Lifespan extension conferred by mitogen-activated protein kinase kinase kinase 5 (*MAP3K5*) longevity-associated gene variation is confined to at-risk men with a cardiometabolic disease

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ABSTRACT

Genetic variants of the kinase signaling gene MAP3K5 are associated with longevity. Here we explore whether the longevity-association involves protection against mortality in all individuals, or only in individuals with aging-related diseases. We tested the strongest longevity associated single nucleotide polymorphism (SNP), rs2076260, for association with mortality in 3,516 elderly American men of Japanese ancestry. At baseline (1991–1993), 2,461 had either diabetes (n=990), coronary heart disease (CHD; n=724), or hypertension (n=1,877), and 1,055 lacked any of these cardiometabolic diseases (CMDs). The men were followed from baseline until Dec 31, 2019. Longevity-associated genotype CC in a major allele homozygote model, and CC+TT in a heterozygote disadvantage model were associated with longer lifespan in individuals having a CMD (covariate-adjusted hazard ratio [HR] 1.23 [95% CI: 1.12–1.35, p=2.5x10⁻⁵] in major allele homozygote model, and 1.22 [95% CI: 1.11–1.33, p=1.10x10⁻⁵] in heterozygote disadvantage model). For diabetes, hypertension and CHD, HR p-values were 0.019, 0.00048, 0.093, and 0.0024, 0.00040, 0.0014, in each respective genetic model. As expected, men without a CMD outlived men with a CMD ($p=1.9 \times 10^{-6}$). There was, however, no difference in lifespan by genotype in men without a CMD (p=0.21 and 0.86, respectively, in each genetic model). In conclusion, we propose that in individuals with a cardiometabolic disease, longevity-associated genetic variation in MAP3K5 enhances resilience mechanisms in cells and tissues to help protect against cardiometabolic stress caused by CMDs. As a result, men with CMD having longevity genotype live as long as all men without a CMD.

INTRODUCTION

Mitogen-activated protein kinase kinase kinase 5 (MAP3K5, also termed ASK1 [apoptosis signal-regulating kinase 1]) is a member of a family of

enzymes involved in kinase signaling cascades in the cell. MAP3K5 plays an important role in cellular responses evoked by changes in environment. These include cell differentiation and survival, apoptosis, innate immune response, and oxidative stress response

[1]. It mediates signal transduction of oxidative stress and receptor-mediated inflammatory signals, such as ones involving tumor necrosis factor and lipopolysaccharide. Its crucial role in the apoptosis signal transduction pathway is mediated by mitochondria-dependent caspase activation. MAP3K5 plays a role in the pathology of a wide range of diseases in which reactive oxygen species (ROS) and/or endoplasmic reticulum stress are causative factors. MAP3K5/ASK1 may influence in vivo insulin action and obesity, and MAP3K5 variants are associated with type 2 diabetes [2]. MAP3K5 may prevent stress-induced disorders and protect from bacterial and viral infection under physiological circumstances. On the contrary, it may exert adverse effects through excessive cellular apoptosis and increased inflammation under some pathological conditions, such as present in neurodegenerative disorders. cardiovascular diseases, inflammatory diseases. and chronic inflammation-induced carcinogenesis [3]. Once activated, MAP3K5/ASK1 acts as an upstream activator of the MKK/JNK and p38 MAPK signal transduction cascades through phosphorylation and activation of several MAP kinase kinases such as MAP2K4/SEK1, MAP2K3/MKK3, MAP2K6/MKK6 and MAP2K7/MKK7. These in turn activate p38 MAPKs and c-jun N-terminal kinases (JNKs). Both p38 MAPK and JNKs control the transcription factor activator protein-1 (AP-1) [4]. While the above pathways are critical for senescence, aging, and age-associated cardiovascular diseases MAP3K5 might not be the cause, but rather a response [5, 6].

In a study of 33 single nucleotide polymorphisms (SNPs) of MAP3K5 in our cohort of American men of Japanese ancestrv (Supplementary Methods. Supplementary Tables 1, 2), alleles of two adjacent SNPs, and haplotypes of these alleles, were associated with longevity in men aged ≥ 95 years [7]. Bonferroni corrected p-values were 0.0043 for the SNP rs2076260 and 0.032 for the SNP rs6904753 in a heterozygous disadvantage model (Supplementary Table 3). Haplotype analysis of these yielded a p-value of 0.00004. Our rationale for studying MAP3K5 was because in mouse liver Map3k5 was differentially expressed in response to caloric restriction [8], a wellknown longevity enhancer.

In the present study we tested the hypothesis that the longevity-associated alleles of *MAP3K5* SNP *rs2076260* mediate their effect on lifespan at least in part by protection against the detrimental effects of aging-related cardiometabolic diseases (CMD), namely diabetes and/or hypertension and/or coronary heart disease (CHD).

RESULTS

Characteristics of subjects

Baseline (1991–1993) characteristics of men in the study for each genotype of *rs2076260*, adjusting, for age, genotype, and prevalence of medical conditions are shown in Supplementary Table 4. By December 31, 2019, 3,480 out of 3,516 subjects had died during the 29 years of follow-up. At baseline, among the 3,516 participants, 29% had been diagnosed with diabetes alone, 53% with hypertension alone, 21% with CHD alone, and 14% with cancer alone. Seventy percent had at least one CMD, and 5.2% had all three CMDs. Mean age at death was 88.6 ± 6.0 years for men with at least one CMD, and 89.4 ± 6.0 years for men who did not have any of the CMDs (p < 0.0001).

MAP3K5 genotype and survival in men with a CMD and men without a CMD

Survival curves for men without a CMD and those with a CMD showed that those who did not have a CMD lived longer (Kaplan-Meier Log-rank $\chi^2 = 24.7$, p =1.9x10⁻⁶). Figure 1(A) shows survival curves for men with a CMD and men without a CMD according to whether they were major allele homozygotes (CC) of rs2076260 or minor allele carriers (CT/TT). Figure 1(B) shows survival curves for the men with a CMD and for men who did not have a CMD according to whether they were heterozygous (CT) or homozygous (CC+TT) for each allele. These curves were determined using a Cox proportional hazard model adding an interaction term of "MAP3K5 with CMD". Only in men with CMD was the longevity-associated genotype associated with greater lifespan than the alternate genotype: p = 0.00004for major allele homozygotes (CC) vs. minor allele carriers (CC/TT), and p = 0.000006 for heterozygotes (CT) vs. homozygotes (CC+TT). Overall, men without a CMD had the longest lifespans. Moreover, importantly, we noted that there was no statistical difference in lifespan for each genotype in men who did not have a CMD. Figure 2 shows forest plots of mortality risk for men with a CMD and men without a CMD for each genetic model.

Age-adjusted baseline variables of men with a CMD and men without a CMD according to genotype (CC, CT, TT) are shown in Supplementary Table 5. None of the variables showed a difference between genotypes. The variables included body mass index (BMI), wait-tohip ratio, fasting plasma glucose, fasting plasma insulin, plasma fibrinogen, white blood cell count, smoking, alcohol intake and physical activity index. Analyses found no evidence of population stratification or admixture in the dataset (data not shown). In men with a CMD, prevalence of diabetes, hypertension and CHD was not statistically different between each genotype.

In men with a CMD, hazard ratios (HR) for association with mortality for just diabetes, just hypertension, just CHD, and for any of these CMDs were statistically significant in two genetic models (Table 1). These were for: (1) major allele homozygotes (*CC*) vs. minor allele carriers (*CT/TT*), and (2) heterozygotes (*CT*) vs. homozygotes of each allele (*CC+TT*). No significant difference was, however, found in two other genetic

models, namely for minor allele homozygotes (TT) vs. major allele carriers (CC/CT), and for an additive model (Supplementary Table 6). Thus, in the 1st model, being homozygous for the major allele conferred strong protection against mortality in men with a CMD. In men without a CMD, however, lifespan was significantly longer irrespective of *MAP3K5* genotype. In the 2nd model, heterozygotes were at a disadvantage in that men who had the *CT* genotype had a significantly higher risk of mortality than men who were homozygous for either the *C* or *T* allele.



Figure 1. Survival curves spanning the period from baseline (1991–1993) to Dec 31, 2019 for men with and men without a CMD according to genotypes of *MAP3K5* SNP *rs2076260*. The survival probabilities were estimated from the Cox proportional hazard model: $h(t) = h(t0) * \exp(\beta 1*Age + \beta 2*BMI + \beta 3*Glucose + \beta 4*CMD + \beta 5*MAP3K5_xx + \beta 6* (CMD*MAP3K5_xx))$, where "xx" is genotype, by fixing age at 75 years, BMI at the mean, 23.5 kg/m², and glucose at the mean, 113 mg/dL (where $\beta 6$ is the effect of the interaction of CMD with *MAP3K5* genotype on mortality, for *CC* vs *CT*/*TT*, i.e., a recessive model, giving $p(\beta 6) = 0.023$). (A) Survival curves for men with a CMD vs. men without a CMD for major allele homozygote (*CC*) vs. minor allele carriers (*CT+TT*) (*p*=0.000041 and *p*=0.95, respectively). (B) Survival curves for men with a CMD vs. men without a CMD for heterozygote disadvantage model, *CT* vs. *CC/TT* (*p*=0.000059 and *p*=0.50, respectively, giving $p(\beta 6) = 0.057$).

Functional annotations

In an attempt to determine how MAP3K5 may influence disease resistance and help to identify biological



Figure 2. Forest plots of mortality risk (hazard ratio and 95% CI), adjusted for age, BMI and glucose at baseline, for men with a CMD and men without a CMD according to genotype of MAP3K5 SNP rs2076260 in each genetic **model.** (A) major allele homozygote (CC) vs. minor allele carriers (CT+TT). (B) heterozygote disadvantage model, CT vs. CC/TT. It can be seen that in men with a CMD who had the longevityassociated genotype, mortality risk was reduced to normal in that it did not differ significantly from the survival curve in men without a CMD. It should be noted that the HRs in Figure 2 differ slightly from those in Table 1. This is because the HRs in Table 1 were obtained from stratified analyses by diabetes, hypertension, CHD, and any CMD (i.e., were separately estimated by disease status). In Figure 2, we compared the HRs for the 4 groups by CMD and MAP3K5 genotype. The HRs and p-values for pairwise comparisons among the 4 groups were estimated in one Cox model.

pathways we examined the following: (1) *MAP3K5* tissue expression, (2) transcription factors (TFs) that might be modified by our sentinel SNP (*rs2076260*), (3) SNPs in linkage disequilibrium (LD) that might modify transcription factor binding, (4) the expression patterns of these TFs, and (5) the location of any *cis*-regulatory elements that are physically linked with our sentinel SNP. Supplementary Figure 1 shows that *MAP3K5* is expressed in most tissues, but notably at high levels in adrenal, ovary, and pituitary.

We screened SNPs in the block of LD containing rs2076260 (Supplementary Figure 2) for functional annotations using the HaploReg database. Seven additional SNPs were in near perfect LD with rs2076260 and are predicted to either modify transcription factor motifs and/or are predicted to alter promoter/enhancer histone marks or DNAse I sensitivity. The latter features are associated with access to regulatory proteins. All **SNPs** were examined in more eight detail (Supplementary Figure 3 and Supplementary Table 7). Five of those SNPs are predicted to modify TF binding and are shown in Supplementary Table 8. Also shown is whether the minor allele is predicted to create or abolish TF binding, the biological pathways, and the tissue expression patterns. While these TFs are expressed in most tissues, they are prominent in lung and thyroid. The biological pathways include: (1)activation of immunoglobulin heavy-chain transcription, (2)repression of GATA4 and GATA6 transcription, (3) unfolded protein response, (4) activation of many muscle-specific, growth factor-induced, and stressedinduced genes, and (5) expression of metallothionein proteins in response to exposure to heavy metals.

Since the MAP3K5 SNPs are in a non-coding region of the genome, they are presumed to affect transcription, whether directly or indirectly, as chromatin modifying units (i.e., cis-regulatory elements) (Supplementary Table 8). Screening of MAP3K5 for expression quantitative loci (eQTLs) using the GTEx portal identified 1,532 entries (not shown) and four splice-site sQTLs, which are variants associated with differential splicing (not shown). Most of the eQTLs in MAP3K5 are differentially expressed in artery (tibial), lung, breast, brain (cerebellar), esophagus (muscularis), spleen, and adipose (visceral) tissues, as defined in the GTEx database. As an aside, it should be noted that dbSNP misclassifies rs2076260 as a mis-sense mutation (https://www.ncbi.nlm.nih.gov/projects/SNP/snp ref.cg i?do not redirect&rs=rs2076260). The base change is actually in the 3rd codon with no change in the encoded amino acid. We believe the confusion arose from sequencing of the opposite strand by some Human Genome Labs, where rs2076260 is a T \rightarrow C change, whereas the complement is $A \rightarrow G$.

Disease	Cox	Genetic	With a CMD		Without a CMD			
(n with, total)	model*	model**	HR (95% CI)	р	HR (95% CI)	р		
Diabetes	1	CT/TT vs. CC	1.18 (1.03–1.36)	0.016	1.11 (1.02–1.22)	0.017		
(990, 2478)	2	CT/TT vs. CC	1.21 (1.05–1.41)	0.011	1.08 (0.98–1.19)	0.10		
Hypertension	1	CT/TT vs. CC	1.18 (1.07–1.30)	0.0013	1.06 (0.95–1.19)	0.27		
(1877, 1639)	2	CT/TT vs. CC	1.22 (1.09–1.36)	0.00041	1.03 (0.92–1.16)	0.60		
CHD	1	CT/TT vs. CC	1.21 (1.02–1.42)	0.026	1.11 (1.02–1.20)	0.017		
(724, 2792)	2	CT/TT vs. CC	1.19 (0.99–1.43)	0.059	1.11 (1.02–1.22)	0.019		
Any CMD	1	CT/TT vs. CC	1.20 (1.09–1.31)	0.000075	0.99 (0.86–1.13)	0.83		
(2461, 1055)	2	CT/TT vs. CC	1.23 (1.12–1.36)	0.000023	0.90 (0.78-1.05)	0.18		
Diabetes	1	CT vs. CC/TT	1.25 (1.10–1.42)	0.00054	1.14 (1.06–1.24)	0.0010		
(990, 2478)	2	CT vs. CC/TT	1.26 (1.10–1.44)	0.0010	1.10 (1.01-1.20)	0.029		
Hypertension	1	CT vs. CC/TT	1.18 (1.08–1.29)	0.00043	1.14 (1.04–1.26)	0.0075		
(1877, 1639)	2	CT vs. CC/TT	1.21 (1.09–1.34)	0.00021	1.08 (0.97-1.20)	0.18		
CHD	1	CT vs. CC/TT	1.34 (1.15–1.55)	0.00012	1.14 (1.05–1.22)	0.0009		
(724, 2792)	2	CT vs. CC/TT	1.33 (1.13–1.57)	0.00058	1.11 (1.02–1.21)	0.012		
Any CMD	1	<i>CT</i> vs. <i>CC/TT</i>	1.21 (1.12–1.31)	0.0000026	1.07 (0.95–1.21)	0.25		
(2461, 1055)	2	CT vs. CC/TT	1.22 (1.12–1.33)	0.0000081	0.98 (0.86–1.13)	0.82		

Table 1. Hazard ratios (HR) by genotype of *MAP3K5* SNP *rs2076260* with total mortality in men with diabetes, CHD, hypertension, and any of these CMDs.

*Cox models: Model 1: Age-adjusted. Model 2: Covariate-adjusted, where covariates adjusted in Cox model were: age (years), BMI (kg/m²), glucose (mmol/l), insulin (mIU/dL), plasma fibrinogen (mg/dl), white blood count ($10^{3}/\mu$ L), smoking (pack-years), alcohol intake (oz/mo), physical activity index, depression, cancer, and stroke.

**Genetic models: *Top half:* Major allele homozygote (*CC*) model. *Bottom half:* Heterozygote disadvantage model.

The longevity-associated SNP *rs2076260* and its nearest neighbor *rs6906753* in LD (D'=0.99) demonstrated significant changes in binding capacities for TFs that included homeobox D10 (HOXD10), POU class 2 homeobox 2 (POU2F2), TATA box binding protein (TBP)-associated factor, RNA polymerase II (TATA), activating transcription factor 3 (ATF3), and Hes related family BHLH transcription factor with YRPW motif 1 (HEY1) (Supplementary Figure 3). The predicted effects of putative functional variants in the SNPs on TF binding and thus tissue expression patterns are shown in Supplementary Table 9.

In order to determine whether *rs2076260* was near any additional regulatory sites and features, we examined the chromatin structures within and surrounding *MAP3K5*. Using the UCSC genome browser we identified 2 long noncoding RNAs (lncRNAs), *MAP3K5-AS1* and *LOC101928429*, located within *MAP3K5* (Supplementary Figure 4 and Supplementary Table 8). From the literature we identified a site, cg21506299, that is differentially methylated in individuals with increased BMI [9]. The above entities were mapped on to the LD heatmap (Supplementary Figure 2) using the LDhap Tool (https://analysistools.cancer.gov/LDlink/?tab=ldmatri x). The locations of these features (shown in Supplementary Figure 2 and summarized in Supplementary Table 8), indicate that the adjacent longevity SNPs (asterisked in Supplementary Table 2) overlap with *MAP3K5-AS1*, that is in LD. The differentially methylated site, cg21506299, overlaps with the lncRNA *LOC101928429* near the promoter (Supplementary Figure 4).

A super-enhancer is a region of the mammalian genome comprising multiple enhancers collectively bound by an array of TFs to drive transcription of genes involved in cell identity. We identified super-enhancers overlapping the *MAP3K5* promoter, using the dbSuper database (<u>https://asntech.org/dbsuper/index.php</u>). These enhancers were identified in various T- and B-cell lines. They are shown in Supplementary Figure 4 and listed in Supplementary Table 10.

We mapped additional potential regulatory sites using the WashU Genome Browser. Supplementary Figure 4 shows the locations of *MAP3K5* relative to *MAP3K5*-*AS1*, *rs2076260*, cg21506299, *LOC101928429*, and super-enhancers. H3K4me3 indicates sites of trimethylation of lysine 4 in histone H3 (green), locations of the H2 histone variant H2A.Z (blue), locations of RNA polymerase II (RNAPII) binding, and CTCF binding. H3K4me3 is associated with sites of open chromatin which are associated with activation of transcription of nearby genes [10]. H2A.Z is associated with regions of genome fluidity [10]. The *MAP3K5* promoter overlaps with H3K4me3 sites as well as H2A.Z sites and is connected, through RNAPII binding/pausing to SNP *rs2076260* as well as *MAP3K5-AS1* and *LOC101928429*. Together these features are predicted to form a *cis*-regulatory unit. This is supported by the locations of CTCF binding sites that generally form insulator domains.

DISCUSSION

The present study has found that the longevityassociated major allele of MAP3K5 SNP rs2076260 is associated with protection against mortality from CMD. As a result, individuals with either one, two or all of the disorders – diabetes, hypertension or CHD – live longer if they have the protective genotype. Our study showed in fact that lifespan of men with a CMD who had the longevity-associated allele was not only longer, but did not differ significantly from lifespan of men without CMD. This indicates that possession of the MAP3K5longevity-associated genotype can mitigate the adverse effects on lifespan of having a CMD.

We also found an association with longevity when we tested the genetic data in a heterozygous disadvantage model. Heterozygote disadvantage is when a heterozygote has a lower overall fitness than either homozygote, and can be a potent driver of population genetic divergence [11].

Until our earlier study [7], MAP3K5 had not previously been linked directly to human aging and longevity. Most studies have, however, involved European populations [10]. According to dbSNP, the frequency of the C allele of rs2076260 in the 1000 Genomes East Asian superpopulation is 0.55, whereas it is only 0.17 in the European super-population (https://www.ncbi. nlm.nih.gov/snp/?term=rs2076260), thus diminishing the chance of detection of an association with longevity in genetic association studies of Europeans. Various genetic factors have been identified that are associated with aging-related risk factors and diseases [12]. In Japanese people, healthy traditional dietary, lifestyle and cultural factors, by reducing risk of CMDs, are thought to contribute to their longer average lifespans [13]. In extreme old age genetic factors become more important than environmental factors in lifespan determination [10]. Our study used a homogeneous population of American men of Japanese ancestry in which there was no evidence of population stratification or admixture [14]. Thus, various favorable factors appear to have contributed to our previous positive genetic finding of an association of SNPs in MAP3K5 with longevity [7].

MAP3K5 is involved in stress response, inflammation, and apoptosis [1]. Disruption of *Map3k5* in mice attenuates left ventricular remodelling [15]. Prolonged activation of p38 or JNK by MAP3K5 results in long-term cellular damage. MAP3K5 inhibition may serve as a therapeutic target, as summarized in studies that have tested the effects of MAP3K5/ASK1 inhibition in cell and animal disease models, as well as in human clinical trials for a variety of diseases [16].

MAP3K5 is expressed in a large number of tissues, most notably adipose tissue, artery, Epstein barr virustransformed lymphocytes, ovary, and pituitary (as documented in the GTEx database: https://gtexportal. org/home/gene/MAP3K5). Moreover, MAP3K5 has several isoforms that are differentially expressed in various tissues (see GTEx database). It is likely that the longevity-associated MAP3K5 genotype is in LD with functional allele(s) of regulatory variant(s) of MAP3K5 that have a stronger effect on gene expression than the alternative genotypes, so boosting the levels of the encoded MAP3K5 protein and leading to higher beneficial biological effects, such as apoptosis. Two longevity-associated SNPs, rs2076260 and its near neighbor rs6904753, in LD with each other, are predicted to influence binding capacities for six transcription factors. The common allele of rs2076260 is predicted to create/enhance binding of factors HOXD10, POU2F2, and TATA, thus increasing MAP3K5 expression, but reduce binding of DMRT1, while the common allele of rs6904753 is predicted to abolish/reduce binding of factors ATF3 and HEY1, which are generally repressive, and thus increase MAP3K5 expression. The additional SNPs in LD may also influence TF binding. Since these TFs are differentially expressed, their specific effect would depend on the tissue.

Data on the tissue expression distributions for the lncRNAs were not available. The lncRNA, MAP3K5-AS1, located within MAP3K5 close to rs2076260, may influence MAP3K5 expression, either negatively by recruiting RNAPII on the negative strand or positively by recruiting other transcription enhancers and/or splicing factors on the positive strand. Since MAP3K5-AS1 is transcribed in the opposite direction as MAP3K5, it operates in effect as an anti-sense RNA, interfering with MAP3K5 transcription, as above. The possibility of a role for MAP3K5-AS1 in the genotypic effect we have observed is therefore of interest. The cg21506299 site differentially methylated in subjects with increased BMI [9], and located in the MAP3K5 promoter, may be of interest in mediating the contrasting genotypic effect found in our men who had a CMD. This is because high BMI is associated with diabetes, hypertension and CHD. Another lncRNA, LOC101928429, at this site

may influence transcription as well. Notable too was that the promoter and both lncRNAs overlap with peaks of H3K4me3 tri-methylation and of peaks of the histone variant H2A.Z, which is associated with regions of genome fluidity in cells [10].

In conclusion, in two closely related genetic models, we show that longevity-associated genotypes of MAP3K5 are associated with virtually complete mitigation of the lifespan shortening effect of having a CMD. Since lifespan was the same irrespective of genotype in men without a CMD, the overall association of genetic variation in MAP3K5 with longevity is contributed entirely by allele-dependent amelioration of the increased mortality risk conferred by CMD. This finding adds to our recent findings of a similar explanation for the longevity association conferred by particular SNPs in FOXO3 [17] and PIK3R5 [18]. The effect does not, however, explain the longevity association we have found for most other genes in our cohort of elderly men [7, 19] (unpublished).

MATERIALS AND METHODS

Study participants

See Supplementary Methods and Supplementary Table 1.

Genotyping

Genotyping methods were as described previously [7] (Supplementary Methods).

Variant search

Variants surrounding *rs2076260* were screened on the RegulomeDB site, which includes known and predicted regulatory elements in the intergenic regions, as well as regions of DNAase hypersensitivity, binding sites for transcription factors, and promoter regions. Sources of these data included public datasets from GEO, the ENCODE project, and published literature [20]. Chromosome 6 locations used the GRCh37.p13 genome build (http://www.gencodegenes.org/releases/19.html).

We also screened the variants using HaploReg, which is a tool for exploring annotations of the noncoding genome at variants on haplotype blocks, such as candidate regulatory SNPs at disease-associated loci [21]. Using LD information from the 1000 Genomes Project, linked SNPs can be visualized along with chromatin state and protein binding annotation from the Roadmap Epigenomics and ENCODE projects, the effect of SNPs on regulatory motifs, and the effect of SNPs on expression from QTL studies. We searched HaploReg version 4.1 for the region between SNPs in LD, *rs9494547* and *rs9494552*, Nov 5, 2015 build, hg38.

Statistical analyses

General linear models were used to compare age-adjusted indirect measurements between groups, and logistic models were used to compare the age-adjusted direct measurements. Cox proportional models were used to assess the association of *MAP3K5* for various genetic models on mortality stratified by disease status, such as by diabetes, by hypertension, by CHD, and by any of CHD, diabetes, or hypertension. The Cox proportional hazard assumption was tested for each Cox model. The effect of interaction of disease with *MAP3K5* genotype on mortality was tested in the Cox model. All statistical analyses were performed using the Statistical Analysis System version 9.4 [22]. Figures were generated using STATA 12 Graphics [23].

AUTHOR CONTRIBUTIONS

B.J.M., R.C., T.A.D., K.H.M., D.C.W., R.C.A., B.J.W. contributed to the study concept and design; R.C. carried out the statistical analyses; K.H.M. supervised recruitment and data collection; T.A.D. supervised the genotyping; B.J.M. drafted the manuscript; R.C., B.J.M., T.A.D., K.H.M., R.C.A., D.C.W. and B.J.W. provided critical input into data interpretation and manuscript preparation.

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CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

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SUPPLEMENTARY MATERIALS

Supplementary Materials and Methods

Study cohort

The present case-control study was conducted as part of the Kuakini Hawaii Lifespan Study and the Kuakini Hawaii Healthspan Study, an embedded cohort study of healthy aging drawn from the original population of the Kuakini Honolulu Heart Program (KHHP) and Honolulu-Asia Aging Study (HAAS) [1, 2]. As such the current study is a case-control sub-study of the KHHP population-based, prospective study of cardiovascular disease among 8,006 Japanese American men that began in 1965. The KHHP participants were recruited during 1965-1968 from 9,877 men aged 45-68 years who had valid contact information from World War II Selective Service records, were born from 1900–1919. and who were living on the island of Oahu in 1965 [3]. Study participants had parents who were both from Japan, mostly the western, central and southern regions [3, 4]. Although 88% of participants were born in Hawaii, there is a theoretical possibility of confounding of case vs. control status for allele frequencies due to geographic origin. Therefore, for certain analyses, cases and controls were stratified by parental prefecture of origin using conditional logistic regression models. Analyses showed no evidence of population stratification in the dataset (data not shown). Subjects were recruited at the same time and place (Oahu) and case and control status only became evident after death or attainment of \geq 95 years of age, meaning there was no apparent reason why genetic background should be different. The KHHP cohort has been described elsewhere [4] and is quite robust for phenotypegenotype associations since the data collection are exceptionally accurate, involving cross validation utilizing an expert Morbidity and Mortality Committee. The Hawaiian Japanese population is from a limited geographic area in Japan, with little outbreeding and, based on the authors' unpublished data, exhibits a smaller degree of genetic diversity than the overall population of Japan.

All participants in the current study were drawn from records of study participants updated to February 2012. Archived phenotypic data and blood samples from Examination 4 of the KHHP (1991–1993), which coincided with the commencement of the HAAS, were used as the baseline for our prior case-control studies. The HAAS was begun as an expansion of the KHHP for the study of neurodegenerative diseases, cognitive function, and other aging phenotypes in elderly persons. Participants included 3,741 men aged 71 to 93 at Examination 4 (mean age 77.9 \pm 4.7 SD years),

approximately half the number of the original KHHP [5], representing 80% of survivors of the original cohort. Our genetics studies have used a quasi-nested casecontrol design and subjects drawn from the KHHP/HAAS population were as described previously [6]. Subjects were followed with regular examinations and blood work until the present, or death. In our prior case-control studies, "cases" (longevity phenotype) were defined as individuals who had survived to at least 95 years of age. As of February 2012, these were in the upper 1% of the 2010 U.S. birth cohort-specific survival [7]. Of those 440 men, 317 had died by February 2012 (mean age at death 97.0 ± 2.1 SD years; range 95-106years) and 123 individuals were still alive (mean age 96.8±1.8 SD years; range 95–107 years). Controls were 374 men of average lifespan randomly selected from KHHP/HAAS cohort as individuals who had died up to the age of 81 years. Mean age at death, 78.1±1.8 SD years (range 72-81 years), accorded with the 3.5 year longer life expectancy of Japanese American men living in Hawaii [8] than the average 1910 U.S. birth cohortspecific survival for middle-aged US men [7].

Procedures performed were in accord with institutional guidelines and were approved by the Institutional Review Board of Kuakini Medical Center. Written informed consent was obtained from all study participants or from family representatives, if participants could not provide consent.

An extensive number of parameters were measured, including hypertension status, total plasma cholesterol level, diabetes, body mass index, cigarette smoking status, alcohol intake, physical activity, and various other parameters. Hypertension was defined as a systolic or diastolic blood pressure of ≥ 160 and ≥ 90 mm Hg, respectively, or was based on the use of antihypertensive medication. То be considered normotensive, systolic and diastolic blood pressures needed to be < 140 and < 90 mm Hg, respectively. Men who were neither normotensive nor hypertensive were classified as having borderline hypertension. Study participants were also classified as having diabetes on the basis of a medical history (physician diagnosed or based on the reported use of insulin or the receipt of oral hypoglycemic therapy). Assessment of overall metabolic output during a typical 24-hour period was based on the use of a physical activity index. The physical activity index was derived by summing the average number of hours per day spent in five different activity levels (basal, sedentary, slight, moderate, and heavy) after each was multiplied by a weighting factor that corresponded to the level of exertion needed to undertake the activity. High levels of the physical

activity index indicate active lifestyles, and low levels indicate inactive lifestyles [9, 10].

Genotyping

The original case-control study (814 subjects) was performed on total leukocyte DNA isolated using the PureGene system (Gentra Systems, Inc.) and quantified using PicoGreen staining (Molecular Probes, Eugene, OR). Tagging SNPs (tSNPs) were genotyped at the University of Hawaii Cancer Center on the Illumina GoldenGate platform (high-throughput genotyping on universal bead arrays). SNP Genotyping of DNA from men with a CMD and men without a CMD was performed on the same platform. The longevity study included 2,900 men who were genotyped using TaqMan[®] reagents (purchased from Applied Biosystems, Thermo Fisher Scientific) for PCR amplification under standard conditions with AmpliTaq Gold[®] DNA polymerase (Perkin-Elmer Corp.). PCR products were detected by TaqMan[®] assay, using a 6-FAM-labeled FRET probe for one allele and a VIC-labeled probe for the other allele, with minor groove binding (MGB) quenchers to enhance assay signal. PCR products were measured using a QuantStudio 12K Flex system. Genotype data were managed through an integrated database sample management-data processing system of proven accuracy. All positive controls on each genotyping plate were evaluated for consistency. SNP call rates exceeded 98%.

We implemented a series of quality control checks based on the Illumina metrics. For inclusion of data for a SNP its call rate had to exceed 0.95 and the Hardy-Weinberg equilibrium *p*-value needed to be >0.01.

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Supplementary Figures



Supplementary Figure 1. Tissue expression pattern of MAP3K5. This gene is expressed at high levels in most tissues, most notably in adrenal, artery, ovary, and pituitary. A value of 50 TPM (transcripts per million) is arbitrarily chosen as cut-off for significance. TPM is a measurement of the proportion of transcripts in the pool of RNA, as measured by RNA-seq.



Supplementary Figure 2. Heatmap of the locations of the longevity-associated SNPs in our original study [11]. Also shown is the location of the differentially methylated site reported by Horsburgh et al. [12] (asterisk), and the location of two long non-coding RNAs (IncRNA), *MAP3K5-AS1* (previously *LOC101928461*) within *MAP3K5* (dotted box), and *LOC101928429* (small solid box near promoter and asterisk). Both IncRNAs are transcribed in the direction opposite (left to right) to that of *MAP3K5* and are referred to as anti-sense RNAs. They may act to interfere with transcription. *MAP3K5* is transcribed from right to left (3' to 5'). *MAP3K5-AS1* overlaps with the block of LD. *MAP3K5-AS1* is in LD with the longevity-associated SNPs we identified previously in *MAP3K5*. Little is, however, known about the IncRNA; the precise location is close to but not exactly the same as the differentially spliced isoform of *MAP3K5*. The LD map for the Japanese population (JPT) is shown in on the left, while that for the Caucasian population (CEU) is shown on the right.

rs2237269 GTF2I



logo: DGRBKKAGG maj: GTGAGGTAGGCC min: GTGAGCTAGGCC

rs2076260 DMRT1

DIVINTI

No logo available

logo: RMWACAWTGTWDCMR maj: TTAAATTGTCTCTAC min: TCAAATTGTCTCTAC

HOXD10



logo: VDBNYMATWAAA maj: TATATTAAATTGT min: TATATCAAATTGT

Pou2f2



logo: VTWTKMAWAWHBND maj: TATATTAAATTGT min: TATATCAAATTGT

TATA CTATAAAAAA logo: WWAWWHDN maj: TATATTAAATTGT min: TATATCAAATTGT

rs6906753 ATF3

logo: BVTGAMKTCA maj: CCCGACGTTA min: CCTGACGTTA



logo: BVTGAMKTCA maj: CCCGACGTTA min: CCTGACGTTA

rs1011969 MEF2



logo: YTWAAWATARCH maj: CTTTAAACAGCCA min: CTTTAAAAAGCCA

TATA



logo: NSYWTAAAAR maj: CTTTAAACAGCCA min: CTTTAAAAAAGCCA

Rs2272887



Supplementary Figure 3. Transcription factor binding sites modified by SNP *rs2076260. rs2076260* and its immediate neighboring SNP, *rs6904753*, are in perfect LD. SNP *rs2076260* is predicted to modify the binding of three transcription factors, HOXD10, Pou2f2, and TATA. The major allele is predicted to create/increase binding of HOXD10, abolish/reduce binding of Pou2f2, and create/increase binding of TATA. For *rs6906753* the major allele would reduce/abolish binding of ATF3 and HEY1 transcription factors. Red rectangles denote the variant SNP nucleotide in the transcription factor canonical sequence. (Abbreviations: maj, major allele; min, minor allele; logo is the canonical recognition site for each. Nucleotide ambiguity codes are: B, not A; V, not T; M, C or A; K, T or G; W, A or T).



Supplementary Figure 4. Possible functional relationships between *rs2076260* and regulatory features in *MAP3K5*. We postulate that *rs2076260* interacts with other *cis*-regulatory elements. Shown by asterisks are the locations of the most significant SNP (*rs2076260*) in *MAP3K5* relative to two long non-coding RNAs, *MAP3K5-AS1* and *LOC101928429*, and the differentially methylated site associated with BMI, cg21506299. The anti-sense lncRNAs as well as SNP *rs2076260* and the promoter proper overlap with peaks of H3K4me4 (green) (see review by Morris et al. [13]), which is commonly associated with sites of activation of transcription of nearby genes, the histone variant H2A.Z (blue), which is associated with regions of genome fluidity (see review Morris et al. [13]). GM12878 (lymphocyte) experiments were chosen for reference. These locations are connected via RNA polymerase II (RNAP0III) and CTCF chromatin looping, indicating that they are likely co-regulated during transcription. *MAP3K5* is transcribed from right to left (purple herring bone). GM12878 (lymphocyte) experiments were chosen for reference. The locations of super-enhancers are shown by solid bars at top.

Supplementary Tables

Supplementary Table 1. Characteristics of cases (n=440) and controls (n=374), ageadjusted (except age and year of birth) used in our prior case-control studies.

Late life examination characteristics	Controls	Cases	р	
Age at examination, y	74.7 ± 2.1	81.5 ± 4.7	2.1E-107*	
Birth year	1916.8 ± 2.0	1909.9 ± 4.7	1.9E–111*	
Anthropometric and physiological				
Height, cm	161.2 ± 5.6	161.3 ± 5.5	0.96	
Weight, kg	59.6 ± 9.6	61.6 ± 8.0	0.020	
Waist to hip ratio	0.95 ± 0.06	0.94 ± 0.05	0.015	
BMI, kg/m ²	23.0 ± 3.1	23.6 ± 2.9	0.040	
Triceps skinfold thickness, mm	10.5 ± 4.1	10.2 ± 4.1	0.40	
Subscapular skinfold thickness, mm	15.5 ± 6.7	16.5 ± 5.5	0.09	
Best forced expiratory volume	1.9 ± 0.49	2.1 ± 0.41	1.5E-06*	
in 1 s, L				
Grip strength, kg	26.9 ± 7.1	31.7 ± 5.2	1.2E-15*	
Blood pressure, systolic, mmHg	151.0 ± 26.3	150.4 ± 22.4	0.079	
Blood pressure, diastolic, mmHg	79.0 ± 12.7	81.6 ± 11.6	0.022	
Ankle-brachial index	1.0 ± 0.20	1.1 ± 0.14	2.1E-14*	
Cognitive (CASI) score	75.6 ± 19.8	87.1 ± 10.6	2.2E-5*	
Hematological and biochemical				
Total cholesterol, mg/dL	183 + 34.2	195 + 32.6	0.00034*	
HDL cholesterol, mg/dL	51.3 + 14.4	51.3+13.1	0.97	
Triglycerides mg/dL	1465 ± 1193	152.3 + 78.7	0.54	
Fasting plasma glucose mg/dL	118.5 ± 36.8	102.0 ± 70.7 108.6 ± 21.5	0.00054*	
Easting plasma insulin mIU/dI	26.2 ± 84.3	13.1 ± 9.5	0.00034	
Plasma fibringen mg/dI	20.2 ± 04.3 331 ± 75.8	13.1 ± 9.3 291 + 52 7	1 5F_10*	
White blood cell count $10^3/\mu$ I	6.98 ± 2.09	5.87 ± 1.50	1.5E 10 1.3E_10*	
Health habits	0.96 ± 2.09	5.67 ± 1.50	1.5E-10	
Current smoker %	13.6	4.0	0.00035*	
Past smoker, %	60.0	4.0	0.00055	
Smaking pack years	00.0	40.3	2 OF 0*	
Alashal approximation approximate	37.2 ± 33.6	17.4 ± 30.3 14.4 ± 20.2	2.9E-9*	
Alconol consumption, ounces/month	29.4 ± 33.0	14.4 ± 29.2	0.00055* 2.5E 5*	
Physical activity index,	29.9 ± 4.0	51.7 ± 4.4	5.3E-3*	
metabolic work/day	20.0		1 05 17*	
Difficulty in walking 0.8 km, %	39.8	6.6	1.0E-1/*	
On diabetes medication, %	18.1	6.9	0.00026*	
Diseases		7 0 0	0.24	
Hypertension, %	74.1	78.8	0.24	
Coronary heart disease, %	26.7	14.2	0.00085	
Stroke history, %	9.3	1.2	7.6E–5*	
Cancer, %	23.8	10.0	8.3E–5*	
Diabetes, %	35.0	21.7	0.00011*	
Emphysema, %	4.8	1.4	0.038	
Bypass history, %	11.9	3.5	0.05	
Angioplasty, %	4.1	1.4	0.37	
Surgery on arteries of the neck, %	1.4	0.69	0.41	
Aorta surgery, %	8.4	0.92	8.7E–7*	
Ankle-brachial index < 0.9. %	24.8	3.7	6.9E–11*	
Sociodemographic				
Education, years	10.2 ± 2.8	10.5 ± 3.5	0.30	
Married, %	75.5	86.9	0.002	

The data shown are age-adjusted. *Indicates that difference remained significant after the p value shown was adjusted by the Bonferroni method for the 41 parameters tested.

Supplementary Table 2. The SNPs genotyped in *MAP3K5* and the minor allele frequency of each in control American men of Japanese ancestry in the Kuakini Honolulu Heart Program, and of Japanese subjects in the dbSNP database.

SNP	КННР	dSNP
rs1570055	0.479	0.488
rs1570054	0.445	0.465
rs12529435	0.181	0.133
rs2327742	0.546	0.488
rs9376222	0.103	0.111
rs7775356	0.297	0.278
rs6905203	0.433	0.442
rs877192	0.256	0.193
rs6910459	0.238	0.221
rs1997703	0.418	0.407
rs2076260*	0.455	0.458
rs12164028	0.345	0.343
rs17723504	0.696	0.200
rs9376207	0.233	0.144
rs6904753*	0.467	0.477
rs9385770	0.479	0.447
rs17796708	0.252	0.244
rs4631311	0.383	0.378
rs6916746	0.118	0.110
rs2272887	0.483	0.477
rs9494547	0.171	0.169
rs13203080	0.241	0.221
rs2237264	0.172	0.148
rs9376211	0.173	0.278
rs13195420	0.189	0.140
rs3765259	0.441	0.401
rs9402838	0.270	0.364
rs9376219	0.183	0.134
rs9494552	0.284	0.273
rs9321564	0.480	0.424
rs4363056	0.409	0.471
rs17723638	0.189	0.156
rs9376221	0.103	0.099

*Denotes those SNPs in a cluster associated with longevity using a dominant model. A cluster was defined as SNPs in close physical proximity and/or belonging to the same block of linkage disequilibrium (LD).

Supplementary Table 3. The two *MAP3K5* SNPs in the case-control study that were associated with longevity after Bonferroni correction and results for the four genetic models used.

SNP	Estimate	S.E.	OR	p	Model*	C_H	р	R_H	р
rs2076260	-0.079	0.098	0.92	0.42	Additive	_	_	_	_
rs2076260	-0.637	0.151	0.53	<.0001	Het vs other	1.95	0.0001	1.81	0.0018
rs2076260	0.4527	0.159	1.57	0.0043	MM vs other	_	_	_	_
rs2076260	0.2987	0.174	1.35	0.086	RR vs other	_	_	_	_
rs6904753	-0.066	0.098	0.936	0.50	Additive	_	_	_	_
rs6904753	-0.417	0.141	0.659	0.0032	Het vs other	1.57	0.0066	1.45	0.040
rs6904753	0.336	0.157	1.40	0.032	MM vs other	_	_	_	_
rs6904753	0.202	0.170	1.22	0.23	RR vs other	_	_	_	_

OR was estimated using a logistic model with long-lived case as the outcome. If OR was significantly > 1, then the genotype comparison indicated a pro-longevity effect.

*Model:

Additive model: gene effect is coded as common homozygote (CC) = 0, heterozygote (CT) =1, minor allele homozygote (TT) = 2 in a linear logistic model.

Het = heterozygote (*CT*).

MM = common homozygote (CC), and RR = minor allele homozygote (TT).

C_H: OR of common homozygote (*CC*) vs. heterozygote (*CT*) with long-lived case.

R_H: OR of minor allele homozygote (TT) vs. heterozygote (CT) with long-lived case.

Other: The other genotype(s).

S.E. = standard error of the mean.

Characteristics	CC	CT	TT	p
n	1000	1733	783	
Age at examination, yr	77.8 ± 4.6	77.6 ± 4.6	77.8 ± 4.7	0.48
Birth year	13.6 ± 4.6	13.8 ± 4.6	13.7 ± 4.7	0.48
Anthropometric and physiological				
Height, cm	161.9 ± 5.7	161.7 ± 5.5	161.6 ± 5.9	0.66
Weight, kg	61.7 ± 9.4	61.2 ± 8.9	61.4 ± 9	0.35
Waist to hip ratio	0.9 ± 0.1	0.9 ± 0.1	0.9 ± 0.1	0.98
BMI, kg/m ²	23.6 ± 3.2	23.4 ± 3	23.5 ± 3.1	0.31
Triceps skinfold thickness, mm	10.1 ± 4	10.1 ± 3.9	10.1 ± 4	0.90
Subscapular skinfold thickness, mm	16.3 ± 6.1	16 ± 5.9	16.4 ± 6.5	0.36
Best forced expiratory volume	2.1 ± 0.5	2.1 ± 0.4	2.1 ± 0.5	0.58
Grip strength, kg	30.4 ± 5.8	30.1 ± 6.2	30.2 ± 6.2	0.55
Blood pressure, systolic, mmHg	148.9 ± 23.9	149.5 ± 23.5	149.5 ± 22.8	0.83
Blood pressure, diastolic, mmHg	79.9 ± 11.3	79.8 ± 11.6	80.3 ± 10.7	0.60
Cognitive (CASI) score	83.3 ± 13.9	82.2 ± 15.3	82.9 ± 13.4	0.14
Hematological and biochemical				
Total cholesterol, mg/dL	191.1 ± 32.2	188.9 ± 32.8	190.3 ± 33.3	0.24
HDL cholesterol, mg/dL	50.7 ± 13.6	51.1 ± 13.1	51 ± 13.5	0.76
Triglycerides, mg/dL	151.2 ± 97.7	147.7 ± 91.5	149.3 ± 95.2	0.63
Fasting plasma glucose, mg/dL	113.9 ± 29.7	113.2 ± 30.4	111.5 ± 27	0.24
Fasting plasma insulin, mIU/dL	15.9 ± 14.7	15.4 ± 13.1	15.3 ± 11.7	0.56
Plasma fibrinogen, mg/dL	305.8 ± 65.8	306.9 ± 63.3	308.4 ± 63.3	0.71
White blood cell count, $10^3/\mu L$	6.3 ± 2.5	6.3 ± 1.7	6.3 ± 2.4	0.96
Health habits				
Current smoker, %	6.5	7	7.3	0.80
Past smoker, %	55.1	56.3	53	0.33
Smoking, pack-years	26.1 ± 34.4	26.9 ± 34.2	23.6 ± 33	0.10
Alcohol consumption, ounces/month	18.7 ± 39.1	18.8 ± 39.9	18.9 ± 43	1.00
Physical activity index, metabolic work/day	31 ± 4.7	30.8 ± 4.5	30.9 ± 4.7	0.55
Difficulty in walking 0.8 km, %	19.4	18.1	18.3	0.65
On diabetes medication, %	11.4	11.6	10.1	0.51
Diseases				
Hypertension (160/95), %	54.2	53.2	52.9	0.83
Coronary heart disease, %	20	20.6	21.3	0.79
Stroke history, %	3.6	4.9	3.9	0.25
Cancer. %	11.7	13.7	14.9	0.13
Diabetes. %	28.9	28.3	28.6	0.94
Emphysema. %	2.7	3.2	2.2	0.40
Bypass history, %	7.6	7.7	5.8	0.19
Angioplasty, %	6	7.3	6.6	0.40
Surgery on arteries of the neck. %	1.3	1.1	0.9	0.74
Aorta surgery. %	3	3.7	2.8	0.40
Ankle-brachial index < 0.9 . %	12.3	12.6	12.5	0.96
Sociodemographic	12.5	12.0	12.0	0.20
Source and a second sec				
Education, years	106+31	105 + 31	106+32	0 53

Supplementary Table 4. Characteristics of all subjects at baseline	e (1991–1993) by MAP3K5 rs2076260
genotype.	

Variable		With CM	D*		Without CMD				
variable	СС	СТ	TT	<i>p**</i>	CC	СТ	TT	р	
n	701	1212	548		299	521	235	-	
Age (years)	77.9 ± 4.6	77.6 ± 4.5	77.9 ± 4.6	0.35	77.7 ± 4.5	77.6 ± 4.7	77.5 ± 4.7	0.82	
BMI (kg/m²)	23.8 ± 3.2	23.6 ± 3	23.7 ± 3.1	0.61	23 ± 3	22.7 ± 3	23.1 ± 3.1	0.20	
Waist to hip ratio	0.95 ± 0.06	0.95 ± 0.06	0.95 ± 0.06	0.60	0.94 ± 0.06	0.93 ± 0.06	0.94 ± 0.05	0.34	
Fasting plasma glucose (mg/dL)	119.2 ± 33.6	118.3 ± 34.7	116 ± 30.8	0.20	101.3 ± 8.4	101.2 ± 8.7	101.5 ± 8.5	0.92	
Fasting plasma insulin (mIU/dL)	20.1 ± 49.6	17.9 ± 24.3	17.7 ± 29.1	0.33	12.1 ± 6.8	12 ± 7	12.5 ± 6.8	0.62	
Plasma fibrinogen (mg/dL)	309.2 ± 65.7	309.3 ± 63.9	314.1 ± 66.5	0.32	297.8 ± 65.4	301.5 ± 61.5	295.1 ± 53	0.38	
White blood cell count $(10^3/\mu L)$	6.4 ± 2.8	6.4 ± 1.7	6.4 ± 2.6	0.96	5.9 ± 1.7	6 ± 1.7	5.9 ± 1.6	0.81	
Smoking (pack-years)	27.5 ± 35.6	27.3 ± 34.2	24.6 ± 34.3	0.28	22.8 ± 31.1	26.0 ± 34.1	21.5 ± 29.8	0.18	
Alcohol intake (oz/mo)	19.2 ± 39.8	20.8 ± 44.9	18.9 ± 43.6	0.64	17.7 ± 37.4	14.3 ± 24.3	18.8 ± 41.5	0.18	
Physical activity index	30.9 ± 4.7	30.7 ± 4.5	30.8 ± 4.7	0.60	31.1 ± 4.5	31.0 ± 4.4	31.1 ± 4.6	0.91	
Depression (%)	9.1	10	12.4	0.18	11.8	10.5	10.1	0.82	
Stroke (%)	4.5	5.6	4.7	0.51	1.7	3.1	2.2	0.39	
Cancer (%)	12.8	12.8	15.3	0.31	9.2	15.9	14	0.026	
Diabetes (%)	41.2	40.4	41.0	0.94	-	-	_	_	
Hypertension (%)	77.3	76	75.5	0.74	-	-	_	_	
CHD (%)	99.9	99.9	100	0.120	_	_	-	_	

Supplementary Table 5. Age-adjusted baseline variables by cardiometabolic disease (CMD)* status and *MAP3K5 rs2076260* genotype.

Values shown are mean ± SD for indirect measures and proportion (%) for direct measurements.

*CMD: diabetes or hypertension or CHD.

***p* from Fishers F-test.

Supplementary Table 6. Hazard ratios (HR) by genotype of *MAP3K5* SNP *rs2076260* with total mortality in men with diabetes, CHD, hypertension, and any of these CMDs in two other genetic models besides the two models shown in Table 1 of the main article.

Discondon (n mith 40401)		Constinue dol**	With a CM	ID	Without a Cl	MD
Disorder (n with, total)	Cox model*	Genetic model***	HR (95% CI)	р	HR (95% CI)	р
Diabetes	1	CC/CT vs. TT	1.11 (0.96-1.29)	0.17	1.06 (0.97-1.17)	0.2091
(990, 2478)	2	CC/CT vs. TT	1.08 (0.92-1.27)	0.34	1.05 (0.94-1.16)	0.4160
Hypertension	1	CC/CT vs. TT	1.03 (0.93-1.15)	0.56	1.12 (1.00-1.26)	0.0515
(1877, 1639)	2	CC/CT vs. TT	1.02 (0.90-1.15)	0.79	1.08 (0.95-1.23)	0.2517
CHD	1	CC/CT vs. TT	1.19 (1.00-1.42)	0.050	1.06 (0.97-1.16)	0.1866
(724, 2792)	2	CC/CT vs. TT	1.19 (0.98-1.45)	0.071	1.02 (0.92-1.13)	0.7077
Any CMD	1	CC/CT vs. TT	1.06 (0.96-1.16)	0.25	1.13 (0.97-1.31)	0.11
(2461, 1055)	2	CC/CT vs. TT	1.03 (0.92-1.14)	0.63	1.10 (0.93-1.30)	0.26
Diabetes	1	Additive	1.03 (0.95-1.12)	0.48	1.02 (0.97-1.08)	0.43
(990, 2478)	2	Additive	1.04 (0.95-1.14)	0.35	1.02 (0.96-1.08)	0.53
Hypertension	1	Additive	1.06 (0.99-1.12)	0.087	0.98 (0.92-1.05)	0.65
(1877, 1639)	2	Additive	1.08 (1.00-1.15)	0.036	0.98 (0.91-1.06)	0.61
CHD	1	Additive	1.01 (0.92-1.11)	0.82	1.02 (0.97-1.07)	0.45
(724, 2792)	2	Additive	1.00 (0.90-1.11)	0.98	1.04 (0.98-1.10)	0.20
Any CMD	1	Additive	1.05 (1.00-1.11)	0.065	0.95 (0.88-1.04)	0.28
(2461, 1055)	2	Additive	1.08 (1.01-1.14)	0.015	0.93 (0.84-1.02)	0.14

*Cox models: Model 1: Age-adjusted. Model 2: Covariate-adjusted, where covariates adjusted in Cox model were: age, BMI, glucose, smoking (pack-year), alcohol intake (oz/mo), physical activity index, depression, cancer, and stroke.

**Genetic models: *Top half:* Minor allele homozygote model. *Bottom half:* Additive model, in which the genetic variables were coded as 0 for genotype *CC*, 1 for *CT* and 2 for *TT*, so that HR represents the additive effect of genotype on mortality.

Supplementary Table 7. Variants in near perfect linkage disequilibrium (LD) with *rs2076260*, and additional information for each.

chr	pos (hg19)	$LD(r^2)$	LD (D')	Variant	Ref/ Alt	ASN freq	EUR freq	Promoter histone marks	Enhancer histone marks	DNAse	Motifs changed
6	136966619	0.81	0.98	rs2237269	G/C	0.37	0.16			HRT, BLD	4 altered motifs
6	136966741	0.93	0.98	rs2237270	T/C	0.37	0.17			-	
6	136968541	0.94	0.99	rs2237271	A/C	0.45	0.17	6 tissues	17 tissues	20 tissues	
6	136969853	0.97	1.00	rs1997703	C/T	0.45	0.18		7 tissues	-	
6	136974283	0.97	0.99	rs6904753	C/T	0.55	0.18		ADRL	-	4 altered motifs
6	136977568	1.00	1.00	rs2076260	T/C	0.56	0.18		6 tissues	MUS, BRN	6 altered motifs
6	136979240	0.84	1.00	rs1011969	C/A	0.55	0.16			GI	Mef2, TATA
6	136980134	0.83	0.99	rs2272887	A/G	0.48	0.16		ESDR, BRN, MUS	5 tissues	Mtf1

All positions are based on genome release GRCh37 released Feb 2009 (UCSC equivalent = hg19). *Abbreviations:* ADRL, adrenal; ESDR, embryonic stem cell derived; BLD, blood; BRN, brain; GI, gastrointestinal; HRT, heart; MUS, muscle.

Supplementary	Table	8.	Summary	of	nucleotide	sequences	and	putative	functional
features for MA	РЗК5.								

Feature	Location (hg19)	Definition
MAP3K5	136,977,568-137,105,225	Gene (transcribed right to left, i.e., 3'-5')
MAP3K5-AS1	136,950,252–136,969,336	LncRNA (transcribed left to right, 5'-3')
rs2076260	136,977,568	Longevity-associated SNP, this study
rs6904753	136,974,283	Neighboring functional SNP
LOC101928429	137,105,185-137,107,192	LncRNA (transcribed left to right, 5'-3')
cg21506299	137,105,225	Site differentially methylated with BMI

*hg19, the genome build describing where features are located. The actual sequences from NCBI/UCSC/Ensembl are identical to these, but their annotations will be different and updated at different frequencies.

Supplementary	/ Table 9.	Putative f	unctional	variants a	nd their	effects or	n transcri	ption of	the common	allele.
Supplementary		I acacive i	anctional	variance a	ind then	chiccus of	i ci anisci i			ancie.

SNP	Result (minor allele)	TF	Biological pathways	Tissue expression*
rs2237269	abolish	GTF2I	Activation of immunoglobulin heavy-chain transcription upon B-lymphocyte activation	Most tissues
rs6904753	create	HEY1	Repressive, induced by the Notch and c-Jun, repressive for GATA4 and GATA6	Lung
	create	ATF3	Unfolded protein response, stress response activating or repressive	Lung, thyroid
rs2076260	create	DMRT1/7	Sex-determining pathway	< 100 FPKM
	abolish	HOXD10	Developmental regulatory system for anterior-posterior axis.	< 100 FPKM
	abolish	POU2F2	Immunoglobulin gene promoter,	< 100 FPKM
	abolish	TATA	RNA polymerase II initiation	< 100 FPKM
rs1011969	create	MEF2	Activates many muscle-specific, growth factor-induced, and stress-induced genes	Most tissues
	create	TATA	Essential for progression of the G1 phase of the cell cycle	Most tissues
rs2272887	create	MTF1	Induces expression of metallothioneins in response to exposure to heavy metals	Most tissues

Data from HaploReg, v4.1. Biological pathway prediction is from GeneCards. Tissue expression patterns are from GTEx. *< 100 FPKM, fragments per kilobase of transcript per million mapped reads (indicates low level of expression).

Supplementary Table 10. Super-enhancers overlapping *MAP3K5*, and their chromosomal location, size and tissue.

ID	Chrom	Start	End	Size	Associated gene	Method	Rank	Cell/tissue
SE_32506	chr6	137070606	137115431	44825	MAP3K5	H3K27ac	52	GM12878
SE_59682	chr6	137063070	137114568	51498	MAP3K5	H3K27ac	79	Ly4
SE_09572	chr6	137071400	137114901	43501	MAP3K5	H3K27ac	436	CD14
SE_11351	chr6	137070304	137115433	45129	MAP3K5	H3K27ac	508	CD20
SE_10763	chr6	137071728	137113378	41650	MAP3K5	H3K27ac	608	CD19 Primary
SE_19826	chr6	137071508	137115095	43587	MAP3K5	H3K27ac	728	CD4p CD25- Il17p PMAstim Th17

Data are from dbSUPER, (<u>https://asntech.org/dbsuper/index.php</u>) positions are according to GRCh37/hg19. ID refers to dbSUPER identification number. For descriptions of methods and rank see Hnisz et al. [14].