Published online 2015 January 14.

Letter

Bioactivity of *Apium petroselinum* and *Portulaca oleracea* Essential Oils as Natural Preservatives

Reza Sharafati Chaleshtori^{1,*}; Mahmoud Rafieian Kopaei²; Elham Salehi¹

¹Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, IR Iran

²Medical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, IR Iran

*Corresponding author: Reza Sharafati Chaleshtori, Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, IR Iran. Tel: +98-3615550021, E-mail: Sharafati33@yahoo.com

Received: May 10, 2014; Revised: August 1, 2014; Accepted: August 8, 2014

Keywords: Antimicrobial Agents; Essential oil; Food Preservatives; Meat

Dear Editor,

Nowadays, essential oils are used in different products such as food as natural antioxidants, antibacterial and flavoring compounds (1). Essential oils are used as a replacement for chemical preservatives such as butylated hydroxytoluene (BHT) and tertiary butyl hydroquinone (TBHQ) that can threaten human health as cancer promoters (2). Nearly 3000 different essential oils are known today, and 300 are available and used commercially (3). They are obtained from aromatic plants and their various parts including roots, stems, leaves, flowers, fruits and seeds (4). The aim of this study was to evaluate the antioxidant activity and antibacterial effects of Apium petroselinum and Portulaca oleracea essential oils on food spoilage bacteria. A. petroselinum and *P. oleracea* were purchased from the local grocery stores of Shahrekord city and authenticated by standard botanic work at the Medical Plants Research Center of Shahrekord University of Medical Sciences, Iran. Essential oils were then extracted by steam distillation for three hours using a Clevenger-type apparatus and dried by adding anhydrous sodium sulfate and stored at 4°C, before being used for the assay. Total phenolic contents in the obtained essence were examined by the Folin Ciocalteu method and the results were expressed as milligrams of gallic acid equivalents (GAEs)/gram of essential oil. The total antioxidant capacities of their essential oils were assayed using β-carotene and linoleic acid (5).

Cultures of Alcaligenes faecalis (PTCC 1624), Providencia rettgeri (PTCC 1512), Serratia marcescens (PTCC 1621), Klebsiella oxytoca (PTCC 1402), Staphylococcus aureus (PTCC 1169), Shigella dysenteriae (PTCC 1188) and Listeria monocytogenes (PTCC 1163) were purchased from the Iranian Research Organization for Science and Technology (IROST). The micro-dilution method in sterile 96-microwell plates was used to obtain the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) (6). Each experiment was done in triplicates. Based on the obtained results, the two essential oils demonstrated antioxidant and antibacterial activities. The most sensitive bacteria were A. faecalis and S. dysenteriae, which are Gram-negative bacteria (Table 1). The results obtained in this work are in agreement with pervious studies. Earlier studies have reported that various bacteria such as S. dysenteriae, E. coli, Helicobacter pylori and S. aureus were sensitive to essential oils and extracts of the above mentioned plants (7, 8). However, the Gram-negative bacteria were more resistant than Gram-positive species, due to restricted diffusion of the hydrophobic compounds by the hydrophilic cell wall structure containing lipo-polysaccharide (LPS) (9). In conclusion, the study results suggest that replacement of synthetic preservatives with natural additives as a good source of healthy compounds, namely phenols (10), can be useful in the food industry to increase shelf life of food products.

Acknowledgements

The authors gratefully thank the Research Deputy and the personnel of the Medical Plants Research Center of Shahrekord University of Medical Sciences, Iran.

Copyright @ 2015, Ahvaz Jundishapur University of Medical Sciences. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

petroselinum and Portulaca oleracea Essential Oils ^a			
	Apium petroseli- num	Portu- laca oleracea	ВНТа
Total Phenolic Content (mg GAEs/g)	8.18 ± 0.45	7.8 ± 0.2	-
Antioxidant Capac- ity, %	45 ± 1.52	56.5 ± 4.19	90.6±3.30
Antibacterial activity			
Alcaligenes faecalis			
MIC mg/mL	1.562	6.25	-
MBC, mg/mL	3.125	12.5	-
Providencia rettgeri			
MIC, mg/mL	6.25	12.5	-
MBC, mg/mL	12.5	25	-
Serratia marcescens			
MIC mg/mL	3.125	12.5	-
MBC, mg/mL	6.25	25	-
Klebsiella oxytoca			
MIC, mg/mL	12.5	12.5	-
MBC, mg/mL	25	25	-
Staphylococcus aureus			
MIC, mg/mL	12.5	12.5	-
MBC, mg/mL	25	25	-
Shigella dysenteriae			
MIC, mg/mL	1.562	6.25	-
MBC, mg/mL	3.125	12.5	-
Listeria monocyto- genes			
MIC, mg/mL	12.5	6.25	-
MBC, mg/mL	25	12.5	-

Table 1. Antioxidant and Antibacterial Characteristics of *Apium* petroselinum and *Portulaca oleracea* Essential Oils ^a

^a Abbreviations: GAEs, gallic acid equivalents; BHT, butylated hydroxytoluene; MIC, minimum inhibitory concentration; MBC, minimum bactericidal concentration

Authors' Contributions

Reza Sharafati-Chaleshtori developed the original idea and the protocol, abstracted and wrote the manuscript. Reza Sharafati-Chaleshtori, Mahmoud Rafieian-Kopaei and Elham Salehi contributed to the development of the protocol, abstracted data, and prepared the manuscript.

Funding/Support

This project was financially supported by the Medical Plants Research Center of Shahrekord University of Medical Sciences, Iran.

References

- 1. Shaaban HAE, El-Ghorab AH, Shibamoto T. Bioactivity of essential oils and their volatile aroma components: Review. *J Essent Oil Res.* 2012;**24**(2):203–12.
- Hyldgaard M, Mygind T, Meyer RL. Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Front Microbiol.* 2012;3:12.
- Burt S. Essential oils: their antibacterial properties and potential applications in foods-a review. Int J Food Microbiol. 2004;94(3):223-53.
- Sanchez E, Garcia S, Heredia N. Extracts of edible and medicinal plants damage membranes of Vibrio cholerae. *Appl Environ Microbiol*. 2010;**76**(20):6888–94.
- Sharafati Chaleshtori R, Sharafati Chaleshtori F, Rafieian Kopae M. Biological characterization of Iranian walnut (Juglans regia) leaves. *Turk J Biol.* 2011;35:635–9.
- Sharafati Chaleshtori R, Rokni N, Razavilar V, Rafieian Kopaei M. The Evaluation of the Antibacterial and Antioxidant Activity of Tarragon (Artemisia dracunculus L.) Essential Oil and Its Chemical Composition. Jundishapur J Microbiol. 2013;6(9).
- Nakhaei Moghaddam M. In vitro anti-bacterial activity of methanolic extract of Apium petroselinum L. seed against clinical isolates of Helicobacter pylori. *Daneshvar Med.* 2010;17:63–70.
- 8. Bae JH. Antimicrobial Effect of Portulaca oleracea Extracts on Food-Borne Pathogens. *J Food Sci Nutr.* 2004;**9**(4):306–11.
- Sulaiman S, Ibrahim D, Kassim J, Sheh-Hong L. Antimicrobial and antioxidant activities of condensed tannin from Rhizophora apiculata barks. J Chem Pharm Res. 2011;3(4):436–44.
- 10. Rafieian-Kopaei M, Baradaran A, Rafieian M. Plants antioxidants: From laboratory to clinic. *J Nephropathol*. 2013;**2**(2):152–3.