#### Neuropeptide-mediated temporal sensory filtering in a primordial nervous system

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### 1 Abstract

2

3 Sensory filtering – prioritizing relevant stimuli while ignoring irrelevant ones – is crucial 4 for animals to adapt and survive in complex environments. While this phenomenon has 5 been primarily studied in organisms with complex nervous systems, it remains unclear 6 whether simpler organisms also possess such capabilities. Here, we studied temporal 7 information processing in Schmidtea mediterranea, a freshwater planarian flatworm with 8 a primitive nervous system. Using long-term behavioral imaging and oscillatory 9 ultraviolet (UV) light stimulations with rhythms matching the timescale of the animal's 10 short-term memory (~minutes), we observed that planarians initially ignored rhythmic 11 oscillations in UV intensity but eventually began tracking them after several cycles, 12 demonstrating sensory filtering. We identified two neuropeptides, knockdown of which 13 eliminated the initial ignoring phase and led to immediate stimulus-tracking, suggesting 14 that these neuropeptides mediate an active sensory gating mechanism preventing response to transient fluctuations in stimuli. Notably, when UV stimulation was coupled 15 16 with synchronous visible light oscillations, the planarians tracked the combined signals 17 immediately, indicating that coherence across sensory modalities can override the initial 18 gating. Our findings demonstrate that even simple nervous systems can filter temporal 19 information and that this mechanism is mediated by neuropeptides. Unlike classical fast-20 acting small-molecule neurotransmitters, neuropeptides provide a slower, sustained, and 21 global form of modulation that allows for more sophisticated control of sensory 22 processing.

# 23 Significance statement

- 24
- 25 We show that simple nervous systems can use specific neuropeptides to achieve sensory
- 26 filtering, a behavior previously thought to require complex brain architecture. This
- 27 neuropeptide-mediated sensory gating mechanism reveals a fundamental role for
- 28 neuropeptides in temporal information processing, offering insights into the mechanistic
- 29 and evolutionary origins of attention-like behaviors.

#### 30 Introduction

31

32 Animals inhabit dynamic environments where they continuously encounter sensory 33 inputs of various modalities and timescales, many of which may be irrelevant to their 34 immediate needs. To navigate these complex conditions, it is beneficial to selectively 35 respond to pertinent signals while filtering out extraneous information (1-3). This 36 filtering also applies to temporal patterns, allowing animals to distinguish transient and persistent stimuli (4). Extensive studies in humans and other mammals have revealed 37 38 complex neural circuits for sensory filtering (5-7). In invertebrates, such processes have 39 been characterized in insects like Drosophila, which also rely on advanced brain 40 structures such as mushroom bodies for sensory filtering (8-10). These observations have 41 led to the notion that sensory filtering involves intricate neural circuits and dynamics (6, 42 11), though the underlying molecular mediators remain largely unknown. This raises an 43 important question: Is sensory filtering exclusive to animals with intricate brains, or can it 44 also arise in simpler organisms with rudimentary neural structures? Addressing this 45 question may uncover core mechanisms of this important neural function and shed light 46 on its evolutionary origins.

47

Here, we studied the freshwater planarian *Schmidtea mediterranea*. The simplicity of its nervous system is such that whether it is a true brain or primitive cephalic ganglia is still contested. Nevertheless, it consists of canonical neural cell types expressing conserved neurotransmitters and receptors (12, 13), and drives basic behaviors such as phototaxis, thigmotaxis, and chemotaxis (14–16). Recently, we found that planarians have short-term memory lasting a few minutes (17), suggesting that they may process temporal information.

55

56 To explore sensory filtering, we examined how planarians respond to oscillatory

57 ultraviolet (UV) light stimulations with minute-scale rhythms, matching the timescale of

58 their memory. Surprisingly, we found that planarians initially ignored rhythmic

59 oscillations but eventually tracked them, while continuing to ignore irregular oscillations.

60 Using RNA interference (RNAi) to perturb key components of neural communications,

61 we identified two specific neuropeptides essential for the initial ignoring. Notably,

62 pairing UV stimulus with concurrently oscillating visible light, planarians followed the

63 rhythm immediately, indicating that coherent multisensory inputs can override the default

- 64 filtering behavior. Overall, these results suggest that even a simple system can filter
- 65 sensory information, which is governed by an active gating mechanism involving
- 66 neuropeptides. This function allows animals to delay tracking of stimulus rhythms only
- 67 after confirming their persistence.
- 68

## 69 **Results**

## 70 Long-term imaging reveals delayed tracking of rhythmic signals

71 To investigate how planarians process temporal information, we employed a long-term 72 imaging platform described in our prior work (17). This setup exposes planarians to 73 controlled stimuli over extended durations to precisely quantify their behavioral 74 responses across many individuals. We subjected planarians to 30-minute trials of 75 sinusoidal UV stimuli, with periods ranging between 2-4 minutes, as the planarian's 76 short-term memory peaks at  $\sim 3 \min (17)$ . We chose sinusoidal waves to avoid 77 discontinuities in the time derivative, which elicit strong aversive response in planarians 78 (17, 18). If planarians responded solely to the current stimulation, we would expect their 79 behavioral activity to track the UV oscillations, peaking in phase with the stimulus (Fig. 80 1A).

81

Upon UV exposure, planarians exhibited an immediate peak in behavioral activity, 82 83 measured by a scalar metric that quantifies high-dimensional behavioral output (17) (Fig. 84 1B). This initial peak did not represent stimulus-tracking, as a similar response occurred 85 upon exposure to constant UV (Fig. S1A). Following the initial reaction, activity levels 86 became relatively constant, indicating that the animals ignored the oscillations in 87 stimulation strength. Surprisingly, after five to six cycles, they started tracking the UV 88 oscillations, displaying clear peaks and troughs with a phase lag relative to the stimulus. 89 This phase lag demonstrates that planarians were not merely reacting to immediate 90 stimulation but were filtering temporal information based on its history. We quantified 91 the phase lags by calculating the time difference between stimulus and activity peaks or

92 troughs, normalized by the UV sine wave's periodicity (Fig. 1C). The lags were

93 consistently more pronounced in the troughs, revealing an asymmetry in the intrinsic

- 94 neural processing delay.
- 95

96 To understand which behaviors contributed to these activity patterns, we used a Hidden 97 Markov Model (HMM) to decompose behaviors into distinct movement types (17). This 98 analysis revealed that roaming and nodding behaviors accounted for the observed activity 99 oscillations, each showing a temporal profile that closely matched overall activity (Fig. 100 **S1B).** To determine whether the delayed stimulus-tracking resulted from a subset of 101 "responder" animals gradually synching, we performed principal component analysis 102 (PCA) on individual worms from multiple trials (Fig. S1C). The analysis showed no 103 distinct subpopulations, indicating that delayed tracking was a behavior consistent across 104 individuals. Additionally, when we aggregated data based on trials, we found no clear 105 evidence of long-term learning or memory across trials (Fig. S1D).

106

107 Reducing oscillation amplitude, while keeping the mean intensity constant, decreases the
108 animals' ability to track stimulations (Fig. 1D; Fig. S1E). At half amplitude, they began
109 tracking the stimulus only after ~8 cycles; at a quarter amplitude, the tracking was lost.

110 Finally, when exposed to UV oscillations with changing periods, planarians exhibited

111 high activity with no stimulus-tracking (Fig. 1E), suggesting that regular rhythmic input

112 is necessary to induce tracking.

113

These findings suggest that planarians ignore transient oscillations before committing to a sustained tracking response to consistent and persistent stimuli. The observed initial delay and phase lag in tracking demonstrate a sophisticated temporal processing behavior of this simple nervous system.

118

## 119 Neuropeptides mediate sensory filtering of oscillating signals

120 We hypothesized that the delayed tracking could result from either active gating or

121 gradual adaptation/learning. If gating were the mechanism, disrupting its molecular

122 mediators should prompt immediate tracking, whereas if adaption were involved,

123 disruption should impair or prevent tracking (Fig. 2A). To test these possibilities, we

124 used RNAi to knock down major neurotransmitters. Disrupting monoamine

125 neurotransmitters did not significantly alter the tracking behavior. For example,

- 126 knockdown of tyrosine hydroxylase (th), which inhibits dopamine synthesis (19), did not
- 127 abolish the ignoring phase though it dampened the amplitude of the activity oscillations
- 128 (Fig. S2A). Knockdown of choline acetyltransferase (*chat*), which blocks acetylcholine
- 129 synthesis (20), shortened the delay and exaggerated the response amplitude, consistent
- 130 with our previous findings that acetylcholine is a major inhibitory neuromodulator in
- 131 planarians (17), but it did not fully eliminate the ignoring behavior (Fig. 2B).
- 132

133 Given the essential roles of neuropeptides in regulating UV responses and short-term

134 memory in planarians (17), we targeted several abundant neuropeptides, including *eye53*,

135 *1020HH*, *spp-1*, and *ppp-1*, expressed in distinct cell types throughout the planarian brain

136 (21–23). Strikingly, knockdown of *ppp-1* or *spp-1* caused immediate and sustained

137 stimulus-tracking (Fig. 2C). Even during the first trial, *ppp-1* and *spp-1* RNAi animals

138 followed the oscillation without any initial delay (Fig. 2D). These results suggest that the

139 ignoring behavior is due to active sensory gating through these two specific

140 neuropeptides. It is worth noting that *ppp-1* and *spp-1* knockdowns did not alter the

141 temporal profiles of responses to short UV pulses (Fig. S2B) or constant UV exposure

142 (Fig. S2C), indicating that their role is specific to processing complex temporal

143 information rather than general UV sensitivity.

144

145 Together, our results demonstrate that specific neuropeptides modulate sensory gating, 146 delaying the tracking of oscillatory inputs. Given that neuropeptides are the largest and 147 most diverse class of signaling molecules and are evolutionarily ancient (24, 25), their 148 function in this primitive nervous system might represent a fundamental mechanism for 149 modulating animal behavior in dynamic environments.

150

## 151 Coherent multisensory inputs override gating

152 In natural environments, sensory inputs from multiple modalities coexist, which require

153 integration for appropriate behavioral responses. For planarians, UV light and visible

154 light are typically concurrent in their natural habitat of shallow water, presenting a natural

155 scenario where dual inputs must be processed together. S. mediterranea possesses distinct

156 ocular and extraocular photoreceptors that detect visible and UV light, respectively (15).

157 This separation allowed us to independently stimulate these two sensory modalities to

158 investigate how planarians integrate inputs sensed differently.

159

160 When exposed to oscillatory visible light (520 nm), planarians showed no stimulus

161 tracking (Fig. 3A), suggesting that visible light alone is insufficient to evoke a tracking

162 response, despite its ability to induce strong negative phototaxis (14). To our surprise,

163 when exposed to simultaneous oscillations of UV and visible light, planarians tracked the

164 combined stimuli without delay, similar to the response observed in *ppp-1* and *spp-1* 

165 knockdown conditions (Fig. 3B). This indicates that coherent multisensory inputs can

166 override the default sensory gating.

167

168 We further tested responses to constant UV light with oscillatory visible light and found

169 that planarians tracked visible light oscillations (Fig. 3C), but with a phase lead, as

170 though predicting upcoming changes (Fig. 3D). In contrast, combining oscillatory UV

171 and constant visible light replicated the delayed tracking observed for UV alone (Fig.

172 **3E**). Lastly, coupling UV and visible light oscillations with a phase shift of  $0.5\pi$ ,

173 eliminated stimulus-tracking (Fig. 3F), highlighting the importance of coherence between

174 sensory modalities for overriding filtering and guiding different behavioral outcomes.

175

These findings demonstrate that the sensory filtering mechanism in planarians is not
limited to UV stimulation alone but can be modulated or overridden by inputs from other
sensory modalities, highlighting the relevance of sensory filtering in differentiating
various types of multimodal signals. These insights may also help understand basic
principles of multisensory integration.

181

#### 182 **Discussion**

183 Our study demonstrates an unexpected simplicity underlying the seemingly complex

184 neural function of sensory filtering: it exists within a simple nervous system and can be

185 disrupted by knocking down individual neuropeptides or overridden by coherent

multisensory inputs. Since neuropeptides are abundantly used in organisms (24–26), our
results may have broad implications.

188

189 We propose two potential mechanisms through which neuropeptides mediate temporal 190 sensory filtering. First, unlike classical neuromodulators, which act rapidly at synapses, 191 neuropeptides can diffuse over long distances and persist in the extracellular space for 192 extended time (27, 28). This widespread diffusion could synchronize neural activity 193 across space and/or time, a process thought to be necessary for selective filtering or 194 attention-like behaviors (6). Alternatively, the observed gating might result from neural 195 circuits dedicated to integrating sensory inputs that are modulated by neuropeptides. The 196 relatively slow action of neuropeptides could naturally provide the nervous system with a 197 memory on the minutes timescale necessary for sensory filtering (9). The inertia to 198 changes in the neural state could prevent the organism from overreacting to transient 199 stimuli, allowing it to focus on persistent and relevant environmental cues. In either case, 200 our work highlights the function of neuropeptides in modulating behaviors that need to 201 evoke long-lasting temporal neural states (29).

202

203 What determines the timescale of filtering? It is plausible that the timescale reflects the 204 ecological significant cues in planarians' natural environment. Planarians lack vision, and 205 their light perception primarily relies on encoding light intensity and wavelength (18). 206 Abrupt changes in illumination may signal immediate threats, such as the sudden 207 appearance of predators or habitat boundaries, prompting instant response; even gradual 208 changes in light intensity, analogous to "looming stimuli" in mouse vision test (30), can 209 indicate a predator approaching or retreating thereby requiring a consistently active 210 response. Persistent rhythmic fluctuations, on the other hand, may convey different 211 information, such as variable light patterns caused by water movement, and demand a 212 different response. By filtering out initial fluctuations in sine waves, planarians can 213 allocate their limited neural resources to different aspects of the environmental cues 214 based on the persistence of temporal patterns.

216 Alternatively, our findings may reveal neural processes that are not directly linked to 217 specific environmental cues but are intrinsic to the operating mechanisms. In small 218 nervous systems, neuropeptides can have disproportionately large effects, potentially 219 inducing slow neural dynamics that influences global brain states. These small peptides 220 (<40 amino acids), packaged in dense core vesicles, are released in response to 221 depolarization at various sites along the neuron (28, 31). Their sizes and structural 222 features contribute to a longer half-life on the scale of hundreds of seconds (28, 32, 33), 223 and the lack of specific reuptake mechanisms means that neuropeptides remain in the 224 extracellular space longer (33). Assuming simple diffusion, we estimate that 225 neuropeptides can travel extracellularly and influence neural activity over distances of 226 >500 µm. While this distance is relatively small in large, complex brains, it is almost the 227 entire dimension of the planarian brain, implicating that peptides can induce global states 228 across the brain. Consistently, in animals of similar or even smaller sizes, such as 229 *Caenorhabditis elegans*, the neuropeptide connectome illustrates how neuropeptides can 230 bridge otherwise disconnected neural circuits, forming a dense and decentralized 231 signaling network (34). Indeed, in C. elegans, most active neurons in the brain participate 232 in continuous coordinated neural activity fluctuations on the scale of ~100 s to represent 233 various behaviors, including sensory-driven action selection (35, 36). 234 235 To determine whether neuropeptide-mediated filtering arises from modulation of neural

236 synchrony, specific circuit dynamics, or a combination of both requires direct

237 measurements of neural activity. Although technical limitations currently preclude *in vivo* 

calcium imaging in planarians (37), our behavioral paradigm using UV sine waves to

239 probe temporal filtering can be applied to other models with advanced genetic and

240 imaging tools (38, 39). Ultimately, understanding how neuropeptide-mediated processes

241 contribute to sensory processing across different species and contexts may inform general

242 principles of sensory filtering and its evolution.

#### 243 Figures

244



245

Figure 1. Planarians exhibit delayed stimulus-tracking in response to UV sinewaves.

A. Schematic of the behavioral measurement to study sensory filtering in planarians.
Two potential outcomes are illustrated: immediate or delayed stimulus-tracking.
B. Behavioral activity of planarians exposed to 30-min UV sine waves with four

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251

different periodicities. Highlighted sections: initial peak, ignoring phase, and stimulus-tracking. Blue lines: median activity; orange lines: stimulus profile.

C. Top: diagram depicting the phase shift calculation. Bottom: phase shift for peaks
and troughs during the last three cycles vs. stimulus periodicities.

255	D. Activity under UV sine waves at half amplitude, showing a longer ignoring phase.
256	The mean intensity of the stimulation is adjusted to match other conditions.
257	E. Exposed to UV cycles with randomly generated periodicities results in no
258	stimulus-tracking.
259	
260	Statistics: In (B, D-E), shaded regions: 95% confidence interval (CI); orange lines:
261	stimulus profile. In (C), dots represent averages of the last three peaks from combined
262	trials and experiments. The box-and-whisker plot shows the distribution of normalized
263	phase shifts: boxes, interquartile range (IQR); bar, median; whiskers, $1.5 \times IQR$ . Sample
264	sizes: in (B), 2.3-min period: 2 batches/135 trials; 2.7-min: 3 batches/104 trials; 3.0-min:
265	7 batches/388 trials; 3.2-min: 3 batches/177 trials; 3.4-min: 2 batches/237 trials; 3.8-min:
266	1 batch/50 trials; 4.0-min: 8 batches/374 trails. In (D), 3 batches/102 trails; and in (E), 3
267	batches/60 trails. Each animal is exposed to a maximum of 10 trials, with each trial

separated by a two-hour interval.



269

#### 270 Figure 2. Neuropeptides regulate temporal sensory filtering.

- A. Schematic illustrating two potential mechanisms for the delayed stimulus-tracking
  that can be distinguished through RNAi experiments: (1) first gradual adaptation,
  or (2) active gating.
- B. Activity of control (left) and *chat* RNAi (right) planarians under UV sine wave
  stimulation with a 3-min period. Blue lines: median activity; orange lines:
  stimulus profile.
- C. Activity of *ppp1* (left) and *spp1* (right) RNAi planarians exposed to sine waves
  with periods of 2.7, 3.0, and 3.3 min.
- D. Activity of *ppp1* (left) and *spp1* (right) RNAi planarians during the first trial of
  UV sine wave stimulation with a 3-min period.
- 281

- 282 Statistics: Shaded regions: 95% CI. Sample sizes: in (A), 2 batches/62 trials for control
- 283 RNAi; 1 batch/70 trials for chat RNAi; in (B), for ppp1 RNAi, 1 batch/50 trials for 2.7-
- 284 min period; 4 batches/169 trials for 3.0-min period; 1 batch/54 trails for 3.3-min period;
- for spp1 RNAi, 1 batch/52 trails for 2.7-min period; 3 batches/98 trails for 3.0-min
- 286 period; 1 batch/57 trials for 3.3-min period.



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### Figure 3. Coherent multisensory inputs override sensory filtering.

- 289 A. Activity under visible light sine wave with a 3-min period. Blue line: median 290 activity; green dashed line: visible light stimulus profile.
- 291 B. Activity in response to coherent UV and visible light stimulations, both applied as 292 sine waves with a 3-min period. Orange line: UV stimulus profile.
- 293 C. Activity in response to a constant UV stimulus, set to the mean intensity of the 294 sine wave stimulus, combined with a visible light sine wave with a 3-min period. 295 Highlighted: stimulus-tracking in the last three cycles. This condition induces a 296 phase lead in tracking behavior, indicated by the gray line.
- 297 D. Quantification showing the flip of phase shifts under conditions shown in (B, 298 blue) vs. (C, red). Box plot displays normalized phase shifts for the last three 299 peaks and troughs.
- E. Activity response to a continuous visible light stimulation combined with a UV 300 301 sine wave with a 3-min period.
- 302 F. Activity response to a UV and visible light sine waves with a  $0.5\pi$  phase shift.
- 303

- 304 Statistics: In (A-C, E-F), shaded regions: 95% CI. In (D), dots represent the average of
- 305 the last three peaks from combined trials and experiments. The box-and-whisker plot
- 306 shows the distribution of normalized phase shifts as in Fig. 1C. Sample sizes: 2
- 307 batches/79 trials (A); 2 batches/92 trials (B); 2 batches/108 trials (C); 2 batches/68 trials
- 308 (E); 2 batches/77 trials (F).

### 309 Materials and Methods

310

311 Animal care and maintenance. Asexual S. mediterranea were maintained in the dark at

- 312 20 °C in water containing 0.5 g/L Instant Ocean Sea Salts and 0.1 g/L sodium
- 313 bicarbonate. Behavior experiments used planarians of ~4 mm in length. Animals were fed
- 314 every 4-7 days and starved a minimum of 4 days before behavioral recording.
- 315

316 *Imaging setup.* The imaging setup, detailed in ref. (17), illuminated animals with an IR

317 light (850 nm) and recorded at 2 frames per second using a Raspberry Pi NoIR camera,

318 ensuring minimal interference with their natural behavior. UV stimuli (365 nm) were

delivered by a custom-built ring of 36 LEDs mounted above the camera to achieve

320 uniform illumination across the dish, and controlled by an Arduino Uno for precise

321 timing and intensity modulation. Visible light (520 nm) was similarly controlled and

delivered using the Adafruit NeoPixel RGB LEDs (model 1586).

323

324 In all stimulation experiments, we maintained a two-hour period of unstimulated, dark

325 time between repetitions of the protocols to prevent any influence between trials or

326 cumulative effects on behavior. A total of 24 hours of data was collected for each

327 experiment, corresponding to 10 trials.

328

329 **RNAi.** Gene knockdowns were performed by feeding animals double-stranded RNA

330 (dsRNA). The dsRNA was synthesized following the standard protocol(21) and fed to the

331 planarians via a liver homogenate at a concentration of  $\sim 100 \text{ ng/}\mu\text{L}$ . Clones for dsRNA

332 synthesis were created using oligonucleotide primers reported in ref. (17) and cloned into

- the vector pJC53.2 (Addgene plasmid ID: 26536) (21). Plasmids containing neuropeptide
- 334 sequences were from ref. (21).

335

336 For the RNAi experiments, animals were fed dsRNA 5-7 times at 4-5 days intervals. For

the controls, animals were fed dsRNA matching the ccdB and camR insert of pJC53.2 in

338 parallel. All animals were then starved for 4 days prior to decapitation, after which the

tails were allowed to regenerate and were imaged after regeneration completed (at 10-15

340 days post-amputation).

341

#### 342 Behavioral activity quantification

- 343 The methodology for quantifying planarian behavior was adapted from ref. (17).
- 344 Behavioral data were processed and analyzed to determine patterns of activity in response
- to UV and visible light stimuli. To assess statistical significance, a bootstrap resampling
- 346 method was employed with 1,000 bootstrap samples, allowing for reliable estimation of
- 347 confidence interval (CI) around the median activity values.
- 348
- 349 Data and code availability. Code for image segmentation is available at
- 350 github.com/samuelbray32/planameterization (https://doi.org/10.5281/zenodo.12697208).
- 351 Code for data analysis and visualization is available at
- 352 github.com/lwyss/timescales\_behavior. We acknowledge the use of ChatGPT for
- assistance in simplifying and annotating code used for plotting the activity data.

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#### 361 **References**

- 362 1. D. E. Broadbent, Task combination and selective intake of information. *Acta Psychol.* 363 (*Amst.*) 50, 253–290 (1982).
- 364 2. R. Desimone, J. Duncan, Neural mechanisms of selective visual attention. *Annu. Rev.* 365 *Neurosci.* 18, 193–222 (1995).
- 366 3. D. van Moorselaar, H. A. Slagter, Inhibition in selective attention. Ann. N. Y. Acad.
  367 Sci. 1464, 204–221 (2020).
- 368 4. A. Kohn, Visual adaptation: physiology, mechanisms, and functional benefits. *J.* 369 *Neurophysiol.* 97, 3155–3164 (2007).
- 5. G. Rees, G. Kreiman, C. Koch, Neural correlates of consciousness in humans. *Nat. Rev. Neurosci.* 3, 261–270 (2002).
- 6. P. N. Steinmetz, *et al.*, Attention modulates synchronized neuronal firing in primate
  somatosensory cortex. *Nature* 404, 187–190 (2000).
- 7. M. Nakajima, L. I. Schmitt, M. M. Halassa, Prefrontal cortex regulates sensory
  filtering through a basal ganglia-to-thalamus pathway. *Neuron* 103, 445–458.e10
  (2019).
- 8. L. Liu, R. Wolf, R. Ernst, M. Heisenberg, Context generalization in *Drosophila* visual learning requires the mushroom bodies. *Nature* 400, 753–756 (1999).
- 9. B. van Swinderen, Attention-like processes in *Drosophila* require short-term memory
  genes. *Science* 315, 1590–1593 (2007).
- 381 10. J. C. Theobald, B. J. Duistermars, D. L. Ringach, M. A. Frye, Flies see second-order
  382 motion. *Curr. Biol.* 18, R464–R465 (2008).
- 11. M. M. Chun, R. Marois, The dark side of visual attention. *Curr. Opin. Neurobiol.* 12, 184–189 (2002).
- 385 12. F. Cebrià, Regenerating the central nervous system: how easy for planarians! *Dev.*386 *Genes Evol.* 217, 733–748 (2007).
- 13. L. S. Wyss, S. R. Bray, B. Wang, Cellular diversity and developmental hierarchy in
  the planarian nervous system. *Curr. Opin. Genet. Dev.* 76, 101960 (2022).
- 14. T. Inoue, H. Hoshino, T. Yamashita, S. Shimoyama, K. Agata, Planarian shows
  decision-making behavior in response to multiple stimuli by integrative brain
  function. *Zool. Lett.* 1, 7 (2015).
- 392 15. N. Shettigar, *et al.*, Discovery of a body-wide photosensory array that matures in an
  393 adult-like animal and mediates eye-brain-independent movement and arousal. *Proc.*394 *Natl. Acad. Sci.* 118, e2021426118 (2021).

395 396	<ol> <li>K. G. Ross, <i>et al.</i>, SoxB1 activity regulates sensory neuron regeneration, maintenance, and function in planarians. <i>Dev. Cell</i> 47, 331–347.e5 (2018).</li> </ol>
397 398 399	<ol> <li>S. R. Bray, L. S. Wyss, C. Chai, M. E. Lozada, B. Wang, Adaptive robustness through incoherent signaling mechanisms in a regenerative brain. <i>Cell Rep.</i> 43, 114580 (2024).</li> </ol>
400 401 402	<ol> <li>N. Shettigar, <i>et al.</i>, Hierarchies in light sensing and dynamic interactions between ocular and extraocular sensory networks in a flatworm. <i>Sci. Adv.</i> 3, e1603025 (2017).</li> </ol>
403 404	19. K. Nishimura, <i>et al.</i> , Reconstruction of dopaminergic neural network and locomotion function in planarian regenerates. <i>Dev. Neurobiol.</i> <b>67</b> , 1059–1078 (2007).
405 406 407	<ol> <li>K. Nishimura, Y. Kitamura, T. Taniguchi, K. Agata, Analysis of motor function modulated by cholinergic neurons in planarian dugesia japonica. <i>Neuroscience</i> 168, 18–30 (2010).</li> </ol>
408 409	21. J. J. Collins, <i>et al.</i> , Genome-wide analyses reveal a role for peptide hormones in planarian germline development. <i>PLoS Biol.</i> <b>8</b> , e1000509 (2010).
410 411	22. T. Inoue, <i>et al.</i> , Morphological and functional recovery of the planarian photosensing system during head regeneration. <i>Zoolog. Sci.</i> <b>21</b> , 275–283 (2004).
412 413	23. M. Khariton, X. Kong, J. Qin, B. Wang, Chromatic neuronal jamming in a primitive brain. <i>Nat. Phys.</i> <b>16</b> , 553–557 (2020).
414 415	<ol> <li>G. Jékely, Global view of the evolution and diversity of metazoan neuropeptide signaling. <i>Proc. Natl. Acad. Sci.</i> 110, 8702–8707 (2013).</li> </ol>
416 417	<ol> <li>O. Mirabeau, JS. Joly, Molecular evolution of peptidergic signaling systems in bilaterians. <i>Proc. Natl. Acad. Sci.</i> 110, E2028–E2037 (2013).</li> </ol>
418 419	<ol> <li>G. Jékely, R. Yuste, Nonsynaptic encoding of behavior by neuropeptides. <i>Curr. Opin. Behav. Sci.</i> 60, 101456 (2024).</li> </ol>
420 421	27. M. P. Nusbaum, D. M. Blitz, E. Marder, Functional consequences of neuropeptide and small-molecule co-transmission. <i>Nat. Rev. Neurosci.</i> <b>18</b> , 389–403 (2017).
422 423	<ol> <li>A. N. van den Pol, Neuropeptide transmission in brain circuits. <i>Neuron</i> 76, 98–115 (2012).</li> </ol>
424 425	29. G. Mountoufaris, <i>et al.</i> , A line attractor encoding a persistent internal state requires neuropeptide signaling. <i>Cell</i> <b>187</b> , 5998–6015.e18 (2024).
426 427	<ol> <li>M. Yilmaz, M. Meister, Rapid innate defensive responses of mice to looming visual stimuli. <i>Curr. Biol.</i> 23, 20, 2011–1015 (2013).</li> </ol>

- 428 31. D. Shakiryanova, A. Tully, R. S. Hewes, D. L. Deitcher, E. S. Levitan. Activity429 dependent liberation of synaptic neuropeptide vesicles. *Nat. Neurosci.* 8, 173–178
  430 (2005).
- 32. S. X. Zhang, *et al.*, Stochastic neuropeptide signals compete to calibrate the rate of
  satiation. *Nature* 1–8 (2024). https://doi.org/10.1038/s41586-024-08164-8.
- 433 33. A. F. Russo, Overview of neuropeptides: awakening the senses? *Headache* 57, 37–46
  434 (2017).
- 435 34. F. Randi F, A. K. Sharma, S. Dvali, A. M. Leifer, Neural signal propagation atlas of
  436 *Caenorhabditis elegans. Nature* 623, 406–614 (2023).
- 437 35. C. M. Chai, H. Park, P. W. Sternberg, Brain-wide bidirectional neuropeptide
  438 modulation of individual neuron classes regulates a developmental decision. *Curr.*439 *Biol.* 32, 3365–3373.e6 (2022).
- 36. S. Kato, *et al.*, Global brain dynamics embed the motor command sequence of *Caenorhabditis elegans*. *Cell* 163, 656–669 (2015).
- 37. R. N. Hall, *et al.*, Heterologous reporter expression in the planarian *Schmidtea mediterranea* through somatic mRNA transfection. *Cell Rep. Methods* 2, 100298
  (2022).
- 38. R. N. Hall, *et al.*, A genetic and microscopy toolkit for manipulating and monitoring
  regeneration in *Macrostomum lignano*. *Cell Rep.* 43, 114892 (2024).
- 39. B. Weissbourd, *et al.*, A genetically tractable jellyfish model for systems and
  evolutionary neuroscience. *Cell* 184, 5854–5868.e20 (2021).

#### **Supplementary Figures** 450







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Time (min)

10 15 30

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#### 452

#### 453 Figure S1. Additional characterization of stimulus-tracking behavior.

454

D

A. Planarian activity under continuous UV stimulation for 30 min, matched in

- 455 average intensity to the sine wave stimulations. Blue line: median activity; orange 456 line: stimulus profile.
- 457 B. Observed probability of HMM states during UV sine wave stimulation with a 3-458 min period. Oscillations (stimulus-tracking) are clear in roaming and nodding 459 states.
- 460 C. PCA of individual animal activity, with each animal represented by a unique color. Data in (B, C, D) is from the 3-min period experiment shown in Fig. 1B. 461
- 462 D. Activity traces grouped by trial number for the first six trials, showing consistent 463 activity patterns with no apparent trial-to-trial differences or long-term learning 464 effects.



- 468 Statistics: In (A, E, D), shaded regions: 95% CI. Sample sizes: 1 batch/48 trails (A); 3
- 469 batches/92 trials (E). In (B), the explained variance by the first two PCs: initial peak
- 470 (0-4 min): PC1 = 26.62%; PC2 = 16.63%; ignoring phase (4-18 min): PC1 =
- 471 16.32%; PC2 = 10.65%; stimulus-tracking (18–30 min): PC1 = 26.40%; PC2 =
- 472 11.68%.
- 473



475 Figure S2. Additional characterization of UV responses in RNAi-treated animals.



- B. Activity of RNAi-treated animals in response to a 30-second UV pulse ending at
  time 0, which show no significant difference across conditions.
- 481 C. Activity of RNAi-treated animals under continuous 30-min UV exposure, with
  482 intensity matching the mean intensity of sine wave stimulations.
- 483

474

484 **Statistics**: Shaded regions: 95% CI. Sample size: in (A), 2 batches/87 trials; in (B), 2

- 485 batches/108 trials for control RNAi; 1 batch/60 trails for *ppp1* RNAi; 1 batch/96 trials for
- 486 spp1 RNAi; in (C), 2 batches/142 trials for control RNAi; 3 batches/136 trails for ppp1

487 RNAi; 2 batches/313 trials for *spp1* RNAi.