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### Case Report

# Chronic Myeloid Leukemia with e19a2 BCR-ABL1 Transcripts and Marked Thrombocytosis: The Role of Molecular Monitoring

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While most patients with chronic myeloid leukemia (CML) express either e13a2 or e14a2 *BCR-ABL1* transcripts, a significant minority expresses variant transcripts, of which e19a2 is the most common. Although considered to have a relatively favourable outcome, reported responses to tyrosine kinase inhibitor (TKI) therapy are variable with molecular monitoring in CML patients with e19a2 *BCR-ABL1* transcripts rarely reported. A case of e19a2 *BCR-ABL1* CML with marked thrombocytosis is described in which the value of molecular monitoring is emphasised during treatment interruptions, dose reductions, and changes. This case serves to demonstrate the requirement for prospective real-time quantitative PCR (RQ-PCR) assays for patients with variant *BCR-ABL1* transcript types and standardisation of such assays to enable modern patient management.

#### 1. Introduction

The BCR-ABL1 fusion gene is the molecular hallmark and causative event of CML. Most CML patients express either e13a2 or e14a2 BCR-ABL1 fusion transcripts but approximately 5% of patients express variant transcripts that may involve fusion of alternative exons, insertions, or breakpoints within exons. Of these variant BCR-ABL1 fusions, the e19a2 is the most common with approximately 50 cases reported to date. CML with e19a2 BCR-ABL1, that encodes a 230 kDa protein [1], was initially reported in neutrophilic CML with a relatively indolent clinical course [2] but has subsequently been reported in typical CML presenting in all phases [3]. While TKI therapy is considered the optimal frontline treatment for CML with quantitative molecular responses predictive of overall and progression-free survival [4], review of the literature indicates that hematological and cytogenetic responses to TKIs are variable in e19a2 BCR-ABL1 CML [3, 5–11], although this may be biased by reporting of cases with atypical features. Molecular responses have rarely been reported with real-time quantitative polymerase chain reaction (RQ-PCR) monitoring thus far documented in only two patients [12, 13], suggesting that this important element of patient management may be forsaken in a significant proportion of patients that express this and other variant transcripts types. Although an elevated platelet count is observed at presentation in a number of CML patients, marked thrombocytosis (platelets >  $1000 \times 10^9/L$ ) is uncommon and often associated with the e19a2 *BCR-ABL1* transcript type [14]. The diagnosis, course, and molecular monitoring of a patient with e19a2 *BCR-ABL1* CML who presented with a marked thrombocytosis is described.

#### 2. Case Report

An eighty-year-old male with a history of Duke's C colon carcinoma in remission presented with an abnormal full

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blood count: Hb 9.9 g/dL, PLT  $1401 \times 10^9$ /L, WBC  $31.7 \times$  $10^9$ /L which comprised neutrophils  $25.2 \times 10^9$ /L, lymphocytes  $2.1 \times 10^9$ /L, monocytes  $0.94 \times 10^9$ /L, eosinophils  $1.75 \times 10^{9}$ /L, basophils  $0.85 \times 10^{9}$ /L, and blasts  $0.1 \times 10^{9}$ /L. The blood film showed thrombocytosis, marked platelet anisocytosis, neutrophilia with left shift, and rouleaux with the patient having an elevated lactate dehydrogenase of 554 IU/L. The bone marrow aspirate was hypercellular with particles present and showed significant myeloid hyperplasia and plentiful megakaryocytes, often present in groups with polymorphic morphology including hypolobulated and mononuclear forms. Cytogenetic studies were not performed but a standard reverse-transcriptase polymerase chain reaction methodology for detection of BCR-ABL1 transcripts [15] showed a single band that was not consistent with the common e13a2 or e14a2 BCR-ABL1 transcripts. Direct sequencing demonstrated the presence of an e19a2 BCR-ABL1 fusion with a final diagnosis of chronic phase CML with a high-risk Sokal score of 3.15.

The patient commenced on imatinib 400 mg od with aspirin 75 mg once daily. After three weeks of therapy, when the platelet count was still greater than  $1000 \times 10^9$ /L, a deep vein thrombosis (DVT) of the left popliteal vein extending into the common femoral vein was diagnosed prompting treatment with low molecular weight heparin. Complete cytogenetic remission was achieved (0/50 Ph+ metaphases) after three months of imatinib therapy with RQ-PCR for e19a2 BCR-ABL1 transcripts performed as previously described [13] demonstrating an initial decrease in BCR-ABL1 transcript level (BCR-ABL1/ABL1 3.8%, Figure 1). Over the following months the patient complained of intermittent abdominal pain and diarrhoea and because of these symptoms, imatinib was initially reduced to 300 mg od with several further treatment interruptions due to the abdominal symptoms and recurrent rash. The patient subsequently underwent gastroenterology investigations and was diagnosed with intermittent small bowel obstruction secondary to adhesions. He underwent a laparotomy but perioperatively had recurrence of DVT and was again fully anticoagulated. As gastrointestinal symptoms persisted after surgery, imatinib was stopped reflected by a rise in BCR-ABL1 transcripts to high levels (Figure 1). Treatment with nilotinib 300 mg bd was instigated but halted after three weeks due to recurrence of both diarrhoea and a rash. Nine months from initial diagnosis the patient was restarted with the best tolerated therapy of imatinib 300 mg od. He continued to have significant symptoms including intermittent diarrhoea and abdominal pains with adherence questionable. The patient was switched to dasatinib 50 mg twice daily 18 months after diagnosis achieving a partial cytogenetic remission at 21 and 24 months (6/50 Ph+ and 12/50 Ph+, resp.) with only a slight molecular response (Figure 1). He remains in hematological remission with relatively improved gastrointestinal symptoms and adherence but at 30 months has worsening cytogenetic and molecular responses: 42/50 Ph+ and clonal evolution in the form of an extra der(22) or an isochromosome of the der(22) and BCR-ABL1/ABL1 100%.

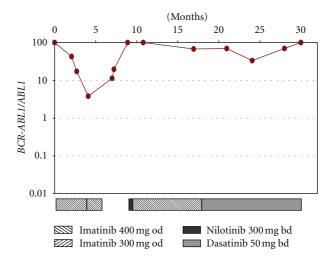


FIGURE 1: RQ-PCR monitoring of e19a2 BCR-ABL1 transcripts.

#### 3. Discussion

Thrombocytosis is a relatively common presenting feature of CML but platelet counts  $>1000 \times 10^9/L$  are rare. An association with e19a2 *BCR-ABL1* transcripts in CML patients and marked thrombocytosis has previously been documented [14], corroborated by the characterisation of a distinctive high platelet count in a transgenic mouse model expressing p230 BCR-ABL1 [16], and supported by the findings in the patient described herein. As to whether the significant thrombocytosis contributed to the DVT in this patient remains unknown: a prompt targeted reduction in platelets may be warranted in such cases to reduce the risk of thrombosis.

E19a2 *BCR-ABL1* is the most common variant transcript type in CML, yet molecular responses to TKI have rarely been evaluated, most likely to the unavailability of commercial plasmids necessary for construction of standard curves. While efforts to harmonise molecular methodologies that quantify e13a2 and e14a2 *BCR-ABL1* transcripts are underway [17], attainment of milestones of TKI therapy below complete cytogenetic remission such as major molecular remission [18] or complete molecular remission cannot be reproducibly assessed in such cases. Effective consensus or standardisation of these RQ-PCR assays, previously performed on only a case per case basis, is required.

This case serves to highlight the requirement for RQ-PCR monitoring, an essential component of modern disease management, for the less common *BCR-ABL1* transcript types during TKI dose reductions, interruptions, or changes in conjunction with conventional cytogenetic analysis.

#### References

[1] G. Saglio, A. Guerrasio, C. Rosso et al., "New type of Bcr/Abl junction in Philadelphia chromosome—positive chronic myelogenous leukemia," *Blood*, vol. 76, no. 9, pp. 1819–1824, 1990.

- [2] F. Pane, F. Frigeri, M. Sindona et al., "Neutrophilic-chronic myeloid leukemia: a distinct disease with a specific molecular marker (BCR/ABL with C3/A2 junction)," *Blood*, vol. 88, no. 7, pp. 2410–2414, 1996.
- [3] B. C. Mondal, S. Majumdar, U. B. Dasgupta, U. Chaudhuri, P. Chakrabarti, and S. Bhattacharyya, "e19a2 BCR-ABL fusion transcript in typical chronic myeloid leukaemia: a report of two cases," *Journal of Clinical Pathology*, vol. 59, no. 10, pp. 1102–1103, 2006.
- [4] T. P. Hughes, A. Hochhaus, S. Branford et al., "Long-term prognostic significance of early molecular response to imatinib in newly diagnosed chronic myeloid leukemia: an analysis from the International Randomized Study of Interferon and STI571 (IRIS)," *Blood*, vol. 116, no. 19, pp. 3758–3765, 2010
- [5] J. J. Lee, H. J. Kim, Y. J. Kim et al., "Imatinib induces a cytogenetic response in blast crisis or interferon failure chronic myeloid leukemia patients with e19a2 BCR-ABL transcripts," *Leukemia*, vol. 18, no. 9, pp. 1539–1540, 2004.
- [6] X. Li, J. Yang, X. Chen et al., "A report of early cytogenetic response to imatinib in two patients with chronic myeloid leukemia at accelerated phase and carrying the e19a2 BCR-ABL transcript," Cancer Genetics and Cytogenetics, vol. 176, no. 2, pp. 166–168, 2007.
- [7] H. Andrikovics, S. Nahajevszky, A. Szilvási et al., "First and second line imatinib treatment in chronic myelogenous leukemia patients expressing rare e1a2 or e19a2 BCR-ABL transcripts," *Hematological Oncology*, vol. 25, no. 3, pp. 143– 147, 2007.
- [8] C. Popovici, A. Charbonnier, O. Gisserot et al., "Y253H mutation appearing in a μ-BCR-ABL (e19a2) CML," *Leukemia Research*, vol. 32, no. 2, pp. 361–362, 2008.
- [9] G. Oshikawa, T. Kurosu, A. Arai, N. Murakami, and O. Miura, "Clonal evolution with double Ph followed by tetraploidy in imatinib-treated chronic myeloid leukemia with e19a2 transcript in transformation," *Cancer Genetics and Cytogenetics*, vol. 199, no. 1, pp. 56–61, 2010.
- [10] S. E. Martin, M. Sausen, A. Joseph, and B. F. Kingham, "Chronic myeloid leukemia with e19a2 atypical transcript: early imatinib resistance and complete response to dasatinib," *Cancer Genetics and Cytogenetics*, vol. 201, no. 2, pp. 133–134, 2010.
- [11] A. Bennour, N. Beaufils, H. Sennana, B. Meddeb, A. Saad, and J. Gabert, "E355G mutation appearing in a patient with e19a2 chronic myeloid leukaemia resistant to imatinib," *Journal of Clinical Pathology*, vol. 63, no. 8, pp. 737–740, 2010.
- [12] M. Cea, G. Cirmena, A. Garuti et al., "A T315I mutation in e19a2 BCR/ABL1 chronic myeloid leukemia responding to dasatinib," *Leukemia Research*, vol. 34, no. 9, pp. e240–e242, 2010
- [13] S. E. Langabeer, S. L. McCarron, P. Carroll, J. Kelly, M. O'Dwyer, and E. Conneally, "Molecular response to first line nilotinib in a patient with e19a2 BCR-ABL1 chronic myeloid leukemia," *Leukemia Research*, vol. 35, no. 9, pp. e169–e170, 2011.
- [14] T. Yamagata, K. Mitani, Y. Kanda, Y. Yazaki, and H. Hirai, "Elevated platelet count features the variant type of BCR/ABL junction in chronic myelogenous leukaemia," *British Journal of Haematology*, vol. 94, no. 2, pp. 370–372, 1996.
- [15] J. J. M. Van Dongen, E. A. Macintyre, J. A. Gabert et al., "Standardized RT-PCR analysis of fusion gene transcripts from chromosome aberrations in acute leukemia for detection of minimal residual disease. Report of the BIOMED-1 Concerted

- Action: investigation of minimal residual disease in acute leukemia," *Leukemia*, vol. 13, no. 12, pp. 1901–1928, 1999.
- [16] K. Inokuchi, K. Dan, M. Takatori et al., "Myeloproliferative disease in transgenic mice expressing P230 Bcr/Abl: longer disease latency, thrombocytosis, and mild leukocytosis," *Blood*, vol. 102, no. 1, pp. 320–323, 2003.
- [17] S. Branford, L. Fletcher, N. C. P. Cross et al., "Desirable performance characteristics for BCR-ABL measurement on an international reporting scale to allow consistent interpretation of individual patient response and comparison of response rates between clinical trials," Blood, vol. 112, no. 8, pp. 3330– 3338, 2008.
- [18] M. Baccarani, J. Cortes, F. Pane et al., "Chronic myeloid leukemia: an update of concepts and management recommendations of European LeukemiaNet," *Journal of Clinical Oncology*, vol. 27, no. 35, pp. 6041–6051, 2009.