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CYP2C9, CYPC19 and CYP2D6 gene profiles and () CrossMark gene susceptibility to drug response and toxicity in Turkish population

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Abstract Pharmacogenetics is a vast field covering drug discovery research, the genetic basis of pharmacokinetics and dynamics, genetic testing and clinical management in diseases. Pharmacogenetic approach usually focuses on variations of drug transporters, drug targets, drug metabolizing enzymes and other biomarker genes. Cytochrome P450 (CYP) enzymes, an essential source of variability in drug-response, play role in not only phase I-dependent metabolism of xenobiotics but also metabolism of endogenous compounds such as steroids, vitamins and fatty acids. CYP2C9, CYP2C19 and CYP2D6 enzymes being highly polymorphic are responsible for metabolism of a variety of drug groups. In the study, it was determined the genotype and allele frequency of CYP2C9*2, CYP2C19*3, CYP2C19*2, CYP2C19*3, CYP2C19*17, CYP2D6*9 and CYP2D6*41, very common and functional single-nucleotide polymorphisms (SNPs), in healthy volunteers. The genotype distributions were consistent with the Hardy-Weinberg equilibrium in the population (p > 0.05). It is believed that the determination of polymorphisms in the enzymes may be beneficial in order to prevention or reduction in adverse effects and death. The recessive allele frequencies of CYP2C9*2, CYP2C19*3, CYP2C19*2, CYP2C19*3, CYP2C19*17, CYP2D6*9 and CYP2D6*41 were 11, 13, 12, 13, 25, 4 and 15%, respectively. According to the obtained results, the carriers of CYP2D6*9 variant allele should be received higher doses of the drugs metabolizing with this enzyme in Turkish population, while the carriers of other variant alleles do not generally have any requirement of dose regimen.

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1. Introduction

CYP enzymes are responsible for phase I metabolism over 90% of drugs and naturally occurring xenobiotics and endogenous substrates. To date, fifty-seven *CYP* genes, which involve three families (CYP1, CYP2 and CYP3) contributing to the oxidative metabolism of various compounds, have been

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detected (Shastry, 2006). CYP2C subfamily has at least four isoforms (CYP2C8, CYP2C9, CYP2C18 and CYP2C19) located on chromosome 10 (Scordo et al., 2004).

CYP2C9, which is greatly polymorphic and the most abundant isoform of CYP2C, metabolizes a variety of drug groups including anticoagulants, anticonvulsants, and non-steroidal anti-inflammatory agents (Sosa-Macías et al., 2010). More than thirty *CYP2C9* variants and sub-variants have been identified. *CYP2C9*2* and *CYP2C9*3*, the most common variants, have effect in decreasing enzyme activity (Alessandrini et al., 2013).

CYP2C19 is responsible for the metabolism of more than 25 clinically important drug groups including a lot of psychotropics, proton pump inhibitors and anticonvulsants. Also, it contributes to the clearance of S-mephenytoin, diazepam, omeprazole, proguanil and *R*-warfarin (Scordo et al., 2004; Rosemary and Adithan, 2007). There are more than 30 *CYP2C19* allelic variants. The most common variants are *CYP2C19*2* and *CYP2C19*3*, which reduce enzyme function. Recently, another variant, *CYP2C19*17*, awakens researchers' interest and is associated with increased enzyme function (Alessandrini et al., 2013).

CYP2D6 is responsible for hydroxylation or demethylation of approximately 25% of clinically important drugs such as antiarrhythmic, psychiatric, antihistaminic and antidepressant (Gardiner and Begg, 2006; Zanger et al., 2004). Also, it is known that CYP2D6 plays important role in the metabolism of the analgesic codeine (Crews et al., 2012). It was identified over one hundred variant alleles of CYP2D6. While some alleles of those cause normal or increased activity in enzyme function (*1, *2 and *35), some lead to decreased activity (*9, *10, *17, *29, and *41) or the absence of enzyme function (*3, *4, *5, and *6) (Broly and Meyer, 1993; Gaedigk et al., 1991; Gaedigk et al., 2003; Sakuyama et al., 2008).

In the present study, it was aimed that the results may provide a helpful support in the optimization of pharmacological therapies in Turkish population by determining their *CYP2C9*, *CYP2C1*9 and *CYP2D6* genotype profiles.

2. Material and methods

Genomic DNA was extracted from whole blood samples of unrelated 160 Turkish healthy volunteers (88 females and 72 males, aged 20–65 years) by High Pure PCR Template Preparation Kit (Roche, Germany) according to the manufacture's protocol. Genotyping of *CYP2C9*2* (rs1799853, 430C > T), *CYP2C9*3* (rs1057910, 1075A > C), *CYP2C19*2* (rs4244285, 681G > A), *CYP2C19*3* (rs4986893, 636G > A), and *CYP2C19*17* (rs12248560, 806C > T) variants was performed by polymerase chain reaction (PCR) restriction fragment length polymorphism (RFLP) methods. The temperature was controlled by a programmable heat block (Gene Amp PCR System 9700; Applied Biosystems, Carlsbad, CA, USA). Restriction enzymes were obtained from New England Biolabs (Hitchin, UK) and Fermentas (Vilnius, Lithuania). The other information about the genetic variants studied is given in Table 1.

Genotyping of *CYP2D6*9* (rs5030656, 2615_2617delAAG) and *CYP2D6*41* (rs28371725, 2988G > A) variants was performed on Roche Light Cycler 480 Real-Time PCR platform. Required DNA purification ensured by using High Pure PCR Product Purification Kit and single nucleotide polymorphism (SNP) analysis was performed by using LightCycler FastStart DNA Master HybProbe and custom designed LightSNiP assay probe (Roche, Germany). All participants provided informed consent and studies were approved by the ethics committee of Istanbul University (2014/1546).

The Hardy-Weinberg equilibrium analysis was performed to compare the observed and expected genotype frequencies of subjects by using the chi-square (χ^2) test. Differences in the *CYP2C9*2*, *CYP2C19*3*, *CYP2C19*2*, *CYP2C19*3*, *CYP2C19*17*, *CYP2D6*9* and *CYP2D6*41* genetic variants between Turkish and other ethnic populations were also assessed by χ^2 test. A *p* value below 0.05 was considered statistically significant throughout the population comparisons. All statistical analyses were performed using Statistical Package for Social Sciences (SPSS) software (Version 17, Chicago, USA).

3. Result and discussion

In the present study, it was determined the genotype and allele frequencies of *CYP2C9*, *CYP2C19* and *CYP2D6* genes in Turkish population. There were no *CYP2C19*3* homozygous variant type (A) and *CYP2D6*9* homozygous wild type (–) (Table 2). There are a lot of publications about CYP2C9, CYP2C19 and CYP2D6 enzyme activities and the relationship between the gene variants and drug-response. However, there are few data on evaluation of genetic profile in Turkish population.

In the present study, it was determined the frequencies of CYP2C9*2 (C) and CYP2C9*3 (C) variant alleles were 11 and 13%, respectively. According to the previous studies, the frequency of CYP2C9*2 (C) was 10, 7, and 28% in European, Caucasian, and Japan, respectively. In Chinese population, CYP2C9*2 (C) was not observed. The frequencies of *CYP2C9**3 (C) were 6, 5.8, 7, 4, and 2.7% in Asian, European, Caucasian, Chinese and Japan, respectively (www.hapmap. org; Allabi et al., 2003; Kimura et al., 1998; Ota et al., 2015; Sugimoto et al., 2007). Bűdi et al. (2015) suggested that it could improve the safety of antiepileptic therapies in vulnerable pediatric patients and prevent patients from misdosing when genotyping CYP2C9*2 and CYP2C9*3. Similarly, Kawai et al. (2014) investigated whether there were any associations with some genetic variants and adverse drug reactions as receiving warfarin therapy which has narrow therapeutic index and is commonly used as anticoagulant. According to the results, the patients who carried CYP2C9*3 variant allele and received warfarin for 30 or more day might have 2-fold risk of major bleeding. In another study, it has been reported the carriers having CYP2C9*2 and CYP2C9*3 had 10-fold lower S-warfarin clearance in contrary to CYP2C9*11 and CYP2C9*33 (Scordo et al., 2002). Higashi et al. (2002) indicated that there was a significant association with the increasing risk of over-anticoagulation and bleeding events in the carriers having CYP2C9*2 and CYP2C9*3, and genotyping of CYP2C9 variants could provide developing dosing protocols rightly and reducing adverse effects in patients being treated warfarin.

In the present study, the allele frequencies of *CYP2C9*2* (C), *CYP2C19*3*(A) and *CYP2C19*17*(T) were 12, 13, and 25%, respectively. The frequencies of *CYP2C19*2* were 15, 17, and 29% in European, Caucasian and Japan, respectively,

SNPs	Primer sequences	Annealing temperature	Restriction enzyme	Digestion products (bp)
CYP2C9 [*] 2	F: 5'gTATTTTggCCTgAAACCCATA 3'	60.6 °C	AvaII	Wild (CC): 454
(rs1799853)	R: 5' ACCCTTggTTTTTCTCAACTC 3'			Mutant (TT): 57, 397
CYP2C9 [*] 3	F: 5' TgCACgAggTCCAgAgATgC 3'	59 °C	NsiI	W (AA): 168
(rs1057910)	R: 5' gATACTATgAATTTggggACTTC 3'			M (CC): 50, 118
CYP2C19 [*] 2	F: 5' AATTACAACCAgAgCTTggC 3'	57.9 °C	SmaI	W (GG): 168
(rs4244285)	R: 5' TATCACTTTCCATAAAAgCAAg 3'			M (AA): 50, 118
<i>CYP2C19[*]3</i>	F: 5' CATTTAgCTTCACCCTgTg 3'	50.6 °C	BsaJI	W (GG): 251
(rs4986893)	R: 5' gggCTTggTCAATATAgAAT 3'			M (AA): 33, 94, 124
<i>CYP2C19[*]17</i>	F: 5' gTgAAgCCTgTTTTATgAA 3'	57.2 °C	SfaNI	W (CC): 165
(rs12248560)	R: 5' gTggCgCATTATCTCTTA 3'			M (TT): 20, 145

SNPs	Genotype	Genotype frequency n (%)	Allele frequency
CYP2C9 [*] 2	C/C	127 (79.36)	C: 0.89
(rs1799853)	C/T	31 (19.38)	T: 0.11
	T/T	2 (1.25)	
<i>CYP2C9[*]3</i>	A/A	114 (80.28)	A: 0.87
(rs1057910)	A/C	18 (12.67)	C: 0.13
	C/C	10 (7.04)	
<i>CYP2C19[*]2</i>	G/G	95 (65.52)	G: 0.88
(rs4244285)	G/A	34 (23.45)	A: 0.12
	A/A	16 (11.03)	
<i>CYP2C19[*]3</i>	C/C	118 (74.68)	A: 0.13
(rs4986893)	Ć/A	40 (25.32)	C: 0.87
	A/A	0 (0)	
<i>CYP2C19[*]17</i>	C/C	98 (64.47)	C: 0.75
(rs12248560)	C/T	33 (21.71)	T: 0.25
× ,	T/T	21 13.81	
<i>CYP2D6[*]9</i>		150 (00.28)	
	AAG/AAG	159 (99.38)	AAG: 0.96
(rs5030656)	(-)/AAG	1 (0.63) 0 (0)	(-): 0.04
¥	(-)/(-)	0 (0)	
<i>CYP2D6[*]41</i>	G/G	112 (73.68)	G: 0.85
(rs28371725)	G/A	35 (23.03)	A: 0.15
	A/A	5 (3.29)	

whereas the distribution of CYP2C19*3 was 11% in Japan. Germany, American, and Caucasian did not have CYP2C19*3 variants. Moreover, the values for CYP2C19*17 were 18, 3, and 1 in Caucasian, Chinese and Japan (Allabi et al., 2003; Ota et al., 2015; Sugimoto et al., 2007; Hamdy et al., 2002; Saber et al., 2014; Santos et al., 2011; Zhou et al., 2009). Kubica et al. (2011) reported CYP2C19*2 had an association about excess of ischemic events such as myocardial infarction and stent thrombosis while CYP2C19*17 variant allele undertakes a critical role for bleeding. In a study, CYP2C19*17 enzyme and its effects were investigated in clopidogreltreated patients with coronary stent placement. It was observed that the carriers CYP2C19*17 heterozygous and homozygous variant alleles had the lowest (ADP)-induced platelet aggregation values (AUmin (area under the curve of arbitrary units): 186) and the highest risk of bleeding events. In

Table 1. The conditions about studied genes in the present study

particular, the risk of bleeding events was approximately 4fold more in the carriers homozygous CYP2C19*17 allelic variant (Sibbing et al., 2010). Moreover, the presence of CYP2C19*17 variant allele in a large cohort of Caucasian subjects was associated with the endoscopically peptide ulcer diseases (Musumba et al., 2013). Mega et al. (2009) showed that the plasma concentration of the active metabolites of clopidogrel and the maximal platelet aggregation decreased in the ratio of 32.4% and 9%, respectively, in the carriers who had at least one CYP2C19 variant contrary to non-carriers. According to this study, CYP2C19*2 genetic variant was associated with higher rate of stent thrombosis. Similarly, the risk of stent thrombosis was an importantly higher in the carriers having at least one CYP2C19*2 variant allele on the contrary to CYP2C19*2 wild type homozygous patients with received clopidogrel. Moreover, CYP2C19*2 variant allele carriers

had \sim 3-fold higher risk of bleeding (Sibbing et al., 2009). It has been suggested an association between *CYP2C19* genotype and an increased risk of cardiovascular events in Japanese patients receiving rabeprazole, a proton pump inhibitor (Hokimoto et al., 2014).

According to the study about risperidone being an atypical antipsychotic drug for treatment of schizophrenia, it was found higher risperidone Cmax (9.66 \pm 2.24 ng/ml), higher AUC0-t (area under the curve) (161.43 \pm 39.31 ng/ml h) and lower clearance in poor CYP2D6 metabolizers. Moreover, it was detected a relationship between adverse effects (such as headache, neurological, psychiatric) and some CYP2C9 and CYP2C19 variants (Cabaleiro et al., 2014). On the other hand, there was no association between the metabolism of olanzapine, an atypical antipsychotic, and CYP2D6 polymorphism (Hägg et al., 2001). Noetzli et al. (2014) investigated the relationship between some genetic variants and donepezil, and is commonly used acetylcholinesterase inhibitor for the treatment of Alzheimer's disease. They observed a significant association with functional alleles of CYP2D6 and donepezil clearance. While its clearance was 32% slower in poor metabolizer, donepezil elimination was 67% faster in ultra-rapid metabolizers. In our population, it was determined the distributions of CYP2D6*9 (AAG) and CYP2D6*41(A) are 96 and 15%, respectively. While the distribution of CYP2D6*9 was 97 and 100% in European and Asian, respectively, the value of CYP2D6*41 was 8 and 4% in Caucasian and China, respectively (www.hapmap.org., Zhou et al., 2009; Gaedigk and Coetsee, 2008). Seven et al. (2014) pointed out that CYP2C9*3 variant allele played an important role as preventing in which drug resistance is developed for epilepsy patients. Also, it was shown that CYP2C9, CYP2C19 and CYP2D6 SNPs could affect the response to antiepileptic drugs.

In conclusion, this study will contribute to represent their genetic profile in terms of important drug-metabolizing enzymes because there was only one study about the genetic profiles of *CYP2C9*, *CYP2C19* and *CYP2D6* in Turkish population. The determination of polymorphisms in mentioned enzymes might provide advantage for dose adjustment of a lot of drugs in order to prevent and reduce adverse effects and even death.

References

- Alessandrini, M., Asfaha, S., Dodgen, T.M., Warnich, L., Pepper, M. S., 2013. Cytochrome P450 pharmacogenetics in African populations. Drug Metabol. Rev. 45 (2), 253–275.
- Allabi, A.C., Gala, J.L., Desager, J.P., Heusterspreute, M., Horsmans, Y., 2003. Genetic polymorphisms of CYP2C9 and CYP2C19 in the Beninese and Belgian populations. Br. J. Clin. Pharmacol. 56, 653– 657.
- Broly, F., Meyer, U.A., 1993. Debrisoquine oxidation polymorphism: phenotypic consequences of a 3-base-pair deletion in exon 5 of the CYP2D6 gene. Pharmacogenetics 3 (3), 123–130.
- Bűdi, T., Tóth, K., Nagy, A., Szever, Z., Kiss, A., Temesvári, M., et al, 2015. Clinical significance of CYP2C9-status guided valproic acid therapy in children. Epilepsia 56 (6), 849–855.
- Cabaleiro, T., Ochoa, D., López-Rodríguez, R., Román, M., Novalbos, J., Ayuso, C., Abad-Santos, F., 2014. Effect of polymorphisms on the pharmacokinetics, pharmacodynamics, and safety of risperidone in healthy volunteers. Hum. Psychopharmacol. 29 (5), 459–469.

- Crews, K.R., Gaedigk, A., Dunnenberger, H.M., Klein, T.E., Shen, D. D., Callaghan, J.T., et al, 2012. Clinical pharmacogenetics implementation consortium (CPIC) guidelines for codeine therapy in the context of cytochrome P450 2D6 (CYP2D6) genotype. Clin. Pharmacol. Ther. 91, 321–326.
- Gaedigk, A., Coetsee, C., 2008. The CYP2D6 gene locus in South African Coloureds: unique allele distributions, novel alleles, and gene arrangements. Eur. J. Clin. Pharmacol. 64, 465–475.
- Gaedigk, A., Blum, M., Gaedigk, R., Eichelbaum, M., Meyer, U.A., 1991. Deletion of the entire cytochrome P450 CYP2D6 gene as a cause of impaired drug metabolism in poor metabolizers of the debrisoquine/sparteine polymorphism. Am. J. Hum. Genet. 48, 943–950.
- Gaedigk, A., Ryder, D.L., Bradford, L.D., Leeder, J.S., 2003. CYP2D6 poor metabolizer status can be ruled out by a single genotyping assay for the -1584G promoter polymorphism. Clin. Chem. 49, 1008–1011.
- Gardiner, S.J., Begg, E.J., 2006. Pharmacogenetics, drug-metabolizing enzymes, and clinical practice. Pharmacol. Rev. 58, 521–590.
- Hägg, S., Spigset, O., Lakso, H., Dahlqvist, R., 2001. Olanzapine disposition in humans is unrelated to CYP1A2 and CYP2D6 phenotypes. Eur. J. Clin. Pharmacol. 57, 493–497.
- Hamdy, S.I., Hiratsuka, M., Narahara, K., El-Enany, M., Moursi, N., Ahmed, M.S., Mizugaki, M., 2002. Allele and genotype frequencies of polymorphic cytochromes P450 (CYP2C9, CYP2C19, CYP2E1) and dihydropyrimidine dehydrogenase (DPYD) in the Egyptian population. Br. J. Clin. Pharmacol. 53, 596–603.
- Higashi, M.K., Veenstra, D.L., Kondo, L.M., Wittkowsky, A.K., Srinouanprachanh, S.L., Farin, F.M., Rettie, A.E., 2002. Association between CYP2C9 genetic variants and anticoagulationrelated outcomes during warfarin therapy. JAMA 287 (13), 1690– 1698.
- Hokimoto, S., Mizobe, M., Akasaka, T., Arima, Y., Kaikita, K., Nakagawa, K., Ogawa, H., 2014. Impact of CYP2C19 polymorphism and proton pump inhibitors on platelet reactivity to clopidogrel and clinical outcomes following stent implantation. Thromb. Res. 133 (4), 599–605.
- International HapMap Consortium 2002–2010, <<u>http://hapmap.ncbi.</u> nlm.nih.gov> (accessed date 14.03.2016).
- Kawai, V.K., Cunningham, A., Vear, S.I., Van Driest, S.L., Oginni, A., Xu, H., Stein, C.M., 2014. Genotype and risk of major bleeding during warfarin treatment. Pharmacogenomics 15 (16), 1973–1983.
- Kimura, M., Ieiri, I., Mamiya, K., Urae, A., Higuchi, S., 1998. Genetic polymorphism of cytochrome P450s, CYP2C19, and CYP2C9 in a Japanese population. Ther. Drug Monit. 20, 243–247.
- Kubica, A., Kozinski, M., Grzesk, G., Fabiszak, T., Navarese, E.P., Goch, A., 2011. Genetic determinants of platelet response to clopidogrel. J. Thromb. Thrombolysis 32 (4), 459–466.
- Mega, J.L., Close, S.L., Wiviott, S.D., Shen, L., Hockett, R.D., Brandt, J., et al, 2009. Cytochrome P-450 polymorphisms and response to clopidogrel. N. Engl. J. Med. 360, 354–362.
- Musumba, C.O., Jorgensen, A., Sutton, L., Van Eker, D., Zhang, E., O'Hara, N., et al, 2013. CYP2C19* 17 gain-of-function polymorphism is associated with peptic ulcer disease. Clin. Pharmacol. Ther. 93 (2), 195–203.
- Noetzli, M., Guidi, M., Ebbing, K., Eyer, S., Wilhelm, L., Michon, A., et al, 2014. Population pharmacokinetic approach to evaluate the effect of CYP2D6, CYP3A, ABCB1, POR and NR112 genotypes on donepezil clearance. Br. J. Clin. Pharmacol. 78 (1), 135–144.
- Ota, T., Kamada, Y., Iwao-Koizumi, K., Murata, S., Kinoshita, K., 2015. Combination analysis in genetic polymorphisms of drugmetabolizing enzymes CYP1A2, CYP2C9, CYP2C19, CYP2D6 and CYP3A5 in the Japanese Population. Int. J. Med. Sci. 12 (1), 78–82.
- Rosemary, J., Adithan, C., 2007. The pharmacogenetics of CYP2C9 and CYP2C19: ethnic variation and clinical significance. Curr. Clin. Pharmacol. 2 (1), 93–109.

- Saber, M.M., Boroumand, M., Behmanesh, M., 2014. Investigation of CYP2C19 allele and genotype frequencies in Iranian population using experimental and computational approaches. Thromb. Res. 133, 272–275.
- Sakuyama, K., Sasaki, T., Ujiie, S., Obata, K., Mizugaki, M., Ishikawa, M., Hiratsuka, M., 2008. Functional characterization of 17 CYP2D6 allelic variants (CYP2D6.2, 10, 14A-B, 18, 27, 36, 39, 47–51, 53–55, and 57). Drug Metab. Dispos. 36, 2460–2467.
- Santos, P.C.J.L., Soares, R.A.G., Santos, D.B.G., Nasciment, R.M., Coelho, G.L.L.M., Nicolau, J.C., et al, 2011. CYP2C19 and ABCB1 gene polymorphisms are differently distributed according to ethnicity in the Brazilian general population. BMC Med. Genet. 12, 13.
- Scordo, M.G., Caputi, A.P., D'Arrigo, C., Fava, G., Spina, E., 2004. Allele and genotype frequencies of CYP2C9, CYP2C19 and CYP2D6 in an Italian population. Pharmacol. Res. 50 (2), 195–200.
- Scordo, M.G., Pengo, V., Spina, E., Dahl, M.L., Gusella, M., Padrini, R., 2002. Influence of CYP2C9 and CYP2C19 genetic polymorphisms on warfarin maintenance dose and metabolic clearance. Clin. Pharmacol. Ther. 72, 702–710.
- Seven, M., Batar, B., Unal, S., Yesil, G., Yuksel, A., Guven, M., 2014. The effect of genetic polymorphisms of cytochrome P450 CYP2C9, CYP2C19, and CYP2D6 on drug-resistant epilepsy in Turkish children. Mol. Diagn. Ther. 18 (2), 229–236.
- Shastry, B.S., 2006. Pharmacogenetics and the concept of individualized medicine. Pharmacogenomics J. 6, 16–21.

- Sibbing, D., Koch, W., Gebhard, D., Schuster, T., Braun, S., Stegherr, J., et al, 2010. Cytochrome 2C19*17 allelic variant, platelet aggregation, bleeding events, and stent thrombosis in clopidogreltreated patients with coronary stent placement. Circulation 121, 512–518.
- Sibbing, D., Stegherr, J., Latz, W., Koch, W., Mehilli, J., Dörrler, K., et al, 2009. Cytochrome P450 2C19 loss-of-function polymorphism and stent thrombosis following percutaneous coronary intervention. Eur. Heart. J. 30, 916–922.
- Sosa-Macías, M., Dorado, P., Alanis-Bañuelos, R.E., Llerena, A., Lares-Asseff, I., 2010. Influence of CYP2D6 deletion, multiplication, −1584C→ G, 31G→ A and 2988G→ a gene polymorphisms on dextromethorphan metabolism among Mexican tepehuanos and mestizos. Pharmacology 86 (1), 30–36.
- Sugimoto, K., Uno, T., Yamazakl, H., Tatelshl, T., 2007. Limited frequency of the CYP2C19*17 allele and its minör role in a Japanese population. Br. J. Clin. Pharmacol. 65 (3), 437–439.
- Zanger, U.M., Raimundo, S., Eichelbaum, M., 2004. Cytochrome P450 2D6: overview and update on pharmacology, genetics, biochemistry. Naunyn Schmiedebergs Arch. Pharmacol. 369, 23– 37.
- Zhou, Q., Yu, X.M., Lin, H.B., Wang, L., Yun, Q.Z., Hu, S.N., Wang, D.M., 2009. Genetic polymorphism, linkage disequilibrium, haplotype structure and novel allele analysis of CYP2C19 and CYP2D6 in Han Chinese. Pharmacogenom. J. 9, 380–394.