

LETTER TO THE JOURNAL

Single-cell transcriptomic atlas reveals immune and metabolism perturbation of depression in the pathogenesis of breast cancer

Epidemiological evidence indicates that major depressive disorder (MDD) may predispose the development and prognosis of breast cancer (BC) in females [1]. However, the mechanisms linking these phenotypes are not fully understood. Chronic stress, a hallmark of depression, has been underscored to affect anti-tumor immunity, tumor metabolic reprogramming, hormone synthesis in BC [2, 3], and increase tumor metastasis [4], but there is a lack of detailed cellular-level characterization of how MDD history affects the tumorigenesis of BC. This study explored the single-cell atlas of multiple tissues from BC patients with and without a history of MDD for characterizing the potential molecular alternations in their tumorigenesis (Figure 1A).

Paired primary tumor tissues ($n = 10$), adjacent normal tissues ($n = 7$), and peripheral blood samples ($n = 10$) were collected from a cohort of 10 BC patients, 5 of whom

had a history of MDD (Supplementary Table S1). All BC patients had estrogen receptor (ER)-positive tumors and were further predicted as Luminal A ($n = 9$) and B subtypes ($n = 1$) (Supplementary Table S2). Further details on patient recruitment, sample handling, and single-cell data analysis are provided in Supplementary Methods. In total, we obtained 224,557 single cells and further annotated them into major cell subsets based on lineage markers and copy number variations [5] (Figure 1B, Supplementary Figure S1A-B). Aneuploid cells in primary tumor tissues were obtained (Supplementary Table S3) to characterize their phenotypic differences in BC patients with MDD history (BC-MDD) or not (BC-Ctrl). The Uniform Manifold Approximation and Projection (UMAP) of unintegrated aneuploid cells revealed intrinsic differences across individual patients (Supplementary Figure S1C). Downstream functional profiling analysis identified distinct immune response pathways in BC-MDD and BC-Ctrl groups and enrichment of the oxidative phosphorylation (OXPHOS) pathway in BC-Ctrl tumors (Supplementary Figure S1D-E). Cellular Gene Set Variation Analysis (GSVA) [6] confirmed the distinct metabolic phenotypes between BC-MDD and BC-Ctrl groups (Figure 1C). Additionally, utilizing pre-defined gene module (GM) signatures of BC tumor cells [7], we observed specific restraint of GM4 and GM6 in BC-MDD (Supplementary Figure S1F-G). GM6 encompasses various antigen presentation genes, aligning with the observed downregulation of major histocompatibility complex class I (MHC-I) class genes in BC-MDD tumor cells (Supplementary Figure S1H).

Upon re-clustering 12,371 normal epithelial cells within primary breast tumor and adjacent normal tissues from the 10 patients, we identified 9 cell clusters, including luminal hormone-responsive (LumHR), luminal secretory (LumSec), and myoepithelial cells [8] (Supplementary Figure S2A-B). Distribution analysis suggested the potential enrichment of LumSec-2 and -3 clusters in primary tumor tissues of BC-Ctrl patients (Supplementary Figure

List of Abbreviations: Basal_SC, single-cell gene signature of basal breast cancer; BC, breast cancer; BC-Ctrl, breast cancer patients in the control group; BC-MDD, breast cancer patients with MDD history; CXCL12, C-X-C chemokine ligand 12; CXCL13, C-X-C motif chemokine ligand 13; CXCR4, C-X-C chemokine receptor 4; DEGs, differentially expressed genes; ER, estrogen receptor; Fibro, Fibroblasts; FN1, fibronectin; GM, gene module; GPR183, G protein-coupled receptor 183; GSVA, gene set variation analysis; HER2, Human Epidermal growth factor Receptor 2; HER2E_SC, single-cell gene signature of HER2 breast cancer; HSPH1, heat shock protein family h member 1; LumA_SC, single-cell gene signature of Luminal A breast cancer; LumB_SC, single-cell gene signature of Luminal B breast cancer; LumHR, Luminal hormone-responsive cell; LumSec, luminal hormone-responsive cell; Lym, Lymphatic endothelial cells; MDD, major depressive disorder; MDK, midkine; MHC-I, Major Histocompatibility Complex Class I; MIF, macrophage migration inhibitory factor; NK, natural killer; OXPHOS, oxidative phosphorylation; PDEF/SPDEF, Prostate-derived ETS factor; SCL38A2, solute carrier family 38 member 2; TCGA-BRCA, the cancer genome atlas-breast invasive carcinoma; UMAP, uniform manifold approximation and projection; Vas, Vascular cell; Vsmc, Vascular smooth muscle cells.

Lingling Wu, Junwei Liu, Yimeng Geng, and Jianwen Fang contributed equally.

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial-NoDerivs](https://creativecommons.org/licenses/by-nc-nd/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2024 The Author(s). *Cancer Communications* published by John Wiley & Sons Australia, Ltd on behalf of Sun Yat-sen University Cancer Center.

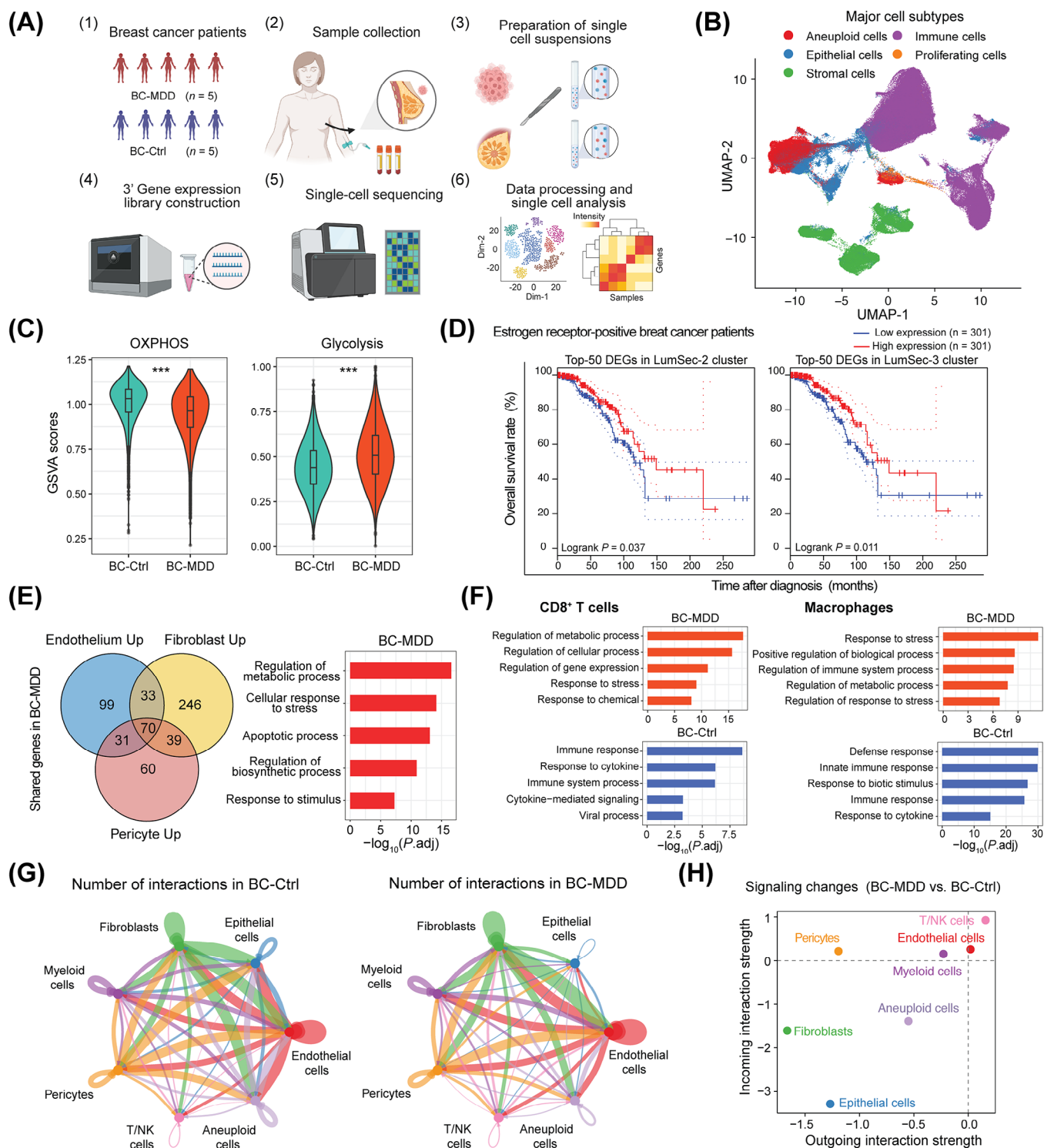


FIGURE 1 scRNA-seq reveals MDD-associated pathogenesis of breast cancer. (A) The experimental design of single-cell analyses in this study. (B) UMAP plot of single cells from tumor tissues, adjacent normal tissues, and peripheral blood samples, colored by major cell types. (C) Comparisons of the enrichment of OXPHOS (left) and glycolysis (right) pathways within aneuploidy cells from BC-Ctrl and BC-MDD groups. (D) Overall survival analyses of the contributions of top-50 highly-expressed gene signatures of LumSec-2 (left) and LumSec-3 (right) clusters in prognosis prediction of the estrogen receptor-positive patients in the TCGA-BRCA cohort. (E) Venn plot of the overlaps between upregulated differentially expressed genes of endothelial, fibroblast, and pericyte cells between BC-Ctrl and BC-MDD sample groups (left) and the pathway enrichment analysis of the shared upregulated genes (right). (F) Pathway enrichment analysis of differentially expressed genes of CD8⁺ T cells (left) and macrophages (right) in the primary tumor tissue of BC-Ctrl and BC-MDD patients. (G) The number of interactions of ligands and receptors across major cell subtypes within BC-Ctrl (left) and BC-MDD (right) samples. (H) The changes of incoming and

S2C-E). Subsequently, we investigated the potential roles of these BC-Ctrl-enriched tumor epithelial cells in tumorigenesis and prognosis. We utilized the top 50 differentially expressed genes (DEGs) of these cell clusters to evaluate their prognostic roles using The Cancer Genome Atlas-Breast Invasive Carcinoma (TCGA-BRCA) datasets [9]. Notably, we revealed that the gene signatures of these cells could only predict the prognosis of ER⁺ but not ER⁻ patients (Figure 1D, Supplementary Figure S2F). Despite the high expression of basal and human epidermal growth factor receptor 2 (HER2) breast cancer phenotypes, the prognostic roles of these cells may be related to the down-regulation of prostate-derived ETS factor (PDEF/SPDEF) [10] (Supplementary Figure S2G-H).

We then integrated and characterized the functional profiling of 25,076 stromal cells within primary breast tumor and adjacent normal tissues, encompassing 7 endothelial cell, 10 fibroblast, and 5 pericyte clusters (Supplementary Figure S3A-E). Frequency distribution analyses identified the tumor-specific cell cluster enrichment but not across patient groups in both tissue sites (Supplementary Figure S3F-H). We then characterized the potential MDD-specific functional changes within stromal cells, and noted consistent functional changes across all major stromal subtypes. We identified 70 upregulated genes shared by all stromal subtypes in BC-MDD patients, with pathway analysis suggested their involvement in metabolic regulation and cellular stress response pathways, consistent with trends observed in individual patients (Figure 1E, Supplementary Figure S3I). Specifically, we found the potential up-regulation of solute carrier family 38 member 2 (*SLC38A2*) and heat shock protein family H member 1 (*HSPH1*) in primary breast tumor and adjacent normal tissues from BC-MDD patients (Supplementary Figure S3J), highlighting the potential roles of amino acid transport and stress responses in MDD-affected BC patients.

To characterize the T cell immune response alterations in BC-MDD patients, we re-integrated 91,615 annotated T/natural killer (NK) cells from primary breast tumors, adjacent normal tissues, and peripheral blood samples. This analysis identified 15 distinct cell clusters (Supplementary Figure S4A-B). Notably, C-X-C motif chemokine ligand 13 (*CXCL13*)⁺ CD4⁺ T cells (CD4-Tem-2) and regulatory T cells were enriched in primary tumors, NK and $\gamma\delta$ T cells were enriched in peripheral blood (Supplementary Figure S4C). However, the frequency comparisons of T/NK cell clusters across patient groups revealed non-significant

differences across all tissues (Supplementary Figure S4D). We then profiled the functional alternations in CD8⁺ T cells in primary tumors with MDD history, which proposed the enrichment of interferon-related genes and cytotoxic genes in BC-Ctrl tumor tissues, validated with individual patients (Supplementary Figure S4E-F). This finding is consistent with the diminished activation of immune response and cytokine response pathways in CD8 T cells in the BC-MDD group (Figure 1F).

We conducted a parallel analysis with 27,563 myeloid cells and identified 16 cell clusters within monocytes, macrophages, and dendritic cells (DCs) (Supplementary Figure S5A-B). Among those, the Macro-m1-1 cell cluster, characterized by high expression of chemokine genes, predominated in primary tumor tissues, alongside the G protein-coupled receptor 183 (*GPR183*)-positive cDC-2 cells (Supplementary Figure S5C). Conversely, we observed no significant distribution variations in myeloid cells across all tissues among patient groups (Supplementary Figure S5D). Exploring the functional modifications of macrophages in primary tumor tissues, we uncovered enrichment of fatty acid-binding genes in the BC-MDD group and interferon-related genes in the BC-Ctrl group, which were validated across individual patients (Supplementary Figure S5E-F). Similar to CD8⁺ T cells, we revealed enrichment of stress response and metabolic-related pathways but with the impairment of innate immune response pathways in the BC-MDD group (Figure 1F).

We next compared the ligand-receptor interactions within primary tumor tissues, and observed differences in interaction number among major cell types between BC-Ctrl and BC-MDD groups (Figure 1G). In the BC-MDD group, the signal strength for interactions showed a significant reduction in epithelial cells, followed by fibroblasts and tumor cells, while signals related to T/NK cells exhibited a slight enhancement (Figure 1H). We then focused on the interaction changes within tumor cells, epithelial cells, and T/NK cells. Within T/NK cells, the C-X-C chemokine receptor 4 and ligand 12 (*CXCR4-CXCL12*) axis and macrophage migration inhibitory factor (MIF) signal were upregulated in the BC-MDD group (Supplementary Figure S6A-B). Meanwhile, we identified the enrichment in signals of midkine (MDK), laminin, and fibronectin (FN1) in the BC-Ctrl group (Supplementary Figure S6C-D). These results suggest altered breast cancer tumor signal interactions associated with a history of MDD.

In summary, our study offers preliminary insights regarding the impacts of MDD history in BC patients. We suggest a potential association between MDD history and a poorer prognosis in BC, characterized by a decrease in specific epithelial cells and impaired immune regulation signals within primary tumors. Furthermore, we propose potential alterations in the metabolic status of BC tumor cells in patients with MDD. Given the constraints of sample size and limited follow-up duration, larger patient cohorts, longitudinal data, and additional validation experiments are necessary to fully understand the complex interplay between MDD history and BC.

DECLARATIONS

AUTHOR CONTRIBUTIONS

Shaohua Hu, Peifen Fu, and Wei Chen conceived and designed the project. Lingling Wu, Yimeng Geng, Xingle Gao, Jianbo Lai, and Shaojia Lu performed psychological assessment and patient screening. Junwei Liu, Weiwei Yin, and Wei Chen analyzed single-cell data. Jianwen Fang, Minya Yao, and Peifen Fu collected patient samples. Junwei Liu, Lingling Wu, Yimeng Geng, and Jianwen Fang wrote the manuscript. All authors read and approved the final manuscript.

ACKNOWLEDGMENTS

We would like to express our gratitude to all the participants and the research team of the First Affiliated Hospital of Zhejiang University for their support.

CONFLICTS OF INTEREST STATEMENT

The authors declare that they have no competing interests.

FUNDING INFORMATION

This work was supported by funding from the Zhejiang Provincial Key Research and Development Program (No. 2021C03107 to S.H.), the Leading Talent of Scientific and Technological Innovation - "Ten Thousand Talents Program" of Zhejiang Province (No. 2021R52016 to S.H.), the National Natural Science Foundation (No. 82172770 to P.F.), and the Fundamental Research Funds for the Central Universities (226-2022-00193, 226-2022-00002).

DATA AVAILABILITY STATEMENT

All data needed to evaluate the conclusions in the paper are present in the paper and/or the Supplementary Materials.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The study was reviewed and approved by the First Affiliated Hospital, Zhejiang University School of Medicine (Approval number: IIT20220235B) and conducted in con-

formity with the International Ethical Guidelines for Research Involving Human Subjects as stated in the Helsinki Declaration. Informed written consent was obtained from all patients in this study.

Lingling Wu^{1,2}

Junwei Liu^{3,4}

Yimeng Geng¹

Jianwen Fang⁵

Xingle Gao¹

Jianbo Lai¹

Minya Yao⁵

Shaojia Lu¹

Weiwei Yin³

Peifen Fu⁵ 

Wei Chen^{3,6,7}

Shaohua Hu^{1,8,9,10,11} 

¹Department of Psychiatry, the First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, Zhejiang, P. R. China

²Department of Psychology, Children's Hospital, Zhejiang University School of Medicine, Hangzhou, Zhejiang, P. R. China

³Key Laboratory for Biomedical Engineering of the Ministry of Education, College of Biomedical Engineering and Instrument Science, Zhejiang University, Hangzhou, Zhejiang, P. R. China

⁴Guangzhou National Laboratory, Guangzhou, Guangdong, P. R. China

⁵Department of Breast Surgery, the First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, Zhejiang, P. R. China

⁶Department of Cardiology of The Second Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou, Zhejiang, P. R. China

⁷School of Basic Medical Science, Zhejiang University, Hangzhou, Zhejiang, P. R. China

⁸The Key Laboratory of Mental Disorder's Management in Zhejiang Province, Hangzhou, Zhejiang, P. R. China

⁹Brain Research Institute of Zhejiang University, Hangzhou, Zhejiang, P. R. China

¹⁰Zhejiang Engineering Center for Mathematical Mental Health, Hangzhou, Zhejiang, P. R. China

¹¹Department of Neurobiology, National Health Commission and Chinese Academy of Medical Sciences Key Laboratory of Medical Neurobiology, School of Brain Science and Brain Medicine, and Ministry of Education Frontier Science Center for Brain Science and Brain-machine Integration, Zhejiang University School of Medicine, Hangzhou, Hangzhou, Zhejiang, P. R. China

Correspondence

Shaohua Hu, Department of Psychiatry, the First
Affiliated Hospital, Zhejiang University School of
Medicine, Hangzhou, Zhejiang, P. R. China.
Email: dorhushaohua@zju.edu.cn

Wei Chen, Key Laboratory for Biomedical Engineering of
the Ministry of Education, College of Biomedical
Engineering and Instrument Science, Zhejiang
University, Hangzhou, Zhejiang, P. R. China.
Email: jackweichen@zju.edu.cn

Peifen Fu, Department of Breast Surgery, the First
Affiliated Hospital, Zhejiang University School of
Medicine, Hangzhou, Zhejiang, P. R. China.
Email: fupeifen@zju.edu.cn

ORCID

Peifen Fu  <https://orcid.org/0000-0002-2988-0502>

Shaohua Hu  <https://orcid.org/0000-0003-0570-670X>

REFERENCES

1. Evans DL, Charney DS, Lewis L, Golden RN, Gorman JM, Krishnan KR, et al. Mood disorders in the medically ill: scientific review and recommendations. *Biol Psychiatry*. 2005;58(3):175-89.
2. Obradovic MMS, Hamelin B, Manevski N, Couto JP, Sethi A, Coissieux MM, et al. Glucocorticoids promote breast cancer metastasis. *Nature*. 2019;567(7749):540-4.
3. Anderson G, Maes M. Oxidative/nitrosative stress and immunoinflammatory pathways in depression: treatment implications. *Curr Pharm Des*. 2014;20(23):3812-47.

4. He X-Y, Gao Y, Ng D, Michalopoulou E, George S, Adrover JM, et al. Chronic stress increases metastasis via neutrophil-mediated changes to the microenvironment. *Cancer Cell*. 2024;42(3):474-86.
5. Gao R, Bai S, Henderson YC, Lin Y, Schalck A, Yan Y, et al. Delineating copy number and clonal substructure in human tumors from single-cell transcriptomes. *Nat Biotechnol*. 2021;39(5):599-608.
6. Hanzelmann S, Castelo R, Guinney J. GSEA: gene set variation analysis for microarray and RNA-seq data. *BMC Bioinformatics*. 2013;14:7.
7. Wu SZ, Al-Eryani G, Roden DL, Junankar S, Harvey K, Andersson A, et al. A single-cell and spatially resolved atlas of human breast cancers. *Nat Genet*. 2021;53(9):1334-47.
8. Kumar T, Nee K, Wei R, He S, Nguyen QH, Bai S, et al. A spatially resolved single-cell genomic atlas of the adult human breast. *Nature*. 2023;620(7972):181-91.
9. Tang Z, Kang B, Li C, Chen T, Zhang Z. GEPIA2: an enhanced web server for large-scale expression profiling and interactive analysis. *Nucleic Acids Res*. 2019;47(W1):W556-W60.
10. Buchwalter G, Hickey MM, Cromer A, Selfors LM, Gunawardane RN, Frishman J, et al. PDEF promotes luminal differentiation and acts as a survival factor for ER-positive breast cancer cells. *Cancer cell*. 2013;23(6):753-67.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.