Hindawi BioMed Research International Volume 2020, Article ID 9807918, 14 pages https://doi.org/10.1155/2020/9807918

Research Article

Development of Prognostic Indicator Based on Autophagy-Related IncRNA Analysis in Colon Adenocarcinoma

Weige Zhou, Shijing Zhang, Hui-biao Li, Zheyou Cai, Shuting Tang, Li-xia Chen, Jian-ying Lang, Zheng Chen, and Xin-lin Chen,

Correspondence should be addressed to Zheng Chen; chenzh68@mail.sysu.edu.cn and Xin-lin Chen; chenxlsums@126.com

Received 14 April 2020; Accepted 30 July 2020; Published 3 September 2020

Academic Editor: Bilal Alatas

Copyright © 2020 Weige Zhou et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

There were no systematic researches about autophagy-related long noncoding RNA (lncRNA) signatures to predict the survival of patients with colon adenocarcinoma. It was necessary to set up corresponding autophagy-related lncRNA signatures. The expression profiles of lncRNAs which contained 480 colon adenocarcinoma samples were obtained from The Cancer Genome Atlas (TCGA) database. The coexpression network of lncRNAs and autophagy-related genes was utilized to select autophagy-related lncRNAs. The lncRNAs were further screened using univariate Cox regression. In addition, Lasso regression and multivariate Cox regression were used to develop an autophagy-related lncRNA signature. A risk score based on the signature was established, and Cox regression was used to test whether it was an independent prognostic factor. The functional enrichment of autophagy-related lncRNAs was visualized using Gene Ontology and Kyoto Encyclopedia of Genes and Genomes. Ten prognostic autophagy-related lncRNAs (AC027307.2, AC068580.3, AL138756.1, CD27-AS1, EIF3J-DT, LINC01011, LINC01063, LINC02381, AC073896.3, and SNHG16) were identified to be significantly different, which made up an autophagy-related lncRNA signature. The signature divided patients with colon adenocarcinoma into the low-risk group and the high-risk group. A risk score based on the signature was a significantly independent factor for the patients with colon adenocarcinoma (HR = 1.088, 95%CI = 1.057 - 1.120; P < 0.001). Additionally, the ten lncRNAs were significantly enriched in autophagy process, metabolism, and tumor classical pathways. In conclusion, the ten autophagyrelated lncRNAs and their signature might be molecular biomarkers and therapeutic targets for the patients with colon adenocarcinoma.

1. Introduction

Colorectal cancer (CRC) ranked third in incidence and second in mortality of all types of cancers worldwide [1]. It is estimated that more than 1.4 million new cases will be diagnosed with CRC, and approximately 53,200 will die of the disease in 2020 [2]. Antineoplastic protocols included endoscopic and surgical local excision, radiotherapy and systemic treatment, local ablative therapies, targeted therapy, immunotherapy, and palliative chemotherapy [3]. Although these

treatments had dramatic progress, the 5-year relative survival rate for colon cancer was 64% [2]. Colon adenocarcinoma (COAD) is the most ordinary histological subtype of CRC [4].

Autophagy, a multistep lysosomal degradation process which promoted metabolic adaptation and nutrient circulation, has been widely studied and demonstrated to be involved in cancer development [5]. In both physiological and pathological situations, autophagy is central to the maintenance of organismal homeostasis. Accordingly,

¹School of Basic Medical Science, Guangzhou University of Chinese Medicine, Guangzhou, China

²The First Affiliated Hospital, Guangzhou University of Chinese Medicine, Guangzhou, China

³The First Clinical College, Guangzhou University of Chinese Medicine, Guangzhou, China

 $^{^4}$ Department of Stomatology, The Third Affiliated Hospital of Sun Yat-sen University, Guangzhou, China

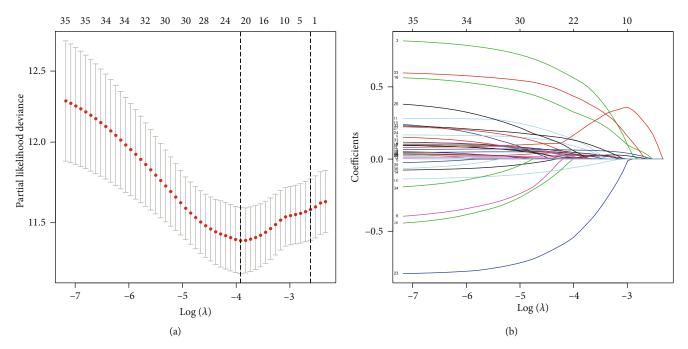


FIGURE 1: Autophagy-related lncRNA selection utilizing Lasso model. (a) Lasso coefficient values of 21 autophagy-related lncRNAs in colon adenocarcinoma. The vertical dashed lines are at the optimal log (lambda) value. (b) Profiles of Lasso coefficients.

disruption of autophagy is highly related to clinically relevant diseases as diverse as cancer, neurodegenerative disease, pathogen infection, and heart disease [6]. Recently, a large number of findings revealed that autophagy had multiple functions in occurrence, maintenance, and development of tumors [7]. Genetic evidence suggested that autophagy was a tumor-suppressor mechanism, and it was also clear that autophagy could promote tumor survival in the response to chemotherapy and under stressful conditions [8]. For the past few years, some researchers strived to find new targeted therapeutic strategies for colon adenocarcinoma by investigating autophagy pathways [9].

Long noncoding RNAs (lncRNAs) were found to perform a wide range of functions in various important biological processes, including cell proliferation and differentiation, genetic regulation of gene expression, RNA attenuation, RNA splicing, protein folding, and microRNA (miRNA) regulation [10]. In CRC, autophagy was known to have dual and contradictory roles in carcinogenesis, but the precise mechanisms resulting in autophagy in cancer were not yet fully verified [11, 12]. lncRNAs were involved in the development, invasion and metastasis, prognosis, and the chemoresistance of colon adenocarcinoma via modulating autophagy [13-16]. These studies focused on single or a few lncRNAs for colon adenocarcinoma [13-16]. The lncRNAs' expression profiles of TCGA database were not performed to explore novel biomarkers for forecasting the prognosis of colon adenocarcinoma. Therefore, we aimed to utilize TCGA database to establish autophagy-related lncRNA signatures and seek new biomarkers to predict the prognosis of the patients with colon adenocarcinoma.

2. Materials and Methods

2.1. Datasets and Sample Extraction. The RNA sequencing (RNA-seq) data about colon adenocarcinoma from The Cancer Genome Atlas (TCGA) database (https:// portal.gdc.cancer.gov/) were obtained. The inclusion criteria were set as follows: (1) patients were diagnosed as colon adenocarcinoma; (2) patients had complete lncRNA data and clinical information. According to the inclusion criteria, 480 patients with colon adenocarcinoma were included. In addition, the patient's complete clinical information was downloaded from TCGA. While screening clinical information, samples with a follow-up time of less than 30 days were excluded. Since the data involved in this study were all from the TCGA database and strictly follow the TCGA publication guidelines (http://cancergenome.nih.gov/abouttcga/policies/ publicationguidelines), there is no need for approval by the ethics committee.

2.2. Screening of lncRNAs and Autophagy-Related Genes. The profiles of lncRNAs were acquired from all the RNA-seq dataset. The total RNA expression data were standardized through log2 transformation. The autophagy-related gene list was downloaded from the Human Autophagy Database (HADb, http://autophagy.lu/clustering/index.html). The correlation between lncRNAs and autophagy-related genes was calculated using the Pearson correlation. The square of correlation coefficient $|R^2| > 0.3$ and P < 0.001 was considered to be autophagy-related lncRNAs. Finally, Cytoscape software 3.7.2 was used to visualize coexpression networks.

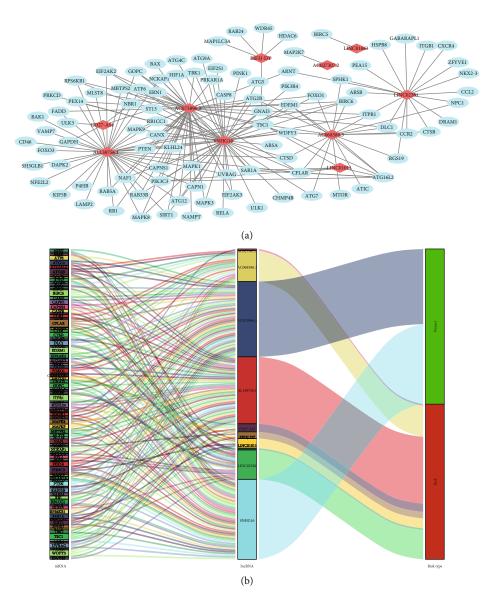


FIGURE 2: The coexpression network and Sankey diagram of prognostic autophagy-related lncRNAs. (a) The coexpression network between prognostic lncRNAs and autophagy-related genes in colon adenocarcinoma. Red diamond nodes represent prognostic lncRNAs, and the sky blue round nodes represent autophagy-related genes. The coexpression network was visualized using Cytoscape 3.7.2 software. (b) Sankey diagram showed the association between prognostic autophagy-related lncRNAs, autophagy-related genes, and risk types.

2.3. Identification of Prognostic Autophagy-Related lncRNAs. In the first place, the prognostic value of autophagy-related lncRNAs was assessed by univariate Cox regression. Autophagy-related lncRNAs with P < 0.05 in univariate analysis were incorporated into least absolute shrinkage and selection operator (Lasso) regression. Then, the results of Lasso were included into a multivariate Cox model in order to establish a risk score. We constructed a risk score based on a linear combination of the autophagy-related lncRNA expression levels multiplied with a regression coefficient (β) : risk score = $\sum_{i=1}^n \beta_i *$ (expression of lncRNA $_i$). Based on the median risk score, the patients were classified into two groups: high-risk and low-risk groups. The survival differences between the two groups were compared using the log-rank test.

TABLE 1: Multivariate Cox results of lncRNAs based on TCGA-COAD data.

lncRNA	Coefficient	HR	95% CI of HR
AC027307.2	0.097	1.102	0.986-1.198
AC068580.3	0.891	2.437	1.378-3.469
AC073896.3	-0.932	0.394	0.231-0.707
AL138756.1	0.205	1.227	0.960-1.455
CD27-AS1	0.083	1.087	0.979-1.179
EIF3J-DT	0.535	1.707	1.153-2.256
LINC01011	0.272	1.313	0.917-1.987
LINC01063	0.573	1.773	1.261-2.560
LINC02381	0.203	1.225	0.947-1.575
SNHG16	-0.132	0.877	0.767-1.035

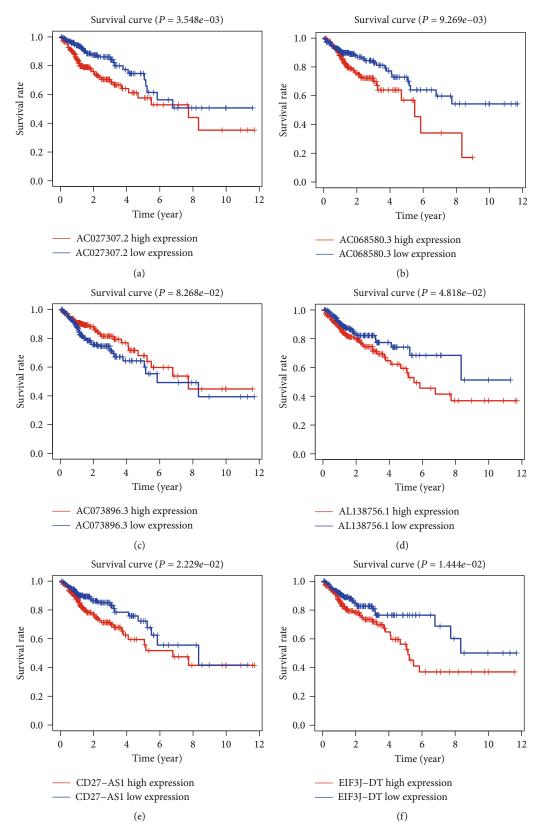


FIGURE 3: Continued.

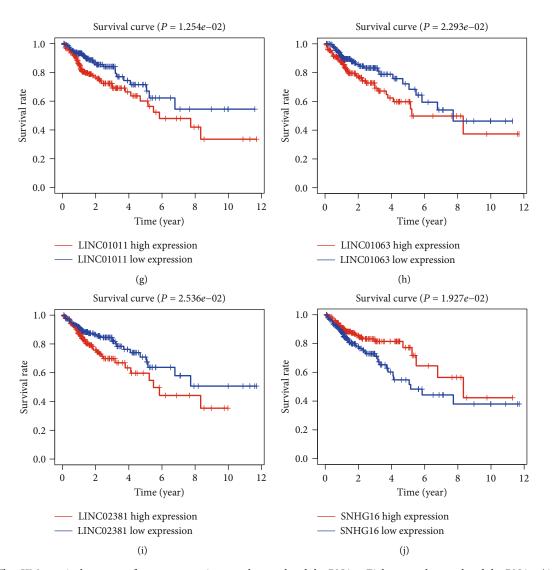


FIGURE 3: The KM survival curves of ten prognostic autophagy-related lncRNAs. Eight autophagy-related lncRNAs (AC027307.2, AC068580.3, AL138756.1, CD27-AS1, EIF3J-DT, LINC01011, LINC01063, and LINC02381) were independent unfavorable factors. Two lncRNAs (AC073896.3 and SNHG16) were independent beneficial factors for colon adenocarcinoma.

- 2.4. Development of Prognostic Model. Cox regression was utilized to build an independent prognostic model. Nomogram was utilized to predict survival for the patients. The index of concordance (C-index), calibration curves, and receiver operating characteristic (ROC) curves were applied to explore the accuracy of the model. The demographic data were included in multivariate Cox regression in order to confirm whether the risk score was an independent indicator for the prognosis.
- 2.5. Functional Analysis. Gene set enrichment analysis (GSEA, http://www.broadinstitute.org/gsea/index.jsp) was utilized to interpret the functional enrichment of gene expression data. We explored the functional enrichment of autophagy-related lncRNAs with a prognostic value and visualized the top 5 Gene Ontology (GO) and the Kyoto

Encyclopedia of Genes and Genomes (KEGG) pathways related to autophagy.

2.6. Statistical Analysis. The Kaplan-Meier method was utilized to generate the survival curves, and the log-rank test was used for comparison. Cox regression and Lasso regression were utilized to estimate the prognostic impact of the autophagy-related lncRNA signature and clinicopathological data. The statistical analyses were conducted in R language (version 3.6). Statistical tests were bilateral, with $P \le 0.05$ being statistically significant.

3. Results

3.1. Construction of a Coexpression Network. A total of 14,142 lncRNAs were identified in TCGA-COAD. A total of 257 autophagy-related genes were obtained from HADb, among

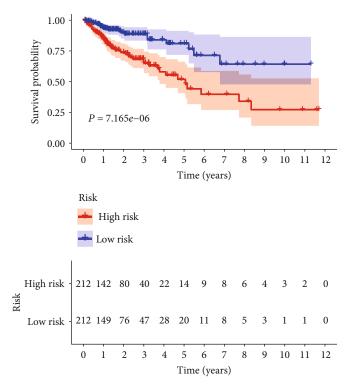


FIGURE 4: The KM survival curve of risk score based on ten autophagy-related lncRNAs.

which 210 genes were expressed in colon adenocarcinoma (Table S1). An autophagy-related gene lncRNA coexpression network was constructed to identify the autophagy-related lncRNAs. Finally, 1,281 autophagy-related lncRNAs were selected ($|R^2| > 0.3$ and P < 0.001).

3.2. Identification of Prognostic Autophagy-Related lncRNA Signature. According to the results of univariate Cox, 48 autophagy-related lncRNAs had a prognostic value for the patients with colon adenocarcinoma (P < 0.05, Table S2). Subsequently, 21 autophagy-related lncRNAs were identified after Lasso regression (Figure 1 and Table S3). Multivariate Cox regression found that ten lncRNAs were independent prognostic factors (Figure 2). Among them, eight lncRNAs (AC027307.2, AC068580.3, AL138756.1, CD27-AS1, EIF3J-DT, LINC01011, LINC01063, and LINC02381) were harmful prognostic factors, and the others (AC073896.3 and SNHG16) were favorable prognostic factors (Table 1 and Figure 3). These ten lncRNAs were utilized to establish an autophagy-related lncRNA signature. The formula of the risk score was as follows: risk score = (0.09742 * AC027307.2) +(0.89061 * AC068580.3) + (0.27224 * LINC01011) + (0.53477)* EIF3J-DT) - (0.13154 * SNHG16) - (0.93184 * AC073896.3) +(0.20468 * AL138756.1) + (0.08307 * CD27-AS1) + (0.20329)* LINC02381) + (0.57267 * LINC01063).

3.3. The Prognostic Influence of the Established Signature. The risk score was significantly associated with the overall survival (OS) of patients with colon adenocarcinoma. The high-risk group had shorter OS compared with the low-

risk group (P = 7.165e - 06, log-rank test) (Figure 4). Cox regression indicated significant prognostic impact of the risk score for the patients with colon adenocarcinoma (Figure 5).

3.4. Clinical Value of the Autophagy-Related lncRNA Signature. Univariate Cox regression revealed that risk score and stage were independent prognostic indicators, and HR of risk score was 1.116 (95% CI: 1.087-1.146, P < 0.001, Table S4, Figure 6(a)). After controlling clinical features, risk score remained an independent prognostic indicator in multivariate analysis (HR = 1.088, 95%CI = 1.057 - 1.120, P < 0.001, Table 2, Figure 6(b)). The areas under the ROC curve corresponding to 1 year, 3 years, and 5 years of survival were 0.723, 0.790, and 0.796, respectively (Figure 6(c)). Risk score, age, and TNM stage were included in the nomogram. As indicated in the nomogram, risk score and TNM stage were the largest contribution to 3and 5-year OS of patients with colon adenocarcinoma (Figure 7(a)). The C-index of the prognostic model was 0.796 (95% CI: 0.739-0.853). The AUC of five-year survival rate showed that risk score (0.798) and stage (0.731) had a certain prediction ability (Figure 7(b)). The risk scores increased with stage, demonstrating that this autophagyrelated lncRNA signature may be related to the progression of colon adenocarcinoma (Table 3).

3.5. Functional Analysis. A total of 263 GO (Table S5) terms and 91 KEGG pathways were obtained (Table S6). In GO analysis, the autophagy-related lncRNAs were mainly

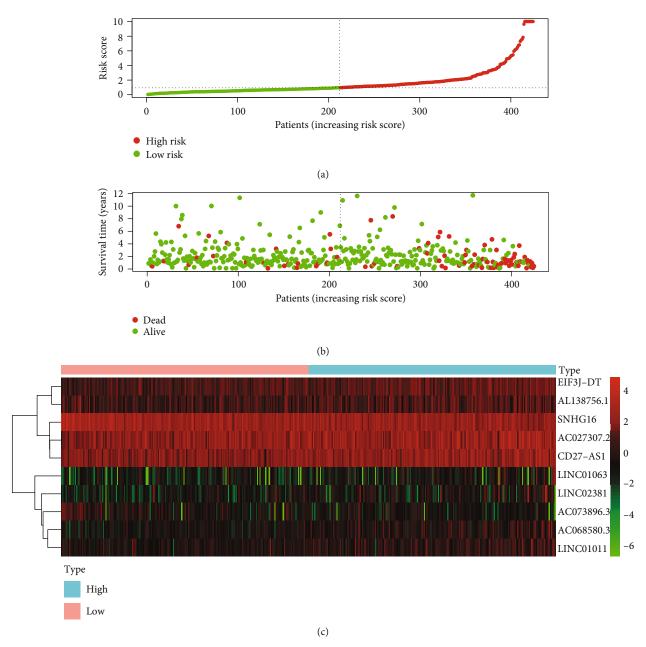


FIGURE 5: The analysis of autophagy-related lncRNA signature for patients with colon adenocarcinoma. (a) The risk score between the high-risk group and the low-risk group. (b) The survival time of the patients. (c) Heat map of ten autophagy-related lncRNAs' expression. The color from green to red reveals a rising tendency from low to high levels.

concentrated in biological processes such as transporting damaged, denatured, or senescent proteins in the cell and organelles to the lysosome for digestion and degradation (Figure 8(a)). KEGG pathways revealed that the lncRNAs were mostly concentrated in the pathway of tumor classical pathways and metabolism (Figure 8(b)). Furthermore, we also found that the gene sets were connected with vital biological processes and functional pathways of tumorigenesis and cancer progression. For instance, leukocyte transendothelial migration, angiogenesis, and hypoxia were closely associated with the invasion and metastasis of cancer.

4. Discussion

Autophagy is a highly regulated process that degrades and recycles cellular components [17]. Dysregulation of autophagy was implicated in many diseases [18]. As a large and heterogeneous subclass of ncRNAs, lncRNAs played an indispensable role in different aspects of tumorigenesis which were considered a new type of biomarkers in cancer diagnosis and prognosis [19]. Most researches focused on the function of specific genes involved in autophagy [20–22]. There are no systematic

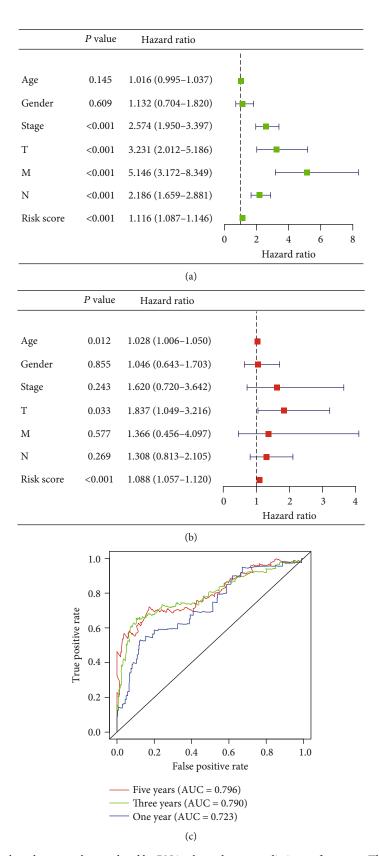


FIGURE 6: Prognostic indicators based on autophagy-related lncRNAs showed great predictive performance. The forest plots for univariate (a) and multivariate (b) Cox regression analysis in colon adenocarcinoma. (c) The areas under the ROC curve about 1 year, 3 years, and 5 years.

TE 2 OF 1 1 1	1 · 1	. 1
TABLE 2: Clinical characteristics and	d risk scores of colon adenocarcinom	a liging multivariate (ox regression
TABLE 2. CHINEUI CHUIUCCCIIStics uni	a fish scores of colon adenocaremoni	a using mainvariate Cox regression.

Variable	В	SE	Z	HR	HR.95L	HR.95H	P value
Age	0.027	0.011	2.516	1.028	1.006	1.050	0.012
Gender	0.045	0.248	0.182	1.046	0.643	1.703	0.855
Stage	0.482	0.413	1.167	1.620	0.720	3.642	0.243
T	0.608	0.286	2.127	1.837	1.049	3.216	0.033
M	0.312	0.560	0.557	1.366	0.456	4.097	0.577
N	0.268	0.243	1.105	1.308	0.813	2.105	0.269
Risk score	0.084	0.015	5.646	1.088	1.057	1.120	< 0.001

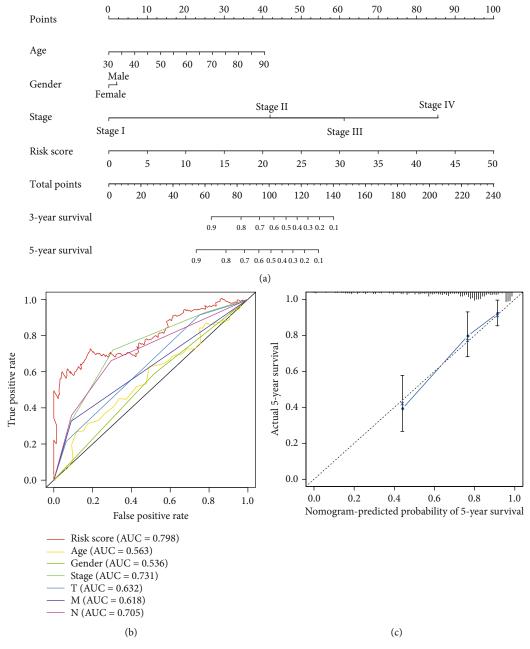


FIGURE 7: The evaluation of prognostic models based on ten autophagy-related lncRNAs. (a) The nomogram of 3-year or 5-year OS based on risk score, age, and TNM stage. (b) The ROC curves analysis based on risk score and the clinicopathologic parameters; (c) calibration plots for evaluating the agreement between the predicted and the actual OS for the prognosis model. The 45° reference line indicates perfect calibration, where the predicted probabilities are consistent with the actual probabilities.

Clinical	n	Risk score		,	ח
		Mean	SD	t	P
Age					
≤65	153	1.568	2.283	-0.566	0.572
>65	225	1.742	3.708		
Gender					
Female	175	1.595	3.670	-0.424	0.672
Male	203	1.738	2.751		
Stage					
I-II	216	1.205	1.243	-2.921	0.004
III-IV	162	2.293	4.617		
T					
T1-2	74	1.038	0.739	-3.574	< 0.001
T3-4	304	1.826	3.540		
M					
M0	317	1.383	1.793	-2.068	0.043
M1	61	3.170	6.702		
N					
N0	225	1.207	1.234	-2.928	0.004
N1-2	153	2.355	4.739		

studies about autophagy-related lncRNA signatures to predict the survival of patients with colon adenocarcinoma. Thus, it was necessary to establish an autophagy-related lncRNA signature to predict the prognosis of patients with colon adenocarcinoma based on the large-scale databases.

In this study, autophagy-related lncRNAs were screened by constructing a coexpression network of lncRNA and autophagy-related genes. Further, Lasso regression and Cox regression were utilized to obtain the following 10 prognostic autophagy-related lncRNAs: AC027307.2, AC068580.3, AL138756.1, CD27-AS1, EIF3J-DT, LINC01011, LINC01063, LINC02381, AC073896.3, and SNHG16. The ten autophagy-related lncRNAs might be prognostic molecular markers of prognosis and potential therapeutic targets for the patients with colon adenocarcinoma.

Six autophagy-related lncRNAs (SNHG16, EIF3J-DT, CD27-AS1, LINC01063, LINC01011, and LINC02381) were reported to be associated with cancer. (1) SNHG16 promoted Hep3B/So cell viability, autophagy, and suppress apoptosis to maintain its resistance to sorafenib by regulating miR-23b-3p [23]. SNHG16 promoted proliferation, migration, invasion, and autophagy of neuroblastoma cells through sponging miR-542-3p and upregulating autophagy-related gene 5 (ATG5) [24]. SNHG16 promoted progression of osteosarcoma and improved cisplatin resistance by sponging miR-16 to upregulate autophagy-related 4B [25]. (2) EIF3J-DT which was also named EIF3J-AS1 has been widely studied and found to be related to many kinds of cancers. Liu et al. revealed that EIF3J-DT could promote proliferation and reduce apoptosis in CRC cells, revealing that EIF3J-DT was a risk factor and possessed oncogenic functions in CRC [26]. EIF3J-DT inversely regulated miR-122e-5p via acting as a competing endogenous RNA in hepatocellular carcinoma (HCC) cells [27]. EIF3J-DT was significantly upregulated in HCC and closely correlated with poor prognosis [28]. Additionally, EIF3J-DT could be used as a therapeutic target and a potential biomarker for the diagnosis and prognosis of buccal mucosa squamous cell carcinoma [29]. (3) Ma et al. found that CD27-AS1 served as a carcinogenic RNA and the regulatory role of CD27-AS1 on CD27 contributes to the melanomagenesis [30]. (4) It was found that increased expression level of LINC01063 was significantly correlated with metastasis and poor prognosis of the patients with colon adenocarcinoma [31]. (5) Fan et al. found that LINC01011 controlled cisplatin sensitivity and mitochondrial fission via suppressing BRCA1 transcription in a tongue SCC model [32]. (6) LINC02381 might have inhibiting effects on CRC tumorigenesis partly through regulating PI3K pathway [33].

For the four remaining autophagy-related lncRNAs (AC027307.2, AC068580.3, AL138756.1, and AC073896.3), there were no studies to report their prognostic roles in cancer. Thus, more researches were necessary to explore how these lncRNAs affect the prognosis of patients with colon adenocarcinoma through autophagy exactly.

A signature based on 10 autophagy-related lncRNAs significantly predicted the prognosis of colon adenocarcinoma patients. Consistent with previous studies, the low-risk group had longer OS than the high-risk group [34, 35]. The areas under the ROC curve corresponding to 1 year, 3 years, and 5 years of survival were 0.723, 0.790, and 0.796. This result indicated that risk score signature had a certain potential in predicting survival. Both univariate and multivariate Cox analyses revealed that the signature could be used as an

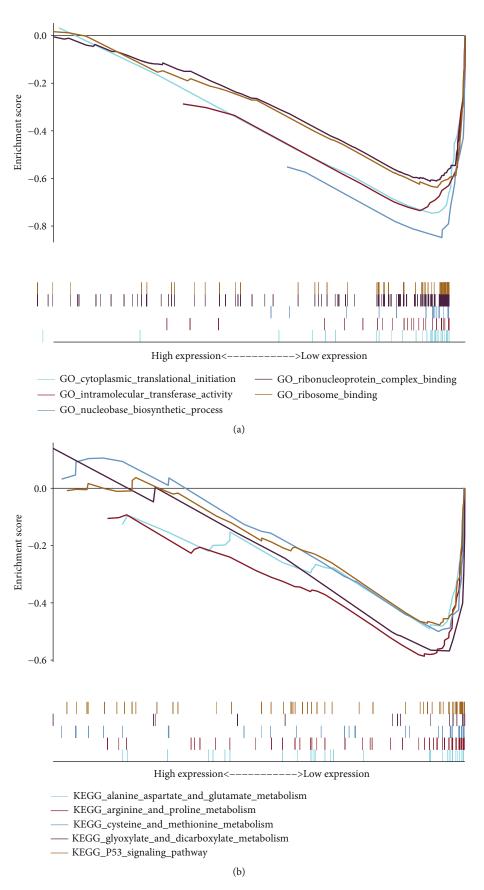


FIGURE 8: The results of functional analysis based on autophagy-related lncRNAs. (a) GO enrichment analysis; (b) KEGG enrichment analysis.

independent prognostic indicator. According to the results of *C*-index, ROC curve, and Calibration curve, the model possessed better discrimination and accuracy, revealing that it might serve as a potential predictive tool for patients with colon adenocarcinoma.

The results of functional enrichment analysis indicated that these prognostic autophagy-related lncRNAs were significantly enriched in biological processes such as autophagy-related, metabolism, and tumor-related pathways. Further, the most important pathways were enriched in autophagy processes in KEGG analysis which involved p53 classical pathways in colon adenocarcinoma. These results help us to explore the mechanism of autophagy-related lncRNAs. The previous study indicated that inhibition of autophagy in CRC cells led to antitumor effects via strengthened apoptosis through p53 and UPR activation [36]. Recently, protopin was proved to exert its antiproliferative activity through stimulating the p53 pathway [37]. There were some research reports on the molecular mechanism of amino acid metabolism and autophagy in colon adenocarcinoma. Amino acids were found to inhibit Raf-1 activation, which interfered with ERK1/2-dependent autophagy control in colon cancer HT-29 cells [38]. Targeted suppression of glutamine metabolism could inhibit the occurrence of colorectal cancer especially when unified with extracellular asparagine depletion and autophagy inhibition [39].

Several limitations existed in the current study. First, the data source of this study is single, and the amount of data included is not large, so the analysis results may have certain deviation. Second, our study is a retrospective study, and more prospective studies will be required to prove the prognostic function of autophagy-related signals. Third, in order to ensure the robustness of the prognostic model, the prognostic model of our established model is required to be further confirmed in other independent cohorts to ensure its robustness. Fourth, the functional experiments should be conducted to further indicate the potential molecular mechanisms for predicting the effect of autophagy-related lncRNAs.

In conclusion, an autophagy-related lncRNA coexpression network provided a valuable source for revealing autophagy-related lncRNA functions in colon adenocarcinoma. Ten autophagy-related lncRNAs were considered to be significantly associated with survival of the patients with colon adenocarcinoma. An autophagy-related lncRNA signature which was composed of ten autophagy-related lncRNAs was used to differentiate patients at different risks, and it was a significantly independent factor for the patients with colon adenocarcinoma. Therefore, the ten autophagy-related lncRNAs and their signature might be molecular biomarkers and therapeutic targets for the patients with colon adenocarcinoma.

Data Availability

All analyzed or generated data were incorporated in this published paper and its supplementary information file.

Conflicts of Interest

The authors declare there are no competing interests.

Authors' Contributions

Weige Zhou conceived and designed the experiments, performed the experiments, analyzed and interpreted the data, contributed reagents/materials/analysis tools, composed or reviewed drafts of the paper, and read and approved the final draft. Shijing Zhang conducted the experiments, analyzed and interpreted the data, wrote or reviewed drafts of the dissertation, read and approved the final draft. Hui-biao Li conducted the experiments, analyzed and interpreted the data, wrote or reviewed drafts of the dissertation, and read and approved the final draft. Zheyou Cai conducted the experiments, analyzed and interpreted the data, contributed the reagents/materials/analysis tools, and read and approved the final draft. Shuting Tang conceived and designed the experiments, analyzed and interpreted the data, and read and approved the final draft. Li-xia Chen analyzed and interpreted the data, contributed the reagents/materials/ analysis tools, and read and approved the final draft. Jian-ying Lang analyzed the data, contributed the reagents/ materials/analysis tools, and read and approved the final draft. Zheng Chen and Xin-lin Chen conceived and designed the experiments. Weige Zhou and Shijing Zhang contributed equally to this work.

Acknowledgments

This study was funded by the National Natural Science Foundation of China (81774451), the Natural Science Foundation of Guangdong Province (2017A030313827), and Science Program for Overseas Scholar (Xinhuo plan) of Guangzhou University of Chinese Medicine (XH20190102).

Supplementary Materials

Supplementary 1. Table S1: The expression profile of autophagy-related gene in colon adenocarcinoma.

Supplementary 2. Table S2 Univariate cox results of ARIncRNAs based on TCGA-COAD data.

Supplementary 3. Table S3 LASSO regression coefficients of twenty-one ARlncRNAs.

Supplementary 4. Table S4 Clinical characteristics and risk scores of colon adenocarcinoma using univariate cox regression.

Supplementary 5. Table S5 Gene set enrichment GO analysis results according to the signature of ten ARlncRNAs (Top 20 p-value).

Supplementary 6. Table S6 Gene set enrichment KEGG analysis results based on the signature of ten ARlncRNAs (Top 20 p-value).

References

[1] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," *CA: a Cancer Journal for Clinicians*, vol. 68, no. 6, pp. 394–424, 2018.

[2] R. L. Siegel, K. D. Miller, A. Goding Sauer et al., "Colorectal cancer statistics, 2020," *CA: A Cancer Journal for Clinicians*, vol. 70, no. 3, pp. 145–164, 2020.

- [3] E. Dekker, P. J. Tanis, J. L. A. Vleugels, P. M. Kasi, and M. B. Wallace, "Colorectal cancer," *Lancet*, vol. 394, no. 10207, pp. 1467–1480, 2019.
- [4] V. Barresi, L. Reggiani Bonetti, A. Ieni, R. A. Caruso, and G. Tuccari, "Histological grading in colorectal cancer: new insights and perspectives," *Histology and Histopathology*, vol. 30, no. 9, pp. 1059–1067, 2015.
- [5] R. K. Amaravadi, A. C. Kimmelman, and J. Debnath, "Targeting autophagy in cancer: recent advances and future directions," *Cancer Discovery*, vol. 9, no. 9, pp. 1167–1181, 2019.
- [6] B. Levine and G. Kroemer, "Biological functions of autophagy genes: a disease perspective," *Cell*, vol. 176, no. 1-2, pp. 11–42, 2019.
- [7] L. Y. Mah and K. M. Ryan, "Autophagy and cancer," Cold Spring Harbor Perspectives in Biology, vol. 4, no. 1, p. a008821, 2012.
- [8] W. T. Tsai, Y. C. Lo, M. S. Wu et al., "Mycotoxin patulin suppresses innate immune responses by mitochondrial dysfunction and p62/sequestosome-1-dependent mitophagy," *The Journal* of Biological Chemistry, vol. 291, no. 37, pp. 19299–19311, 2016.
- [9] P. Mokarram, M. Albokashy, M. Zarghooni et al., "New frontiers in the treatment of colorectal cancer: autophagy and the unfolded protein response as promising targets," *Autophagy*, vol. 13, no. 5, pp. 781–819, 2017.
- [10] T. R. Mercer, M. E. Dinger, and J. S. Mattick, "Long non-coding RNAs: insights into functions," *Nature Reviews. Genetics*, vol. 10, no. 3, pp. 155–159, 2009.
- [11] R. Mathew, V. Karantza-Wadsworth, and E. White, "Role of autophagy in cancer," *Nature Reviews. Cancer*, vol. 7, no. 12, pp. 961–967, 2007.
- [12] F. Burada, E. R. Nicoli, M. E. Ciurea, D. C. Uscatu, M. Ioana, and D. I. Gheonea, "Autophagy in colorectal cancer: an important switch from physiology to pathology," World J Gastrointest Oncol, vol. 7, no. 11, pp. 271–284, 2015.
- [13] M. Wang, D. Han, Z. Yuan et al., "Long non-coding RNA H19 confers 5-Fu resistance in colorectal cancer by promoting SIRT1-mediated autophagy," *Cell Death & Disease*, vol. 9, no. 12, p. 1149, 2018.
- [14] W. Zhang, W. Yuan, J. Song, S. Wang, and X. Gu, "LncRNA CPS1-IT1 suppresses EMT and metastasis of colorectal cancer by inhibiting hypoxia-induced autophagy through inactivation of HIF-1alpha," *Biochimie*, vol. 144, pp. 21–27, 2018.
- [15] Y. Si, Z. Yang, Q. Ge et al., "Long non-coding RNA Malat1 activated autophagy, hence promoting cell proliferation and inhibiting apoptosis by sponging miR-101 in colorectal cancer," Cellular & Molecular Biology Letters, vol. 24, no. 1, 2019.
- [16] Z. Guo, C. Zhou, X. Zhong et al., "The long noncoding RNA CTA-941F9.9 is frequently downregulated and may serve as a biomarker for carcinogenesis in colorectal cancer," *Journal of Clinical Laboratory Analysis*, vol. 33, no. 9, p. e22986, 2019.
- [17] S. N. Devenport and Y. M. Shah, "Functions and implications of autophagy in colon cancer," *Cell*, vol. 8, no. 11, p. 1349, 2019.
- [18] B. Levine and G. Kroemer, "Autophagy in the pathogenesis of disease," *Cell*, vol. 132, no. 1, pp. 27–42, 2008.
- [19] K. T. Jin, Z. B. Lu, J. Q. Lv, and J. G. Zhang, "The role of long non-coding RNAs in mediating chemoresistance by modulating autophagy in cancer," *RNA Biol*, pp. 1–14, 2020.

[20] S. Mo, W. Dai, W. Xiang et al., "Prognostic and predictive value of an autophagy-related signature for early relapse in stages I-III colon cancer," *Carcinogenesis*, vol. 40, no. 7, pp. 861–870, 2019.

- [21] J. Gil, D. Ramsey, P. Pawlowski et al., "The influence of tumor microenvironment on ATG4D gene expression in colorectal cancer patients," *Medical Oncology*, vol. 35, no. 12, p. 159, 2018.
- [22] Z. Huang, J. Liu, L. Luo et al., "Genome-wide identification of a novel autophagy-related signature for colorectal cancer," *Dose Response*, vol. 17, no. 4, p. 1559325819894179, 2019.
- [23] Z. Jing, X. Ye, X. Ma et al., "SNGH16 regulates cell autophagy to promote sorafenib resistance through suppressing miR-23b-3p via sponging EGR1 in hepatocellular carcinoma," *Cancer Medicine*, vol. 9, no. 12, pp. 4324–4338, 2020.
- [24] Y. Wen, X. Gong, Y. Dong, and C. Tang, "Long non coding RNA SNHG16 facilitates proliferation, migration, invasion and autophagy of neuroblastoma cells via sponging miR-542-3p and upregulating ATG5 expression," *Oncotargets and Therapy*, vol. Volume 13, pp. 263–275, 2020.
- [25] Y. Liu, S. Gu, H. Li, J. Wang, C. Wei, and Q. Liu, "SNHG16 promotes osteosarcoma progression and enhances cisplatin resistance by sponging miR-16 to upregulate ATG4B expression," *Biochemical and Biophysical Research Communications*, vol. 518, no. 1, pp. 127–133, 2019.
- [26] D. Liu, H. Zhang, J. Cong et al., "H3K27 acetylation-induced lncRNA EIF3J-AS1 improved proliferation and impeded apoptosis of colorectal cancer through miR-3163/YAP1 axis," *Journal of Cellular Biochemistry*, vol. 121, no. 2, pp. 1923– 1933, 2019.
- [27] X. Yang, B. Yao, Y. Niu et al., "Hypoxia-induced lncRNA EIF3J-AS1 accelerates hepatocellular carcinoma progression via targeting miR-122-5p/CTNND2 axis," *Biochemical and Biophysical Research Communications*, vol. 518, no. 2, pp. 239–245, 2019.
- [28] J. X. Gu, X. Zhang, R. C. Miao et al., "Six-long non-coding RNA signature predicts recurrence-free survival in hepatocellular carcinoma," *World Journal of Gastroenterology*, vol. 25, no. 2, pp. 220–232, 2019.
- [29] K. D. Patel, H. H. Vora, T. I. Trivedi et al., "Transcriptome profiling and pathway analysis in squamous cell carcinoma of buccal mucosa," *Experimental and Molecular Pathology*, vol. 113, p. 104378, 2020.
- [30] J. Ma, P. Xu, S. Guo, Q. Shi, and C. Li, "1199 The regulatory role of lncRNA CD27-AS1 on CD27 contributes to melanomagenesis," *The Journal of Investigative Dermatology*, vol. 138, no. 5, pp. S204–S204, 2018.
- [31] Y. Lin, X. Pan, Z. Chen, S. Lin, and S. Chen, "Identification of an immune-related nine-lncRNA signature predictive of overall survival in colon cancer," *Frontiers in Genetics*, vol. 11, p. 318, 2020.
- [32] S. Fan, T. Tian, X. Lv et al., "IncRNA CISAL inhibits BRCA1 transcription by forming a tertiary structure at its promoter," *iScience*, vol. 23, no. 2, p. 100835, 2020.
- [33] M. Jafarzadeh, B. M. Soltani, M. Soleimani, and S. Hosseinkhani, "Epigenetically silenced LINC02381 functions as a tumor suppressor by regulating PI3K-Akt signaling pathway," *Biochimie*, vol. 171-172, pp. 63-71, 2020.
- [34] J. Lv, Y. Guo, L. Yan, Y. Lu, D. Liu, and J. Niu, "Development and validation of a five-lncRNA signature with prognostic value in colon cancer," *Journal of Cellular Biochemistry*, vol. 121, no. 8-9, pp. 3780–3793, 2020.

- [35] H. Yang, H. C. Lin, H. Liu et al., "A 6 lncRNA-based risk score system for predicting the recurrence of colon adenocarcinoma patients," *Frontiers in Oncology*, vol. 10, p. 81, 2020.
- [36] K. Sakitani, Y. Hirata, Y. Hikiba et al., "Inhibition of autophagy exerts anti-colon cancer effects via apoptosis induced by p53 activation and ER stress," *BMC Cancer*, vol. 15, no. 1, 2015.
- [37] Y. Son, Y. An, J. Jung et al., "Protopine isolated from Nandina domestica induces apoptosis and autophagy in colon cancer cells by stabilizing p53," *Phytotherapy Research*, vol. 33, no. 6, pp. 1689–1696, 2019.
- [38] S. Pattingre, C. Bauvy, and P. Codogno, "Amino acids interfere with the ERK1/2-dependent control of macroautophagy by controlling the activation of Raf-1 in human colon cancer HT-29 cells," *The Journal of Biological Chemistry*, vol. 278, no. 19, pp. 16667–16674, 2003.
- [39] J. Li, P. Song, L. Zhu et al., "Synthetic lethality of glutaminolysis inhibition, autophagy inactivation and asparagine depletion in colon cancer," *Oncotarget*, vol. 8, no. 26, pp. 42664–42672, 2017.