

CASE STUDY

Open Access

IL-17A is increased in the serum and in spinal cord CD8 and mast cells of ALS patients

Milan Fiala^{1*}, Madhuri Chattopadhyay², Antonio La Cava³, Eric Tse¹, Guanghao Liu¹, Elaine Lourenco³, Ascia Eskin⁴, Philip T Liu⁵, Larry Magpantay⁶, Stephen Tse¹, Michelle Mahanian¹, Rachel Weitzman¹, Jason Tong¹, Caroline Nguyen¹, Tiffany Cho¹, Patrick Koo¹, James Sayre⁷, Otoniel Martinez-Maza⁶, Mark J Rosenthal¹, Martina Wiedau-Pazos⁸

Abstract

The contribution of inflammation to neurodegenerative diseases is increasingly recognized, but the role of inflammation in sporadic amyotrophic lateral sclerosis (sALS) is not well understood and no animal model is available. We used enzyme-linked immunosorbent assays (ELISAs) to measure the cytokine interleukin-17A (IL-17A) in the serum of ALS patients (n = 32; 28 sporadic ALS (sALS) and 4 familial ALS (fALS)) and control subjects (n = 14; 10 healthy subjects and 4 with autoimmune disorders). IL-17A serum concentrations were 5767 ± 2700 pg/ml (mean \pm SEM) in sALS patients and 937 ± 927 pg/ml in fALS patients in comparison to 7 ± 2 pg/ml in control subjects without autoimmune disorders ($p = 0.008$ ALS patients vs. control subjects by Mann-Whitney test). Sixty-four percent of patients and no control subjects had IL-17A serum concentrations > 50 pg/ml ($p = 0.003$ ALS patients vs. healthy subjects by Fisher's exact test). The spinal cords of sALS (n = 8), but not control subjects (n = 4), were infiltrated by interleukin-1 β - (IL-1 β -), and tumor necrosis factor- α -positive macrophages (co-localizing with neurons), IL-17A-positive CD8 cells, and IL-17A-positive mast cells. Mononuclear cells treated with aggregated forms of wild type superoxide dismutase-1 (SOD-1) showed induction of the cytokines IL-1 β , interleukin-6 (IL-6), and interleukin-23 (IL-23) that may be responsible for induction of IL-17A. In a microarray analysis of 28,869 genes, stimulation of peripheral blood mononuclear cells by mutant superoxide dismutase-1 induced four-fold higher transcripts of interleukin-1 α (IL-1 α), IL-6, CCL20, matrix metalloproteinase 1, and tissue factor pathway inhibitor 2 in mononuclear cells of patients as compared to controls, whereas the anti-inflammatory cytokine interleukin-10 (IL-10) was increased in mononuclear cells of control subjects. Aggregated wild type SOD-1 in sALS neurons could induce in mononuclear cells the cytokines inducing chronic inflammation in sALS spinal cord, in particular IL-6 and IL-17A, damaging neurons. Immune modulation of chronic inflammation may be a new approach to sALS.

Background

Amyotrophic lateral sclerosis (ALS) is a paralyzing neurodegenerative disease, characterized by the loss of upper and lower motor neurons. A majority of cases are sporadic (sALS) and their cause remains unknown. Less than 10% of ALS cases are familial (fALS) with 20% of these cases linked to various mutations in the Cu/Zn mutant superoxide dismutase 1 (SOD-1) gene [1]. SOD-1 is an ubiquitous small cytosolic metalloenzyme that catalyzes the conversion of superoxide anions to

hydrogen peroxide [2]. A subset of familial ALS cases is characterized by mutant SOD-1 protein aggregates in neuronal inclusions [3], which have toxic properties and occur selectively in motor neurons. Recently, inclusions with misfolded SOD-1 forms [4] and a wild-type SOD-1 sharing aberrant conformation and pathogenic pathway with mutant SOD-1 [5] have also been identified in sporadic ALS spinal cord motor neurons, suggesting the possibility that misfolded SOD-1 auto antigens stimulate inflammation in sporadic ALS as well.

SOD-1 mutations have diverse effects on the structure, functional activity and native stability of SOD-1, but a common pathway has been proposed through the formation of SOD-1 aggregates in the spinal cords of

* Correspondence: Fiala@mednet.ucla.edu

¹Department of Medicine, David Geffen School of Medicine at UCLA and VA Greater Los Angeles Healthcare System, 650 Charles E. Young Dr. South, Los Angeles, CA 90095-1735, USA

Full list of author information is available at the end of the article

patients expressing SOD-1 mutations [6]. Emerging evidence suggests that protein misfolding and aggregation might be a common pathophysiological link between sALS and fALS. In symptomatic transgenic mice that over express mutant SOD-1, a number of misfolded forms of SOD-1 are present in the spinal cords including those that expose regions of SOD-1 normally buried such as the dimer interface, and some of these forms have been found in patients. An altered SOD-1 species was found within the anterior horns of sALS patients that likely originated from misfolded wild type SOD-1 [7], and oxidation of wild type SOD-1 produced a misfolded protein with toxic properties of mutant SOD-1 [8]. Recently, abnormally folded SOD-1 has been detected in the spinal cord inclusions of a subset of sALS patients [4]. Structural studies of the inclusions found in the spinal cords of transgenic ALS mice show that they are largely composed of SOD-1 fibrils [9,10]. These forms likely occur due to a lack of bound metal cofactors, such as copper and/or zinc, and the normal inter subunit disulfide bond, the posttranslational modifications that are critical for the exceptionally high stability and solubility of SOD-1. Soluble SOD-1, upon removal of bound metals, can be rapidly converted to amyloid fibrils by the reduction of the intramolecular disulfide bond, even in a small fraction of the protein [11].

Increased serum and CSF concentrations of cytokines in neurodegenerative diseases, such as Huntington disease [12] and Parkinson disease [13], are considered important in the disease pathogenesis even before the disease onset. In addition, non-neuronal glial cells contribute to ALS disease mechanisms [14], which is supported by transgenic mouse studies. Inflammatory cytokines, prostaglandin E2 and leukotriene B4, inducible nitric oxide synthase and NO were found in astrocytes from the G93A-SOD-1 mouse, an important model of human fALS [15]. Furthermore, adult microglia from mutant SOD-1 transgenic mice released tumor necrosis factor- α [16], which may stimulate IL-6 production from astrocytes and microglia leading to reactive gliosis in pathophysiological processes in the CNS [17]. However, the role of cytokines is not well understood in sALS patients, although previous studies highlighted a number of abnormal chemokines and cytokines, including CCL2 (MCP-1), interleukin-6 (IL-6), tumor necrosis factor- α (TNF- α), and recently, interleukin-17 (IL-17) and interleukin-23 (IL-23) in patients [18]. As recently suggested [19], some of these immune factors such as IL-6, and interleukin-13 (IL-13) positive T-cells, and IL-17A described herein, may be also useful as blood biomarkers for ALS.

The active role played by the immune system in ALS is revealed by the disrupted blood-brain barrier and the

presence of activated macrophages, microglia, dendritic cells, T cells and mast cells in the spinal cord of ALS patients [20,21]. Positron emission studies show microglial activation during all stages of the disease in the motor cortex and other areas of the brain correlating with disease severity [22]. Transgenic mice expressing mutant SOD-1 also show evidence of extensive microglial activation, such as the increased expression of pro-inflammatory factors including transforming growth factor- β 1, TNF- α and macrophage-colony stimulating factor [23]. Therefore, the pathway by which misfolded or aggregated SOD-1 triggers an inflammatory response is crucial to the role of inflammation in ALS.

Here, we report the cytokines induced by wild type and mutant SOD-1 proteins in peripheral blood mononuclear cells of ALS patients and extend the relevance of these studies by examining the immunopathology in the spinal cord of confirmed ALS patients (deceased). To our knowledge, these studies are the first to investigate how aggregated WT SOD-1 can trigger IL-1 β , IL-6 and IL-23 responses in human mononuclear cells, the cytokines participating in the induction of IL-17A. Our results suggest that the activation of chronic inflammation, including the IL-17A mediated pathway, a signature of autoimmune diseases such as multiple sclerosis [24], is also critical in ALS.

Methods

Study population

Blood specimens for cytokine testing were obtained under UCLA Institutional Review Board-approved protocols. Written informed consent was obtained from all patients or their representatives according to the Declaration of Helsinki. 32 ALS patients, 28 with sporadic ALS (mean age 57.8 years) and 4 familial ALS subjects, who do not have a known SOD-1 mutation (mean age 67 years), 10 normal controls (mean age 56.6 years), and 4 subjects with autoimmune disease (mean age 58.7 years) were enrolled in this study via the UCLA ALS clinic (Table 1). The patients were diagnosed as probable or definite ALS by the revised EL Escorial criteria [25] and electromyography. The mean score of the ALS functional rating scale of sALS patients was 32.3 ± 2.1 months and mean disease duration was 24.5 ± 3 months.

Spinal cord tissues and fluid

Immunohistochemistry was performed on paraffin-embedded thoracic spinal cord tissues of 8 sporadic ALS patients, 3 Alzheimer disease patients and 4 other subjects without neuropathology (Table 2), which were provided by the UCLA Brain Bank (four ALS, 3 Alzheimer disease, and 1 case without neuropathology) and the UCLA National Neurological AIDS Bank (NNAB)

Table 1 Demographic and cytokine information

	sALS	fALS	Control	Autoimmune disorder ^a
n	28	4	10	4
Age (years)	57.8 (2.5)	67.0 (4.7)	56.6 (9.6)	58.7 (13.0)
M/F ratio	14/14	2/2	7/3	4/0
Duration (months)	24.5 (3.0)	16.0 (4.2)	N/A	N/A
FRS	32.3 (2.1)	31.5 (2.7)	N/A	N/A
IL-17A ^b	64.2%*	75.0%	0.0%*	100.0%
IL-17A ^c	5767.4 (2700.4)	937.3 (927.8)	7 (4.8)	9993.6 (1865.5)
IL-17A ^d	1145.4 (317.9) ⁺	984 (581.6)	6.7 (2.1) ⁺	2815 (861.7)
IL-17A ^e	1126.0 (320.1) ⁺	1124.0 (594.9)	7.1 (2.4) ⁺	2074.0 (709.8)

Data are expressed as mean (± standard error of mean)

^a Patients with eczema or asthma

^b IL-17% positive in the serum (sALS vs. healthy controls, $p = 0.003$)

^c IL-17 (pg/mL) serum level (ALS patients vs. normal controls, $p = 0.008$)

^d IL-17 (pg/mL) PBMC supernatant level (without stimulation)

^e IL-17 (pg/mL) PBMC supernatant level (with stimulation by fibrillar WT SOD-1 (2 µg/mL))

* $p < 0.003$ sALS vs. control

⁺ $p < 0.01$ sALS vs. control

(four ALS cases and 3 cases without neuropathology). UCLA NNAB also provided frozen lumbar and thoracic spinal cord sections of 3 ALS patients and 3 control non-ALS subjects without neuropathology.

Immunohistochemistry and immunofluorescence

Paraffin-embedded sections were deparaffinized, peroxidase activity was blocked with 3% hydrogen peroxide in methanol for 10 min, and subjected to heat-induced antigen retrieval at 95°C for 25 min. After dual endogenous enzyme block, they were stained by primary and secondary antibodies using Dakocytomation Envision[®]

System. Frozen sections were fixed with 4% paraformaldehyde, permeabilized with 1% Triton X-100, blocked with 1% bovine serum albumin, incubated with primary antibodies overnight at 4°C and secondary antibodies for 1 h at 37°C [26]. Primary antibodies were rabbit or mouse anti-CD3, anti-CD4, anti-CD8, anti-CD68 (Dako), goat anti-IL-1 β (Santa Cruz), goat anti-caspase 3 (Santa Cruz), mouse anti-IL-6 (Cymbus), mouse anti-IL-10 (DNAX), mouse anti-IL-17A (R&D Systems), goat anti-IL-17A (C20) (Santa Cruz), mouse anti-TNF- α (SantaCruz), mouse monoclonal anti-mast cell tryptase, clone AA1 (DakoCytomation), mouse anti-granzyme B, clone GrB-7(Chemicon), normal mouse or goat IgG. Secondary anti-mouse, anti-rabbit, or anti-goat antibodies were ALEXA-595 or ALEXA-488-conjugated (Invitrogen). Phalloidin was FITC or TRITC conjugated (Sigma). The preparations were independently examined by two observers (MF and GL) using the Olympus Research microscope with Hamamatsu camera or using Leica SP5X White Light Laser Confocal microscope.

Table 2 Demographic and diagnostic data of deceased patient providing spinal cord tissues

Subject	Age (at death)	Sex	Pathological Diagnosis
1	75 years	female	Sporadic amyotrophic lateral sclerosis
2	83 years	male	Sporadic amyotrophic lateral sclerosis
3	83 years	male	Sporadic amyotrophic lateral sclerosis
4	50 years	male	Sporadic amyotrophic lateral sclerosis
5	66 years	male	Sporadic amyotrophic lateral sclerosis
6	64 years	female	Sporadic amyotrophic lateral sclerosis
7	64 years	female	Sporadic amyotrophic lateral sclerosis
8	54 years	male	Sporadic amyotrophic lateral sclerosis
9	67 years	male	Alzheimer disease, Braak stage VI
10	90 years	male	Braak stage VI
11	76 years	male	Braak stage I
12	37 years	male	Angiosarcoma of pleura, no metastases
13	78 years	male	No neuropathology
14	62 years	female	No neuropathology
15	46 years	Male	No neuropathology

SOD-1 protein preparation and fibrillation

Wild-type and mutant SOD-1 (G37R, G93A, D101N) were expressed in *Saccharomyces cerevisiae* and purified using a combination of ammonium sulfate precipitation and hydrophobic interaction, ion exchange and size exclusion chromatography on Sephadex G75 column [27,28,11]. Purified SOD1, also known as "As-isolated SOD1" (AI SOD-1), was demetallated to generate APO-SOD-1 using multiple rounds of dialysis against EDTA. The metal content of APO- and AI- SOD1 was verified by Inductively-coupled plasma mass spectrometry (ICP-MS). Typically, AI SOD1 contained 2.5 equivalents of zinc and 0.5 equivalents of copper, while APO-SOD-

1 contained 0.5-0.8 equivalents of each metal per dimer. To convert apo-SOD-1 to fibrils, 50 μ M protein was incubated in 10 mM potassium phosphate (pH 7) and 5 mM DTT in a total volume of 200 μ L in a chamber of a 96-well plate, including a polytetrafluoroethylene (PTFE) ball (1/8 inch diameter) and 40 μ M thioflavin T [11]. The cytokine and plate was constantly agitated in a Fluoroskan microplate instrument (Thermo) at 37°C and fibril formation was monitored by thioflavin T fluorescence using λ_{ex} of 444 nm and λ_{em} of 485 nm. Fibril formation was indicated by a sigmoidal growth in fluorescence and verified by electron microscopy. For co-incubation with PBMC's, the SOD1 fibrils were precipitated by centrifugation at 16,000g for 15 minutes and resuspended in 10 mM potassium phosphate at pH 7. In some cases, fibrils were sonicated for 10 minutes in a bath sonicator at room temperature. To exclude contamination with endotoxin, endotoxin concentrations were determined using the Limulus amoebocyte lysate assay (LAL assay) (Associates of Cape Cod) by a quantitative kinetic assay. The endotoxin levels, 0.0448 ± 0.0000 pg/ml in the wild type superoxide dismutase 1, and 0.32 ± 0.08 pg/ml in the G37R superoxide dismutase 1, were below the concentrations active as a pyrogen.

Cytokine assays in fluids

Peripheral blood mononuclear cells were separated from heparin-an anticoagulated blood by Ficoll-Hypaque gradient centrifugation. The supernatant media of overnight cultures stimulated using the indicated SOD-1 protein (2 μ g/ml), amyloid- β 1-42 (2 μ g/ml) or medium with DMSO (1:1,000) were tested by multiplexed bioassays for human cytokines, the High Sensitivity Human Cytokine Panel - Pre-mixed 13 Plex (Millipore). The Luminescence platform assay panel simultaneously quantified supernatant concentrations of human IL-1 β , interleukin-2 (IL-2), interleukin-4 (IL-4), interleukin-5 (IL-5), IL-6, interleukin-7 (IL-7), interleukin-8 (IL-8), interleukin-10 (IL-10), interleukin-12 (IL-12), IL-13, interferon- γ , granulocyte-macrophage colony stimulating factor, TNF- α . Assay results, expressed in pg/ml, were obtained using a Bio-RAD BioPlex 200 dual laser, flow-based sorting and detection analyzer. IL-17A cytokine was assayed in peripheral blood mononuclear cell supernatant and serum by a human-specific ELISA (DuoSet) kit (R&D Systems). IL-23 was assayed by the Human IL-23 immunoassay (R&D Systems).

Whole-genome expression analysis was performed with the Affymetrix Gene Array ST 1.0. RNA from each sample was prepared using manufacturer recommended protocols and the Qiagen RNeasy columns. Each sample was labeled using standard protocols and reagents from Affymetrix. Probes were fragmented and hybridized

to the Affymetrix Human Gene 1.0 ST Array. Raw cell files generated from the Affymetrix Expression Console software were loaded into GeneSpring GX 10.0.2 software (Agilent) for analysis. We used the Robust Multi-array Analysis (RMA) probe summarization algorithm, with a transcript level of CORE.

Statistical testing was performed using the statistical software SPSS, Version 10.0, as follows: IL-17A in serum and IL-23 by non-parametric Mann-Whitney and Wilcoxon tests; supernatant cytokines by paired t-test analysis; the proportion of positive tests by the Fisher's exact test.

Results

Interleukin-17A in the blood of ALS patients

The cytokine concentrations of IL-17A were assayed in 28 sporadic and 4 familial ALS patients, 10 normal control subjects, and 4 subjects suffering from episodes of eczema or asthma. Mean IL-17A serum concentrations of sALS patients (5767 ± 2700 pg/ml) were significantly higher ($p = 0.008$ by Mann-Whitney and Wilcoxon tests) than those in normal controls without autoimmune manifestations (7 ± 4.8 pg/ml) (Table 1). Mean IL-17A serum concentrations in fALS patients were 937 ± 927 pg/ml. The level of IL-17A was increased above the highest observed level in control subjects of 40 pg/ml in 64% of sALS patients, 75% fALS patients, 100% subjects with immune disorders and 0% normal controls ($p = 0.003$ sALS vs. healthy controls). The production of IL-17A in sALS PBMC's was constitutive, i.e., unlike other cytokines, it was not increased by SOD-1 stimulation over 18 hr: not stimulated 1145 ± 317.9 pg/ml; stimulated with fibrillar wild type SOD-1, $1,126 \pm 320$ pg/ml. In a cross-sectional analysis, IL-17 concentrations were highest in early patients and appeared to decrease with duration of illness and FRS (N.S.) (Figure 1A, B). However, the serum and PBMC supernatant concentrations of IL-17A in two patients followed over three months appeared to be fluctuating (data not shown).

Immunopathology of the ALS spinal cord

T cells, macrophages/microglia and mast cells were the immune cells found in ALS and control spinal cords. The ALS spinal cord was infiltrated in patchy fashion by T cells and mast cells in the gray matter, and diffusely by macrophages/microglia in the gray and white matter (Figure 2).

In the gray matter, IL-17A was expressed in patchy fashion on several cell types: (1) CD8-positive CD3-positive T cells, (2) CD8-negative CD3-positive T cells, (3) mast cell tryptase-positive mast cells. Some CD3 cells expressed granzyme B or caspase-3 (Figure 3).

The gray and the white matter were diffusely infiltrated by CD68-positive macrophages/microglia. These

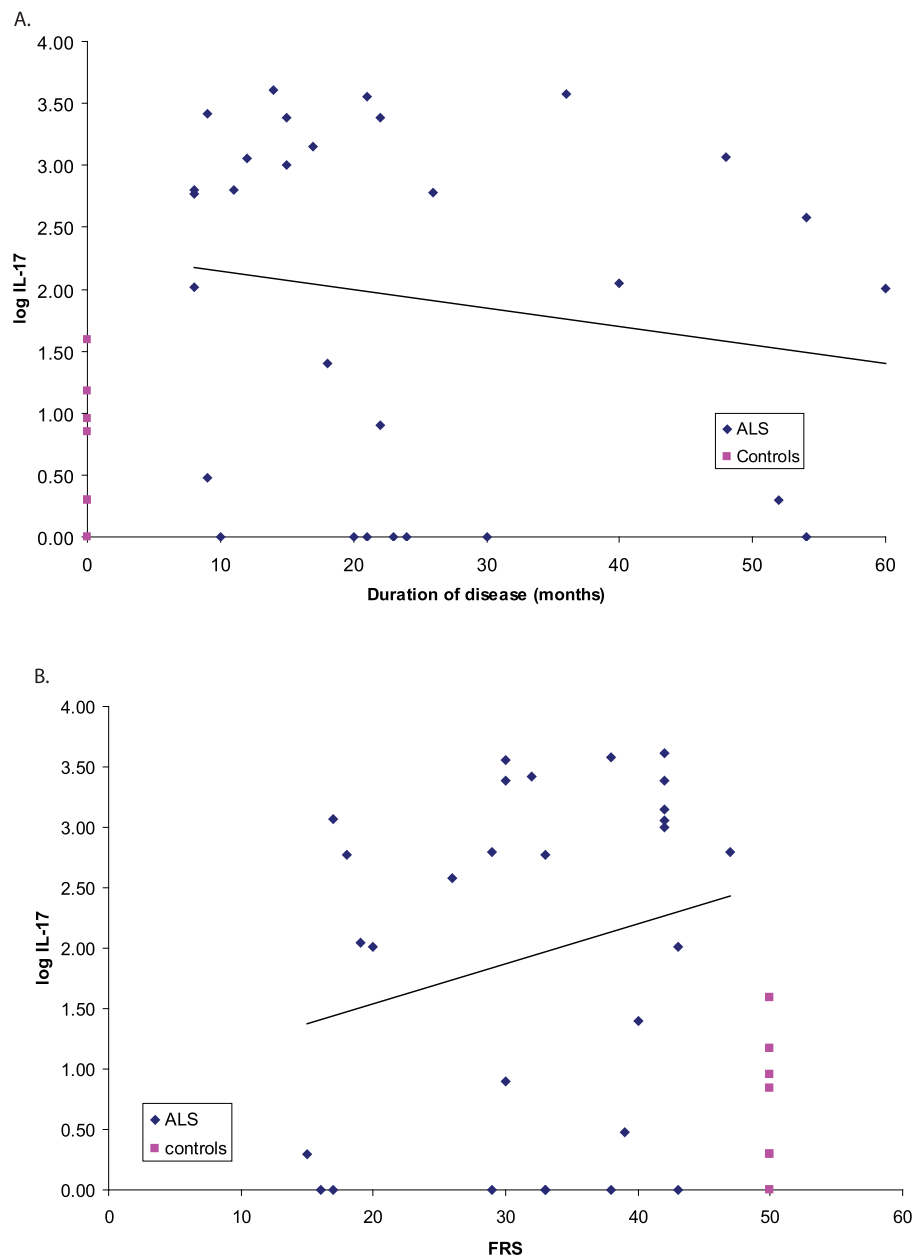


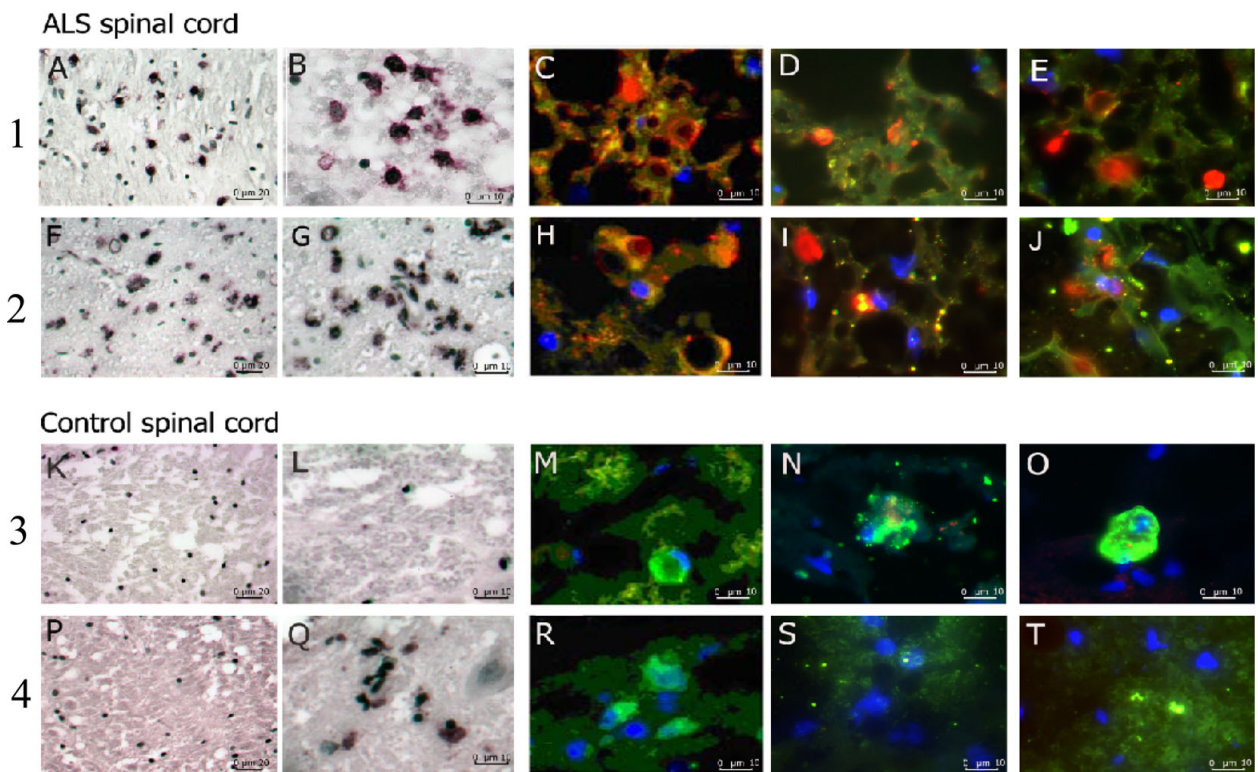
Figure 1 Relation of blood IL-17A levels to disease duration and ALS functional rating scale (FRS). **(A)** Duration vs. log IL-17, correlation = -0.16, $p = 0.38$. **(B)** FRS vs. log IL-17, correlation = 0.23, $p = 0.22$.

cells were IL-1 α and TNF- α -positive (Figure 2 and 4), and many co-localized with neurons (Figure 5).

In summary: Two observers examined 87 slides stained by CD3, CD8, IL-17A, CD68, IL-1 β , TNF- α and mast cell tryptase antibodies. IL-17A/CD3 T cells were found in five of 8 ALS spinal cords but not in any AD or control spinal cords; macrophages in 4 ALS spinal cords and two AD spinal cords; and mast cells in two ALS and three AD spinal cords (Figure 2U).

Induction of IL-1 β , IL-6 and IL-23 cytokines by fibrillar wild type and mutant SOD-1

To clarify whether certain physical forms of SOD-1 could induce the cytokines IL-1 β , IL-6 and IL-23, which can polarize T cells into the TH17 subset, we tested their induction by three forms of wild type and mutant SOD-1: as-isolated (AI), demetallated (APO), and fibrillar (Figure 6). These preparations had no significant levels of endotoxin (see Methods). IL-1 β and IL-6 are



U. Summary	ALS	AD	Control
CD 8 ^a	4/8	0/3	0/4
IL-17/ CD 8 ^a	4/8	0/3	0/4
IL-17/ CD 3 ^a	5/8	0/3	0/4
CD 68 ^a	4/8	2/3	0/3
IL-1β/ CD 68 ^a	3/3	ND	0/3
TNF-α/ CD 68 ^a	3/5	ND	0/3
Mast Cell Tryptase ^a	2/8	3/3	1/3

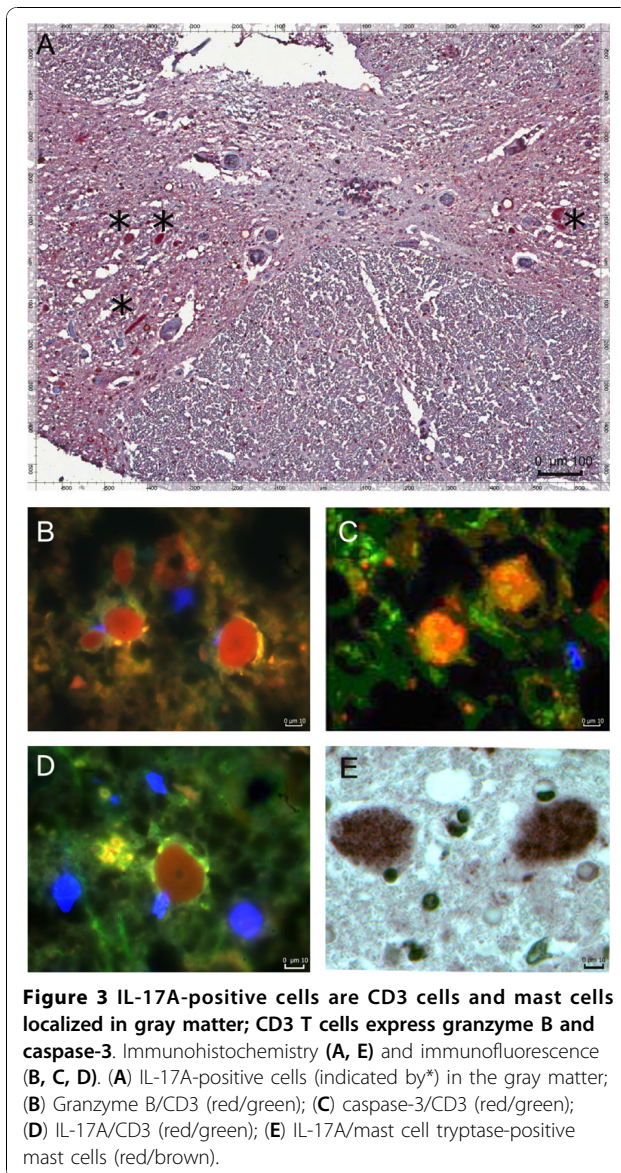
a. Number positive/Total number examined

Figure 2 IL-17A/CD8 T cells and TNF-α/CD68 cells infiltrate ALS spinal cord tissues. Sections of thoracic spinal cord of five ALS patients (A to J) and 5 control subjects (two Alzheimer disease patients (K, L, P, Q) and 3 patients without neurological disease (M, N, O, R, S, T) were stained by CD8 antibody (A to E), CD3 antibody (K to O) and CD68 antibody (F to J; P to T) using immunohistochemistry (IHC) (A, B, F, G, K, L, P, Q) or immunofluorescence (IFA) (C, D, E, H, I, J, M, N, O, R, S, T). The significant results were: **1.** CD8 in ALS gray matter: focal aggregates of IL-17A/CD8 T cells (red/brown by IHC (A, B) and red/green by IFA (C, D, E)) in 4/8 ALS spinal cords; **2.** CD68 in ALS gray and white matter: diffuse infiltration by TNF-α/CD68 cells (red/brown by IHC (F, G), and red/green by IFA (H, I, J)) in 4/8 ALS spinal cords; **3.** CD3 in control gray matter: Lack of IL-17A on CD3 cells (no red staining by IHC (K, L) and IFA (M, N, O)) in 3 AD and 4 control spinal cords; **4.** CD68 in control gray and white matter: No TNF-α on CD68 cells (no red staining by IHC (P, Q), and IFA (R, S, T)) in 3 control spinal cords; IL-17A/mast cell tryptase-positive cells (Fig. 4E) were found in three of 5 ALS and 3 of 3 AD spinal cords. Staining experiments with normal mouse or goat IgG and a secondary antibody were negative with all specimens.

the central pro-inflammatory cytokines and their induction by different physical forms of SOD-1 identifies these forms as responsible for inflammation. The dose-response experiment with PBMC's of two ALS patients and the wild type SOD-1 proteins showed induction by the fibrillar and APo forms, but not the AI form; and the experiment with the G37R and D101N SOD-1

mutant SOD-1 proteins, showed induction by the AI and APO forms, but not the fibrillar form.

IL-23 production was increased by stimulation with fibrillar SOD-1 of ALS patients' PBMC's (n = 6; mean ± S.E.M; not stimulated 52 ± 11.2 pg/ml, stimulated 123 ± 24.5 pg/ml; Wilcoxon p value = 0.0156) and control subjects' PBMC's (n = 3; mean ± S.E.M; not stimulated



27.6 ± 2.9 pg/ml, stimulated 188 ± 50 pg/ml; Wilcoxon $p = .1$) (data not shown).

To clarify the spectrum of cytokine responses in ALS patients, we tested induction of 13 supernatant cytokines in a sample of 29 patients by sham, amyloid- β , AI and fibrillar forms of wild type SOD-1, and the AI form of G37R SOD-1. The fibrillar wild type SOD-1 and AI G37R SOD-1, but not A β or AI wild type SOD-1, significantly increased six of the 13 tested cytokines in the supernatant media: IL-1 β , IL-6, IL-7, TNF- α , granulocyte-macrophage colony stimulating factor, and IL-10 (Figure 7). Stimulation of control PBMC's with SOD-1 forms induced the same six cytokines (data not shown).

Transcriptional stimulation by SOD-1 increases inflammatory cytokines in patients and the anti-inflammatory cytokine IL-10 in control subjects

The responses to mutant SOD-1 were analyzed at the transcriptional level by microarray hybridization. The analysis of 28,869 transcript cluster identifications in peripheral blood mononuclear cells of 2 controls and 3 patients (stimulated 18 hr by mutant SOD-1) showed strong stimulation of 7 cytokines (IL-10, IL-23A, granulocyte macrophage colony stimulating factor, IL-1 β , IL-1 α , IL-6 and IL-7) in both patients and control subjects (Figure 8). Of these, the increase in transcription of IL-1 α and IL-6 was ~ four-fold higher in patients compared to controls. In agreement with the constitutive production of IL-17A, its mRNA was not stimulated by G37R SOD-1. The chemokines CCL2, CXCL1, CXCL2, and CXCL3 were transcribed at a high level in patients as well as controls at baseline and after stimulation. G37R SOD-1 stimulated more CCL20 (5.5 fold), matrix metalloproteinase 1 (4.5fold), and tissue factor pathway inhibitor 2 (11.7 fold) in PBMC's of patients, as compared to control subjects. On the other hand, the anti-inflammatory cytokine IL-10 mRNA was stimulated more (2.0 fold) in PBMC's of control subjects, as compared to patients.

Discussion

Our results show that, in a cross-sectional study, 64% of sALS and fALS subjects have strongly increased serum concentrations of the cytokine IL-17A, compared to normal subjects, and the concentrations of IL-17A fluctuate, which could result in false-negative results in some subjects. The spinal cord of deceased sALS patients show a milieu in which polarization of CD3 cells to IL-17A-producing cells can develop in response to products of macrophages, T cells and mast cells; including IL-1 β , TNF- α , IL-6, IL-23, and probably eicosanoids (Figure 2 and 6). The cytokine IL-17A is pathogenic in inflammatory and autoimmune diseases such as multiple sclerosis [24], psoriasis, inflammatory bowel disease, systemic lupus erythematosus, and rheumatoid arthritis [29]. CD8 cells in gray matter might have a role in tissue destruction by cytotoxic cytokines, the cytotoxic molecules granzyme B, and nitric oxide (NO), resembling the role of CD8 cells in multiple sclerosis [24]. Although IL-17A is expressed on CD4 cells in the animal model of multiple sclerosis, experimental allergic encephalitis [24], IL-17A is expressed also on other cells, such as macrophages in asthma [30], CD8 cells in Behcet disease and psoriasis [31], and mast cells in rheumatoid arthritis synovium.

The ALS spinal cord is infiltrated by IL-17A-positive T cells and IL-17A-positive mast cells in gray matter and by TNF- α -positive macrophages/microglia in gray

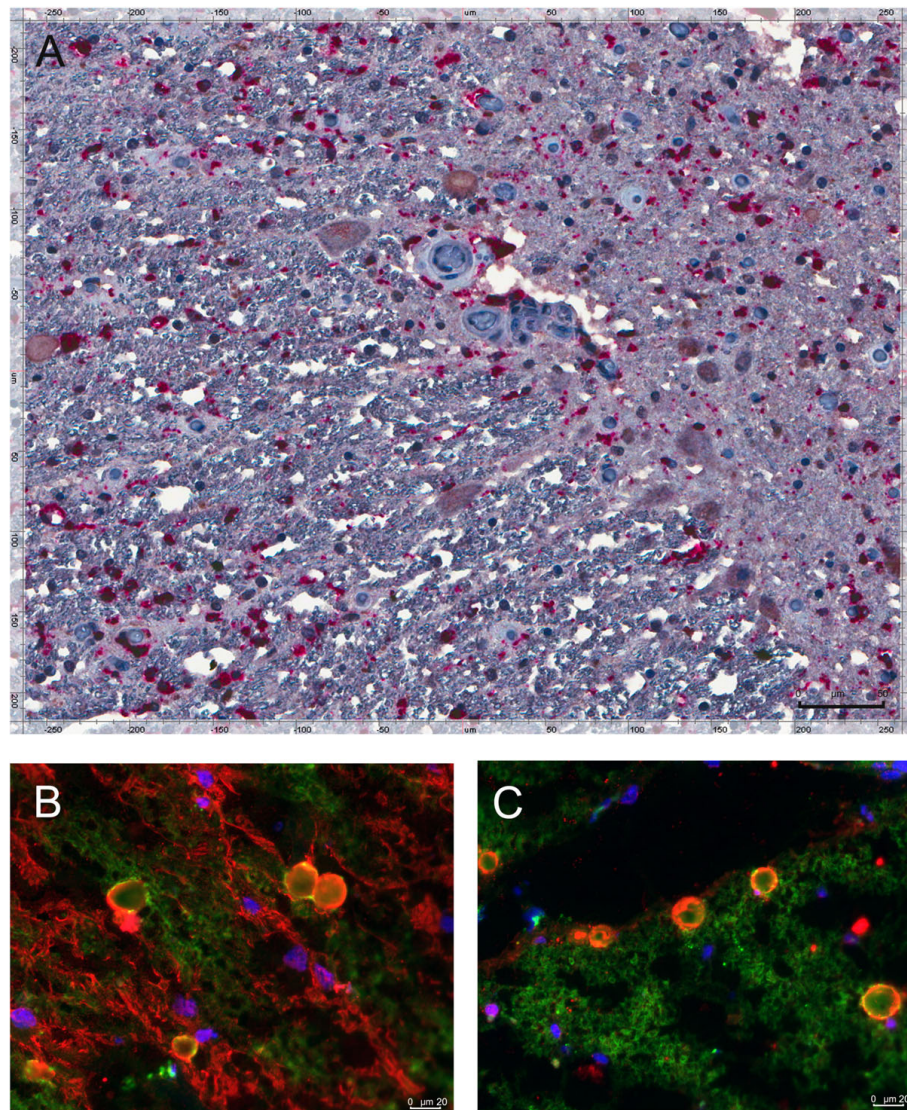


Figure 4 IL-1 β /CD68 macrophages infiltrate both the gray and the white matter of the ALS spinal cord. Immunohistochemistry (A); immunofluorescence (B, C). (A) IL-1 β /CD68 (red/brown) macrophages in the gray and the white matter; (B, C) IL-1 β /CD68 (red/green) macrophages.

and white matter (Figure 2, 3, 4). Macrophages/microglia were found to co-localize with neurons, reminiscent of previously demonstrated large phagocytic cells surrounding atrophic neurons [32]. Although the identification of these cells as macrophages or microglia is not possible since both are CD68-positive, blood-derived macrophages may penetrate into the ALS spinal cord, as suggested by their presence around the vessels with disrupted ZO-1 junctions [32], and into Alzheimer disease brain, as shown by their invasion across brain endothelial cells with disrupted ZO-1 junction [33].

To clarify the induction of IL-17A in ALS patients, we focused attention on IL-1 β , IL-6 [34], and IL-23, which are known to induce IL-17A and can be produced by

macrophages and/or dendritic cells (Figure 6). The development of IL-17A-producing TH17 cells is initiated by transforming growth factor- β and IL-6, which induce phosphor-STAT-3 and the transcription factor ROR γ t, and is stabilized and expanded by the cytokines IL-21 and IL-23 [35,36]. Human TH17 cell differentiation requires IL-6, IL-1 β and IL-21 or IL-23 [37]. In human studies, transforming growth factor- β has not been found to be essential [38]. In a recent mouse study, TH17 cells, which were induced by IL-1 β , IL-6 and IL-23, were more pathogenic than those induced in presence of transforming growth factor- β [39]. The cytokines and chemokines required for TH17 polarization, IL-1 α , IL-6, and CCL20, and matrix

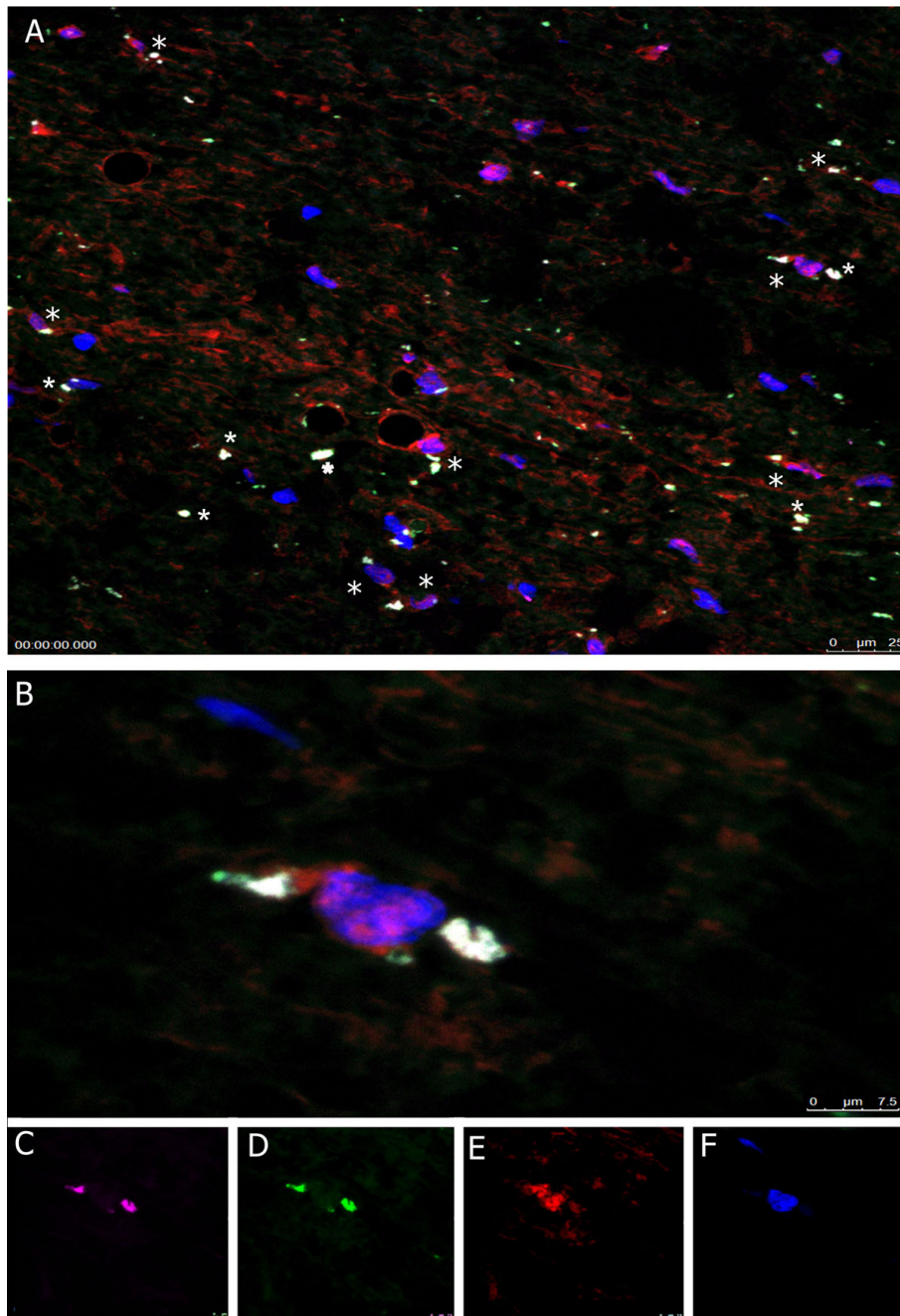
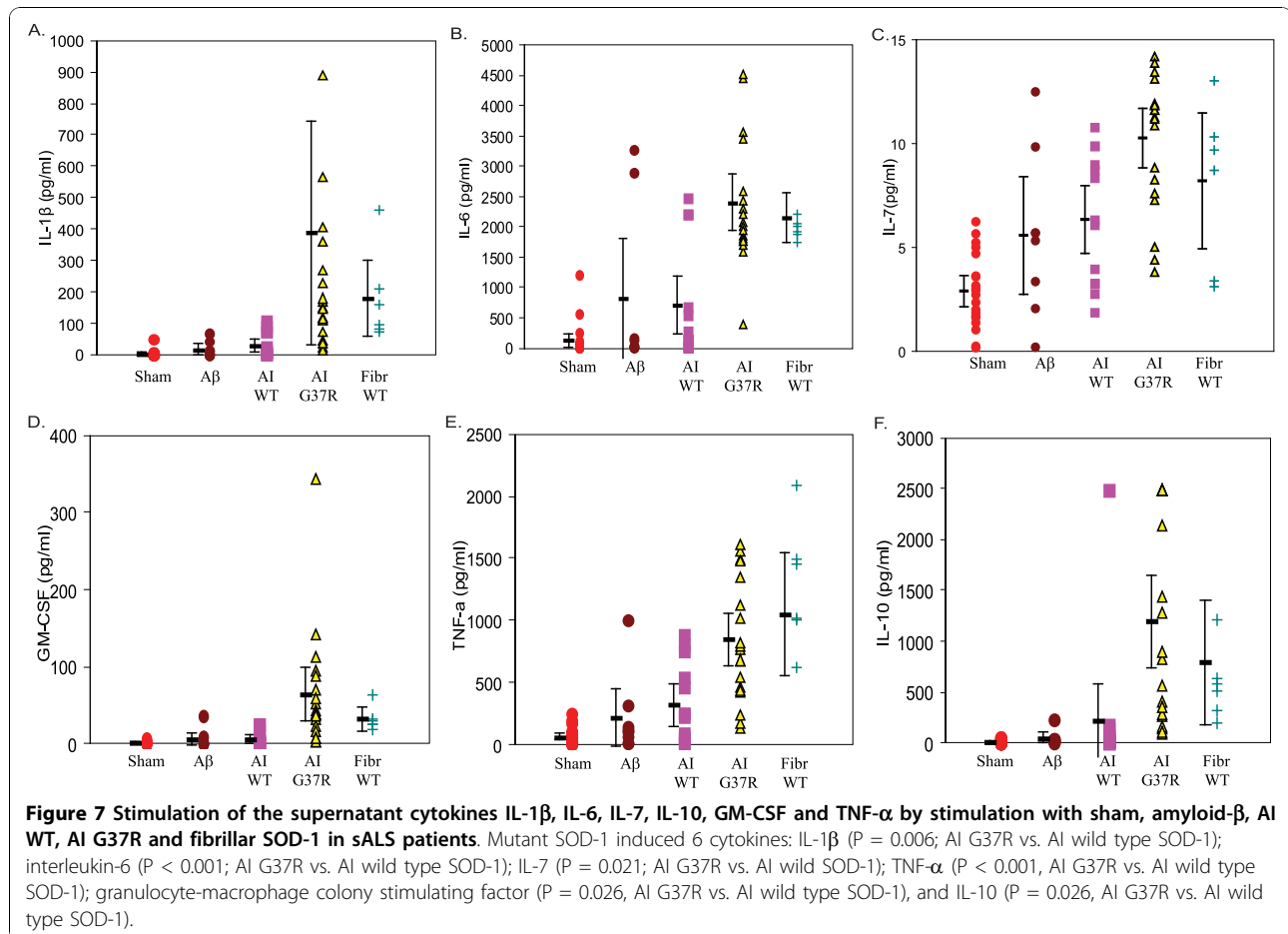
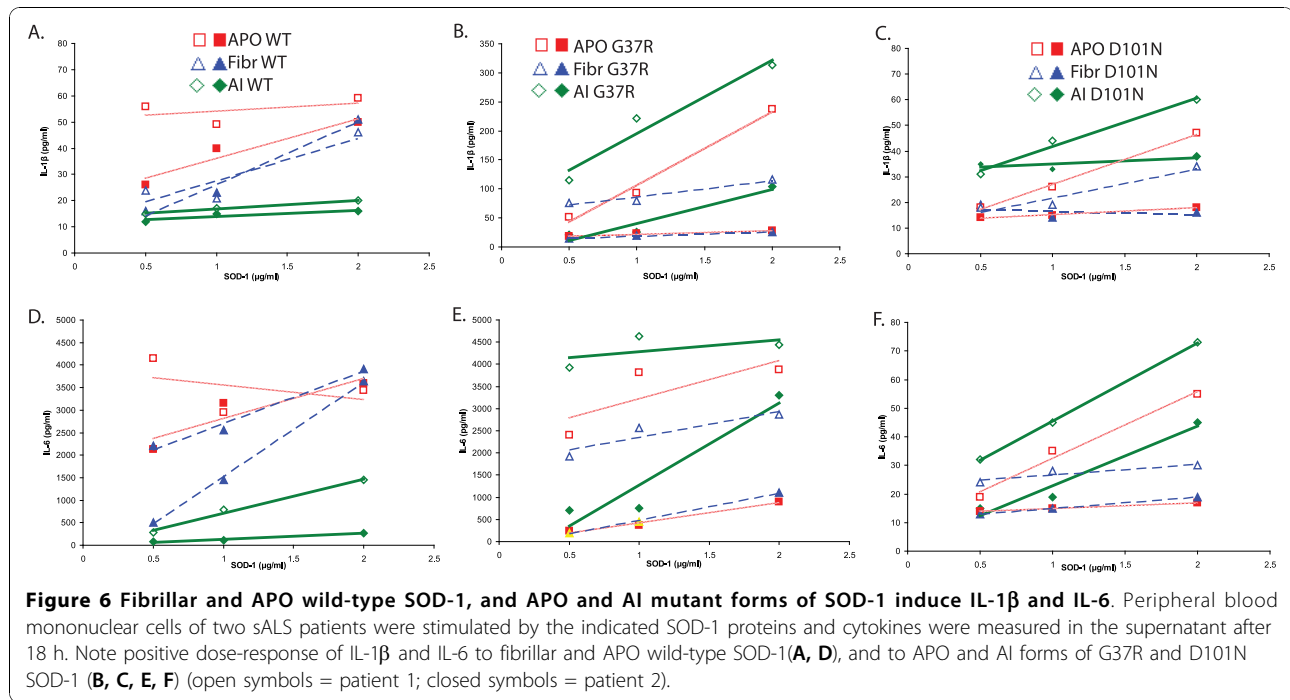


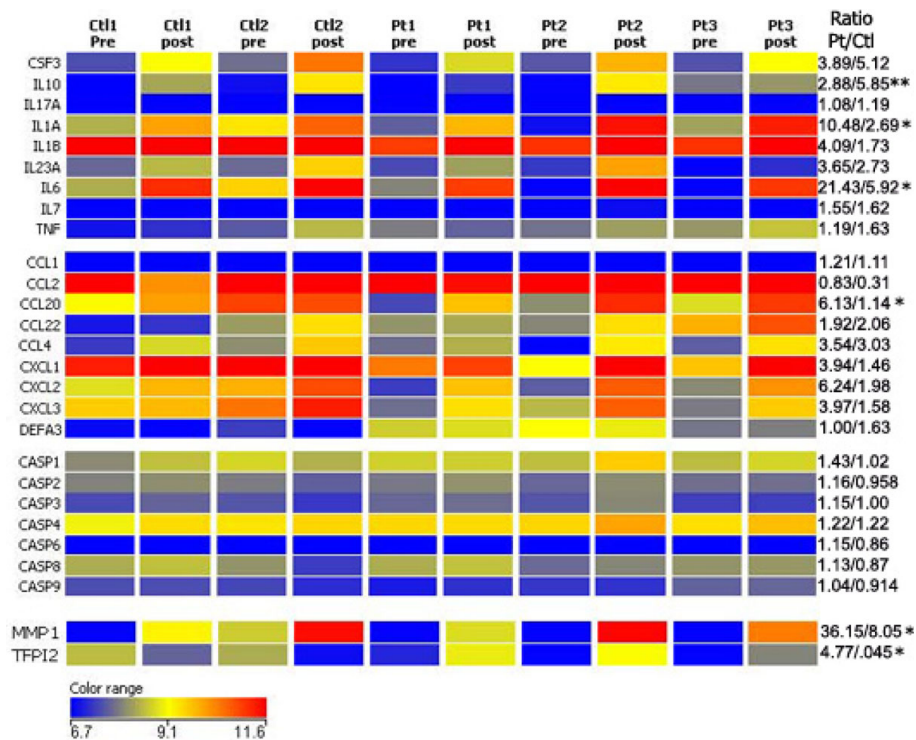
Figure 5 TNF- α /CD68 macrophages overlie neurons. Confocal microscopy of ALS spinal cord. **(A)** TNF- α /CD68 (magenta/green) macrophages overlie many NeuN/DAPI (red/blue) neurons and appear white (*) (40 \times); **(B)** Two macrophages (magenta/green) overlie a neuron (red/blue) (100 \times); **(C to F)** TNF- α (magenta); CD68 (green); neu-N (red); DAPI (blue).

metalloproteinase 1, were transcriptionally stimulated more in ALS patients than in controls (Figure 8).

The presence of IL-17A in mast cells in the spinal cord of patients with ALS and Alzheimer disease (Figure 3) has not been previously reported. Mast cells together with macrophages produce eicosanoids [40], which are important in polarization of the TH17 subset [41]. Mast

cells are emerging as master regulators with bi-functional role in both innate and adaptive immunity [42]. In the setting of autoimmunity, mast cells have a role in the initiation of the pathological immune response in experimental allergic encephalomyelitis through modulation of regulatory T cells into pathogenic Th17 cells [43]. Mast cells foster inflammation through the





* 4 fold increase in patients > controls
 ** 2 fold increase in controls > patients

Figure 8 Transcriptional upregulation by G37R SOD-1 of cytokines, chemokines, MMP1 and TFPI2. The values after Robust Multi-array Analysis (RMA) are displayed for patients and control subjects and the ratio of values (patient/control) for each gene is shown. Note the cytokines (*) increased and those (**) decreased in patients in comparison to controls.

production of IL-6 and the shift of regulatory T cells to TH17 cells [44].

Fibrillar and APO forms of wild type SOD-1, but not the AI form, induced the key cytokines, IL-1 β and IL-6, indicating their crucial role in inflammation of sALS patients (Figure 6). These autoantigens are likely present in the inclusions with non-native/misfolded forms of SOD-1, which are present in sporadic ALS spinal cords [4], and might be released from live or dying neurons [45,46] and be presented to autoimmune T cells by macrophages and dendritic cells.

Whole-genome expression analysis revealed that stimulation by SOD-1 increased in mononuclear cells of both patients and controls the transcription of cytokines, chemokines and matrix metalloproteinases. In patients' cells, however, the pro-inflammatory cytokines IL-1 α and IL-6 were enhanced more and the anti-inflammatory cytokine IL-10 was enhanced less than in controls' cells (Figure 8). The chemokines expressed at a high level even before stimulation include CCL2 (MCP-1), CXCL1 (GRO α), and CXCL3 (GRO γ). The chemokine CCL20 (MIP-3 α), a chemoattractant for CCR6, the marker of Th17 cells [47], was increased by SOD-1

stimulation more in patients' than in controls' cells. Matrix metalloproteinase 1, an effector of tissue remodeling [35], and tissue factor pathway inhibitor 2 were strongly stimulated in patients' cells, suggesting global pathology in ALS [48]. In agreement with the constitutive production of IL-17A in PBMC's, no increase in the transcription of IL-17A upon 18-hr SOD-1 stimulation was observed.

Conclusions

On the basis of these and previous observations [3,14,20], we propose the following immunopathogenesis of sporadic ALS (Figure 9): (a) aggregated wild type SOD-1 may induce chemokines and eicosanoids in a variety of neural cells, such as microglia, astrocytes and neurons; (b) The chemokines (CXCL1, CXCL3 and CCL2) and leukotrienes attract monocytes and T cells into the neuropil; (c) Aggregated SOD-1 stimulates macrophages and microglia to produce eicosanoids, tumor necrosis factor- α , IL-1 β , and IL-6, which (d) stimulate T cells to produce IL-6, IL-23 and other cytokines and chemokines; (e) IL-1 β , IL-6, CCL20 and IL-23 polarize CD8 (and CD4) T cells and mast cells to

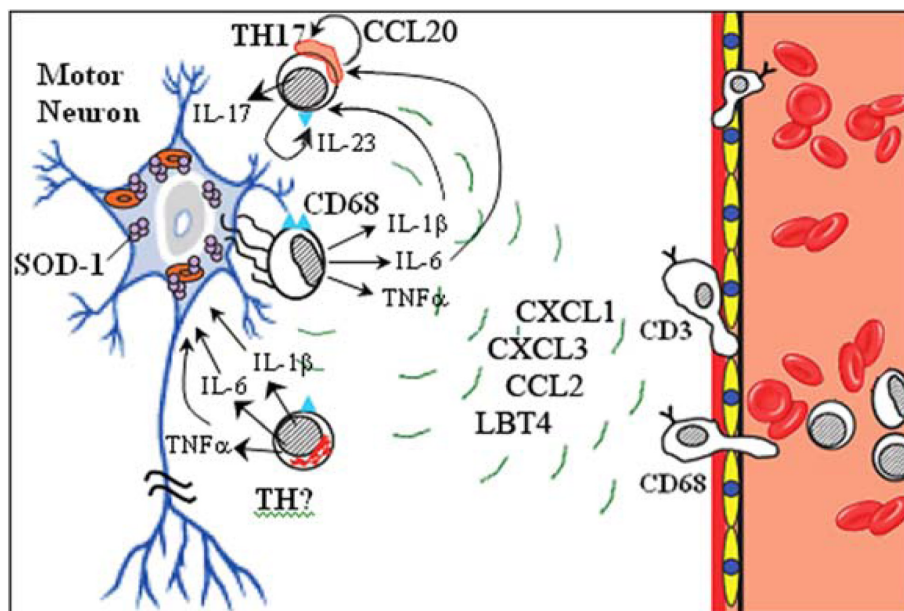


Figure 9 Putative model of autoimmune inflammation in the ALS spinal cord. Neurons, astrocytes and microglia induce chemokines when stimulated by aggregated superoxide dismutase 1. The chemokines (CXCL1, CXCL3 and CCL2) and leukotrienes attract monocytes to the neuropil. Aggregated SOD-1 stimulates macrophages and microglia to produce tumor necrosis factor- α , IL-1 α , IL-1 β , IL-6, and GM-CSF, which stimulate T cells to produce IL-23 and other cytokines. The cytokines IL-1 α , IL-1 β , IL-6, CCL20 and IL-23 polarize T cells and mast cells to produce IL-17A.

produce IL-17A; (f) CD8 T cells expressing granzyme B together with macrophages, mast cells, complement, NO, and other effector mechanisms cause neuronal apoptosis. Our results show a higher expression of IL-17A but lower expression of IL-10 in patients than controls, suggesting that the activation of regulatory T pathways is suppressed in patients, further suggesting a higher vulnerability of ALS patients to IL-17A-mediated damage. The discovery of IL-17A explains the chronic nature of inflammation in the ALS spinal cord and offers a new approach to therapy by immune modulation of inflammatory cytokines.

Acknowledgements

We thank P. M. Murphy and H. Vinters for review of the manuscript, the UCLA Brain Bank and the National Neurological AIDS Bank (NNAB) (funded by NS 38841 and MH 083500) for providing the tissues, the UCLA Muscular Dystrophy Core Center (funded by AR057230) for Microarray Data Analysis, and Nang Doan, UCLA Department of Pathology, for immunochemical staining. The Schwab family and the Vickter Foundation supported MW-P.

Author details

¹Department of Medicine, David Geffen School of Medicine at UCLA and VA Greater Los Angeles Healthcare System, 650 Charles E. Young Dr. South, Los Angeles, CA 90095-1735, USA. ²Department of Chemistry and Biochemistry, UCLA, 10833 Le Conte Ave, Los Angeles, 90024, USA. ³Department of Medicine, David Geffen School of Medicine at UCLA, 10833 Le Conte Ave, Los Angeles, 90024, USA. ⁴Department of Human Genetics, David Geffen School of Medicine at UCLA, 10833 Le Conte Ave, Los Angeles, 90024, USA. ⁵Department of Microbiology, Immunology and Molecular Genetics, David Geffen School of Medicine at UCLA, 10833 Le Conte Ave, Los Angeles, 90024, USA. ⁶Departments of Obstetrics & Gynecology and Microbiology,

Immunology & Molecular Genetics, David Geffen School of Medicine at UCLA, 10833 Le Conte Ave, Los Angeles, 90024, USA. ⁷Department of Biostatistics, UCLA School of Public Health, 10833 Le Conte Ave, Los Angeles, 90024, USA. ⁸Department of Neurology, David Geffen School of Medicine at UCLA, 10833 Le Conte Ave, Los Angeles, 90024, USA.

Authors' contributions

MF designed and supervised the study, analyzed the data, and wrote the paper. MC prepared SOD-1 proteins and helped to write the paper. AL, OM-M, MJR helped to write the paper. AE analyzed microarray data. ET, GL, EL, PTL, LM, ST, MMR, WJT, CN, TC, and PK performed the immune studies. JS performed statistical analysis. MWW examined the patients and helped to write the paper. All authors have read and gave approval for the publication of this final version of the manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 30 September 2010 Accepted: 9 November 2010

Published: 9 November 2010

References

1. Rosen DR, Siddique T, Patterson D, Figlewicz DA, Sapp P, Hentati A, Donaldson D, Goto J, O'Regan JP, Deng HX, et al: **Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis.** *Nature* 1993, **362**:59-62.
2. Bruijn LI, Miller TM, Cleveland DW: **Unraveling the mechanisms involved in motor neuron degeneration in ALS.** *Annu Rev Neurosci* 2004, **27**:723-749.
3. Chattopadhyay M, Valentine JS: **Aggregation of Copper-Zinc Superoxide Dismutase in Familial and Sporadic ALS.** *Antioxid Redox Signal* 2009, **11**(7):1603-14.
4. Forsberg K, Jonsson PA, Andersen PM, Bergemalm D, Graffmo KS, Hultdin M, Jacobsson J, Rosquist R, Marklund SL, Brannstrom T: **Novel antibodies reveal inclusions containing non-native SOD1 in sporadic ALS patients.** *PLoS One* 5:e11552.

5. Bosco DA, Morfini G, Karabacak NM, Song Y, Gros-Louis F, Pasinelli P, Goolsby H, Fontaine BA, Lemay N, McKenna-Yasek D, et al: **Wild-type and mutant SOD1 share an aberrant conformation and a common pathogenic pathway in ALS.** *Nat Neurosci* 2010, **13**(11):1396-403.
6. Shaw BF, Valentine JS: **How do ALS-associated mutations in superoxide dismutase 1 promote aggregation of the protein?** *Trends Biochem Sci* 2007, **32**:78-85.
7. Gruzman A, Wood WL, Alpert E, Prasad MD, Miller RG, Rothstein JD, Bowser R, Hamilton R, Wood TD, Cleveland DW, et al: **Common molecular signature in SOD1 for both sporadic and familial amyotrophic lateral sclerosis.** *Proc Natl Acad Sci USA* 2007, **104**:12524-12529.
8. Kabashi E, Valdmanis PN, Dion P, Rouleau GA: **Oxidized/misfolded superoxide dismutase-1: the cause of all amyotrophic lateral sclerosis?** *Ann Neurol* 2007, **62**:553-559.
9. Basso M, Massignan T, Samengo G, Cheroni C, De Biasi S, Salmons M, Bendotti C, Bonetto V: **Insoluble mutant SOD1 is partly oligoubiquitinated in amyotrophic lateral sclerosis mice.** *Journal of Biological Chemistry* 2006, **281**:33325-33335.
10. Sasaki S, Warita H, Murakami T, Shibata N, Komori T, Abe K, Kobayashi M, Iwata M: **Ultrastructural study of aggregates in the spinal cord of transgenic mice with a G93A mutant SOD1 gene.** *Acta Neuropathol* 2005, **109**:247-255.
11. Chattopadhyay M, Durazo A, Sohn SH, Strong CD, Gralla EB, Whitelegue JP, Valentine JS: **Initiation and elongation in fibrillation of ALS-linked superoxide dismutase.** *Proc Natl Acad Sci USA* 2008, **105**:18663-18668.
12. Bjorkqvist M, Wild EJ, Thiele J, Silvestroni A, Andre R, Lahiri N, Raibon E, Lee RV, Benn CL, Soulet D, et al: **A novel pathogenic pathway of immune activation detectable before clinical onset in Huntington's disease.** *J Exp Med* 2008, **205**:1869-1877.
13. Hofmann KW, Schuh AF, Saute J, Townsend R, Fricke D, Leke R, Souza DO, Portela LV, Chaves ML, Rieder CR: **Interleukin-6 serum levels in patients with Parkinson's disease.** *Neurochem Res* 2009, **34**:1401-1404.
14. Boillee S, Vande Velde C, Cleveland DW: **ALS: a disease of motor neurons and their nonneuronal neighbors.** *Neuron* 2006, **52**:39-59.
15. Hensley K, Abdel-Moaty H, Hunter J, Mhatre M, Mou S, Nguyen K, Potapova T, Pye QN, Qi M, Rice H, et al: **Primary glia expressing the G93A-SOD1 mutation present a neuroinflammatory phenotype and provide a cellular system for studies of glial inflammation.** *J Neuroinflammation* 2006, **3**:2.
16. Weydt P, Yuen EC, Ransom BR, Moller T: **Increased cytotoxic potential of microglia from ALS-transgenic mice.** *Glia* 2004, **48**:179-182.
17. Gruol DL, Nelson TE: **Physiological and pathological roles of interleukin-6 in the central nervous system.** *Mol Neurobiol* 1997, **15**:307-339.
18. Rentzos M, Rombos A, Nikolaou C, Zoga M, Zouvelou V, Dimitrakopoulos A, Alexakis T, Tsoutsou A, Samakovi A, Michalopoulou M, Evdokimidis J: **Interleukin-17 and interleukin-23 are elevated in serum and cerebrospinal fluid of patients with ALS: a reflection of Th17 cells activation?** *Acta Neurol Scand* 2010, **122**(6):425-9.
19. Turner MR, Kiernan MC, Leigh PN, Talbot K: **Biomarkers in amyotrophic lateral sclerosis.** *Lancet Neurol* 2009, **8**:94-109.
20. Graves M, Fiala M, Dinglasan L, NQ L, Sayre J, Chiappelli F, C vK, Vinters H: **Inflammation in amyotrophic lateral sclerosis spinal cord and brain is mediated by activated macrophages, mast cells and T cells.** *Amyotrophic lateral sclerosis* 2004, **5**:1-7.
21. Henkel JS, Engelhardt JI, Siklos L, Simpson EP, Kim SH, Pan T, Goodman JC, Siddique T, Beers DR, Appel SH: **Presence of dendritic cells, MCP-1, and activated microglia/macrophages in amyotrophic lateral sclerosis spinal cord tissue.** *Ann Neurol* 2004, **55**:221-235.
22. Turner MR, Cagnin A, Turkheimer FE, Miller CC, Shaw CE, Brooks DJ, Leigh PN, Banati RB: **Evidence of widespread cerebral microglial activation in amyotrophic lateral sclerosis: an [11C](R)-PK11195 positron emission tomography study.** *Neurobiol Dis* 2004, **15**:601-609.
23. Elliott JL: **Cytokine upregulation in a murine model of familial amyotrophic lateral sclerosis.** *Brain Res Mol Brain Res* 2001, **95**:172-178.
24. Tzartos JS, Friese MA, Craner MJ, Palace J, Newcombe J, Esiri MM, Fugger L: **Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis.** *Am J Pathol* 2008, **172**:146-155.
25. Brooks BR, Miller RG, Swash M, Munsat TL: **El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis.** *Amyotrophic Lateral Scler Other Motor Neuron Disord* 2000, **1**:293-299.
26. Zaghi J, Goldenson B, Inayathullah M, Lossinsky AS, Masoumi A, Avagyan H, Mahanian M, Bernas M, Weinand M, Rosenthal MJ, et al: **Alzheimer disease macrophages shuttle amyloid-beta from neurons to vessels, contributing to amyloid angiopathy.** *Acta Neuropathol* 2009, **117**:111-124.
27. Goto JJ, Gralla EB, Valentine JS, Cabelli DE: **Reactions of hydrogen peroxide with familial amyotrophic lateral sclerosis mutant human copper-zinc superoxide dismutases studied by pulse radiolysis.** *J Biol Chem* 1998, **273**:30104-30109.
28. Doucette PA, Whitson LJ, Cao X, Schrif V, Demeler B, Valentine JS, Hansen JC, Hart PJ: **Dissociation of human copper-zinc superoxide dismutase dimers using chaotrope and reductant. Insights into the molecular basis for dimer stability.** *J Biol Chem* 2004, **279**:54558-54566.
29. Kirkham BW, Lassere MN, Edmonds JP, Juhasz KM, Bird PA, Lee CS, Shnier R, Portek IJ: **Synovial membrane cytokine expression is predictive of joint damage progression in rheumatoid arthritis: a two-year prospective study (the DAMAGE study cohort).** *Arthritis Rheum* 2006, **54**:1122-1131.
30. Song C, Luo L, Lei Z, Li B, Liang Z, Liu G, Li D, Zhang G, Huang B, Feng ZH: **IL-17-producing alveolar macrophages mediate allergic lung inflammation related to asthma.** *J Immunol* 2008, **181**:6117-6124.
31. Raychaudhuri SK, Raychaudhuri SP: **Scid mouse model of psoriasis: a unique tool for drug development of autoreactive T-cell and th-17 cell-mediated autoimmune diseases.** *Indian J Dermatol* 2005, **55**:157-160.
32. Graves MC, Fiala M, Dinglasan LA, Liu NQ, Sayre J, Chiappelli F, van Kooten C, Vinters HV: **Inflammation in amyotrophic lateral sclerosis spinal cord and brain is mediated by activated macrophages, mast cells and T cells.** *Amyotrophic Lateral Scler Other Motor Neuron Disord* 2004, **5**:213-219.
33. Fiala M, Liu QN, Sayre J, Pop V, Brahmandam V, Graves MC, Vinters HV: **Cyclooxygenase-2-positive macrophages infiltrate the Alzheimer's disease brain and damage the blood-brain barrier.** *Eur J Clin Invest* 2002, **32**:360-371.
34. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, Weiner HL, Kuchroo VK: **Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells.** *Nature* 2006, **441**:235-238.
35. Awasthi A, Kuchroo VK: **Th17 cells: from precursors to players in inflammation and infection.** *Int Immunol* 2009, **21**:489-498.
36. Cornelissen F, Mus AM, Asmawidjaja PS, van Hamburg JP, Tocker J, Lubbers E: **Interleukin-23 is critical for full-blown expression of a non-autoimmune destructive arthritis and regulates interleukin-17A and RORgammat in gammadelta T cells.** *Arthritis Res Ther* 2009, **11**:R194.
37. Manel N, Unutmaz D, Littman DR: **The differentiation of human T(H)-17 cells requires transforming growth factor-beta and induction of the nuclear receptor RORgammat.** *Nat Immunol* 2008, **9**:641-649.
38. Acosta-Rodriguez EV, Napolitani G, Lanzavecchia A, Sallusto F: **Interleukins 1beta and 6 but not transforming growth factor-beta are essential for the differentiation of interleukin 17-producing human T helper cells.** *Nat Immunol* 2007, **8**:942-949.
39. Ghoreschi K, Laurence A, Yang XP, Tato CM, McGeachy MJ, Konkel JE, Ramos HL, Wei L, Davidson TS, Bouladoux N, et al: **Generation of pathogenic T(H)17 cells in the absence of TGF-beta signalling.** *Nature* 2010, **467**:967-971.
40. Boyce JA: **Mast cells and eicosanoid mediators: a system of reciprocal paracrine and autocrine regulation.** *Immunol Rev* 2007, **217**:168-185.
41. Sheibanie AF, Khayrullina T, Safadi FF, Ganea D: **Prostaglandin E2 exacerbates collagen-induced arthritis in mice through the inflammatory interleukin-23/interleukin-17 axis.** *Arthritis Rheum* 2007, **56**:2608-2619.
42. Galli SJ, Grimaldeston M, Tsai M: **Immunomodulatory mast cells: negative, as well as positive, regulators of immunity.** *Nat Rev Immunol* 2008, **8**:478-486.
43. Piconese S, Gri G, Tripodo C, Musio S, Gorzanielli A, Frossi B, Pedotti R, Pucillo CE, Colombo MP: **Mast cells counteract regulatory T-cell suppression through interleukin-6 and OX40/OX40L axis toward Th17-cell differentiation.** *Blood* 2009, **114**:2639-2648.
44. Tripodo C, Gri G, Piccaluga PP, Frossi B, Guarnotta C, Piconese S, Franco G, Vetri V, Pucillo CE, Florenza AM, et al: **Mast cells and Th17 cells contribute to the lymphoma-associated pro-inflammatory microenvironment of angioimmunoblastic T-cell lymphoma.** *Am J Pathol* 2010, **177**:792-802.
45. Zhao W, Beers DR, Henkel JS, Zhang W, Urushitani M, Julien JP, Appel SH: **Extracellular mutant SOD1 induces microglial-mediated motoneuron injury.** *Glia* 2008, **58**:231-243.

46. Urushitani M, Sik A, Sakurai T, Nukina N, Takahashi R, Julien JP: **Chromogranin-mediated secretion of mutant superoxide dismutase proteins linked to amyotrophic lateral sclerosis.** *Nat Neurosci* 2006, **9**:108-118.
47. Hirota K, Yoshitomi H, Hashimoto M, Maeda S, Teradaira S, Sugimoto N, Yamaguchi T, Nomura T, Ito H, Nakamura T, et al: **Preferential recruitment of CCR6-expressing Th17 cells to inflamed joints via CCL20 in rheumatoid arthritis and its animal model.** *J Exp Med* 2007, **204**:2803-2812.
48. Bossolasco P, Cova L, Calzarossa C, Servida F, Mencacci NE, Onida F, Polli E, Lambertenghi Delilieri G, Silani V: **Metalloproteinase alterations in the bone marrow of ALS patients.** *J Mol Med* 2010, **88**(6):553-64.

doi:10.1186/1742-2094-7-76

Cite this article as: Fiala et al.: IL-17A is increased in the serum and in spinal cord CD8 and mast cells of ALS patients. *Journal of Neuroinflammation* 2010 **7**:76.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

