



Immune responses to bioengineered organs

Jordi Ochando^a, Dominique Charron^{b,c}, Pedro M. Baptista^{d,e},
and Basak E. Uygun^f

Purpose of review

Organ donation in the United States registered 9079 deceased organ donors in 2015. This high percentage of donations allowed organ transplantation in 29 851 recipients. Despite increasing numbers of transplants performed in comparison with previous years, the numbers of patients that are in need for a transplant increase every year at a higher rate. This reveals that the discrepancy between the demand and availability of organs remains fundamental problem in organ transplantation.

Recent findings

Development of bioengineered organs represents a promising approach to increase the pool of organs for transplantation. The technology involves obtaining complex three-dimensional scaffolds that support cellular activity and functional remodeling through tissue recellularization protocols using progenitor cells. This innovative approach integrates cross-thematic approaches from specific areas of transplant immunology, tissue engineering and stem cell biology, to potentially manufacture an unlimited source of donor organs for transplantation.

Summary

Although bioengineered organs are thought to escape immune recognition, the potential immune reactivity toward each of its components has not been studied in detail. Here, we summarize the host immune response toward different progenitor cells and discuss the potential implications of using nonself biological scaffolds to develop bioengineered organs.

Keywords

allogeneicity, bioengineered organs, extracellular matrix, stem cells

INTRODUCTION

Organ donation in the United States registered 9079 deceased organ donors in 2015 according to the 'Newsletter Transplant', (September 2016, Vol. 21). This high percentage of donations allowed organ transplantation in 29 851 recipients: 24 225 kidney, 7127 liver, 2819 heart, 2072 lung, 947 pancreas and 141 small bowel. Despite increasing numbers of transplants performed in comparison with previous years (2014 = 28 523 transplant recipients), the number of patients that are in need for a transplant increases every year at a higher rate, and more than 11 900 people in the United States are currently on the waiting list for a life saving organ (<https://www.unos.org/>). As a result, about 13 000 people in the United States alone die or become sick every year while waiting for a transplantable organ (<https://optn.transplant.hrsa.gov/>). In addition, 22 people die every day on average from the lack of available organs for transplant (<https://www.donatelife.net/>), which reveals a fundamental problem in organ transplantation: discrepancy between the demand and availability of organs.

This situation has been the major driving force behind the rise of organ engineering approach to create transplantable grafts that may potentially be used as alternatives to donor organs. In organ engineering, human cadaveric or animal organs are

^aDepartment of Oncological Sciences, Icahn School of Medicine at Mount Sinai, New York, New York, USA, ^bHLA&MEDECINE 'Jean Dausset' Laboratory Network and Transplantex, Hôpital Saint-Louis, Inserm U 976 Université Paris-Diderot, Paris, France, ^cJiao Tong University, Shanghai, China, ^dInstituto de Investigacion Sanitaria de Aragon (IIS Aragon), ^eCIBERehd, Zaragoza, Spain and ^fDepartment of Surgery, Center for Engineering in Medicine, Massachusetts General Hospital, Harvard Medical School, The Shriners Hospitals for Children, Boston, Massachusetts, USA

Correspondence to Jordi Ochando, Department of Oncological Sciences, Icahn School of Medicine at Mount Sinai, New York, United States. E-mail: jordi.ochando@mssm.edu

Curr Opin Organ Transplant 2017, 22:79–85

DOI:10.1097/MOT.0000000000000378

This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

KEY POINTS

- Bioengineered organs represent a potential source of unlimited donor grafts for transplantation.
- The immune response to bioengineered organs remains largely unknown.
- Nonsel self ECM and stem cells used to build bioengineered organs trigger immune responses.
- Common immunosuppressive therapy may not control the immunogenicity against bioengineered organs.

perfused with a detergent solution to remove the cells leaving behind the extracellular matrix (ECM) scaffold that still maintains the microarchitecture and the composition of the native organ. The resulting structure can be later repopulated with healthy cells – adult, progenitor or pluripotent stem cell derived – to engineer transplantable and functional organ substitutes [1]. This strategy has been shown to be feasible for heart [2^{***}], liver [3], lung [4,5] and kidney [6] using animal or human cadaveric organs in a number of studies since 2008. Although it is theoretically possible to create nonimmunogenic organs by using patient-derived cells for repopulating scaffolds, the specific in-vivo immune reactivity to different subsets of stem cells and to the different components of the ECM scaffolds is not fully understood.

Here, we review the current knowledge on immune reactivity of stem cells that are potentially used to engineer transplantable organ grafts and explore potential therapies that may be employed to minimize the immune responses to bioengineered organs.

IMMUNE RESPONSE TO STEM CELLS

Stem cells have the unique capacity to self-renew and differentiate into specialized cell types, therefore providing a mean to restore tissues and functions altered by diseases and/or senescence. They are particularly suited for regenerative medicine. Among them, there are embryonic stem cells (ESCs); various primary cell types including endothelial, mesenchymal, cardiac progenitor and stem cells collectively termed adult stem cells; and induced pluripotent stem cells (iPSCs). Historically, there has been a relative lack of concern for the immune reactivity of stem cells due to the almost dogmatic concept that ESCs are immune privileged. Recently, however, stem cells immunogenicity has become a major concern, as the immune privileged status of ESCs has been questioned [7]. Although stem cells may have a demonstrable low immunogenic profile in their

undifferentiated state, their progeny express human leukocyte antigen (HLA) molecules at their surface, which makes them immunogenic. Even self-derived autologous iPSCs are exposed to genetic instability and epigenetic reprogramming which may alter their immune neutrality [8]. Given that obtaining autologous iPSC derived from often elderly diseased patients represents a difficult task, adult stem cells of nonself allogeneic origin have become a more realistic and pragmatic therapeutic choice [9]. The translational development of stem cell therapies requires taking into account the immune status of the cells as well as of the patient, and the immune potential of the therapeutic product in a manner similar to organ transplantation. An immunologically educated choice of the therapeutic cells as well as a pretreatment and posttreatment immune monitoring plan will need to be implemented to ensure the optimal clinical success.

HUMAN EMBRYONIC STEM CELLS

The proposed immune privilege status of human ESCs (hESCs) originates from the observation that undifferentiated hESCs do not induce allogeneic T-cell proliferation in in-vitro MLR assays [10]. However, murine ESCs (mESCs) transplanted in an allogeneic recipient are rejected, and repeated injections of allogeneic mESCs are rejected faster likely through immune memory mechanisms [11]. The mechanisms of rejection are still unclear because, despite major histocompatibility complex (MHC) class I expression of the inner cell mass of blastocyst-stage embryo was confirmed by immunohistochemistry, mESCs were not sensitive to natural killer (NK) cell killing [12]. Although the in-vitro and in-vivo ESC derivatives express low levels of MHC class I molecules, IFN treatment results in MHC class I increase on both ESCs and their derivatives, which may also increase their immunogenicity [12]. These initial experiments anticipated various strategies to overcome the allogeneicity of ESCs, which included the development of a universal cell model devoid of MHC class I proteins. Several groups have proposed to knockout $\beta 2m$ and observed that the loss of MHC class I on those hESCs render them resistant to $CD8^+$ T-cell rejection [13]. This strategy was extended to the inducible expression of MHC class II by deleting its master regulator the CIITA (class II trans-activator) [14[■]]. Although the combined knockout of $\beta 2m$ and CIITA are effective from an immune standpoint, it is unlikely that these engineered cells and their derivatives will be used therapeutically as any cell lacking MHC will not be eliminated when infected by a virus or undergo an oncogenic event. Other approaches to obtain immune insensitive ESCs that

include the creation of nuclear transfer human hESC cell lines along with common immunosuppressive regimen or costimulatory blockade for the induction of tolerance has been proposed [15–17].

INDUCED PLURIPOTENT STEM CELLS

Although autologous in origin, the iPSCs display a yet unpredictable array of genetic and epigenetic modifications leading to phenotypic abnormalities likely to be recognized as such by the immune system of the recipient. Although the main concern is tumorigenicity, the intricate immunogenicity has to be taken in account for their clinical development. The expression of MHC molecules and other molecules, such as Tapasin or transporter associated with antigen processing 1, which are required for proper expression of MHC class I, are low in iPSCs as a result of the reprogramming procedure. An initial report stressed the capacity of iPSCs to elicit specific syngeneic T-cell responses in C57BL/6 mice [8]. Although further studies claimed that tissue (skin) derived from iPSCs may have low immunogenic profile, the expression and stability of low expression profile of MHC molecules cannot be guaranteed particularly within inflammatory and/or hypoxic environment. In contrast to autologous iPSCs, allogeneic iPSCs are likely to drive an allogeneic immune response, as even low levels of MHC expression are sufficient to be recognized by the host immune system by the direct or indirect allorecognition pathways. The immunogenic potential of allogeneic iPSCs is likely to vary according to the reprogramming protocol, where some procedures may minimize the genetic and/or the epigenetic variability and instability. The search for a transcriptional signature associated with lower immunogenicity would be indeed of value in selecting the optimal iPSCs for therapy. Therefore, the allogeneicity of iPSCs should be avoided or controlled to achieve an effective and nondetrimental use of iPSCs. The potential immunological response of the host cells may be lowered by common immunosuppressants that regulate effector mechanisms during T-cell responses. Thus, the capacity for allogeneicity is intrinsic to any cell expressing the MHC molecules, whereas the consequences at the effector T-cell level may be modulated or suppressed. Furthermore the use of autologous iPSCs being a long and expensive process, the use of iPSCs from HLA homozygous donors appears to be a more realistic development in human regenerative medicine.

CARDIAC STEM CELLS

Transplantation of cardiac stem/progenitor cells represents a promising source of cells for organ

bioengineering. The proof-of-concept obtained in animal models showing attenuated left ventricular remodeling and improved ventricular function after injection of various types of cardiac stem cells (CSCs) led to test different cell populations including autologous adult CSCs or cardiosphere-derived cells in meaningful clinical trials. However, the biological limitation of the autologous setting and the logistical constraints call upon the use of allogeneic products. The mechanism(s) regulating the behavior of allogeneic CSCs has been only recently investigated with the aim of evaluating the risk and validating their clinical potential. We investigated the immune status of allogeneic hCSCs in the prospect of using them clinically, as the allogeneic option was favored for logistic reasons (principally off-the-shelf availability and quality control). hCSCs express low levels of MHC class I molecules, which can be upregulated under inflammatory and hypoxic conditions mimicking the local environment of ischemia-reperfusion following organ transplantation. Under these conditions, we observed that hCSCs express significant levels of the costimulatory molecule programmed death ligand 1 (PDL1) and IL10, which results in immune regulation [18]. This tolerogenic phenomenon was shown to be the result of a cell contact-dependent expansion and activation of regulatory T cells through the PD1–PDL1 pathway, which was amplified under inflammatory conditions. In addition, hypoxic hCSCs are also able to downregulate NK cell–mediated cytotoxicity and to polarize NK cells toward secretion of anti-inflammatory cytokines IL10 and IL13 rather than inflammatory IFN γ and TNF α [19]. We therefore proposed the benefits of using allogeneic cells maintained through hypoxic conditions with the potential paracrine effect on endogenous cardiogenesis [9]. This promoted the entry of allogeneic hCSC to an ongoing clinical trial.

Cardiosphere-derived cells constitutively express MHC classes I and II molecules in an IFN γ -inducible manner but do not express CD80/86 costimulatory molecules. This immune profile would prevent NK cell–mediated cytotoxicity and favor anergic T-cell responses. A recent report showed, in a rat model of acute myocardial infarction, that two consecutive injections of allogeneic cardiosphere-derived cells would not elicit a detectable immune response and produce a cardiac benefit without sign of rejection or humoral immune sensitization [20^{*}]. The proliferative response and cytokine release were identical in autologous and allogeneic settings, therefore providing experimental argument that there may be no need of immunosuppressive therapy. These results are in line with the concept of allogeneic-driven benefit and the mechanism described above for

adult hCSC. A cautionary note, however, is that this favorable immune status will need to be maintained for a sufficient length of time to allow the beneficial cardiac effect and should not be compromised by recurrent infectious or inflammatory conditions.

ADULT STEM CELLS (TISSUE STEM CELLS)

Adult or, as often called, tissue stem cells have been identified throughout the years in several organs and tissues, including liver [21–23], lungs [24,25], bone marrow [26–31], blood vessels [32–34], skeletal muscle [35–38], intestine and colon [39], skin [40–42] and heart [43,44], just to mention a few. In tissues with slow cell turnover (like the heart, brain or liver), they are found quiescent in specific areas of each tissue designated as stem cell niche and are activated after injury. In organs with faster cellular turnover (like the skin or the intestine), they are constantly dividing and generating more differentiated progeny [45] to quickly replace short living mature cells.

In the past 3 decades, tissue engineers and stem cell biologists have been exploring the most adequate ways to harness the inherent power of these cells to create new tissues and treat human diseases. However, our knowledge of their immunogenicity upon transplantation is still quite incomplete. Particularly, in the field of bioengineered organs, our basic science and clinical experience is still quite limited. Nonetheless, some experimental work on their behavior *in vitro* and *in vivo* upon cellular transplantation is providing new insights into the specific mechanisms that are regulating host immune's response. Adult and fetal liver stem cells are a good example of this.

The required procedures to isolate liver stem cells from fetal and postnatal donors and their inherent biology are generally known today [21,46^{***}]. In addition, even more primitive stem cells have been identified throughout the human biliary tree stem cells (hBTSCs) with the potential to generate hepatocytes, cholangiocytes, pancreatic islets and intestinal cells [22,47]. Independently of their donor origin, they seem to have low immunogenicity, and it is feasible to transplant them without the use of immunosuppression into human patients [48,49^{*}]. The lack of signs of rejection and/or allergy without any immunosuppressive treatment, seems to correlate with marginal or absence of expression of HLA classes I and II antigens both in hepatic and biliary tree stem cells from fetal liver [50–52].

Also, Riccio *et al.* [50] data suggest that hBTSCs could modulate the T-cell response through the secretion of FasL, which impacts the lymphocyte Fas/FasL pathway by producing 'early' apoptosis in CD4⁺ and CD8⁺ T cells. In addition, Maraldi *et al.* [52]

proposed that hepatocyte growth factor secreted by hBTSC also exerts a cytoprotective role by stimulating apoptosis in these same human immune cells. Finally, Bruno *et al.* [51] have shown that hBTSC can inhibit T-cell proliferation by releasing prostaglandin E₂, impair dendritic cell differentiation from monocytes and inhibit NK-cell degranulation. Hence, all these immunomodulatory processes hint at some of the core mechanisms how these human fetal liver and hBTSC (despite their apparent different origin) control immune surveillance toward themselves. Though, the critical question that remains is what exactly happens *in vivo* after cell transplantation that maintains these cells viable in the target organ for months [53]. This question, combined with an expected slight increase in the levels of expression of HLA type I during the stem cell differentiation process suggest a more complex mechanism of immunomodulation, which is not completely understood and whose maintenance is still quite unknown in differentiated stem cells seeded in bioengineered organs. Hence, without further supporting research, it is quite conceivable that unlike primitive stem cells, long-term protection from immune system-mediated killing could be an overconfident outlook in the future development of bioengineered organ-based therapeutics.

MESENCHYMAL STEM CELLS

Despite the current difficulties in predicting the immune response to bioengineered organs, the increasing clinical use of allogeneic MSCs with their immunomodulatory properties is probably a game changer. Their use in bioengineered organs has the advantage of adding an important 'plastic' stromal cell population with the potential to generate important cell lineages and shape the tissue (smooth muscle cells, 'fibroblasts' and pericytes). In addition, their true potential also resides in their ability to induce immune tolerance to the bioengineered tissues/organs [54]. This review is not specifically addressing their nomenclature or functional characteristics, their origin or prolific heterogeneity. Nevertheless, in spite of the common properties of MSCs listed by the International Society for Cellular Therapy [55], important differences and molecular signatures have been observed between MSCs that were derived from different tissues [56,57]. This helps us to postulate the use of tissue-specific MSCs in organ bioengineering to harness MSCs's unique tissue identity for effective organ reconstruction. Yet, when considering trophic and immunomodulation alone, it seems far easier and reasonable to rely on 'universal donor' allogeneic MSCs to gain these effects [54]. Although it is unclear at this point

whether allogeneic MSCs in the bioengineered organs will stimulate immune tolerance or will persist following transplantation [58,59].

CONCLUSION

The evolving paradigm of tissue/organ bioengineering presents new challenges to bioengineers, when addressing the large numbers of cells and resources needed to make bioengineered solid organs a clinical reality. The commercial implementation of scale-out approaches to produce autologous therapies has been difficult to implement due to the regulatory landscape, inherent costs and complexity [60]. Hence, some of the present-day interrogations and debate are focused on the adoption of autologous versus allogeneic cells in the bioengineering of tissues and organs, with the anticipated consequences in the patient's life due to the required immunosuppressive therapies when the 'allogeneic option' is favored. However, considering the current reality and despite some recent progress [43,44,46²²], it is virtually impossible to generate the necessary billions of cells to bioengineer a human liver [61] or a heart [2²²] from isolated adult stem cell. Consequently, hESC and hiPSC represent a more realistic alternative available to fulfill this endeavor. This represents a potential problem, as the innate and the cellular immune response to stem cells are not fully understood. In addition, the humoral immune response following injection of allogeneic stem cells needs to be taken into account [62]. The presence of anti-HLA antibodies after hESC and hiPSC therapy and preformed antibodies prior to injection might compromise and limit the survival and the efficacy of the bioengineered organ. Hence, unknown consequences of the potential immunogenicity of these bioengineered organs created with allogeneic hESC and hiPSC-derived cells is gaining relevance and shifts the focus of the debate from the use of autologous cells to the inherent control of their immunogenicity.

The immune reactivity to decellularized scaffolds that are composed of ECM, also need to be taken into account, especially when derived from allogeneic or xenogeneic tissues. The immune reactivity of ECM has been studied in the past to conclude that ECM of xenogeneic origin is immunogenic. Allaire *et al.* [63] demonstrated in 1997 that the immunogenicity of arterial xenografts is supported by the ECM, which suggests that interspecies, but not an intraspecies graft antigenicity is induced by ECM transplant [64]. A recent review from Keane and Badylak [65²²] summarizes the current immunological implications associated with the use of biological scaffolds. Indeed, several reports describe that each

of the ECM components activate the innate immune response: proteoglycans (heparan sulfate, chondroitin sulfate and keratan sulfate) [66]; nonproteoglycan polysaccharide (hyaluronic acid) [67²²]; fibers (collagen and elastin) [68,69] and others (fibronectin and laminin) [70]. The immune reactivity against the ECM is of special interest in the context of chronic rejection, as this process is not prevented by current immunosuppressive therapy and involves multiple immunological processes in which graft ECM is slowly destroyed, whereas graft blood vessels are obstructed by the deposition of collagen [71].

Understanding how decellularized–recellularized organs maintain their function while inhibiting undesirable immune responses *in vivo* has critical implications to successfully develop this innovative technology. It remains unknown if common immune suppressive therapy, such as rapamycin or cyclosporine, is sufficient to overcome the allo/xenogeneic barrier of the transplanted bioengineered organs *in vivo*. Regardless, one should not lose perspective that sometime in a not-so-distant future, bioengineered organs will increase the pool of available organs for transplantation, but at the cost of immunosuppression until patient's specific hiPSC are easy and inexpensive to generate and produce. Therefore, immune-monitoring strategies for bioengineered organs will hopefully need to be developed in the near future.

Acknowledgements

We would like to thank Patricia Conde for her assistance with the study.

Financial support and sponsorship

The work was supported by the National Institutes of Health (R00DK088962) to B.E.U., by H2020-MSCA-IF-2014-660554 and ISCIII PI15/00563 to P.M.B., and SAF SAF2013-48834-R/SAF2016-80031-R to J.O.

Conflicts of interest

B.E.U. has a financial interest in Organ Solutions, LLC, which is reviewed and arranged by Massachusetts General Hospital and Partners HealthCare in accordance with their conflict of interest policies. The remaining authors have no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Badylak SF, Taylor D, Uygun K. Whole-organ tissue engineering: decellularization and recellularization of three-dimensional matrix scaffolds. *Annu Rev Biomed Eng* 2011; 13:27–53.

2. Ott HC, Matthiesen TS, Goh SK, *et al*. Perfusion-decellularized matrix: using nature's platform to engineer a bioartificial heart. *Nat Med* 2008; 14:213–221.
- This is the first article describing the use of organ decellularization/recellularization approach to develop bioengineered organs.
3. Uygun BE, Soto-Gutierrez A, Yagi H, *et al*. Organ reengineering through development of a transplantable recellularized liver graft using decellularized liver matrix. *Nat Med* 2010; 16:814–820.
4. Petersen TH, Calle EA, Zhao L, *et al*. Tissue-engineered lungs for in vivo implantation. *Science* 2010; 329:538–541.
5. Ott HC, Clippinger B, Conrad C, *et al*. Regeneration and orthotopic transplantation of a bioartificial lung. *Nat Med* 2010; 16:927–933.
6. Song JJ, Guyette JP, Gilpin SE, *et al*. Regeneration and experimental orthotopic transplantation of a bioengineered kidney. *Nat Med* 2013; 19:646–651.
7. Drukker M, Benvenisty N. The immunogenicity of human embryonic stem-derived cells. *Trends Biotechnol* 2004; 22:136–141.
8. Zhao Z, Zhang ZN, Rong Z, Xu Y. Immunogenicity of induced pluripotent stem cells. *Nature* 2011; 474:212–215.
9. Al-Daccak R, Charron D. Allogenic benefit in stem cell therapy: cardiac repair and regeneration. *Tissue Antigens* 2015; 86:155–162.
10. Li L, Baroja ML, Majumdar A, *et al*. Human embryonic stem cells possess immune-privileged properties. *Stem Cells* 2004; 22:448–456.
11. Swijnenburg RJ, Schrepfer S, Cao F, *et al*. In vivo imaging of embryonic stem cells reveals patterns of survival and immune rejection following transplantation. *Stem Cells Dev* 2008; 17:1023–1029.
12. Drukker M, Katz G, Urbach A, *et al*. Characterization of the expression of MHC proteins in human embryonic stem cells. *Proc Natl Acad Sci U S A* 2002; 99:9864–9869.
13. Lu P, Chen J, He L, *et al*. Generating hypoimmunogenic human embryonic stem cells by the disruption of beta 2-microglobulin. *Stem Cell Rev* 2013; 9:806–813.
14. Chen H, Li Y, Lin X, *et al*. Functional disruption of human leukocyte antigen II in human embryonic stem cell. *Biol Res* 2015; 48:59.
- The authors optimized human ESCs (hESCs) for clinical application through gene modification of the HLA II defective hESCs via deleting class II trans-activator.
15. Gallegos TF, Sancho-Martinez I, Izpissua Belmonte JC. Advances in cellular reprogramming: moving toward a reprieve from immunogenicity. *Immunol Lett* 2013; 155:14–17.
16. Grinnemo KH, Sylven C, Hovatta O, *et al*. Immunogenicity of human embryonic stem cells. *Cell Tissue Res* 2008; 331:67–78.
17. Lui KO, Waldmann H, Fairchild PJ. Embryonic stem cells: overcoming the immunological barriers to cell replacement therapy. *Curr Stem Cell Res Ther* 2009; 4:70–80.
18. Louden L, Boukouaci W, Borlado LR, *et al*. Allogenicity of human cardiac stem/progenitor cells orchestrated by programmed death ligand 1. *Circ Res* 2013; 112:451–464.
19. Boukouaci W, Louden L, Siewiera J, *et al*. Natural killer cell crosstalk with allogeneic human cardiac-derived stem/progenitor cells controls persistence. *Cardiovasc Res* 2014; 104:290–302.
20. Reich H, Tseliou E, de Couto G, *et al*. Repeated transplantation of allogeneic cardiosphere-derived cells boosts therapeutic benefits without immune sensitization in a rat model of myocardial infarction. *J Heart Lung Transplant* 2016. [Epub ahead of print]
- The authors described that repeat dosing of allogeneic cardiosphere-derived cells in immunocompetent rats is well tolerated and effective.
21. Schmelzer E, Zhang L, Bruce A, *et al*. Human hepatic stem cells from fetal and postnatal donors. *J Exp Med* 2007; 204:1973–1987.
22. Cardinale V, Wang Y, Carpino G, *et al*. Multipotent stem/progenitor cells in human biliary tree give rise to hepatocytes, cholangiocytes, and pancreatic islets. *Hepatology* 2011; 54:2159–2172.
23. Cardinale V, Wang Y, Carpino G, *et al*. The biliary tree – a reservoir of multipotent stem cells. *Nat Rev Gastroenterol Hepatol* 2012; 9:231–240.
24. Kajstura J, Rota M, Hall SR, *et al*. Evidence for human lung stem cells. *N Engl J Med* 2011; 364:1795–1806.
25. Rock JR, Onaitis MW, Rawlins EL, *et al*. Basal cells as stem cells of the mouse trachea and human airway epithelium. *Proc Natl Acad Sci U S A* 2009; 106:12771–12775.
26. Spangrude GJ, Heimfeld S, Weissman IL. Purification and characterization of mouse hematopoietic stem cells. *Science* 1988; 241:58–62.
27. Smith C, Gasparetto C, Collins N, *et al*. Purification and partial characterization of a human hematopoietic precursor population. *Blood* 1991; 77:2122–2128.
28. Storms RW, Trujillo AP, Springer JB, *et al*. Isolation of primitive human hematopoietic progenitors on the basis of aldehyde dehydrogenase activity. *Proc Natl Acad Sci U S A* 1999; 96:9118–9123.
29. Punzel M, Ho AD. Divisional history and pluripotency of human hematopoietic stem cells. *Ann N Y Acad Sci* 2001; 938:72–81; discussion 81–72.
30. Friedenstein AJ, Piatetzky Sli, Petrakova KV. Osteogenesis in transplants of bone marrow cells. *J Embryol Exp Morphol* 1966; 16:381–390.
31. Caplan AL. Mesenchymal stem cells. *J Orthop Res* 1991; 9:641–650.
32. Caplan AL. All MSCs are pericytes? *Cell Stem Cell* 2008; 3:229–230.
33. James AW, Zara JN, Corselli M, *et al*. An abundant perivascular source of stem cells for bone tissue engineering. *Stem Cells Transl Med* 2012; 1:673–684.
34. Crisan M, Corselli M, Chen WC, Peault B. Perivascular cells for regenerative medicine. *J Cell Mol Med* 2012; 16:2851–2860.
35. Mauro A. Satellite cell of skeletal muscle fibers. *J Biophys Biochem Cytol* 1961; 9:493–495.
36. Laguens R. Satellite cells of skeletal muscle fibers in human progressive muscular dystrophy. *Virchows Arch Pathol Anat Physiol Klin Med* 1963; 336:564–569.
37. Sakai Y. Experimental studies on the role of satellite cells in regeneration of rat skeletal muscle fibers. *Acta Pathol Jpn* 1977; 27:305–320.
38. Mitchell KJ, Pannérec A, Cadot B, *et al*. Identification and characterization of a nonsatellite cell muscle resident progenitor during postnatal development. *Nat Cell Biol* 2010; 12:257–266.
39. Barker N, van Es JH, Kuipers J, *et al*. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* 2007; 449:1003–1007.
40. Barrandon Y, Green H. Three clonal types of keratinocyte with different capacities for multiplication. *Proc Natl Acad Sci U S A* 1987; 84:2302–2306.
41. Jones PH, Watt FM. Separation of human epidermal stem cells from transit amplifying cells on the basis of differences in integrin function and expression. *Cell* 1993; 73:713–724.
42. Rochat A, Kobayashi K, Barrandon Y. Location of stem cells of human hair follicles by clonal analysis. *Cell* 1994; 76:1063–1073.
43. Beltrami AP, Barlucchi L, Torella D, *et al*. Adult cardiac stem cells are multipotent and support myocardial regeneration. *Cell* 2003; 114:763–776.
44. Hsieh PC, Segers VF, Davis ME, *et al*. Evidence from a genetic fate-mapping study that stem cells refresh adult mammalian cardiomyocytes after injury. *Nat Med* 2007; 13:970–974.
45. Wodarz D. Effect of stem cell turnover rates on protection against cancer and aging. *J Theor Biol* 2007; 245:449–458.
46. Huch M, Gehart H, van Bostel R, *et al*. Long-term culture of genome-stable bipotent stem cells from adult human liver. *Cell* 2014; 160:299–312.
- This study identified the conditions that allow long-term expansion of primary adult human liver stem cells.
47. Semeraro R, Carpino G, Cardinale V, *et al*. Multipotent stem/progenitor cells in the human foetal biliary tree. *J Hepatol* 2012; 57:987–994.
48. Khan AA, Shaik MV, Parveen N, *et al*. Human fetal liver-derived stem cell transplantation as supportive modality in the management of end-stage decompensated liver cirrhosis. *Cell Transplant* 2010; 19:409–418.
49. Cardinale V, Carpino G, Gentile R, *et al*. Transplantation of human fetal biliary tree stem/progenitor cells into two patients with advanced liver cirrhosis. *BMC Gastroenterol* 2014; 14:204.
- This report represents proof of the concept that stem cells are a suitable and feasible for cell therapy in humans.
50. Riccio M, Carnevale G, Cardinale V, *et al*. The Fas/Fas ligand apoptosis pathway underlies immunomodulatory properties of human biliary tree stem/progenitor cells. *J Hepatol* 2014; 61:1097–1105.
51. Bruno S, Grange C, Tapparo M, *et al*. Human liver stem cells suppress T-cell proliferation, NK activity, and dendritic cell differentiation. *Stem Cells Int* 2016; 2016:8468549.
52. Maraldi T, Guida M, Beretti F, *et al*. Human biliary tree stem/progenitor cells immunomodulation: role of hepatocyte growth factor. *Hepato Res* 2016. [Epub ahead of print]
53. Khan AA, Shaik MV, Parveen N, *et al*. Human fetal liver-derived stem cell transplantation as supportive modality in the management of end-stage decompensated liver cirrhosis. *Cell Transplant* 2010; 19:409–418.
54. Ankrum JA, Ong JF, Karp JM. Mesenchymal stem cells: immune evasive, not immune privileged. *Nat Biotechnol* 2014; 32:252–260.
55. Dominici M, Le Blanc K, Mueller I, *et al*. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006; 8:315–317.
56. Crisan M, Yap S, Castella L, *et al*. A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell* 2008; 3:301–313.
57. Zhao H, Feng J, Ho TV, *et al*. The stroma provides a niche for mesenchymal stem cells of craniofacial bones. *Nat Cell Biol* 2015; 17:386–396.
58. Toma C, Wagner WR, Bowry S, *et al*. Fate of culture-expanded mesenchymal stem cells in the microvasculature: in vivo observations of cell kinetics. *Circ Res* 2009; 104:398–402.
59. Lee RH, Pulin AA, Seo MJ, *et al*. Intravenous hMSCs improve myocardial infarction in mice because cells embolized in lung are activated to secrete the anti-inflammatory protein TSG-6. *Cell Stem Cell* 2009; 5:54–63.
60. Houd P, Ginty P, Chandra A, Williams DJ. Manufacturing models permitting roll out/scale out of clinically led autologous cell therapies: regulatory and scientific challenges for comparability. *Cytotherapy* 2014; 16:1033–1047.
61. Baptista PM, Siddiqui MM, Lozier G, *et al*. The use of whole organ decellularization for the generation of a vascularized liver organoid. *Hepatology* (Baltimore, MD) 2011; 53:604–617.
62. Charron D, Suberbielle-Boissel C, Tamouza R, Al-Daccak R. Anti-HLA antibodies in regenerative medicine stem cell therapy. *Hum Immunol* 2012; 73:1287–1294.
63. Allaire E, Mandet C, Bruneval P, *et al*. Cell and extracellular matrix rejection in arterial concordant and discordant xenografts in the rat. *Transplantation* 1996; 62:794–803.
64. Allaire E, Bruneval P, Mandet C, *et al*. The immunogenicity of the extracellular matrix in arterial xenografts. *Surgery* 1997; 122:73–81.

65. Keane TJ, Badylak SF. The host response to allogeneic and xenogeneic biological scaffold materials. *J Tissue Eng Regen Med* 2015; 9:504–511. This is an excellent revision of the immune response to the extracellular matrix (ECM) components of nonself origin.
66. Taraballi F, Corradetti B, Minardi S, *et al.* Biomimetic collagenous scaffold to tune inflammation by targeting macrophages. *J Tissue Eng* 2016; 7:2041731415624667.
67. Todd JL, Wang X, Sugimoto S, *et al.* Hyaluronan contributes to bronchiolitis obliterans syndrome and stimulates lung allograft rejection through activation of innate immunity. *Am J Respir Crit Care Med* 2014; 189:556–566. The authors describe that accumulation of the ECM component hyaluronan contributes to bronchiolitis obliterans syndrome associated with lung allograft rejection.
68. Bayrak A, Pruger P, Stock UA, Seifert M. Absence of immune responses with xenogeneic collagen and elastin. *Tissue Eng Part A* 2013; 19: 1592–1600.
69. Boeer U, Buettner FF, Klingenberg M, *et al.* Immunogenicity of intensively decellularized equine carotid arteries is conferred by the extracellular matrix protein collagen type VI. *PLoS One* 2014; 9: e105964.
70. Garcia-Nieto S, Johal RK, Shakesheff KM, *et al.* Laminin and fibronectin treatment leads to generation of dendritic cells with superior endocytic capacity. *PLoS One* 2010; 5:e10123.
71. Dormond O, Dufour M, Seto T, *et al.* Targeting the intragraft microenvironment and the development of chronic allograft rejection. *Hum Immunol* 2012; 73:1261–1268.