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CASE REPORT

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Gastric and colon metastasis from breast cancer: case report, review of the literature, and possible underlying mechanisms

J Carlos Villa Guzmán¹ J Espinosa¹ R Cervera¹ M Delgado² R Patón³ JM Cordero García⁴

¹Department of Medical Oncology, ²Department of Pathology, ³Department of Gastroenterology, ⁴Department of Nuclear Medicine, University Ciudad Real General Hospital, Ciudad Real, Spain

Correspondence: J Carlos Villa Guzmán Department of Medical Oncology, University Ciudad Real General Hospital, Obispo Rafael Torjia St, 13002 Ciudad Real, Spain Tel +34 I 696 658 482 Fax +34 926 27 85 79 Email jvillaguzman1@yahoo.es

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Abstract: Gastrointestinal metastases from breast cancer are not common. We present a 58-yearold female diagnosed with lobular breast cancer some years before whose relapses were gastric and colonic mucosal. Simultaneous metastases are extremely rare. To our knowledge, no cases of initial dual affectation have been reported. The patient also showed gastritis by *Helicobacter pylori*. Invasive lobular breast carcinoma is the most frequent special type of breast cancer and carries some specific molecular alterations such as loss of expression of E-cadherin. Although underlying mechanisms of metastasization are not entirely known, chemokines as well as inflammatory events seem to be implicated in this process. Interaction between chemokines and their receptors frequently induces cell migration. We hypothesize that *H. pylori*, inflammatory cells, and chemokines may create a favorable environment attracting tumor cells. **Keywords:** chemokines, gastrointestinal metastases, *H. pylori*, lobular carcinoma

Introduction

Breast cancer is the most frequent malignancy among women. Approximately 16,000 new cases are diagnosed each year in Spain. The 5-year survival rate is constantly increasing, which implies a growing prevalence in almost all developed countries. Modern adjuvant treatment regimens have yielded a reduction in relapses and higher cure rates.

Relapses have been reported in 30%–80% of patients in spite of surgery, chemotherapy, and radiotherapy, but are rare in early-stage breast cancer.¹ The most common sites of distant metastases are in bone, liver, lung, and brain.

Lobular carcinoma is less likely to involve the gastrointestinal tract. The incidence of extrahepatic gastrointestinal tract metastases observed in autopsy studies varies from 4% to 18%, with the most commonly affected organ being the stomach, followed by colon and rectum.^{2,3} Nevertheless, coexisting solitary metastases to both stomach and colon are extremely rare.

Case report

A 58-year-old female smoker was diagnosed with pulmonary thromboembolism during her first pregnancy, and hypothyroidism and hypertension twenty years ago. She underwent quadrantectomy and lymphadenectomy in May 2006. Pathologic analysis from breast tissue showed a 2.5 cm invasive lobular carcinoma (ILC), grade III, and 20 of 25 axillary nodes were affected. Immunohistochemical results were as follows: estrogen receptor (ER)-positive, progesterone receptor (PR)-positive, Her-2/NEUnegative, p53 and isolated cells stained for Ki67. Body computed tomography and bone scan were negative for advanced disease.

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In June 2009, serum markers CEA and CA15.3 began to rise. Chest X-rays, abdominal ultrasound, bilateral mammography, brain magnetic resonance imaging, and positron emission tomography scan did not reveal metastatic disease. A closer follow-up was done. In January 2010, the patient revealed dyspeptic symptoms such as nausea and epigastric pain. She underwent an esophagogastroscopy that demonstrated a granular mucosa with an adenomatous pattern. In the antrum, prepyloric fold did not modify with blowing, and biopsies were taken from all suspected areas (Figure 1). Pathology revealed mucous gastric metastases from a lobular carcinoma of the breast. Immunohistochemical examination was carried out using primary antibodies manufactured by Dako Denmark A/S (Glostrup, Denmark). Mucous gastric metastases were positive for cytokeratin-19 and hormonal receptors (Figure 2). Of note, there was a chronic gastritis caused by *Helicobacter pylori*. A colonoscopy was performed in February 2009, which showed a polyp that was subsequently excised, and a pathology report showed metastases from a lobular breast cancer, also positive for cytokeratin-7 and hormonal receptors (Figures 3 and 4). Both colonic and gastric biopsies were negative for Erb2/ NEU protein.

Because of this gastrointestinal progression, capecitabine 1,250 mg/m² every 12 hours (14 days, every 21) was initiated. Before treatment, CEA was higher than 100 ng/mL (<2.5)



Figure I Endoscopic image of gastric mucosa with granular mucosa with adenomatous pattern.

and CA15.3 exceeded 200 U/mL (<32.4). After six courses of chemotherapy, CEA decreased to 23 ng/mL, while CA15.3 was elevated near the end of chemotherapy and the patient was asymptomatic. A new esophagogastroscopy showed similar findings. A new positron emission tomography scan revealed a hypermetabolic lesion in the second lumbar vertebra. Pancytopenia developed, so a bone marrow biopsy was performed, revealing metastasis in this location. Therefore, a new line of chemotherapy with paclitaxel–carboplatin– bevacizumab plus zoledronic acid was started. Despite several chemotherapy and hormone therapy lines consisting of cisplatin–gemcitabine, vinorelbine, liposomal doxorubicin, and other aromatase inhibitors, the patient developed brain metastases and finally died in August 2011.

Discussion

Single gastric⁵ and colon¹ metastases and metastases in both locations⁶ from breast cancer have been described, but, to our knowledge, this is the first case of a synchronous spread to stomach and colon as the first and unique initial manifestation of metastatic breast cancer.

The incidence of gastric metastases from breast cancer in long-term series and autopsies ranges between 2% and 18%.⁷ Lobular breast cancer is the predominant histology found in gastrointestinal tract metastases. The affinity of lobular breast cancer cells for this system is well known.^{3,8} Distant spread may occur many years after the initial treatment for breast cancer.⁵ Other rare sites are gynecologic organs⁹ and the peritoneum.¹ Ductal tumors tend to metastasize to the liver, the lung, and the brain,^{5,11} although some cases of intestinal metastases have been reported.^{8,11}

Symptoms from gastrointestinal metastases are inexpressive.^{1,5,7} Serum marker CA15.3 may be elevated, as in our case.¹ The endoscopic pattern usually is confounded with a gastrointestinal cancer. The most frequent pattern of spread is diffuse infiltration.^{12–14} The median time to development of metastases is approximately 4 or 5 years.¹³

Metastases to colon could imitate primary colon cancers,^{1,8} may acquire a similar aspect to linitis,¹⁵ or Crohn's disease,¹ and may even be a solitary adenomatous polyp.¹⁶ Our patient had a solitary metastatic localization to an adenomatous polyp in the colon as well as stomach infiltration. Although breast cancer metastasis in stomach and colon may coexist,⁶ it has never been described as the first manifestation of dissemination.

Treatment of gastrointestinal metastases from breast cancer is under discussion. Systemic therapy is the first option, and surgical treatment is considered in cases with

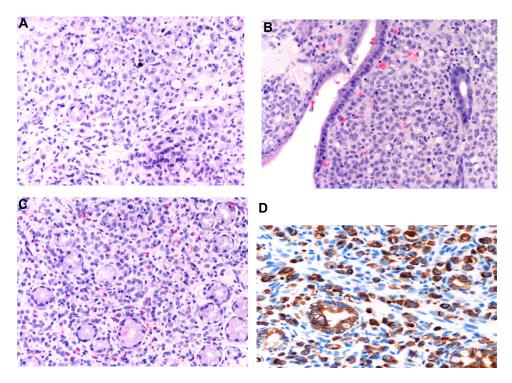


Figure 2 Hematoxylin and eosin staining of the gastric specimen demonstrating strands of invasive cells in the mucosa. Notes: (A) Lobular breast cancer cells individually dispersed in a single-cell-file pattern around gastric glands (10× magnification). (B) Discohesive lobular breast cancer cell pattern in gastric specimen (10× magnification). (C) Gastric mucosa infiltrated by lobular breast cancer cells adopting the typical "Indian file" pattern (10× magnification). (D) Immunohistochemical staining for cytokeratin CK19 demonstrating breast cancer cells in gastric mucosa (20× magnification).

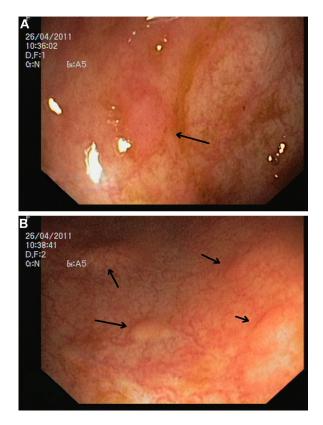


Figure 3 Endoscopic image of colonic mucosa. Notes: (A) Colonic mucosa erosion (arrow). (B) Several polyps in colonic mucosa (arrows).

obstruction or bleeding.³ There is a low response rate to chemotherapy in ILC patients.¹⁷ The median survival rate is 2 years,^{1,3} which implies a poor prognosis, particularly for gastric metastasis.³ Hormonotherapy and chemotherapy were used to treat our patient. Surgical treatment was not necessary.

Imaging and endoscopic findings are not enough to differentiate primary from secondary gastrointestinal lesions.^{3,5} Biopsy and pathology are essential for diagnosis. Deep biopsies must be performed with endoscopy,^{5,7} and histology must be compared with the primary breast cancer.¹⁸ Sometimes, microscopy may differ between primary and metastatic disease, but immunohistochemical analysis may be the only consistent method for differentiating these entities.7 Pathologic criteria include infiltration of the serosal, muscular, and submucosal layers by cells, typically in an "Indian file" pattern,11 resulting in a signet ring appearance.19 ER and PR positivity may suggest breast origin, but these are positive in 32% and 12%, respectively, of patients with primary gastric cancer. However, the expression of ultimategeneration antibodies against ER α is reliable and useful in differentiating gastric metastasis from breast cancer.5 Her-2 extracellular domain (ECD) expression may help in the differential diagnosis of primary gastric cancer vs metastatic

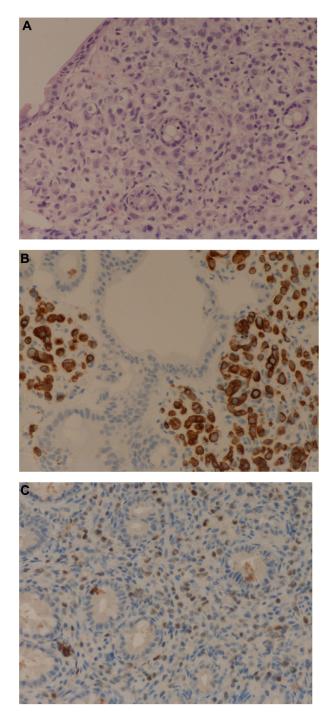


Figure 4 Biopsy of colonic mucosa.

Notes: (**A**) Hematoxylin and eosin staining demonstrating strands of invasive cells in the mucosa (10× magnification). (**B**) Immunohistochemical staining for cytokeratin CK7 demonstrating breast cancer cells in colonic mucosa (20× magnification). (**C**) Immunohistochemical staining for estrogen receptors demonstrating breast cancer cells in colonic mucosa (20× magnification).

breast carcinoma. Absence of ECD is related to lobular breast cancer. Both markers, the presence of ER α and the absence of ECD expression, may reliably distinguish these entities.²⁰ In our case, both specimens from stomach and colon were compared with the primary surgical piece removed some years before and were found to match. Breast lesions were characterized by staining for CK7 or CK19 (Figures 2 and 3) and hormone receptors. Cytoplasmic positivity for gross cystic disease fluid protein-15 (GCDFP-15) is useful in identifying breast cancer metastasis to gastric mucosa,⁵ but, despite its high specificity (90%), its sensitivity is low (50%).³ We did not use this immunohistochemical marker. Positivity for ER, PR, CK7, and CDFP15, as well as negativity for CA19-9, CK20, and CDX2, suggest a breast origin of a gastric lesion,^{3,8,9,11,19} particularly in a patient with a past history of breast cancer. Staining for p53 and Erb2/NEU are usually negative,¹ as in our case.

ILCs are characterized by rather discohesive cells individually dispersed or arranged in a single-cell-file pattern immersed in a fibrous stroma.²¹ This lack of cohesion could be responsible for the development of distant metastases. Most ILCs are of histological grade I/II, express hormone receptors, and do not overexpress HER2 protein like our patient. The pleomorphic variant of ILCs is reported to have an aggressive clinical behavior.

Beta-catenin plays a central role in the ECD/catenin cell-cell adhesion complex, and it may be involved in cellular signaling pathways. Simultaneously, loss of E-cadherin and alpha-, beta-, and gamma-catenin are important steps in the formation of lobular carcinoma in situ, as a precursor of invasive lobular breast cancer. Inactivation of E-cadherin through CDH1 gene mutation, loss of heterozygosity of chromosomal region 16q22.1, and/or gene promoter methylation is an early event reported in more than 95% of ILCs.²² Moreover, loss or downregulation of CDH1 is associated with tumor dedifferentiation and leads to different events: disruption of the tissue architecture, loss of adhesive properties and proliferation-suppressive function and gain in cellular motility increasing of invasive properties and dissemination of epithelial malignant cells. Of interest, ILC has been reported in families with hereditary diffuse gastric cancer that carry germline inactivation mutations of CDH1.23

From an immunohistochemistry point of view, ILCs are not positive for CK20 and express CK7, CK18, thrombospondin (TSP), and integrin alpha-V.³¹ ILC cells also express survivin, cathepsin B, TPI1, SPRY1, SCYA14, TFAP2B, osteopontin, HLA-G, and CHC1.

With regard to genomic considerations, there are some genes overexpressed in ILCs that code proteins involved in cell adhesion like *VWF*, *ELN*, *DPT*, and *EMCN*. There is upregulation of *SFRP1*, *TGFBR2*, and *IGF1* genes. On the other hand, there are genes overexpressed in classic

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and pleomorphic ILCs implicated in actin cytoeskeleton remodeling/signaling and cell adhesion networks such as ANKDR28 and AFF1.²¹

It is worth mentioning the new role of chemokines in mechanisms of metastasization. Chemokines are a super-family of chemotactic cytokines present in organs that act as specific modulators in leukocyte migration to sites of inflammation, but are also involved in the initiation and promotion of carcinogenesis by providing growth and angiogenic factors. There are two families that represent the bulk of known chemokines: CC (the first two cysteines are adjacent to one another) and CXC (the first two cysteines are separated by one amino acid).²⁴

Chemokines interact with cell-surface receptors, present in tumor cells, which are members of a large superfamily of seven transmembrane G-protein-coupled cell surface receptors (GPCRs). Interaction between chemokines and their receptors induces migration of cells and mediates inflammatory and tumor cell migration. These chemokines stimulate certain cells to express their receptors by autocrine and paracrine mechanisms. More than 40 chemokines and 18 receptors are currently known.²⁴

Chemokine receptors CXCR4 and CXCR7 are highly expressed in breast cancer cells, and they are responsible for the chemotaxis to certain target organs such as lymph nodes. CXCL12 (also named SDF-1alpha) and CCL21 (or 6Ckine) bind CXCR4 and CXCR7.²⁵ The CXCL12/CXCR4 pathway is implicated in the mobilization, trafficking, and homing of cancer stem cells into metastatic sites.²⁶

CXCR4 is implicated in vascularization by initiating and sustaining tumor formation. Signaling through chemokine receptors mediates actin polymerization and pseudopodia formation, favoring invasiveness. CXCR4 also stimulates the production of matrix metalloproteases.²⁷

CXCL12 induces synthesis of metalloprotease MT1-MMP,²⁷ modulates integrin expression, and promotes tumor cell adhesion by attaching cells to extracellular matrix proteins of the basement membrane or to ligands on other cells.²⁸ CXCL12 can also regulate tumor cell apoptosis by activating NF- κ B, which in turn inhibits tumor necrosis factor- α (TNF- α) production. Activation of NF- κ B can sensitize cancer cells to CXCL12 stimulation through upregulation of CXCR4 expression. Methylation of CXCL12 promoter in the colonic epithelium favors metastases of tumors in the colon, but further studies are necessary to confirm this hypothesis. Breast tumor cells entering vascular or lymphatic circulation may migrate and adhere mainly to areas expressing CXCL12, therefore migrating to different organs.²⁴

Carcinoma cells recruit normal fibroblasts into tumor masses. Fibroblasts produce growth factors, extracellular matrix molecules, and metalloproteases and secrete soluble factors with proinflammatory and suppressant effects. Fibroblasts activate and turn into carcinoma-associated fibroblasts (CAFs). These cells contribute to tumorigenesis and metastasis and are recruited by transforming growth factor beta (TGF- β), platelet-derived growth factor (PDGF), fibroblast growth factor 2 (FGF2), and vascular endothelial growth factor (VEGF) released by tumor cells. In primary tumor and metastatic breast cancer, CAFs produce CXCL12 that promotes tumor growth and enhances angiogenesis through CXCR4 expressed on carcinoma cells and endothelial progenitor cells.²⁹ Cytokines and chemokines derived from CAFs induce immune suppression and facilitate invasion and transendothelial migration. Interestingly, both stroma and cancer cells can produce CXCL12.

Furthermore, hypoxia induces CXCR4 expression and CXCL12 production, sensitizes tumor cells to CXCL12 signals, and promotes tumor metastasis.^{24,28} CXCL12 attracts vascular endothelial cells and plasmacytoid dendritic cells into the tumor environment, and these cells induce neoan-giogenesis through production of IL-8 and TNF- β . CXCL12 expression seems to be regulated by hypoxia and a hormone-triggered signal pathway. The CXCL12/CXCR4-mediated tumor cell proliferation is regulated by estrogen signaling. In our case, the tumor expressed both ER and PR, which makes this pathway feasible.

It is known that interaction between CCL21 and CCR7 plays a crucial role in lymphocytes homing to secondary lymphoid organs through lymphatic vessels. The mechanism by which CCL21 mediates migration of tumor cells to lymph nodes is similar to the mechanism of the lymphocytes' homing effect. This mechanism may underlie gastric metastasis in *H. pylori* gastritis by acting as an organ with a great inflammatory property similar to lymph nodes in our patient.

Finally, there is a chemokine subgroup growth-related oncogene (GRO): GRO-2 (CXCL-2), GRO-3 (CXCL-3), and interleukin-8 (CXCL-8). These molecules are overexpressed in colonic adenomas and act as angiogenic factors, attract and activate neutrophils, and stimulate endothelial cell migration. GRO-2 (CXCL-2) and GRO-3 (CXCL-3) bind to the receptor CXCR2, present in neutrophils and endothelial cells, activating them through the NF- $\kappa\beta$ pathway. GRO-2 is upregulated in ER α -positive breast cancer patients, and this fact correlates with shorter relapse-free survival.³⁰ Colonic metastasis in our patient was found on a polyp. It is possible that the polyp was an adenoma previously, and breast cancer cells may have homed over a previous adenoma by this chemokine pathway.

Moreover, there is an interesting role of inflammation. Our patient was diagnosed with gastritis related to H. pylori. We hypothesize a possible interaction between the chemokine receptors of circulating breast cancer stem cells and bacterial or inflammatory derived chemokines. It has been reported that there is a relationship between inflammation and tumorigenesis in prostate cancer, where inflammation can induce protumorigenic CXCL16 and CXCR6,³¹ attracting tumor cells. So in our case, it can be hypothesized that chronic inflammation due to H. pylori infection may have attracted tumor cells to the stomach and colon because of the action of chemokines. Under favorable circumstances, due to the influence of the same or other cytokines and growing/angiogenic factors, these cells may grow and develop metastatic disease. The degree of cellular infiltration by H. pylori seems to be lower in the corpus than in the antrum.⁵⁰ Our patient had gastric infiltration in the antrum, where H. pylori was also detected.

Infection with *H. pylori* causes chronic active gastritis, characterized by inflammatory cells infiltrating the gastric epithelial layer and the underlying lamina propria. *H. pylori* infection is associated with increased rates of expression of CXC chemokines (IL-8, GRO), CC chemokines, normal T-cells, and macrophage inflammatory protein 1-alpha (MIP-1 α), as well as increased mucosal levels of IL-8. Some of these upregulations are directly controlled by NF- κ B.³² It is possible that all these cytokines induced by *H. pylori* may attract and activate tumor cells to home to gastric mucosae.

It is feasible that tobacco consumption may enhance inflammation in gastritis caused by *H. pylori* by inducing GRO and CXCL5 mRNA production. Our patient was a heavy smoker, so this may explain in part the localization of mucosal metastases. Tobacco seems to increase oxygen free radicals from the PMNs and decrease mucus content. Additionally, nicotine may play a role in the induction of CXC chemokine expression, but further studies must be undertaken to confirm this hypothesis.

In colon, metastases may be mediated by receptors of stem cell chemokines in colon bacterial walls. Colon bacteria may produce proteins that mediate inflammation and control metalloprotease production in colonic mucosa in a similar way to *H. pylori*. Molecules influencing polyp production such as GRO2 and GRO3 may contribute to attract tumor cells. It has been reported that human colon epithelial cells release galectin-3, which can activate NF- $\kappa\beta$ in colonic

lamina propria fibroblasts and induce the secretion of IL-8 from these cells.³³

Finally, proteinase-activated receptor (PAR) mediates the cellular actions of serine proteinases. PAR1 and PAR2 are widely distributed in the gastrointestinal tract, and their activation in gastrointestinal inflammation results in proinflammatory reactions mediated by IL-6, IL-8, and prostaglandins. PAR2 activation induces NF- $\kappa\beta$ and AP-1-dependent IL-8 production in association with activation of p38 MAPK and ERK1/2.³⁴ This may be another mechanism favoring tumor growing in inflamed mucus.

Conclusion

Gastrointestinal metastases occur in some cases of lobular breast cancer, but dual infiltration of gastric and colonic regions has not been previously described as the first manifestation of spread. This surprising behavior may involve a mechanism in which chemokines and inflammatory phenomena might be implicated.

The expression of chemokine receptors could determine the type of tumor that metastasizes, whereas the site of dissemination is dictated by expression of the chemokine ligand. Expression of chemokines is passive, and metastatic behavior may be encoded in the primary cancer cell genetic profile.

Further studies are needed to identify the true role of chemokine receptors in metastatic phenomena.

Disclosure

The authors report no conflicts of interest in this work.

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