

Research

Open Access

Virtual cooperativity in myoglobin oxygen saturation curve in skeletal muscle *in vivo*

Akitoshi Seiyama*

Address: Division of Physiology and Biosignaling, Osaka University Graduate School of Medicine, 2-2 Yamadaoka, Suita, Osaka 565-0871, Japan

Email: Akitoshi Seiyama* - aseiyama@phys1.med.osaka-u.ac.jp

* Corresponding author

Published: 24 January 2006

Received: 23 June 2005

Dynamic Medicine 2006, 5:3 doi:10.1186/1476-5918-5-3

Accepted: 24 January 2006

This article is available from: <http://www.dynamic-med.com/content/5/1/3>

© 2006 Seiyama; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Myoglobin (Mb) is the simplest monomeric hemoprotein and its physicochemical properties including reversible oxygen (O_2) binding in aqueous solution are well known. Unexpectedly, however, its physiological role in intact muscle has not yet been established in spite of the fact that the role of the more complex tetrameric hemoprotein, hemoglobin (Hb), in red cells is well established. Here, I report my new findings on an overlooked property of skeletal Mb.

Methods: I directly observed the oxygenation of Mb in perfused rat skeletal muscle under various states of tissue respiration. A computer-controlled rapid scanning spectrophotometer was used to measure the oxygenation of Mb in the transmission mode. The light beam was focused on the thigh (quadriceps) through a 5-mm-diameter light guide. The transmitted light was conducted to the spectrophotometer through another 5-mm-diameter light guide. Visible difference spectra in the range of 500–650 nm were recorded when O_2 uptake in the hindlimb muscle reached a constant value after every stepwise change in the O_2 concentration of the buffer.

Results: The O_2 dissociation curve (ODC) of Mb, when the effluent buffer O_2 pressure was used as the abscissa, was of a sigmoid shape under normal and increased respiratory conditions whereas it was of rectangular hyperbolic shape under a suppressed respiratory condition. The dissociation curve was shifted toward the right and became more sigmoid with an increase in tissue respiration activity. These observations indicate that an increase in O_2 demand in tissues makes the O_2 saturation of Mb more sensitive to O_2 pressure change in the capillaries and enhances the Mb-mediated O_2 transfer from Hb to cytochrome oxidase (Cyt. aa₃), especially under heavy O_2 demands.

Conclusion: The virtual cooperativity and O_2 demand-dependent shifts of the ODC may provide a basis for explaining why Mb has been preserved as monomer during molecular evolution.

Background

Mb is a monomeric hemoprotein with a molecular weight of 17 kDa, carrying a single oxygen (O_2)-binding site per molecule. It is located near the contractile elements and cell membranes in the red skeletal and cardiac muscles of

vertebrates [1]. Previously, Millikan [2,3] proposed the following three possible physiological functions for Mb: (a) an O_2 store during temporary deficits in O_2 supply, (b) an intracellular O_2 transport agent and (c) an intracellular catalyst. Among them, the first function has traditionally

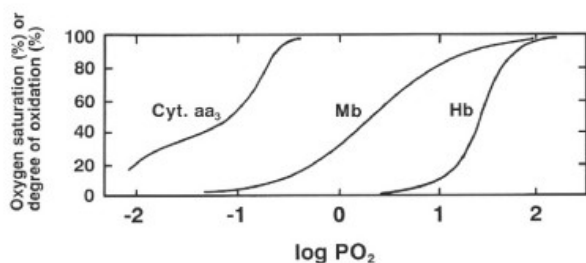


Figure 1

Oxygen dissociation curves (ODCs) for Hb (whole blood) and Mb and oxidation curve for Cyt. aa₃ (at 37°C). PO₂, partial pressure of oxygen in mmHg. Data from Imai [36].

been accepted. In the muscles of a beating heart and exercising skeletal muscles, Mb acts as a short-term O₂ store (i.e., an O₂ buffer), tiding the muscles over from one contraction to the next. The rich Mb content in skeletal muscles in aquatic mammals is considered to provide a long-term O₂ store during diving. However, this role of Mb, at least in human, is not significant because its oxygen storage capacity is so low that the total oxygen bound to Mb is exhausted within ca. 5.5 s after being cut off from the O₂ supply [4]. The second function, called "facilitated O₂-diffusion by Mb", was based on findings in *in vitro* experiments [5,6]. The conditions required for this facilitated diffusion to occur are [7]: (a) existence of deoxygenated Mb in a certain fraction (or certain low intracellular partial pressure of O₂), (b) existence of a spatial gradient of oxygenated Mb concentration as a driving force for translational diffusion of Mb, and (c) sufficient mobility of the oxygenated Mb to permit diffusion. Although this mechanism has been widely accepted, several discrepancies remain unresolved [8-12]. As for the third function, Doe-ller and Wittenberg [13] proposed the occurrence of Mb-mediated oxidative phosphorylation in heart myocytes under aerobic conditions. However, Mb concentration is not closely related to the oxidative capacity of muscles, that is, the concentration is higher in skeletal muscles (~0.5 mmole/kg wet wt.) than in heart muscles (~0.25 mmole/kg wet wt.) [7].

Thus, the physiological roles of Mb have not yet been established. Recently, alternative functions of (d) O₂ sensing and (e) nitric oxide scavenging were proposed [14]. Another recent paper [15] seemed to have totally scrambled the past long-term disputes about the physiological significance of Mb. It was shown using gene-knockout technology that mice without Mb are fertile, exhibit normal exercise capacity, and have a normal ventilatory response to low O₂ levels, suggesting that Mb is not essential for apparently normal cardiovascular and musculo-skeletal function in a terrestrial, homoiothermic

mammal. However, it has been reported that the disruption of Mb results in the activation of multiple compensatory mechanisms such as increases in Hb concentration, hematocrit, coronary flow, coronary reserve, and capillary density [16]. Further, a Mb-like hemoprotein, neuroglobin, has been found in the vertebrate brain [17] contrary to the long-held belief that Mb is restricted to vertebrate cardiomyocytes and oxidative skeletal myofibers. These studies imply that further investigations are required to reveal the physiological role of Mb in intact organs.

In contrast to Mb, which shows a rectangular hyperbolic ODC, the vertebrate Hb, a tetramer carrying four O₂ binding sites, shows a sigmoid ODC that is described in terms of a four-step cooperative O₂ binding. It is widely accepted that the sigmoid ODC enables Hb to transport O₂ with high efficiency: it is nearly fully saturated with O₂ in the lungs and it unloads O₂ sensitively depending on decreases in the partial pressure of oxygen (PO₂) in peripheral tissues. Here, no convincing explanation has been given for the question: does the hyperbolic ODC of Mb have any physiological adequacy or reasonability? The Bohr effect of Hb (pH dependence of O₂ affinity) has physiological significance, in that it enhances O₂ unloading from Hb in the capillaries where pH tends to decrease and in that it increases the solubility of CO₂ as bicarbonate in the venous blood through deoxygenation-induced uptake of protons by Hb. In contrast, Mb lacks the Bohr effect and it had long been believed that Mb was a totally non-allosteric protein, although recently lactate, a metabolic product, was found to cause a right-shift of the ODC for horse and sperm whale Mbs [18].

It is well established that the O₂ affinity of Mb is higher than that of Hb but lower than that of Cyt. aa₃, as known from the relative positions of the ODCs for Mb and Hb and the oxidation curve for Cyt. aa₃ (Fig. 1). This fact led one to the idea that Mb acts as an intracellular O₂ transfer agent from Hb (vascular space) to Cyt. aa₃ (mitochondria). Here, one must not overlook an important fact. The three curves in Fig. 1 are drawn with the same PO₂ scale. Therefore, they give O₂ saturation (Y) or the degree of oxidation for the individual proteins when dissolved in the same solution and are in equilibrium with oxygen at the given PO₂ value. However, *in vivo*, they sense different PO₂ values due to the presence of a PO₂ gradient along the path from the inside of red cells to the mitochondria in myocytes. Thus, the relative positions of the three curves in Fig. 1 must be considered with this precaution, and direct *in vivo* observations of Y or the degree of oxidation for these three individual proteins are required to get insight into their ensemble functional roles. Recently, using ¹H nuclear magnetic resonance spectroscopy, Mole et al. [19] and Richardson et al. [20] directly observed Y

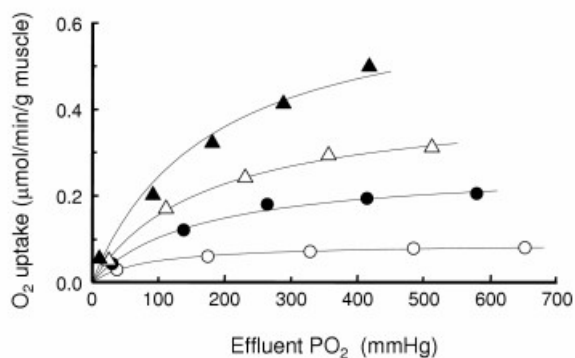


Figure 2
Steady-state O₂ uptake rate (V) of perfused rat hindlimb muscles as a function of effluent buffer PO₂. The rat hindlimb was perfused with Krebs-bicarbonate buffers containing no additive (●) as control, 0.4 mM of KCN (○) for suppression of muscle respiration, and 5 μM (△) or 10 μM (▲) 2,4-dinitrophenol for stimulation of muscle respiration. Symbols express observed points. Each plot is the mean of experiments using three animals, and the errors for each data point are less than the size of symbols. The solid lines were calculated using a rectangular hyperbolic curve: $V = V_{max}(PO_2/P_{V50}) / \{1 + (PO_2/P_{V50})\}$. The values of P_{V50} and V_{max} are given in Table 1 which also includes the maximal values of influent and effluent PO₂.

for Mb in human skeletal muscles under exercise of different intensities or during breathing of air with different O₂ contents. In these studies, Mb was used as an indicator of intracellular PO₂, and no attention was paid to the relation between Mb saturation and capillary PO₂.

In the present study, we directly measured Y for Mb in isolated rat hindlimb muscles, perfused with a Hb-free medium, under vigorous changes in respiration conditions. We plotted the Y values as a function of buffer PO₂ and found that the apparent ODC thus plotted for skeletal muscle Mb was rectangular hyperbolic under a suppressed metabolic activity condition but it became sigmoid under enhanced metabolic activity conditions, realizing virtually cooperative O₂ binding by monomeric Mb.

Methods

Muscle perfusion

All experimental procedures were performed according to the institutional guidelines for animal care and use of the Committee for Animal Care of Osaka University and the Japanese Physiological Society. Male Wistar rats (250 to 300 g body weight, N = 12) fed on a commercial diet were used. Rats were anesthetized with sodium pentobarbital (30 mg/kg body wt., intraperitoneal injection). Prepara-

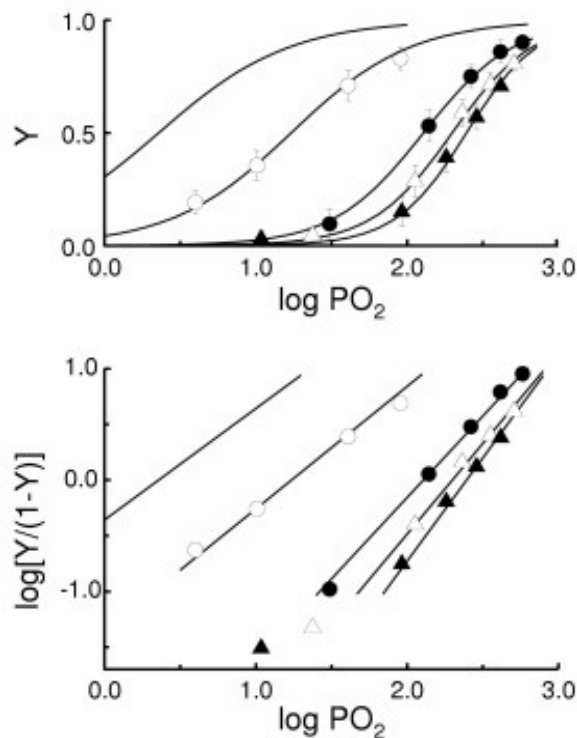


Figure 3
Apparent ODCs for Mb in perfused muscle at various steady-state O₂ uptake levels. PO₂ is the same as in Fig. 2. **A**, ODCs as O₂ saturation (Y) plotted against log PO₂. The lines were calculated from the Hill equation (see below). Each ODC was obtained from three animals (three muscle preparations). Symbols (mean ± SD) express observed points and their meaning is the same as in Fig. 2. The lines without symbols are the ODCs for Mb in non-respiring muscle [21]. **B**, Apparent ODCs as expressed by the Hill plot which is based on the linearized Hill equation [25]: $\log \{Y/(1 - Y)\} = n (\log PO_2 - \log P_{Y50})$. The slope of the plot (n) was constant and expressed as n_{app} in the present paper. The n_{app} and intercept values obtained from the Hill plots are listed in Table 2.

tion of the isolated rat hindlimb and the perfusion apparatus were described previously [21,22]. Surgery was modified from those of Ruderman et al. [23] and Shiota et al. [24]. After a midline abdominal incision, the superficial epigastric vessels were ligated. The abdominal wall was then incised from the pubic symphysis to the xiphoid process. The spermatic, testis, and inferior mesenteric arteries and veins were ligated, and the spermatic, the testis, and part of the descending colon were excised, together with contiguous adipose tissue. The caudal artery and internal iliac artery and vein were also ligated. Ligatures were placed around the neck of the bladder, the coagulating gland and the prostate gland. While carefully

Table 1: Values of muscle perfusion parameters and Mb oxygenation parameters in various tissue respiration states

Respiration state:	Suppressed (0.4 mM KCN)	Control	Enhanced (5 μ M DNP ^a)	Enhanced (10 μ M DNP)
Influent PO ₂	700	700	700	700
Effluent PO ₂	652 \pm 14	579 \pm 6	512 \pm 15	418 \pm 18
V _{max} ^b	0.09	0.27	0.42	0.68
P _{V50} ^c	83	160	170	180

a, 2,4-dinitrophenol; b, Maximal value of steady-state O₂ uptake rate (V) at infinite influent PO₂ (in μ mol/min/g muscle); c, effluent PO₂ at V = half V_{max} (in mmHg). Values of V_{max} and P_{V50} were obtained from solid lines shown in Figure 2.

removing the skin covering the lower half of the animal, the vessels that supply the subcutaneous region were ligated. Then, the inferior epigastric, iliolumbar and renal arteries and veins were ligated as well as the coeliac axis and portal vein. Further, a ligature was also placed around the tail. A hemoglobin-free Krebs-bicarbonate buffer (NaCl, 115 mM; KCl, 5.9 mM; MgCl₂, 1.2 mM; NaH₂PO₄, 1.2 mM; Na₂SO₄, 1.2 mM; NaHCO₃, 25 mM; CaCl₂, 2.5 mM; glucose, 10 mM; pH 7.4) containing 4% (w/v) polyvinylpyrrolidone (PVP-40T; average M.W., 40,000; Sigma) was perfused from the abdominal aorta in the flow-through mode at a constant flow rate of 1.0 ml/min/g muscle. Perfusate and muscle temperature were maintained at 25 \pm 0.5 °C. The effluent was collected from the inferior vena cava in order to measure the O₂ uptake rate. PO₂ in the influent and the effluent buffers was monitored with an oxygen electrode. The rate of O₂ uptake was calculated from the flow rate and the difference in O₂ concentration between the influent and the effluent buffers. Before each measurement, the rat hindlimb was perfused with the buffer equilibrated with 95% O₂ + 5% CO₂ for 30 min. Then, the O₂ concentration in the perfusate was decreased stepwise by mixing a buffer equilibrated with 95% O₂ + 5% CO₂ and another equilibrated with 95% N₂ + 5% CO₂, and the measurement was started. As required, potassium cyanide or 2,4-dinitrophenol was infused to modify the O₂ uptake rate of the perfused muscle. During each measurement of about 60 min, the perfusion pressure remained nearly constant at 73–78 mmHg. All chemicals used were of analytical reagent grade.

Spectrophotometric measurement of myoglobin oxygenation

A computer-controlled rapid scanning spectrophotometer (USP-501, Unisoku, Osaka, Japan) was used to measure the oxygenation of Mb in the transmission mode [21,22]. The light beam was focused on the thigh (quadriceps) through a 5-mm-diameter light guide. The transmitted light was conducted to the spectrophotometer through another 5-mm-diameter light guide. Visible difference spectra in the range of 500–650 nm were recorded when O₂ uptake in the hindlimb muscle reached a constant value after every stepwise change in the O₂ concentration of the buffer.

Analysis of data

Changes in the O₂ uptake rate were analyzed using a rectangular hyperbolic curve equation: $V = V_{\max}(\text{PO}_2/P_{V50}) / \{1 + (\text{PO}_2/P_{V50})\}$. Here, the maximal rate of O₂ uptake (V_{max}) and effluent buffer PO₂ at half maximal O₂ uptake (P_{V50}) were obtained from the slope (1/V_{max}) and the ordinate intercept (P_{V50}/V_{max}) of the Hanes-Woolf plot (effluent PO₂/V vs. effluent PO₂). Changes in oxygen saturation of Mb (Y) were analyzed using the Hill equation [25], $Y = \text{PO}_2^n / (\text{PO}_2^n + P_{Y50}^n)$, where P_{Y50} is PO₂ at half saturation of Mb (Y₅₀) and n is the Hill coefficient. In the original Hill equation, n was treated as a constant. This equation expressed the ODC of Mb well but not the ODC of Hb because the Hill plot for Hb deviated from a straight line at both extremes. To make the Hill plot applicable to Hb, Wyman [26] extended the equation by linearizing it in the form: $\log \{Y/(1 - Y)\} = n (\log \text{PO}_2 - \log P_{Y50})$ where n was treated as a variable. This extension allowed cooperativity measured by n to vary depending on Y.

Results

Oxygen uptake by perfused muscle in different respiration states

Figure 2 shows the steady-state O₂ uptake rate (V) of a perfused muscle. The respiration rate of the muscle was varied by controlling mitochondrial respiration activity by about 7.5-fold (compare the V_{max} values described below) from a suppressed state with an inhibitor (KCN) of mitochondrial respiration to enhanced states with two levels of an uncoupler (2,4-dinitrophenol) of mitochondrial respiration. Three preparations of muscle were used for the experiments in each mitochondrial activity state. The actual PO₂ values of the influent and effluent buffers at the maximal O₂ inflow rate are listed in Table 1. Changes in the value of V were well expressed by a rectangular hyperbolic curve as a function of effluent buffer PO₂ (Fig. 2). Table 1 also gives the estimated V_{max} and P_{V50} obtained from these data as described in Materials and Methods. V_{max} and P_{V50} became larger by approximately 7.5-fold and 2-fold, respectively, for the maximal increase in respiration activity. With elevation of respiration activity, the critical PO₂, at which O₂ uptake of perfused hindlimb muscle starts to decrease, increased to higher values. This indicates that, under higher respiration activity, O₂ supply

Table 2: Linear regression parameters for Hill's plot of myoglobin oxygenation in perfused rat hindlimb muscle, $\log [Y/(1-Y)] = k + n_{app} \cdot \log [\text{effluent PO}_2]$

Respiration state:	Suppressed (0.4 mM KCN)	Control	Enhanced (5 μ M DNPa)	Enhanced (10 μ M DNP)
Slope, n_{app} *	1.10 \pm 0.10	1.46 \pm 0.06	1.63 \pm 0.15	1.85 \pm 0.05
Intercept, k *	-1.36 \pm 0.13	-3.08 \pm 0.13	-3.75 \pm 0.34	-4.43 \pm 0.12

* 95% confidence level.

to the perfused muscle was limited even at very high influent PO_2 (~ 700 mmHg). This situation occurred because the flow rate of the perfusate and the capillary PO_2 were controlled independently of the respiration activity state so that the PO_2 gradient between the perfusate and the mitochondria became larger at higher respiration states.

Relationship between effluent buffer PO_2 and Mb oxygenation in perfused muscle

Figure 3A shows ODCs for Mb in the perfused muscle. Here, Y is plotted against effluent buffer PO_2 . These ODCs are apparent in the sense that the PO_2 is not the value at the location where Mb is working. The curve was shifted to the right and became steeper as muscle respiration activity was enhanced. These oxygenation data were further expressed by means of the Hill plot ($\log [Y/(1-Y)]$ vs. $\log \text{PO}_2$), yielding linear plots (Fig. 3B). The effluent buffer PO_2 at half saturation (P_{Y50}) and the slope of the Hill plot (the Hill coefficient, n) obtained from these plots are listed in Table 2, where n is expressed as n_{app} (apparent n). The P_{Y50} value became larger with an increase in muscle respiration activity. The $\log P_{Y50}$ value was nearly linearly related to the $\log P_{V50}$ value (not shown). The n_{app} value also increased from 1.10 in the suppressed respiration activity state to 1.85 in the 7.5-fold enhanced respiration activity state.

Since this virtual cooperativity is of particular interest, its relation to O_2 uptake rate was further examined. Figure 4 shows the dependence of n_{app} on V at the half O_2 saturation point of Mb (V_{Y50}). The n_{app} value asymptotically increased from unity for the non-respiring state to 2.23 at infinite V_{Y50} . These results indicate that the apparent ODC of Mb in the perfused muscle is transformed from a hyperbolic curve to a sigmoid curve depending on the magnitude of tissue respiration. Effect of the Hill coefficient (n) on ratio of substrate (or ligand) concentrations necessary to change enzyme activity from 90% to 10% of maximal can be expressed with a parameter, $R (= 81^{1/n})$ [27]. Here, the O_2 transport efficiency (EO_2) was estimated as ratio of the parameter at $n_{app} = 1$ to that at a given value of n_{app} (Fig. 4 inset). Figure 5 shows the effect of muscle respiration on the O_2 gradient between effluent and the perfused tissue. Assuming the effluent buffer PO_2 approximates the

capillary PO_2 , the calculated O_2 gradient from capillary to cytoplasmic space (ΔPO_2) is plotted against V_{Y50} . Here, the P_{50} value of Mb in the perfused muscle was 2.3 mmHg [21]. ΔPO_2 increased with the increase in V_{Y50} . This result indicates the presence of a large O_2 diffusion barrier between capillary lumen and cytoplasmic space.

Discussion

In the present study, by using computer-controlled rapid scanning fiber-optic spectrophotometry, I directly measured Y for Mb in isolated rat hindlimb muscles under extensive changes in respiration rate caused by mitochondrial activity control or perfusate PO_2 control. It is assumed that capillary PO_2 may be approximated by effluent PO_2 in the present experiment, and I plotted the Y values as a function of effluent buffer PO_2 . Thereby, I expected that this treatment enabled a meaningful comparison of the ODCs for Mb and Hb. I found that thus plotted apparent ODC for skeletal muscle Mb was hyperbolic under a suppressed metabolic activity condition whereas it became sigmoid under enhanced metabolic activity conditions, realizing virtually cooperative oxygenation of the monomeric Mb.

It is generally accepted that cooperative O_2 binding by Hb is advantageous for efficient O_2 transfer from the alveolar gas to red cells and from red cells to peripheral tissues. Based on the Hill equation, Graby and Meldon [28] showed that an n value (here, n is a constant) of 1.5 to 2.0 is more favourable for minimizing the change in blood flow under resting conditions than the normal n value of 2.5 to 3.0, whereas an n value as large as 3 is beneficial for a large amount of O_2 extraction under vigorous exercise. Kobayashi et al. [29] showed that, under resting conditions, O_2 release from Hb becomes most sensitive to PO_2 change at $Y = 38\%$ where cooperativity measured by n (here, n is a variable of PO_2) is not maximal, whereas it becomes less sensitive at the mixed venous blood PO_2 where Y is around 70% and cooperativity is nearly maximal. These reports indicate that, under resting conditions, the blood reserves an O_2 transport capacity to meet possible increases in O_2 demand, e.g. under exercise conditions, and the sigmoid character of ODC becomes more important under such conditions. This situation is real-

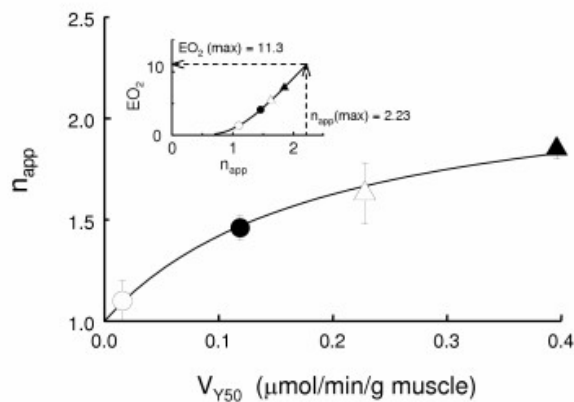


Figure 4

Relationship between n_{app} and steady-state O_2 uptake rate at $Y = 50\%$ (V_{Y50}). The V_{Y50} values were obtained from the hyperbolic curves in Fig. 2. The symbols (mean \pm SD) are as in Fig. 2. The solid line, which was calculated from the equation: $n_{app} = 1 + 1.23 \cdot V_{Y50} / (0.193 + V_{Y50})$, simulates the observed dependence. The maximal value of n_{app} at infinite V_{Y50} is 2.23, which was obtained using the Hanes-Woolf plot, $V_{Y50} / (n_{app} - 1)$ vs. V_{Y50} . The inset figure shows the relationship between n_{app} and O_2 transport efficiency (EO_2) (see Text).

ized by maintaining Y at a rather high level (70%) below which the Y value drops sharply upon PO_2 decrease within the very steep middle portion of ODC.

The present study has clearly shown that the apparent ODC for Mb in intact skeletal muscle is sigmoid, the n_{app} value being 1.46 under the control condition (Table 2) and 2.23 under the maximal respiratory condition (Fig. 4). These n_{app} values greater than unity imply that the muscle Mb binds O_2 in a virtually cooperative manner with variation of effluent buffer PO_2 . This phenomenon implies that the sensitivity of Y for Mb to vessel PO_2 change becomes higher for increased O_2 demands than for normal O_2 demand. In addition to this effect, the rightward shift of the ODC upon increases in oxygen demand will undoubtedly enhance O_2 unloading from Mb. These effects will facilitate Mb-mediated O_2 transfer from Hb to Cyt. aa₃, especially for heavy O_2 demands. Based on the Hill equation, the O_2 transport efficiency of Mb in the perfused muscle is estimated to increase ca. 4-fold under the control condition and ca. 11-fold under maximally respiring condition (Fig. 4 inset).

The Mbs isolated from body wall or radular muscle of a limited number of annelidan and molluscan species are dimers and show some cooperativity in oxygen binding ($1 < n < 2$) but no Bohr effect [30]. The physiological signif-

icance of these dimeric Mbs is unknown. As shown in the present study, the ODC of monomeric Mb can exhibit virtual cooperativity and O_2 demand-dependent shifts. The virtually cooperativity and O_2 demand-dependent shifts of Mb oxygenation *in vivo* are probably common features at least for vertebrate Mbs, and this may provide a basis for explaining why the vertebrate Mb has been preserved as a monomer during molecular evolution.

The virtual cooperativity in Mb oxygenation observed in the present study is explained in terms of the PO_2 gradient along the O_2 diffusion path. If the tissue O_2 demand was null, then the PO_2 gradient would be absent and the apparent ODC for Mb would be identical with the real ODC for Mb in solution. At a steady state with a certain level of O_2 demand a PO_2 gradient develops across red cell membrane, blood plasma, capillary wall, sarcolemma and sarcoplasm, making the PO_2 sensed by Mb lower than the capillary PO_2 . Then, the apparent ODC will be shifted toward the right because a capillary PO_2 value higher than the PO_2 value sensed by Mb is needed to maintain the same Y value as that which occurs in the absence of a PO_2 gradient. When the tissue O_2 demand is kept constant, the ratio of capillary PO_2 to sarcoplasm PO_2 will become larger at low capillary PO_2 than at high capillary PO_2 . This will cause a more extensive rightward shift of the apparent ODC in the low saturation range than in the high saturation range, making the curve steeper than the real one. Increase in tissue oxygen demand will enhance this mechanism and make the curve more right-shifted and sigmoid. All the apparent ODCs observed in the present study are shifted toward the right compared to the real one measured for Mb in solution (Fig. 3).

At present, detailed explanations for this cooperative mechanism is difficult. However, it could be argued that heterogeneous oxygenation in tissue [31] and in single myocytes [32] might be responsible in part for the shift and the shape change of the Mb ODC, and might also enhance intercellular O_2 transfer, i.e., re-distribution of O_2 among adjacent myocytes, although we adopted high and constant flow rate perfusion conditions (i.e., about 50 times higher than normal blood flow) and, thus, the perfused vessels of muscle were always passively dilated.

Unfortunately, it is not practical to use a Hb solution or a red cell suspension as the perfusate in our experiments because the absorption spectra for Hb and Mb are too similar and independent observations of Mb oxygenation are not feasible, especially when the concentration of Hb is much higher than that of Mb. To overcome the problem that the O_2 solubility of the buffer is much smaller than that of a Hb solution or a red cell suspension two strategies were employed: one was to make the PO_2 of the influent buffer as high as that of water vapor-saturated O_2 (ca.

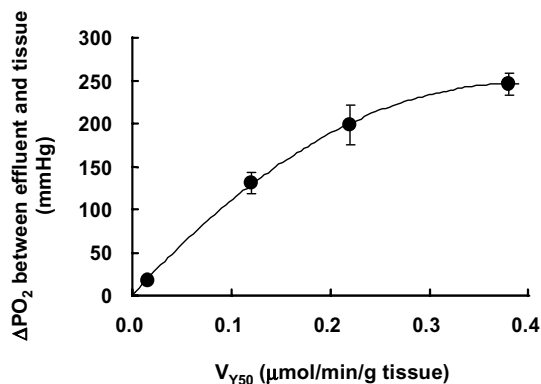


Figure 5

Correlation between O_2 consumption at 50% of Mb oxygenation (V_{Y50}) and calculated O_2 gradient from vascular to cytoplasmic space in perfused hindlimb muscle (ΔPO_2). To calculate ΔPO_2 , $\text{PO}_2 = 2.3$ mmHg at $Y = 50\%$ of Mb in the perfused muscle was used [21]. $\Delta\text{PO}_2 = -1635 * V_{Y50} + 1269$ ($r^2 = 0.9995$).

700 mmHg) and the other was to use a high flow rate for the buffer, which was about 50 times higher than that of normal blood flow. As a result, the inflow of O_2 was about 5 times larger than that of the tissue O_2 consumption at the control metabolic rate. The large O_2 diffusion barrier (see Fig. 5) and the high PO_2 of the influent buffer (and consequently, the high capillary PO_2) are an additional (and probably, major) cause for the rightward shift of the apparent ODC of Mb. The apparent ODCs of Mb in the control and enhanced respiration activity states (Fig. 3A) are right-shifted compared to the whole blood ODC (Fig. 1). One may suppose that Mb cannot work when the capillary PO_2 is in the physiological range (40 to 100 mmHg) because its O_2 saturation is too low to function, as judged from Fig. 3A. However, the actual apparent ODCs for Mb in muscles with blood circulation will be shifted much more toward the left compared to those shown in Fig. 3A, and Mb can be saturated with O_2 to practical levels. The important point is that the difference in *in vivo* O_2 saturation between Hb and Mb is not so large as that expected from the ODCs in Fig. 1. In fact, Y for Mb in working muscles is less than around 50% [19,20,31-33]. Red blood cell (RBC) in perfusion buffer appears to exert considerable effects on intracellular oxygenation in the beating heart [34], probably due to the facilitated O_2 transfer by RBC motion within capillary lumen [35]. Therefore, the virtually cooperative oxygenation of Mb might be only demonstrated in organs perfused with RBC-free medium. However, it is well known that the blood flow in the capillary bed is not constant and frequently only plasma flow

is observed. In this case, the virtually cooperative oxygenation of Mb may play a significant role for O_2 transfer from capillary to mitochondria.

In summary, I found that the ODC for Mb in intact skeletal muscle is sigmoid and right-shifted. This virtually cooperative O_2 binding by Mb and the right-shift of ODC become more marked as tissue respiration activity is increased. Hence, increase in O_2 demand in tissues makes the O_2 saturation of Mb more sensitive to capillary PO_2 change and enhances Mb-mediated O_2 transfer from red cell to mitochondria. The virtual cooperativity and O_2 demand-dependent shifts of ODC may give a basis for explaining why Mb has been preserved as a monomer during molecular evolution. Preservation of a monomeric structure may be required to retain multi-functional role of Mb *in vivo*.

Acknowledgements

This work was supported in part by a research grant from the Ministry of Education, Science and Culture of Japan.

References

1. Kagen LJ: *Myoglobin* New York, London: Columbia University Press; 1973.
2. Millikan GA: **Experiments on muscle haemoglobin *in vivo*; the instantaneous measurement of muscle metabolism.** *Proc R Soc Lond B Biol Sci* 1937, **123**:218-241.
3. Millikan GA: **Muscle hemoglobin.** *Physiol Rev* 1939, **19**:503-523.
4. Wyman J: **Facilitated diffusion and the possible role of myoglobin as a transport mechanism.** *J Biol Chem* 1966, **241**:115-121.
5. Wittenberg JB: **Oxygen transport: a new function proposed for myoglobin.** *Biol Bull* 1959, **117**:402-403.
6. Scholander PF: **Oxygen transport through hemoglobin solutions.** *Science* 1960, **131**:585-590.
7. Wittenberg JB: **Myoglobin-facilitated oxygen diffusion: Role of myoglobin in oxygen entry into muscle.** *Physiol Rev* 1970, **50**:559-636.
8. Jones DP, Kennedy FG: **Intracellular O_2 gradients in cardiac myocytes. Lack of a role for myoglobin in facilitation of intracellular diffusion.** *Biochem Biophys Res Commun* 1982, **105**:419-424.
9. Kennedy FG, Jones DP: **Oxygen dependence of Mitochondrial function in isolated rat cardiac myocytes.** *Am J Physiol* 1986, **250**:C374-C383.
10. Livingston DJ, LaMar GN, Brown WD: **Myoglobin diffusion in bovine heart muscle.** *Science* 1983, **220**:71-73.
11. Jurgens KD, Peters T, Gros G: **Diffusivity of myoglobin in intact skeletal muscle cells.** *Proc Natl Acad Sci USA* 1994, **91**:3829-3833.
12. Paradopoulos S, Jurgens KD, Gros G: **Diffusion of myoglobin in skeletal muscle cells-dependence on fiber type, contraction and temperature.** *Pflügers Arch* 1995, **430**:519-525.
13. Doeller JE, Wittenberg BA: **Myoglobin function and energy metabolism of isolated cardiac myocytes: effect of sodium nitrite.** *Am J Physiol* 1991, **261**:H53-H62.
14. Garry DJ, Meeson A, Yan Z, Williams RS: **Life without myoglobin.** *Cell Mol Life Sci* 2000, **57**:896-898.
15. Garry DJ, Ordway GA, Lorenz JN, Radford NB, Chin ER, Grange RW, Bassel-Duby R, Williams RS: **Mice without myoglobin.** *Nature* 1998, **395**:905-908.
16. Godecke A, Fogel U, Zanger K, Ding Z, Hirchenhain J, Decking UKM, Schrader J: **Disruption of myoglobin in mice induces multiple compensatory mechanisms.** *Proc Natl Acad Sci USA* 1999, **96**:10495-10500.
17. Burmester T, Weich B, Reinhardt S, Hankeln T: **A vertebrate globin expressed in the brain.** *Nature* 2000, **407**:520-523.

18. Giardina B, Ascenzi P, Clementi ME, De Sanctis G, Rizzi M, Coletta M: **Functional modulation by lactate of myoglobin.** *J Biol Chem* 1996, **271**:16999-17001.
19. Mole PA, Chung Y, Tran T-K, Sailusuta N, Hurd R, Jue T: **Myoglobin desaturation with exercise intensity in human gastrocnemius muscle.** *Am J Physiol* 1999, **277**:R173-R180.
20. Richardson RS, Noyszewski RA, Kendrick KF, Leigh JS, Wagner PD: **Myoglobin O₂ desaturation during exercise. Evidence of limited O₂ transport.** *J Clin Invest* 1995, **96**:1916-1926.
21. Seiyama A, Maeda N, Shiga T: **Optical measurement of perfused rat hindlimb muscle with relation of oxygen metabolism.** *Jpn J Physiol* 1991, **41**:49-61.
22. Seiyama A, Kosaka H, Maeda N, Shiga T: **Effect of hypothermia on skeletal muscle metabolism in perfused rat hindlimb.** *Cryobiology* 1996, **33**:338-346.
23. Ruderman NB, Houghton CRS, Hems R: **Evaluation of the isolated perfused rat hindquarter for the study of muscle metabolism.** *Biochem J* 1971, **124**:639-651.
24. Shiota T, Sugano M: **Characteristics of rat hindlimbs perfused with erythrocyte- and albumin-free medium.** *Am J Physiol* 1986, **251**:C78-C84.
25. Hill AV: **The possible effects of the aggregation of the molecules of haemoglobin on its dissociation curves.** *J Physiol* 1910, **40**:4-7.
26. Wyman J: **Linked functions and reciprocal effects in hemoglobin: A second look.** *Advan Protein Chem* 1964, **19**:223-286.
27. Taketa K, Pogell BM: **Allosteric inhibition of rat liver fructose 1,6-diphosphatase by adenosine 5'-monophosphate.** *J Biol Chem* 1965, **240**:651-662.
28. Garbyb L, Meldon J: *The Respiratory Functions of Blood* New York, London: Plenum Medical Book; 1977.
29. Kobayashi M, Kimura S, Ishigaki KI, Makino N, Imai K: **Significance of oxygen affinity of fetal and adult human hemoglobins.** *Zool Sci* 1996, **13**:661-664.
30. Suzuki T, Imai K: **Evolution of myoglobin.** *Cell Mol Life Sci* 1998, **54**:979-1004.
31. Gayeski TEJ, Honig CR: **O₂ gradients from sarcolemma to cell interior in red muscle at maximal V_{O₂}.** *Am J Physiol* 1986, **251**:H789-H799.
32. Takahashi E, Doi K: **Regulation of oxygen diffusion in hypoxic isolated cardiac myocytes.** *Am J Physiol* 1996, **271**:H1734-H1738.
33. Wittenberg BA, Wittenberg JB: **Transport of oxygen in muscle.** *Annu Rev Physiol* 1989, **51**:857-878.
34. Schenkman KA, Beard DA, Ciesielski WA, Feigl EO: **Comparison of buffer and red blood cell perfusion of guinea pig heart oxygenation.** *Am J Physiol* 2003, **285**:H1819-1825.
35. Seiyama A, Shiga T: **Oxygen transfer in peripheral organs: Researches on intact organs with optical techniques.** *Adv Exerci Sports Physiol* 1998, **4**:37-49.
36. Imai K: **Oxygen transport and its regulation.** *Rinsho Kensa (in Japanese)* 1986, **30**:363-371.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

