

Chimeric antigen receptor T-cell therapy following autologous transplantation for secondary central nervous system lymphoma

A case report

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Abstract

Rationale: Chimeric antigen receptor (CAR) T-cell therapy is effective in treating relapsed and refractory B-cell non-Hodgkin lymphoma. However, because of the mortality risk associated with immune effector cell-associated neurotoxicity syndrome and pseudoprogression, patients with central nervous system (CNS) involvement are less likely to receive CAR T-cell therapy.

Patients concerns: We report a case of a 61-year-old, male patient with intravascular large B-cell lymphoma who suffered a CNS relapse after standard chemotherapy.

Diagnosis: A diagnosis of intravascular large B-cell lymphoma with CNS involvement was made.

Interventions: We treated the patient using CAR T-cell therapy following a conditioning regimen consisting of thiotepa and busulfan and autologous stem cell transplantation. Although he experienced grade 1 cytokine release syndrome, no other serious adverse events, such as immune effector cell-associated neurotoxicity syndrome or pseudoprogression, were observed.

Outcomes: The patient achieved complete remission after the CAR T-cell infusion.

Lessons: CAR T-cell therapy following autologous stem cell transplantation is a viable option for relapsed/refractory lymphoma with CNS infiltration. Further clinical studies are warranted to verify its safety and efficacy.

Abbreviations: ASCT = autologous stem cell transplantation, BuTT = thiotepa and busulfan, CAR = chimeric antigen receptor, CNS = central nervous system, CR = complete remission, CRS = cytokine release syndrome, DLBCL = diffuse large B-cell lymphoma, HDC = high-dose chemotherapy, HD-MTX = high-dose methotrexate, ICANS = immune effector cell-associated neurotoxicity syndrome, IVLBCL = intravascular large B-cell lymphoma, MRI = magnetic resonance imaging, R/R = relapsed/ refractory, WBRT = whole brain radiotherapy.

Keywords: autologous stem cell transplantation, case report, central nervous system relapse, chimeric antigen receptor T-cell therapy, intravascular large B-cell lymphoma

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1. Introduction

The therapeutic efficacy of chimeric antigen receptor (CAR) Tcell therapy has revolutionized the treatment of relapsed/ refractory (R/R) B-cell non-Hodgkin lymphoma.^[1] However, CAR T-cell therapy can have potentially severe side effects, such as cytokine release syndrome (CRS) and CAR T-cell therapyrelated immune effector cell-associated neurotoxicity syndrome (ICANS).^[2] The incidence of CRS and ICANS in patients with hematological malignancies is significantly higher than in those with other solid malignancies.^[3] Due to the mortality risk associated with ICANS, patients with central nervous system (CNS) involvement are excluded from almost all clinical trials of CAR T-cell therapy. Although there are some reports of the application of CD19-targeted CAR T-cells in CNS lymphoma treatment,^[4-6] there are no reports of an increased incidence of ICANS in patients with CNS lymphoma so far. The efficacy and safety of CAR T-cell therapy in patients with CNS involvement have not been established yet due to the rarity of these patients.

Herein, we reported a case of a CNS relapse of intravascular large B-cell lymphoma (IVLBCL) successfully treated with autologous stem cell transplantation (ASCT) followed by CAR T-cell therapy. This study was approved by the Ethics Committee of Tokyo Metropolitan Cancer and Infectious Diseases Center, Komagome Hospital. Written informed consent was obtained from the patient for publication of this case report.

2. Case report

A previously healthy, 61-year-old, male patient presented with fever and dyspnea. On admission, a blood test revealed a marked elevation of lactate dehydrogenase (1450 U/L normal: 106–211 U/L) and interleukin 2 receptor (8800 U/mL normal: 122–496 U/ mL). Computed tomography revealed consolidation in the upper lobe. IVLBCL was diagnosed based on an analysis of bone marrow, lung biopsy, and skin biopsy specimens. Brain magnetic

resonance imaging (MRI) revealed no abnormalities, and the patient was not considered to have lymphoma with CNS involvement. He achieved complete remission (CR) after 6 cycles of cyclophosphamide, doxorubicin, vincristine, and prednisone and 8 cycles of rituximab plus 2 cycles of high-dose methotrexate (HD-MTX). One month later, he experienced a CNS relapse with brain lesions detected by positron emission tomography and computed tomography. MRI revealed a $25 \text{ mm} \times 20 \text{ mm} \times 34$ mm mass in the right basal ganglia and other smaller lesions in the brain (Fig. 1). No lymphoma cells were found in cerebrospinal fluid, nor were any lesions detected outside the CNS. Owing to the difficulty of curing a CNS relapse by salvage chemotherapy even with ASCT, the addition of CAR T-cell therapy after ASCT was planned. He underwent 3 cycles of HD-MTX and rituximab followed by 3 cycles of HD-MTX/cytarabine. Lymphocyte apheresis for CAR T-cell manufacturing was performed during the salvage chemotherapy. Subsequent MRI showed a partial response. A peripheral blood stem cell harvest was performed, and the patient received a conditioning regimen consisting of BuTT (thiotepa 5 mg/kg on days -7 to -6, busulfan 3.2 mg/kg on days -5 to -4) and ASCT on day 0. Although MRI after the ASCT was unable to detect the residual lesions, the patient was still considered to have residual CNS disease. He received lymphodepleting chemotherapy consisting of fludarabine 25 mg/m²/day and cyclophosphamide 250 mg/m²/day on days -7 to -5 and an infusion of anti-CD19 CAR T-cells (tisagenlecleucel) on day 0. Four hours after the CAR T-cell infusion, a fever, which failed to respond to broad-spectrum antibiotics or supportive treatment, developed without hypotension or hypoxia, leading to the diagnosis of grade 1 CRS. Although the patient was treated with tocilizumab 8 mg/kg/day on days 2 and 3, the symptoms continued. Finally, he was given methylprednisolone 2 mg/kg/ day from day 3, and his temperature normalized by day 5. Thereafter the methylprednisolone was tapered, then stopped on day 8. No neurotoxic events were observed during the follow-up

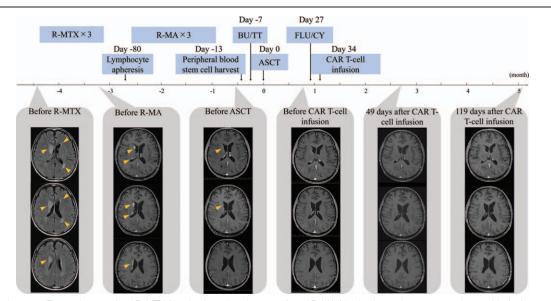


Figure 1. Clinical course: The patient received R-MTX (rituximab and methotrexate) and R-MA (rituximab, methotrexate, and cytarabine) after a central nervous system (CNS) relapse. He received BuTT (busulfan and thiotepa) as the conditioning regimen followed by ASCT (autologous stem cell transplantation). Posttransplantation, he received lymphodepleting chemotherapy consisting of FLU/CY (fludarabine and cyclophosphamide) and an infusion of anti-CD19 CAR (chimeric antigen receptor) T-cells. Brain MRI after the CAR-T cell infusion demonstrated resolution of the CNS lesions, indicating a complete response. The arrowheads show the presence of lymphoma lesions in the brain. MRI = magnetic resonance imaging.

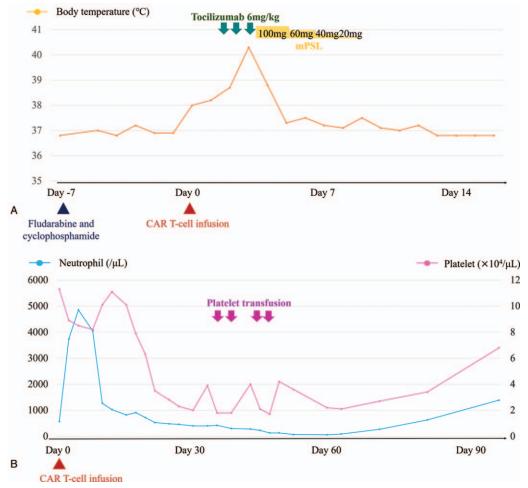


Figure 2. Clinical course after CAR T-cell infusion: Fever \geq 38°C without hypotension or hypoxia developed after CAR (chimeric antigen receptor) T-cell infusion, corresponding to grade 1 cytokine release syndrome. The patient was treated with tocilizumab and methylprednisolone (mPSL) (A). Grade 4 neutropenia and thrombocytopenia developed after CAR T-cell infusion (B).

period. Grade 4 neutropenia developed from day 23 and improved by day 78. No serious infections were observed. Grade 4 thrombocytopenia developed from day 36 and improved by day 47 (Fig. 2). One month after the CAR T-cell infusion, brain MRI showed a complete absence of brain lesions, indicating a CR (Fig. 1). Eight months after the CAR T-cell infusion, the patient remains in CR.

3. Discussion

The patient in the present study had R/R B-cell non-Hodgkin lymphoma with a CNS relapse and failed to achieve a CR with salvage chemotherapy, high-dose chemotherapy (HDC) or ASCT. After CAR T-cell therapy, he finally achieved durable CR without any severe adverse events, such as ICANS. To the best of our knowledge, the present report is the first to demonstrate the safety and efficacy of CAR T-cell therapy following ASCT in patients with CNS involvement.

Previous studies reported the risk of CNS relapse in diffuse large B-cell lymphoma (DLBCL) to be approximately 5%.^[7–9] IVLBCL is characterized by clinically aggressive behavior and a high rate of CNS involvement at diagnosis. Several, previous studies have examined IVLBCL patients with a CNS relapse, and a retrospective study reported that the risk of CNS recurrence at 3 years was 25% among 82 patients without CNS involvement at the initial diagnosis.^[10–13]

The incidence of CNS involvement secondary to DLBCL is considerably higher in patients with certain, high-risk, clinical features at diagnosis. Multivariate risk models, such as CNS-IPI, are effective in predicting the risk of CNS recurrence.^[14] Identifying patients with a high risk of CNS relapse allows us to offer appropriate prophylaxis during the first line therapy. Intravenous and intrathecal prophylaxis is most frequently used, but the evidence is equivocal as to its effectiveness.^[15] Intravenous HD-MTX can be safely administered in combination with standard chemoimmunotherapy and decreases the risk of CNS recurrence in patients with high-risk DLBCL. However, there is currently no standard form of CNS-prophylaxis.^[15]

The prognosis of patients with a CNS relapse is still poor, and there is as of yet no consensus on the optimal salvage treatment. The median overall survival after CNS relapse is reportedly about 7 months.^[16,17] The data supporting the efficacy of HDC and ASCT in patients with DLBCL with secondary CNS lymphoma are limited although some studies have shown favorable survival outcomes (the median DFS and overall survival for patients with CR at ASCT was 36 and 33 months, respectively).^[18–21] The

optimal HDC regimen for secondary CNS lymphoma is also unknown. Conditioning regimens containing agents that can effectively penetrate the blood-brain barrier, such as BuTT, are reportedly associated with promising disease control for relapsed CNS lymphoma. Some retrospective analyses reported an association between the thiotepa/busulfan-based conditioning therapy with ASCT for secondary CNS lymphoma and progression-free survival at years 2 and 3 of 76.1% to 93%, respectively.^[18,19] Despite the use of the HDC regimen and ASCT in the treatment of CNS lesions, a complete cure for secondary CNS lymphoma is still elusive. CAR T-cell therapy may therefore augment these therapies to improve the possibility of CR.

The occurrence of lethal neurotoxicity after CAR T-cell therapy prevents most patients with a CNS lesion from receiving this treatment. Abramson et al^[4] reported the first case of CNS lymphoma treated with CAR T-cell therapy, which induced CR without ICANS. A retrospective analysis of tisagenlecleucel for the treatment of R/R B-cell non-Hodgkin lymphoma with secondary CNS involvement showed that none of the patients experienced CAR T-cell-related neurotoxicity.^[5] In addition, the rate of ICNAS did not increase in 5 patients treated with axicabtagene ciloleucel for R/R NHL with secondary CNS involvement.^[22]

The pathogenesis of ICANS is not completely understood but may be related to cytokine-mediated toxicity resulting from endothelial dysfunction, increased blood-brain barrier permeability, and failure to protect cerebrospinal fluid from high concentrations of systemic cytokines.^[23] The most significant risk factor of ICANS is a history of severe CRS. A high tumor burden is also associated with ICANS development.^[24,25] In the present case, ICNAS did not occur after CAR T-cell therapy partly because the ASCT after BuTT was able to reduce the tumor burden sufficiently.

Pseudoprogression, a well-defined phenomenon in other immunotherapies for solid tumors, is characterized by a temporary enlargement of the tumors due to immune cell infiltration and is also observed during CAR T-cell therapy. However, this enlargement can cause local compression,^[26] and CAR T-cell therapy in the presence of substantial residual disease can be fatal in cases of CNS invasion. Combined with the fact that a durable response to CAR T-cell therapy is associated with a low baseline tumor burden,^[27] reducing the tumor burden as much as possible before CAR T-cell therapy, especially in cases of CNS involvement, is crucial.

HDC and ASCT are the current standard of care for R/R DLBCL. However, relapse rates are still high. Some reports demonstrated the feasibility of CAR T-cell therapy a couple of days after ASCT for R/R B-cell non-Hodgkin lymphoma, [28-30] unlike the present case, in which CAR T-cell therapy was performed about 1 month after ASCT. The tumor burden decreases as a result of myeloablative conditioning, and the immunosuppressive microenvironment is attenuated after ASCT. Consecutive administration of ASCT and CAR-T cells eradicates the residual disease and reduces the recurrence rate. In fact, CAR T-cell therapy following ASCT exhibited a higher rate of CR in patients with R/R large B-cell lymphoma than CAR T-cell therapy alone.^[28] However, the therapy was associated with a high incidence of reversible neurotoxicity, with 10 of 15 patients in a phase I clinical study experiencing grade 3 to 4 neurotoxicity, which may have been caused by the high-dose conditioning and pegfilgrastim administered immediately before the CAR T-cell infusion.^[29] We believe that CAR T-cell therapy after an interval following ASCT can increase the response rate by reducing the tumor volume with less neurotoxicity.

Whole brain radiotherapy (WBRT) for CNS lymphoma prolongs progression-free survival.^[31] However, the major impediment to using WBRT is the high risk of late-delayed neurotoxicity, which manifests as progressive leukoencephalopathy with cognitive deterioration often leading to severe dementia and death. In an attempt to minimize neurotoxicity, WBRT is often avoided altogether in the treatment of patients with lymphoma with CNS involvement.^[32] CAR T-cell therapy following ASCT may be a viable alternative to WBRT in the treatment of CNS lymphoma.

4. Conclusion

CAR T-cell therapy was safely administered after ASCT to a patient with secondary CNS lymphoma. CAR T-cell therapy following ASCT is a promising treatment for patients with lymphoma with CNS involvement. Vigilant follow-up of the present patient and further studies are needed to determine whether CAR T-cell cell therapy following ASCT is indeed a viable therapeutic option for R/R B-cell non-Hodgkin lymphoma with CNS infiltration.

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References

- Neelapu SS, Locke FL, Bartlett NL, et al. Axicabtagene ciloleucel CAR Tcell therapy in refractory large B-cell lymphoma. N Engl J Med 2017;377:2531–44.
- [2] Schuster SJ, Bishop MR, Tam CS, et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. N Engl J Med 2019;380: 45–56.
- [3] Yu WL, Hua ZC. Chimeric antigen receptor T-cell (CAR T) therapy for hematologic and solid malignancies: efficacy and safety-a systematic review with meta-analysis. Cancers (Basel) 2019;11:47doi:10.3390/ cancers11010047.
- [4] Abramson JS, McGree B, Noyes S, et al. Anti-CD19 CAR T cells in CNS diffuse large-B-cell lymphoma. N Engl J Med 2017;377:783–4.
- [5] Frigault MJ, Dietrich J, Martinez-Lage M, et al. Tisagenlecleucel CAR Tcell therapy in secondary CNS lymphoma. Blood 2019;134:860–6.
- [6] Tu S, Zhou X, Guo Z, et al. CD19 and CD70 dual-target chimeric antigen receptor T-cell therapy for the treatment of relapsed and refractory primary central nervous system diffuse large B-cell lymphoma. Front Oncol 2019;9:1350doi: 10.3389/fonc.2019.01350.
- [7] Hollender A, Kvaloy S, Nome O, Skovlund E, Lote K, Holte H. Central nervous system involvement following diagnosis of non-Hodgkin's lymphoma: a risk model. Ann Oncol 2002;13:1099–107.

- [8] Kanemasa Y, Shimoyama T, Sasaki Y, et al. Central nervous system relapse in patients with diffuse large B cell lymphoma: analysis of the risk factors and proposal of a new prognostic model. Ann Hematol 2016;95:1661–9.
- [9] Shimazu Y, Notohara K, Ueda Y. Diffuse large B-cell lymphoma with central nervous system relapse: prognosis and risk factors according to retrospective analysis from a single-center experience. Int J Hematol 2009;89:577–83.
- [10] Anghel G, Petrinato G, Severino A, et al. Intravascular B-cell lymphoma: report of two cases with different clinical presentation but rapid central nervous system involvement. Leuk Lymphoma 2003;44:1353–9.
- [11] Imai H, Kajimoto K, Taniwaki M, et al. Intravascular large B-cell lymphoma presenting with mass lesions in the central nervous system: a report of five cases. Pathol Int 2004;54:231–6.
- [12] Ohno T, Sakamoto T, Mizumoto C, et al. Leukemic and meningeal relapse of CD5+ intravascular large B-cell lymphoma with downmodulation of CD20 after rituximab therapy. Int J Hematol 2006;84:74–8.
- [13] Shimada K, Murase T, Matsue K, et al. Central nervous system involvement in intravascular large B-cell lymphoma: a retrospective analysis of 109 patients. Cancer Sci 2010;101:1480–6.
- [14] Schmitz N, Zeynalova S, Nickelsen M, et al. CNS international prognostic index: a risk model for CNS relapse in patients with diffuse large B-cell lymphoma treated with R-CHOP. J Clin Oncol 2016; 34:3150–6.
- [15] Abramson JS, Hellmann M, Barnes JA, et al. Intravenous methotrexate as central nervous system (CNS) prophylaxis is associated with a low risk of CNS recurrence in high-risk patients with diffuse large B-cell lymphoma. Cancer 2010;116:4283–90.
- [16] Bromberg JE, Doorduijn JK, Illerhaus G, et al. Central nervous system recurrence of systemic lymphoma in the era of stem cell transplantationan International Primary Central Nervous System Lymphoma Study Group project. Haematologica 2013;98:808–13.
- [17] Lee MY, Kim HS, Lee JY, et al. Efficacy and feasibility of autologous stem cell transplantation in patients with diffuse large B-cell lymphoma with secondary central nervous system involvement. Int J Hematol 2015;102:678–88.
- [18] Oh DH, Chua N, Street L, Stewart DA. Treatment of patients with secondary central nervous system lymphoma with high-dose busulfan/ thiotepa-based conditioning and autologous stem cell transplant. Leuk Lymphoma 2016;57:28–33.
- [19] Qualls D, Sullivan A, Li S, et al. High-dose thiotepa, busulfan, cyclophosphamide, and autologous stem cell transplantation as upfront consolidation for systemic non-Hodgkin lymphoma with synchronous central nervous system involvement. Clin Lymphoma Myeloma Leuk 2017;17:884–8.

- [20] Welch MR, Sauter CS, Matasar MJ, et al. Autologous stem cell transplant in recurrent or refractory primary or secondary central nervous system lymphoma using thiotepa, busulfan and cyclophosphamide. Leuk Lymphoma 2015;56:361–7.
- [21] Patil S, Spencer A, Schwarer A, et al. Disease status at autologous stem cell transplantation and the cell of origin phenotype are important predictors of outcome in patients with neurologic (central nervous system) relapse of diffuse large B-cell lymphoma undergoing autologous stem cell transplantation. Leuk Lymphoma 2009;50:1964–8.
- [22] Ghafouri S, Timmerman J, Larson S, Mead MD. Axicabtagene Ciloleucel CAR T-cell therapy for relapsed/refractory secondary CNS non-Hodgkin lymphoma: comparable outcomes and toxicities, but shorter remissions may warrant alternative consolidative strategies? Bone Marrow Transplant 2020;56:974–7.
- [23] Gust J, Hay KA, Hanafi LA, et al. Endothelial activation and blood-brain barrier disruption in neurotoxicity after adoptive immunotherapy with CD19 CAR-T cells. Cancer Discov 2017;7:1404–19.
- [24] Freyer CW, Porter DL. Cytokine release syndrome and neurotoxicity following CAR T-cell therapy for hematologic malignancies. J Allergy Clin Immunol 2020;146:940–8.
- [25] Rice J, Nagle S, Randall J, Hinson HE. Chimeric antigen receptor T cellrelated neurotoxicity: mechanisms, clinical presentation, and approach to treatment. Curr Treat Options Neurol 2019;21:40doi: 10.1007/ s11940-019-0580-3.
- [26] Wang J, Hu Y, Yang S, et al. Role of fluorodeoxyglucose positron emission tomography/computed tomography in predicting the adverse effects of chimeric antigen receptor T cell therapy in patients with non-Hodgkin lymphoma. Biol Blood Marrow Transplant 2019;25:1092–8.
- [27] Locke FL, Rossi JM, Neelapu SS, et al. Tumor burden, inflammation, and product attributes determine outcomes of axicabtagene ciloleucel in large B-cell lymphoma. Blood Adv 2020;4:4898–911.
- [28] Liu W, Huang W, Lv R, et al. Anti-CD19 CAR T therapy following autologous HSCT may be safe and effective in patients with refractory large B-cell lymphoma. Blood 2019;134(Suppl 1):784–1784.
- [29] Sauter CS, Senechal B, Rivière I, et al. CD19 CAR T cells following autologous transplantation in poor-risk relapsed and refractory B-cell non-Hodgkin lymphoma. Blood 2019;134:626–35.
- [30] Wang X, Popplewell LL, Wagner JR, et al. Phase 1 studies of central memory-derived CD19 CAR T-cell therapy following autologous HSCT in patients with B-cell NHL. Blood 2016;127:2980–90.
- [31] Omuro A, Taillandier L, Chinot O, et al. Primary CNS lymphoma in patients younger than 60: can whole-brain radiotherapy be deferred? J Neurooncol 2011;104:323–30.
- [32] Omuro AM, Ben-Porat LS, Panageas KS, et al. Delayed neurotoxicity in primary central nervous system lymphoma. Arch Neurol 2005;62: 1595–600.