Variation (<http://www.1000genomes.org/ensembl-browser> accessed 27 March 2013).

#### KINDRED 6

A 42-year-old Scottish woman presented with punctate callosities affecting her palms and soles, which were present since early adulthood (Figure 1g and h).



**Figure 2. Family tree and microsatellite markers showing linkage patterns in kindred 6.** Two regions of microsatellite repeats were screened: one marker within intron 5 and a second marker within intron 1 of *AAGAB*. Numbers indicate the length (in base pairs) of PCR products for each marker from each allele. The microsatellite marker of intron 1 is absent from one allele of *AAGAB* in PPK1-affected individuals, indicating a genomic deletion resulting in hemizyosity.

The proband's mother was affected to a milder degree, and her 15-year-old son showed early signs of punctate keratoderma on the plantar surfaces. Sequencing of all exons of *AAGAB* in these three cases failed to identify any mutations. However, microsatellite linkage analysis in the pedigree was consistent with involvement of the *AAGAB* locus (Figure 2). Hemizygous inheritance of a microsatellite marker within intron 1 of *AAGAB* indicated a genomic deletion. A reduction in the gene dosage for *AAGAB* exon 1 in the proband compared with her unaffected father was confirmed using semiquantitative fluorescent PCR (Supplementary Figure S5a and b online), whereas exons 2 and 3 were amplified in equal quantities in the proband and control (Supplementary Figure S5c and d online). The precise size of this genomic deletion involving exon 1/intron 1 of *AAGAB* has not been defined, but it is consistent with the resultant haploinsufficiency of *AAGAB*.

It has previously been shown that knockdown of p34 in HaCaT cells *in vitro* leads to increased stabilization of the EGFR protein, a receptor tyrosine kinase (Pöhler *et al.*, 2012). In addition to EGFR, keratinocytes express several other receptor tyrosine kinases (RTKs). To further investigate the functional effects of *AAGAB* mutations, we tested the effect of gene knockdown on the transmembrane protein Axl, an RTK that is highly expressed in skin and is involved in signaling pathways to cellular proliferation (Postel-Vinay and Ashworth, 2012). We investigated the

effect of p34 knockdown on Axl in HaCaT keratinocytes and found a significant increase in the amount of Axl protein in cells treated with *AAGAB*-specific small interfering RNA (Supplementary Figure S6 online; methods are fully described in online Supplementary Tables S1 and S2 online).

These six kindreds demonstrate replication of a newly reported genetic cause for punctate PPK1, further confirming *AAGAB* as a causative gene. *AAGAB* mutations result in hereditary deficiency of p34, leading to hyperproliferative hyperkeratosis through increased growth factor signaling, which is thought to result from impairment in the endocytosis and recycling of EGFR proteins (Pöhler *et al.*, 2012).

A total of 13 predicted loss-of-function mutations have now been reported; these are listed in Supplementary Table S3 online. We have identified recurrent mutations resulting in frameshift and deleterious splice site changes, as well as previously unreported variants. There is no apparent genotype–phenotype correlation within this small group of patients, but our clinical observation is that environmental factors and personal skin care regimes affect the degree of plantar hyperkeratosis. The PPK1 families reported here demonstrate some diversity in the clinical appearance of plantar hyperkeratosis, whereas the punctate palmar keratoses have a more similar appearance in each kindred (Figure 1).

An association with the development of malignancy in PPK1 patients has

been reported, including breast and colonic adenocarcinoma (Bennion and Patterson, 1984; Stevens *et al.*, 1996). It is not clear whether there is truly an over-representation of malignancy in PPKP1 cases, and malignant neoplasms were not prevalent in the six kindreds reported here.

Hyperkeratosis of the plantar skin localized to the weight-bearing areas may be explained by mechanical trauma, but the extremely focal distribution of hyperkeratosis on the palmar skin in the disorder is striking. The mechanism(s) responsible for producing this very focal disease on the background of a mutation expressed throughout the palmar skin are likely to be of clinical relevance, but they remain to be defined. It is possible that a second mutation has occurred within each focal area of hyperkeratosis. This was not identified by microdissection and sequence analysis (Pöhler *et al.*, 2012), but further investigation is warranted. A clearer understanding of the genetic and/or environmental mechanisms by which focal keratoses arise may give further insight into the role of p34 in controlling cell division. Because of the role of EGFR and Axl in various malignancies, Axl-specific inhibitors are under development (Postel-Vinay and Ashworth, 2012) and are potentially of relevance to this rare but distressing keratoderma.

The p34 protein encoded by AAGAB has been functionally implicated in the biology of intracellular transport of membrane-bound, clathrin-coated vesicles. We hypothesized that a possible link between an inherited defect in vesicle transport and epidermal hyperproliferation might involve RTKs, which are turned over by endocytosis via clathrin- and AP2-dependent processes

(Ceresa, 2006; Rappoport and Simon, 2009). The upregulation of Axl protein expression after small interfering RNA knockdown of AAGAB demonstrates the involvement of p34 in RTK turnover in addition to EGFR. The identification of molecular mechanisms in palmoplantar keratoderma serves to improve our understanding of the pathogenesis of cutaneous hyperkeratosis. It may also offer insight into the control of keratinization in physiological conditions, as well as more prevalent inflammatory skin diseases characterized by hyperproliferation and hyperkeratosis.

#### CONFLICT OF INTEREST

The authors state no conflict of interest.

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#### SUPPLEMENTARY MATERIAL

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#### REFERENCES

- Bennion SD, Patterson JW (1984) Keratosis punctate palmaris et plantaris and adenocarcinoma of the colon. *J Am Acad Dermatol* 10:587–91
- Ceresa BP (2006) Regulation of EGFR endocytic trafficking by rab proteins. *Histol Histopathol* 21:987–93
- Gao M, Yang S, Li M *et al.* (2005) Refined localization of a punctate palmoplantar keratoderma gene to a 5.06-cM region at 15q22.2-15q22.31. *Br J Dermatol* 152:874–8
- Giehl KA, Eckstein GN, Pasternack SM *et al.* (2012) Nonsense mutations in AAGAB cause punctate palmoplantar keratoderma type Buschke-Fischer-Brauer. *Am J Hum Genet* 91:754–9
- Martinez-Mir A, Zlotogorski A, Londono D *et al.* (2003) Identification of a locus for type 1 punctate palmoplantar keratoderma on chromosome 15q22-q24. *J Med Genet* 40:872–8
- Pöhler E, Mamai O, Hirst J *et al.* (2012) Haploinsufficiency for AAGAB causes clinically heterogeneous forms of punctate palmoplantar keratoderma. *Nat Genet* 44:1272–6
- Postel-Vinay S, Ashworth A (2012) AXL and acquired resistance to EGFR inhibitors. *Nat Genet* 44:835–6
- Rappoport JZ, Simon SM (2009) Endocytic trafficking of activated EGFR is AP-2 dependent and occurs through preformed clathrin spots. *J Cell Sci* 122:1301–5
- Stevens HP, Kelsell DP, Leigh IM *et al.* (1996) Punctate palmoplantar keratoderma and malignancy in a four generation family. *Br J Dermatol* 134:720–6
- Zhang XJ, Li M, Gao TW *et al.* (2004) Identification of a locus for punctate palmoplantar keratoderma at chromosome 8q24.13-8q22.21. *J Invest Dermatol* 122:1121–5



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