

Emppen Current status and future potential of predictive biomarkers for immune checkpoint inhibitors in gastric cancer

Byung Woog Kang,¹ Ian Chau ¹

To cite: Kang BW, Chau I. Current status and future potential of predictive biomarkers for immune checkpoint inhibitors in gastric cancer. ESMO Open 2020;5:e000791. doi:10.1136/ esmoopen-2020-000791

Received 16 April 2020 Revised 17 May 2020 Accepted 25 May 2020

C Author (s) (or their employer(s)) 2020. Re-use permitted under CC BY-NC. No commercial re-use. Published by BMJ on behalf of the European Society for Medical Oncology.

¹Department of Oncology/ Hematology, Kyungpook National University Hospital, School of Medicine, Kyungpook National University, Daegu, Republic of Korea

²Department of Medicine, Royal Marsden Hospital, London and Surrey, UK

Correspondence to Dr Ian Chau; ian.chau@rmh.nhs.uk

ABSTRACT

Immunotherapy is revolutionising cancer treatment and has already emerged as standard treatment for patients with recurrent or metastatic gastric cancer (GC). Recent research has been focused on identifying robust predictive biomarkers for GC treated with immune checkpoint inhibitors (ICIs). The expression of programmed cell death protein-ligand-1 (PD-L1) is considered a manifestation of immune response evasion, and several studies have already reported the potential of PD-L1 expression as a predictive parameter for various human malignancies. Meanwhile, based on comprehensive molecular characterisation of GC, testing for Epstein-Barr virus and microsatellite instability is a potential predictive biomarker. Culminating evidence suggests that novel biomarkers, such as the tumour mutational burden and gene expression signature, could indicate the success of treatment with ICIs. However, the exact roles of these biomarkers in GC treated with ICIs remain unclear. Therefore, this study reviews recent scientific data on current and emerging biomarkers for ICIs in GC, which have potential to improve treatment outcomes.

INTRODUCTION

The prognosis for metastatic or recurrent gastric cancer (GC) remains very poor, making it the third leading cause of cancerrelated death worldwide.¹ However, the recent development of immune checkpoint inhibitors (ICIs) targeting the programmed cell death protein-1 (PD-1) and programmed cell death protein-ligand-1 (PD-L1) pathways has produced improved outcomes for GC and already been successfully incorporated into clinical practice.² In particular, checkpoint inhibition with anti-PD1 monoclonal antibodies, such as pembrolizumab and nivolumab, has led to durable and significant responses in a minority of GCs.^{3–5} As a result, interest has increased in selecting the right patient population to benefit such ICIs, along with further exploration of immunotherapy. Notably, various tumourrelated and host-related factors with a critical impact on systemic immune functions may influence the response to ICIs.⁶ Moreover, a significant proportion of patients do not respond to these therapies, and there can

also be a threat of unpredictable immunerelated adverse events (AEs) and even severe toxicity.⁷⁸ Therefore, identifying more robust predictive biomarkers is critical to optimise treatment with ICIs, while avoiding unnecessary treatment of patients who could develop life-threatening or life-altering AEs.

GC is a heterogeneous and complex disease.⁹ Thus, various approaches, such as molecular classification, have already been proposed for the subhistological exploration of GC as a potential tool for more effective therapeutic strategies. The Cancer Genome Atlas Research Network (TCGA) suggested a comprehensive molecular characterisation of 295 GCs using various platforms, and proposed four distinct subtypes: Epstein-Barr virus (EBV)-positive, microsatellite unstable (microsatellite instability (MSI)), genomically stable and tumours with chromosomal instability.¹⁰ More recently, the Asian Cancer Research Group (ACRG) described four subtypes: MSI, microsatellite stable (MSS)/ epithelial-to-mesenchymal transition, MSS/ TP53-positive and MSS/TP53-negative.¹¹ While the above subtypings show some overlapping molecular aberrations, MSI was identified as a subtype by both TCGA and ACRG.²⁹ MSI is a molecular marker of a defective function of the DNA mismatch repair (MMR) system that recognises and repairs the erroneous insertion, deletion and misincorporation of bases that can arise during DNA replication and recombination, as well as repairing some forms of DNA damage.¹² It is also well known that MSI-high (MSI-H) tumours exhibit a high tumour mutational burden (TMB), neoantigen load and immune infiltration, making them respond well to ICIs.¹³ Meanwhile, the distinct characteristics of EBV-positive GC is the overexpression of PD-L1 and PD-L2.¹⁴ The driving features of PD-L1 positivity in EBV-positive GC can also be effectively targeted with immunotherapy, similar to the MSI-H subtype. Interestingly, in a recent phase II trial by Kim et al,





pembrolizumab showed a promising efficacy in patients with MSI-H and EBV-positive tumours.¹⁵ Accordingly, based on a better molecular characterisation of GC, this review focuses on the current and emerging biomarkers for ICIs that would facilitate precision medicine.

Clinical evidences of ICIs in GC

Several phase II and III trials have recently investigated the PD-1 and PD-L1 blockade in GC, as summarised in table 1.4516-19 Pembrolizumab is an IgG4 human antibody targeting PD-1, thereby interfering with the interaction between PD-1 and PD-L1.²⁰ In the phase Ib KEYNOTE-012 trial, 39 patients with PD-L1-positive GC or gastrooesophageal junction (GEI) cancer were enrolled in both Asian and non-Asian countries.²¹ The objective response rate (ORR) was 22% and durable responses were seen with a median duration of responses of 40 weeks. The subsequent phase II multicohort KEYNOTE-059 trial (cohort 1) enrolled 259 patients with recurrent or metastatic GC and GEI cancer who received two pretreated lines of chemotherapy.⁴ Here, the ORR was 11.6% and median overall survival (OS) was 5.6 months. Following these two trials, the randomised phase III KEYNOTE-061 trial compared pembrolizumb monotherapy with paclitaxel in patients with PD-L1-positive GC that had progressed on first-line flouropyrimidine and platinum doublet chemotherapy.¹⁷ While patients with a PD-L1 status were initially unselected, PD-L1-positive patients with a combined positive score (CPS) of 1 or higher were included in the latter part of the study. The CPS is the number of PD-L1 staining cells, including tumour cells, lymphocytes and macrophages, divided by the total number of viable tumour cells, while a tumour proportion score (TPS) is the percentage of viable tumour cells showing partial or complete membrane staining, relative to all viable tumour cells present in the sample.²² Approximately 67% of patients were found to have PD-L1-positive tumours using the CPS. Pembrolizumab did not meet its primary end point of a longer OS and progression-free survival (PFS) in patients with PD-L1-positive tumours. Notwithstanding, it is worth noting that patients who expressed PD-L1 CPS of 10 or higher exhibited a better benefit from treatment with pembrolizumab in post hoc analyses, although these subgroup analyses should be interpreted with caution.

Subsequently, the KEYNOTE-062 was a large randomised first-line clinical trial of 763 patients with advanced GC or GEJ cancer who were randomly assigned to one of three arms: pembrolizumab at 200 mg every 3 weeks for up to 2 years, pembrolizumab plus chemotherapy (cisplatin and 5-fluorouracil or capecitabine) or placebo plus chemotherapy.²³ Pembrolizumab was non-inferior to chemotherapy for OS in patients with CPS 1 or higher. No survival benefit was observed with the addition of pembrolizumab to chemotherapy compared with chemotherapy alone in this study.

Nivolumab, also an IgG4 antibody, is very similar in structure to pembrolizumab, except that nivolumab binds to

the N-terminal loop on the PD-1 molecule, while pembrolizumab binds to the C'D loop.²⁴ The phase III ATTRAC-TION-2 (ONO-4538-12) trial compared nivolumab with a placebo in 493 Asian patients with unresectable or recurrent GC that was refractory to or intolerant of at least two previous standard chemotherapy regimens.⁵ The results showed a significantly prolonged OS for the nivolumab group with a 1-year OS rate of 27.3% vs 11.6%. More recently, the long-term survival was reported showing 2-year survival rates of 10.6% for nivolumab and 3.2% for placebo.²⁵ Nivolumab also showed a significant advantage compared with the placebo in terms of PFS and the radiological objective response. Therefore, these results provide randomised evidence that nivolumab is a valid approach to improving the clinical outcomes for patients with GC in a third-line and subsequent-line setting. However, the overall clinical value of ATTRACTION-02 was partially limited by several important issues.^{26 27} The patient population was only Asian and PD-L1 positivity, reported at a low frequency of 14% as this was assessed as TPS rather than CPS, was not associated with the survival outcomes. Plus, there was no comparative data on quality of life. A phase I/II CHECKMATE-032 study also reported that nivolumab and nivolumab plus ipilimumab provide a durable antitumour activity in heavily pretreated Western patients with chemotherapy-refractory GC, GEJ cancer and oesophageal cancer.¹⁹ In particular, the clinical benefit of nivolumab monotherapy was consistent with that reported for Asian patients in the ATTRACTION-02 study. Yet, similar to the ATTRACTION-02 study, there was no association of PD-L1 positivity according to TPS with survival outcomes. Therefore, ongoing randomised controlled trials of ICIs, including pembrolizumab and nivolumab in earlier line treatment need to unify assessment of PD-L1 expression and create more accurate profiles of AEs in GC.

Avelumab is an IgG1 antibody which binds to the front beta-sheet of PD-L1 and possesses PD-1/PD-L1 blockade activity with antibody-dependent cell-mediated cytotoxicity (ADCC).²⁸ There are some differences between PD-1 inhibition and PD-L1 inhibition, as PD-1 targeting therapeutic antibodies including pembrolizumab and nivolumab block the PD-1/PD-L1 or PD-1/PD-L2 interaction to restore tumor-specific T-cell reactivity without mediating ADCC.²⁴ The JAVELIN Gastric 300 trial, the first study to compare avelumab with standard chemotherapy in third-line treatment for GC, did not achieve its primary end point of improving OS or the secondary end points of PFS and ORR.¹⁸ This negative finding may be attributed to the usage of the active comparator in the control arm. In addition, there are possible reasons, including the different drug biding sites, heterogeneity of tumour biology and methodology of PD-L1 testing. Notwithstanding, fewer patients experienced AEs with avelumab than with chemotherapy, although researchers found no evidence of clinical benefit compared with commonly used chemotherapy in any of the examined subgroups, including the tumour PD-L1 expression status.

Kang BW. Chau I. <i>ESMO Open</i> 2020: 5 :e000791	. doi:10.1136/esmoopen-2020-000791
Rang DW, onau I. 20110 Opon 2020,0.00001 01	. 001.10.1100/0011000001 2020 000101

Table 1 Sel	ected pha	se II and III studies ev	/aluatir	immune gr	checkpoint i	nhibitors in gastric cancer							
Agents	Target	Study name	Phase	Setting	PD-L1 status	Treatment arms	Patient number	Geographic region	Primary end points	RR (%)	PFS	SO	References
Pembrolizumab	PD-1	KEYNOTE-059 (cohort 1)	_	Third-line or later	Unselected	Pembrolizumab	259*	Global	RR	11.6	2	5.6	4
Pembrolizumab	PD-1	KEYNOTE-061	=	Second- line	Positive	Pembrolizumb Paclitaxel	196 199	Global	PFS, OS	16 14	1.5 4.1	9.1 8.3	17
Nivolumab	PD-1	ATTRACTION-02 (ONO-4538-12)	=	Third-line or later	Unselected	Nivolumab Placebo	330 163	Asian	SO	11.2 0	1.6 1.5	7.5 5.1	Q
Nivolumab	PD-1	ATTRACTION-04 I (part 1)	/	First-line	Unselected	Nivolumab/SOX or CAPOX	40	Asian	RR	65.8	9.5	Not reached	16
Nivolumab	PD-1	CHECKMATE-032 I	E.	Third-line or later	Unselected	Nivolumab Nivolumab1/Ipilimumab3† Nivolumab3/Ipilimumab1†	59 49 52	Western	RR	12 24 8	1.4 1.6 1.6	6.2 6.9 4.8	19
Avelumab	PD-L1	JAVELIN Gastric 300	=	Third-line	Unselected	Avelumab Physician's choice‡	272 133	Global	SO	2.2 4.3	1.4 2.7	4.6 5	18
*Among 259 patie +Nivolumeh 1 mo	ents, 148 (57. //cr plus ipilin	1%) were PD-L1 positive. mimsh 3 ma/ca /NIN/01 .IDI/	, <i>neve</i> (8	lovia sheeks	ha by/back	us inilimumah 1 ma/ka (NIIVO3+ID	1) even 3 we	o A Q					

TNvolumab 1 mg/kg plus iplimumab 3 mg/kg (NIVC1+IPI3) every 3 weeks, nivolumab 3 mg/kg plus iplimumab 1 mg/kg (NIVC3+IPI1) every 3 weeks. ‡Paclitaxel 80 mg/m² on days 1, 8 and 15 or irinotecan 150 mg/m² on days 1 and 15, each of a 4-week treatment cycle. CAPOX, capeciabine and oxaliplatin; OS, overall survival; PD-1, programmed cell death protein-1; PD-L1, programmed cell death protein-ligand 1; PFS, progression-free survival; RR, response rate; SOX, S-1 and oxaliplatin.

These results might support the potential of avelumab for combination or maintenance therapy. Notwithstanding, the recent JAVELIN Gastric 100 trial with maintenance avelumab therapy following initial chemotherapy in GC produced disappointing results.²⁹ Although there was more efficacy seen with avelumab in an exploratory analysis of 137 PD-L1-positive patients with a CPS \geq 1 (14.9 vs 11.6 months), no significant difference with respect to OS was observed between the avelumab maintenance and continued chemotherapy groups (10.4 vs 10.9 months, p=0.177).

As discussed above, several clinical trials are currently ongoing with various strategies.³⁰ ³¹ The phase III ATTRACTION-04 (NCT02746796) trial is evaluating nivolumab plus standard chemotherapy (oxaliplatin plus either S-1 or capecitabine) versus chemotherapy alone in Asian patients.¹⁶ The phase II component (part 1) of this study showed chemotherapy plus nivolumab led to an ORR of 65.8% and median PFS of 9.5 months, while the subsequent phase III trial (part 2) is evaluating the survival outcomes. Another phase III trial, CHECKMATE-649 (NCT02872116), has completed recruitment with over 2000 patients randomising patients to oxaliplatin-based chemotherapy alone or in combination with nivolumab or nivolumab plus ipilimumab.³² The third arm of this study terminated recruitment early due to early safety signal.

Antiangiogenic treatment, such as ramucirumab and bevacizumab, is also generating recent interest. As several studies have suggested that simultaneously blocking angiogenesis and PD-1 pathways induces synergistic antitumour effects, especially involving the control of the tumour microenvironment (TME).33 34 Researchers including the current authors noted that ramucirumab in combination with pembrolizumab (JVDF) showed a manageable safety profile with favourable antitumour activity in patients with previously treated GC.³⁵ Consequently, the phase I/II NivoRam (NCT02999295) trial has evaluated the safety and tolerability of the addition of ramucirumab to nivolumab in patients with GC as secondline therapy.⁷ As an alternative antiangiogenic and multitargeted kinase inhibitor, a phase I trial of regorafenib plus nivolumab is also exploring in GC (NCT03406871).³⁶ This trial demonstrated an ORR of 44% in GC and ICIspretreated patients with GC achieved a partial response (PR) in 11/25 patients. Interestingly, of the seven patients who had received prior anti-PD-1/PD-L1 treatment, three achieved an objective response.

Another interesting phase II trial investigating the role of pembrolizumab plus trastuzumab combined with chemotherapy in patients with previously untreated human epidermal growth factor receptor 2-positive tumours is currently ongoing (NCT02954536).³⁷ Preliminary results from this study showed 48% of patients experienced reduction in target lesions after one dose of pembrolizumab/trastuzumab before oxaliplatin/capecitabine was introduced in second cycle. The high ORR of 89% was coupled with median PFS of 13 months. This

has led to the current phase III KEYNOTE-811 study randomising patients to chemotherapy plus trastuzumab with or without pembrolizumb (NCT03615326).³⁸ Thus, these therapeutic strategies including combination treatment represent a true opportunity in the contemporary treatment of GC and may produce further success, when considering integrative genomic data.

Biomarkers for ICIs in GC

The identification and validation of reliable biomarkers are important to facilitate precise patient selection and increase the clinical benefit from ICIs. An overview of the predictive roles of biomarkers obtained from tissue or blood and their characterisation in the management of GC is briefly summarised in table 2.

PD-L1 expression

Testing for PD-L1 expression by IHC is the current standard in most solid tumours and several studies have already assessed the clinical outcomes according to the PD-L1 expression status in GC. Nevertheless, different antibodies are being used for IHC to assess with different performance and different scoring criteria for PD-L1 expression. A recent multicentre study (Blueprint PD-L1 IHC Comparison Project) attempted to compare the performance of each IHC PD-L1 assay in lung cancer.³⁹ Three assays including 22C3 for pembrolizumab, 28-8 for nivolumab and SP263 for durvalumab were found to be comparable to each other in the staining of tumour tissue, whereas SP142 for atezolizumab was found to be less sensitive. However, further study is required to carefully validate these assays in GC.

As noted above, in the KEYNOTE-059 trial, PD-L1 expression was assessed using the PDL1 IHC 22C3 pharmDx assay and measured using a CPS.⁴ This trial demonstrated a higher ORR (15.5% vs 6.4%) in patients with high PD-L1 expression, defined as CPS \geq 1. Both PFS and OS were also more prolonged in this group.⁴⁰ Although pembrolizumab in a second-line trial (KEYNOTE-061) did not significantly prolong OS, greater benefits were seen in tumours with higher PD-L1 expression (CPS ≥ 10 , ORR=25%; CPS ≥ 1 , ORR=16%; CPS <1, ORR=2%).¹⁷ However, the ATTRACTION-02 and JAVELIN Gastric 300 trials showed no clinical improvement for PD-L1-positive tumours as they used TPS rather than CPS.^{18 27} These differences may also have been due to the use of different cut-off points and scoring systems, a lack of standardisation of the assays and testing platforms, the heterogeneous nature of PD-L1 expression in tumours, intratumoural/ intertumoural heterogeneity and intraobserver/interobserver variability.⁴¹⁻⁴³ For instance, the ATTRACTION-02 study retrospectively evaluated PD-L1 expression using a PD-L1 IHC 28-8 pharmDx assay, defined as the TPS, while PD-L1 expression was prospectively assessed in tumour cells, tumour-associated lymphocytes and macrophages using a 22C3 pharmDx assay in KEYNOTE-061.^{17 27} As such, PD-L1 positivity was only reported in about 15% patients using TPS with nivolumab, whereas it is generally

lable 2 Ove	erview of c	andidate biomarkers associat	ing with response	to immune checkpoint inhibitors in gastrie	c cancer
	Sample				
Biomarkers	source	Methods	Treatment	Recent results in gastric cancer	References
PD-L1	Tumour	IHC	Pembrolizumab	Expression of PD-L1 ≥1 associated with better clinical efficacy (ORR, mRD)	4
			Pembrolizumab	Expression of PD-L1 CPS of 10 or higher associated with better clinical efficacy (OS, ORR)	17
			Pembrolizumab	Expression of PD-L1 associated with higher response rate	15
EBV positivity	Tumour	In situ hybridisation	Pembrolizumab	EBV positivity associated with higher response rate	15
MSI-H	Tumour	MSI testing or IHC	Nivolumb	MSI-H associated with clinical efficacy (ORR, DCR, OS)	19
			Pembrolizumab	MSI-H associated with better clinical efficacy (ORR)	4
			Pembrolizumab	MSI-H associated with better clinical efficacy (ORR, OS)	17
			Pembrolizumab	MSI-H associated with better clinical efficacy (ORR)	15
ТМВ	Tumour or blood	WES or targeted sequencing	Toripalimab	TMB associated with better clinical efficacy (ORR, OS)	50
TILs	Tumour	Image analysing software or manually counted	ICIs	Presence of TILs associated with better clinical efficacy in various solid tumours, but very limited data in GC	78
GEP	Tumour	Multigene profiling	Pembrolizumab	IFN-gamma (6-gene) signature associated with better clinical efficacy (ORR, PFS)	21
			Pembrolizumab	T-cell inflamed (18-gene) signature associated with better clinical efficacy (ORR, PFS)	4
Gut microbiota	Stool	Culture or molecular technique (sequencing/ metagenomics)	ICIs	Various species associated with enhancement and IRAEs of ICIs in various solid tumours	94
NLR	Blood	Complete blood count	ICIs	Increased NLR correlated with DCR and OS*	110
			Nivolumab	Decreased change of NLR associated with better survival	111

*Sixty-seven patients had tumours from the stomach.

CPS, combined positive score; DCR, disease control rate; EBV, Epstein-Barr virus; GC, gastric cancer; GEP, gene expression profiling; ICIs, immune checkpoint inhibitors; IFN, interferon; IHC, immunohistochemistry; IRAEs, immune-related adverse events; mRD, median response duration; MSI-H, microsatellite instability; NLR, neutrophil-to-lymphocyte ratio; ORR, overall response rate; OS, overall survival; PD-L1, programmed cell death protein-ligand 1; PFS, progression-free survival; TILs, tumour-infiltrating lymphocytes; TMB, tumour mutational burden.

between 60% and 70% using CPS ≥ 1 as the cut-off. More recently, using the CHECKMATE-032 study data, tumours were re-scored using CPS and there was better correlation between nivolumab treatment and survival even at the CPS ≥ 5 level.⁴⁴ Thus, one of the coprimary patient population in the first-line CHECKMATE-649 study, that has recently completed recruitment, is including a subpopulation of patients with PD-L1 CPS $\geq 5.$ ³²

EBV positivity

EBV status is also emerging as a potential biomarker for personalised treatment strategies in GC.¹⁴ In situ

hybridisation detection of EBV-encoded small RNA in tumour cells is generally recommended to identify EBVassociated GC (EBVaGC).³⁰ The incidence of EBVaGC varies from 1% to 30% in different regions with an average of 10% worldwide.⁴⁵ Nevertheless, this subgroup is associated with better prognosis, thus less frequently found in advanced or metastatic setting. In particular, EBV-positive tumours frequently display PD-L1/2 overexpression, and occasional immune cell signalling activation.^{10 14} Several research groups found that the level of PD-L1 expression ranging from approximately 34% to 92% of EBVaGC with variable results between studies.²⁰ PD-L1 positivity has also been significantly associated with a poorer prognosis than PD-L1 negativity in EBVaGC. Furthermore, EBV triggers a significantly higher infiltration of CD8+ T cells in TME.⁴⁶ In previous studies of 120 patients with EBV-positive cancer, the current authors showed that high levels of tumour-infiltrating lymphocytes (TILs) were associated with a favourable prognosis, while intratumoural PD-L1 positivity with a worse prognosis.⁴⁷

In the phase I JAVELIN Solid Tumour trial where avelumab was shown to be beneficial for a patient with metastatic GC, it is worth noting that EBV-positive tumours with a low mutation burden and MSI tumours with a high mutation burden had statistically significantly higher tumour lymphocytic infiltration when compared with MSS tumours.⁴⁸ The strength of immune-mediated signalling signatures in EBV-positive tumours also represents a T-cell-inflamed TME.^{10 49} These findings support the concept that ICIs can be used in patients with GC with EBV by suppressing the PD-1 pathway in tumour cells and allowing immune activation. A recent phase II trial by Kim et al demonstrated improved efficacy associated with pembrolizumab in patients with EBV-positive tumours.¹⁵ This study enrolled 61 patients with pretreated GC. In a subgroup analysis, pembrolizumab monotherapy as salvage treatment showed that all six EBV-positive patients with GC attained PR (ORR=100%) with a median duration of 8.5 months. However, in another study, 4 out of 55 patients considered EBV-positive were treated with toripalimab.⁵⁰ Only one PR (25%) was observed with two stable and one progressive diseases. Patients with PR was also PD-L1-positive. These contrasting results with pembrolizumab could be due to toripalimab rather than EBVaGC as a predictive biomarker for ICI.

Microsatellite instability-high

MMR deficiency is generally characterised by a failure to repair DNA replication-associated errors, leading to the accumulation of mutations in microsatellite regions of the genome.⁵¹ These phenomena are known as MSI.⁵² Currently, two different methods have been validated for detecting MSI-H.⁵³ The MMR status is assessed by IHC staining to measure the expression levels of the proteins involved in DNA MMR, and a polymerase chain reactionbased exam also tests the length of repetitive DNA that are known as microsatellite in the normal and tumour tissues. While there are discrepancies between the IHC of MMR protein expression and MSI test results, the overall concordance in the two tests is high.⁵² The incidence of MSI in GC varies between countries, being relatively high in approximately 5%–30% of Western patients.⁵¹ MSI-H GC is commonly associated with intestinal type, female sex, older age, lack of lymph node metastases and onset in the distal stomach.⁵⁴ To date, multiple retrospective studies and limited prospective studies have reported on a positive association between MSI-H and a better prognosis in resectable GC.⁵⁵ For example, the MAGIC study reported that patients with MSI-H tumours have

superior survival compared with patients with MSS/MSIlow (MSI-L) tumours when treated with surgery alone and conversely have inferior survival to patients with MSS/MSI-L tumours when treated with perioperative chemotherapy plus surgery.⁵⁶ However, similar to EBV status, patients with MSI-H had better prognosis, thus only 4%–5% of patients with metastatic GC would be MMR-deficient. The prognostic and predictive values of the MSI status on the survival of patients with metastatic GC remain a subject of debate.⁵²

Theoretically, in the presence of MMR deficiency, undetected DNA replication errors, leading to a tumour with a high mutational burden, reproduce various neoantigens that stimulate T-cell activation and tumour infiltration by immune cells. KEYNOTE-012 trial reported that MSI-H tumours showed a partial response in two out of four patients, regardless of PD-L1 expression.²¹ A subgroup analysis of KEYNOTE-059 revealed an ORR of 57.1% for patients with MSI-H GC.⁴ In KEYNOTE-061 and more recently reported KEYNOTE-062, there was a substantially enhanced survival benefit in patients treated with pembrolizumab compared with chemotherapy.^{17 23} Similar to the results for EBV-positive tumours, the clinical study by Kim et al also showed that MSI-H tumours responded particularly well to pembrolizumab monotherapy (ORR=85.7%).¹⁵ In the CHECKMATE-032 trial that assessed the efficacy of another PD-1 monoclonal antibody nivolumab, the ORR was 29% for the MSI-H group vs 11% for the MSI-L group or MSS group.¹⁹ Therefore, this evidence highlights the potential of MSI-H as a predictor of the response to ICIs in GC.

Of note, whereas MSI-H/MMR deficiency is the most consistent predictor of efficacy to ICIs in GC, a substantial portion of MSI-H GC still has unsatisfactory outcomes even with ICIs. The degree of MSI and resultant mutation load, in part, might explain the variable response to PD-1 blockade in MMR-deficient tumours.⁵⁷ Tumours sensitive to PD-1 antibodies showed a loss or a reduction in tumour allele frequency of missense (non-synonymous single-nucleotide variant) and indel mutations after PD-1 treatment suggestive of immune editing of tumour cells

TMB and neoantigen

TMB may be a potential biomarker of outcomes with ICIs in multiple solid tumour types.⁴¹ Generally, cells have a number of repair pathways to maintain their genome stability.¹³ The mutational load acquired by defective DNA repair pathways frequently alters protein function and expression, resulting in the formation of neoantigens that serve a source of antitumour immune response. Therefore, it is reasonable to hypothesise that tumours with a high mutational load are more likely to produce neoantigens and increase immunogenicity.⁵⁸ In turn, this course of reaction induces a more intensified immune response, resulting in tumours becoming more sensitive to treatment with ICIs.⁴¹ Although tumour-specific neoantigens with high clonality are more predictable and beneficial for the response to ICIs, accurate measurement of these neoantigens is known to be expensive and timeconsuming.⁵⁹ In this situation, TMB could be a good approach for indirectly evaluating the neoantigen load. TMB is defined by the total number of somatic nonsynonymous mutations per coding area of the tumour DNA.⁵⁸ Several studies have already demonstrated the predictive impact of TMB in lung cancer and melanoma. One early study by Yarchoan et al observed a significant correlation between TMB and ORR for anti-PD-1 or anti-PD-L1 therapy.⁶⁰ Rizvi *et al* also reported that patients with TMB>50th percentile exhibited an improved durable clinical benefit rate and PFS versus those with lower TMB.⁶¹ This benefit was also seen in the CHECKMATE-227 study that included 299 patients with advanced non-small-cell lung cancer (NSCLC) who received a combination of nivolumab and ipilimumab as the first-line metastatic setting.⁶² A significantly prolonged PFS was reported for the patients with higher TMB treated with the combination treatment, irrespective of the expression of PD-L1. Likewise, a large-scale study across multiple cancer types found a significant association between TMB and the clinical outcome.^{63–67} These findings can also provide a novel strategy for subgroups with high TMB, considering that the measurement of the mutational load is a critical factor for therapeutic success.

However, for patients with GC, there is still insufficient evaluation and conflicting results on the utility of TMB as a biomarker of the response to ICIs.⁶⁵ Interestingly, Wang et al performed a TMB analysis of 54 patients with chemorefractory GC who were treated with toripalimab as a monotherapy in a prospective phase Ib/II clinical trial.⁵⁰ In this study, TMB-high (TMB-H) patients responded significantly better than the TMB-low patients (ORR 33.3% vs 7.1%, p=0.017). A survival benefit has also been demonstrated for patients with high TMB (OS 14.6 vs 4.0 months, p=0.038). Similar correlation was also found between TMB-H according to circulating tumour DNA and better survival when treated with pembrolizumab in GC.¹⁵ In light of recent approaches, this close relationship between TMB and clinical outcomes also points to the possibility of TMB as a predictive biomarker in patients with GC. However, in the study with regorafenib plus nivolumab, due to small sample size, TMB did not correlate with response or PFS to this combination.³⁶

Despite its identified significant predictive role, there are still many challenges in precisely estimating TMB. First, it is difficult to apply the protocol, including wholeexome sequencing or targeted sequencing panels using next-generation sequencing, to clinical practice due to various problems, such as the sample amount, cost, sensitivity, coverage and analysis time.^{8 30} Second, a standardised cut-off value for TMB has not yet been clearly established, since many studies have reported a wide range of cutoffs for different tumour types.⁵⁸ Thus, given the variety of TMB cutoffs, assays related with TMB in clinical studies should be interpreted cautiously. Moreover, the availability of tumour sampling to detect TMB is commonly limited and TMB may present temporal variability. A novel blood-based TMB approach could be considered as an alternative method, as the advantages of repeating sampling during treatment could provide information of a dynamic immune reaction.⁸ Plus, this approach is less invasive and enables investigators to document the evolution of TMB. Interestingly, in a study by Gandara *et al*, blood-based TMB was correlated with tissue-based TMB and showed a longer PFS in patients with metastatic NSCLC who received treatment with atezolizumab.⁶⁸ In summary, TMB could be a novel and independent biomarker that reflects the therapeutic effects of ICIs in GC. However, its accuracy in predicting the efficacy of ICIs varies among studies and still needs to be explored for GC.

Tumour infiltrating lymphocytes

The immune microenvironment of tumours is now recognised as an important determinant for understanding the relationship between a patient's immune system and their cancer, informing prognosis and guiding immunotherapy like ICIs.⁶⁹ Tumour cells are typically surrounded by infiltrating inflammatory cells, such as cytotoxic T cells, helper T cell subsets, regulatory T cells, tumor-associated macrophages, dendritic cells and myeloid lineage leucocytes.⁷⁰ Among these, differentiated lymphocytes, referred to as TILs, are considered a manifestation of the host immune response against tumour cells and seem to play an important role in various human malignancies.⁷¹ Several studies have already reported the potential of TILs as a predictive parameter.⁷² A recent in vitro study proposed that subpopulation of a CD8+ T cells is involved in mediating tumour control and responds to checkpoint blockades.⁷³ In breast cancer, TILs have been shown to predict patient outcomes and responses to ICIs.⁷⁴ Plus, the presence of stromal TILs has been associated with an improved ORR in patients with triple-negative breast cancer receiving pembrolizumab.⁷⁵ TILs have also been investigated as a predictive biomarker for breast cancer and could predict the efficacy of atezolizumab.⁷⁶ Recent research by Loupakis et al demonstrated a significant correlation between a high number of TILs and clinical responses and survival benefit in a large data set of patients with MSI-H metastatic colorectal cancer treated with ICIs.⁷⁷ These clinical benefits were also consistent with another meta-analysis, indicating that TILs are associated with improved prognosis predictions for OS.⁷⁸

Despite such evidence that TILs contribute to determining prognosis, the exact predictive value of TILs in GC treated with ICIs remains unclear.⁷² In particular, the detection of TILs could be a key biomarker for the treatment of TIL-rich tumours, such as EBV-positive or MSI-H, considering that ICIs could become an important part of the cancer armamentarium in these GC subsets.⁷¹ However, doubts remain on the methodology of interpreting TILs and the cut-off values for TILs in GC. In contrast to breast cancer, there is no current consensus on estimating TILs in GC specifically.⁷⁰ Plus, it is essential to elucidate the precise predictive role of each lymphocyte subset including, regulatory T cells, which could play a role in immunosuppression and tumour progression.⁷⁹ Moreover, the invasive margin or central infiltration could have a different density of T cells, leading to variable results. In malignant melanomas, the density of CD8+ T cells at the tumour edge has been shown to predict the response to pembrolizumab rather than the density at the tumour centre.⁸⁰ Galon *et al* recently developed an immunoscore based on the density, location, phenotype and functionality of T cells in colorectal cancer and found these immune infiltrates to be a better predictor of survival than TNM classification.⁸¹ Thus, the application of this approach in clinical practice would seem to be quite feasible for determining the response to ICIs for GC. Therefore, further innovative attempts regarding TILs could assist in discovering an effective biomarker for predicting efficacy of ICIs in GC.

Gene expression signatures

Several studies on gene expression profiling (GEP) are currently attempting to predict the response to ICIs in various types of cancers.⁸² In particular, immune gene signatures, such as IFN-gamma signalling and activated T cells, could have potential as predictive markers of ICI responses.⁴¹ Recent GEP revealed that an IFN-gammarelated gene profile obtained from baseline tumour tissue was predictive of the ORR and PFS in patients with melanomas treated with pembrolizumab.⁸³ Auslander et al also reported that a novel immune-predictive score (IMPRES) was significantly correlated with a better response to ICIs, suggesting that GEP could be incorporated in enhancing therapy response.⁸⁴ Similarly, additional studies have demonstrated a link between GEP and ICI responses in lung cancer.^{85 86} As described above. the KEYNOTE-012 trial for GC investigated the use of a six-gene IFN-gamma signature that was previously identified to predict the response in melanomas. Although this gene signature did not meet significance due to the small number of enrolled patients, a trend was seen associating the responders and the IFN-gamma signature.²¹ For further exploration of the association between this gene signature and patient outcomes, the KEYNOTE-059 trial analysed the association of the T-cell inflamed 18-gene signature with response and survival in GC.⁴ Higher T-cell inflamed score was associated with an improved likelihood of response to pembrolizumab and improved prognosis for GC, providing a strong rationale for clinical trials using GEP in patients with GC receiving ICIs.

GEP signatures have also been evaluated to correlated response with nivolumab in the CHECKMATE-032 study showing a potential predictive role for response with a 4-gene inflammatory signature incorporating CD274 (PD-L1), CD8A, lymphocyte activating 3 and signal transducer and activator of transcription 1.^{19 44} Consequently, it is increasingly evident that GEP is a promising option for selecting patients with GC who could benefit from ICIs. Thus, further well-designed and randomised studies

of large numbers of cases are needed to evaluate the role of GEP as a potential biomarker for ICIs in GC.

Gut microbiota

The relationship between microbiota and clinical responses to ICIs in GC is another ongoing area of research, with several studies exploring how microbiota may affect the therapeutic efficacy of immunotherapy.⁶ Gut microbiota play a fundamental role in the maintenance of host physiology and immune homeostasis, interacting with epithelial cells and stromal cells to modulate multiple vital functions.⁸⁷ They can also regulate barrier function, pathogen control and cell metabolism. In addition, Helicobacter pylori infection can contribute to the establishment of a persistent infection through the creation of an immunosuppressive microenvironment.⁸⁸ Chronic H. pylori infection results in gastric carcinogenesis and T-cell hyporesponsiveness and induced PD-L1 expression.⁸⁹ Interestingly, several studies reported that T cells exposed to H. pylori had an impaired ability to proliferate and H. pylori-positive tumours showed higher PD-L1 expression, leading to downregulate immune surveillance mechanisms.^{90 91} For this reason, there is much speculation that gut microbiota could affect the therapeutic efficacy of immunotherapy, particularly ICIs, and there is already accumulating evidence support this in preclinical studies.⁹²

Regarding the effects of ipilimumab, in a mouse model, the antitumour response was found to depend on the gut microbiota including Bacteroides fragilis or Bacteroides thetaiotaomicron.93 It was also demonstrated that tumours in antibiotic-treated or germ-free mice did not respond to this ICI. Currently, several clinical studies have reported a link between gut microbiota and ICI responses across multiple human cohorts.94 Although most studies only included fewer than 50 patients, the results from melanomas showed that the efficacy of anti-PD-1 therapy was influenced by gut microbiota.^{95 96} In a recent study conducted by Pinato et al that included 196 patients, prior antibiotic therapy and the response to ICI therapy were associated with OS, independent of the tumour site, disease burden and performance status.⁹⁷ Another research group analysed the clinical predictors of outcome in 76 patients with GC and 85 patients with oesophageal cancer treated with ICIs.⁹⁸ There was no difference in outcomes between patients treated with antibiotics during or in the 2 months preceding ICI treatment versus those who were not. However, decreased OS was observed among those patients who received antibiotics in the 30 days prior to commencing ICIs. This phenomenon indicates that the use of antibiotics may adversely modify the gut microbiota, thereby impairing the antitumour immunity and response to ICIs. In addition to their modulating effects on ICIs, gut microbiota may also be involved in immune-related AEs.⁹⁹ Recent evidence found that an abundance of Bacteroidetes was correlated with a low frequency of ipilimumab-induced colitis.¹⁰⁰ Therefore, when taken together, these findings suggest that gut microbiota may be relevant to the efficacy and toxicity of ICIs. However, the vast majority of studies have been retrospective in nature, with a limited ability to characterise the sample population. Thus, understanding the exact relationship between gut microbiota and the immune response remains limited.

Neutrophil-to-lymphocyte ratio

Besides the above-mentioned biologic and molecular biomarker, laboratory parameters reflecting the condition of systemic inflammation are relatively economical to evaluate, easily measurable, repeatable and ready to use in daily clinical practice.¹⁰¹ There is increasing evidence that the neutrophil-to-lymphocyte ratio (NLR) can be an effective prognostic marker as well as predictive indicator related to ICIs for various solid tumours.^{102 103} In fact, neutrophils are already known to be associated with detrimental outcomes in cancer, participating in different stages of the oncogenic process including tumour growth, invasion and metastases, while lymphocytes might affect a favourable impact on their tumour inhibiting properties.^{104 105} Several large studies including melanoma, NSCLC and genitourinary cancer treated with ICIs found that a high NLR resulted in worse OS and PFS across various types of malignancies.¹⁰⁶⁻¹⁰⁹ Recently, Li et al prospectively collected data from discovery and validation cohorts among 160 patients with non-colorectal gastrointestinal cancer receiving ICIs.¹¹⁰ They found that the NLR level was significantly correlated with reduced OS, which is also consistent with other previous studies. In addition, Ota et al demonstrated that changes in the NLR values from those at 30 or 60 days after first-dose nivolumab were associated with significantly shorter PFS and OS in patients with GC.¹¹¹ Thus, for GC, these findings provide supporting evidence that the NLR may contribute to determining the predictive value of ICIs. Interestingly, some attempts have also been made to assess the correlation between the NLR and pseudoprogression/hyperprogression. A recent retrospective study of 25 patients with NSCLC treated with anti-PD-1/PD-L1 therapy reported that the pre-treatment and post-treatment NLRs were useful in distinguishing between pseudoprogression and true-progression.¹¹² More recently, another study examined a database of 263 patients with NSCLC and showed that immunophenotyping the peripheral blood CD8+ T lymphocytes was associated with hyperprogression and survival outcomes. Although the studies to date have been small and retrospective, the NLR may be useful for predicting therapeutic effects, especially as an early response marker.¹¹³

CONCLUSIONS AND FUTURE PERSPECTIVES

Immunotherapy has begun to revolutionise cancer treatment and already emerged as standard treatment for patients with recurrent or metastatic GC. Research has also been focused on finding robust predictive biomarkers for GC treated with ICIs. First, PD-L1 expression by IHC has been widely implemented as a predictive marker for ICIs to identify patients with GC who are more likely to benefit from the therapy. However, the relationship between PD-L1 expression and the therapeutic effect of nivolumab/ avelumab still remains unclear, and pembrolizumab requires a clear PD-L1 threshold for effective prediction using a validated CPS score. To obtain convincing data, more precise and standardised methods are also needed to analyse PD-L1 expression. Meanwhile, in the era of molecular classification, accumulating evidence shows that EBVpositive and MSI-H tumours are the most immunogenic GC subtypes and ICIs have achieved an enhanced benefit in these GC subsets. Therefore, in the case of recurrent or metastatic GC, testing for EBV and MSI status should be considered, plus the related impact needs to be addressed in ongoing trials for earlier treatment settings. Third, novel biomarkers, such as TMB, TILs and GEP that exhibit a host cellular immune response against tumours, have also shown encouraging results in GC. However, the challenge remains to apply the data generated from validated clinical trials to clinical practice in order to provide precision immunotherapy. Plus, gut microbiota have been identified as another attractive biomarker for ICIs, with a recognised influence on host immunity and cancer. Clearly, a better understanding of the interaction between the microbial network and antitumour immunity will help to select patients who are more likely to respond to ICIs. Finally, NLR has also been investigated for their potential to be integrated as predictive markers of the response to ICIs, especially as this parameter is relatively more cost-effective and easier to measure. In summary, it is foreseeable that these emerging biomarkers will eventually shift the treatment paradigm of GC. Thus, to optimise this great opportunity, further high-quality evidence with standardised methods and proper patient selection are needed to discover reliable predictive biomarkers for ICIs in GC.

Acknowledgements The authors would like to acknowledge National Health Service funding to the National Institute for Health Research Biomedical Research Centre at the Royal Marsden NHS Foundation Trust and The Institute of Cancer Research.

Contributors All authors contributed and approved the final manuscript.

Funding The authors have not declared a specific grant for this research from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests IC: Advisory Board: Eli-Lilly, Bristol-Meyers Squibb, MSD, Bayer, Roche, Merck-Serono, Five Prime Therapeutics, AstraZeneca, Oncologie International, Pierre Fabre Research funding: Eli-Lilly, Janssen-Cilag, Sanofi Oncology, Merck-Serono honorarium: Eli-Lilly

Patient consent for publication Not required.

Provenance and peer review Not commissioned; externally peer reviewed.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, any changes made are indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

ORCID iD

lan Chau http://orcid.org/0000-0003-0286-8703

Open access

REFERENCES

- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018;68:394–424.
- 2 Chau I. Clinical development of PD-1/PD-L1 immunotherapy for gastrointestinal cancers: facts and hopes. *Clin Cancer Res* 2017;23:6002–11.
- 3 Chénard-Poirier M, Smyth EC. Immune checkpoint inhibitors in the treatment of gastroesophageal cancer. *Drugs* 2019;79:1–10.
- 4 Fuchs CS, Doi T, Jang RW, et al. Safety and efficacy of pembrolizumab monotherapy in patients with previously treated advanced gastric and gastroesophageal junction cancer: phase 2 clinical KEYNOTE-059 trial. JAMA Oncol 2018;4:e180013.
- 5 Kang Y-K, Boku N, Satoh T, et al. Nivolumab in patients with advanced gastric or gastro-oesophageal junction cancer refractory to, or intolerant of, at least two previous chemotherapy regimens (ONO-4538-12, ATTRACTION-2): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017;390:2461–71.
- 6 Helmink BA, Khan MAW, Hermann A, et al. The microbiome, cancer, and cancer therapy. Nat Med 2019;25:377–88.
- 7 Lote H, Cafferkey C, Chau I. PD-1 and PD-L1 blockade in gastrointestinal malignancies. *Cancer Treat Rev* 2015;41:893–903.
- 8 Arora S, Velichinskii R, Lesh RW, et al. Existing and emerging biomarkers for immune checkpoint immunotherapy in solid tumors. Adv Ther 2019;36:2638–78.
- 9 Serra O, Galán M, Ginesta MM, et al. Comparison and applicability of molecular classifications for gastric cancer. Cancer Treat Rev 2019;77:29–34.
- 10 Cancer Genome Atlas Research Network. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature* 2014;513:202–9.
- 11 Cristescu R, Lee J, Nebozhyn M, *et al.* Molecular analysis of gastric cancer identifies subtypes associated with distinct clinical outcomes. *Nat Med* 2015;21:449–56.
- 12 Larrea AA, Lujan SA, Kunkel TA. Snapshot: DNA mismatch repair. Cell 2010;141:730.
- 13 Liebl MC, Hofmann TG. Identification of responders to immune checkpoint therapy: which biomarkers have the highest value? J Eur Acad Dermatol Venereol 2019;33:52–6.
- 14 Kang BW, Baek DW, Kang H, et al. Novel therapeutic approaches for Epstein-Barr virus associated gastric cancer. Anticancer Res 2019;39:4003–10.
- 15 Kim ST, Cristescu R, Bass AJ, et al. Comprehensive molecular characterization of clinical responses to PD-1 inhibition in metastatic gastric cancer. Nat Med 2018;24:1449–58.
- 16 Boku N, Ryu M-H, Kato K, et al. Safety and efficacy of nivolumab in combination with S-1/capecitabine plus oxaliplatin in patients with previously untreated, unresectable, advanced, or recurrent gastric/gastroesophageal junction cancer: interim results of a randomized, phase II trial (ATTRACTION-4). Ann Oncol 2019;30:250–8.
- 17 Shitara K, Özgüroğlu M, Bang Y-J, et al. Pembrolizumab versus paclitaxel for previously treated, advanced gastric or gastrooesophageal junction cancer (KEYNOTE-061): a randomised, openlabel, controlled, phase 3 trial. *Lancet* 2018;392:123–33.
- 18 Bang Y-J, Ruiz EY, Van Cutsem E, et al. Phase III, randomised trial of avelumab versus physician's choice of chemotherapy as third-line treatment of patients with advanced gastric or gastrooesophageal junction cancer: primary analysis of javelin gastric 300. Ann Oncol 2018;29:2052–60.
- 19 Janjigian YY, Bendell J, Calvo E, *et al.* CheckMate-032 study: efficacy and safety of nivolumab and nivolumab plus ipilimumab in patients with metastatic esophagogastric cancer. *J Clin Oncol* 2018;36:2836–44.
- 20 Seo AN, Kang BW, Kwon OK, et al. Intratumoural PD-L1 expression is associated with worse survival of patients with Epstein-Barr virus-associated gastric cancer. Br J Cancer 2017;117:1753–60.
- 21 Muro K, Chung HC, Shankaran V, et al. Pembrolizumab for patients with PD-L1-positive advanced gastric cancer (KEYNOTE-012): a multicentre, open-label, phase 1B trial. Lancet Oncol 2016;17:717–26.
- 22 Kulangara K, Zhang N, Corigliano E, et al. Clinical utility of the combined positive score for programmed death ligand-1 expression and the approval of pembrolizumab for treatment of gastric cancer. Arch Pathol Lab Med 2019;143:330–7.
- 23 Tabernero J, Van Cutsem E, Bang Y-J, et al. Pembrolizumab with or without chemotherapy versus chemotherapy for advanced gastric or gastroesophageal junction (G/GEJ) adenocarcinoma: the phase III KEYNOTE-062 study. JCO 2019;37:LBA4007–LBA07.
- 24 Picardo SL, Doi J, Hansen AR. Structure and optimization of checkpoint inhibitors. *Cancers* 2019;12:38.

- 25 Chen L-T, Satoh T, Ryu M-H, et al. A phase 3 study of nivolumab in previously treated advanced gastric or gastroesophageal junction cancer (ATTRACTION-2): 2-year update data. Gastric Cancer 2020;23:510–9.
- 26 Chau I. Checkpoint inhibition: an attraction in advanced gastric cancer? *Lancet* 2017;390:2418–9.
- 27 Satoh T, Kang Y-K, Chao Y, et al. Exploratory subgroup analysis of patients with prior trastuzumab use in the ATTRACTION-2 trial: a randomized phase III clinical trial investigating the efficacy and safety of nivolumab in patients with advanced gastric/gastroesophageal junction cancer. Gastric Cancer 2020;23:143–53.
- 28 Liu K, Tan S, Chai Y, et al. Structural basis of anti-PD-L1 monoclonal antibody avelumab for tumor therapy. Cell Res 2017;27:151–3.
- 29 Moehler MH, Dvorkin M, Ozguroglu M, et al. Results of the javelin gastric 100 phase 3 trial: avelumab maintenance following firstline (1L) chemotherapy (CTX) vs continuation of CTX for HER2– advanced gastric or gastroesophageal junction cancer (GC/GEJC). JCO 2020;38:278–78.
- 30 Kwak Y, Seo AN, Lee HE, et al. Tumor immune response and immunotherapy in gastric cancer. J Pathol Transl Med 2020;54:20–33.
- 31 Togasaki K, Sukawa Y, Kanai T, et al. Clinical efficacy of immune checkpoint inhibitors in the treatment of unresectable advanced or recurrent gastric cancer: an evidence-based review of therapies. Onco Targets Ther 2018;11:8239–50.
- 32 Moehler MH, Janjigian YY, Adenis A, et al. CheckMate 649: a randomized, multicenter, open-label, phase III study of nivolumab (NIVO) + iplimumab (IPI) or nivo + chemotherapy (CTX) versus CTX alone in patients with previously untreated advanced (AdV) gastric (G) or gastroesophageal junction (GEJ) cancer. JCO 2018;36:TPS1 92–TPS92.
- 33 Butters O, Young K, Cunningham D, et al. Targeting vascular endothelial growth factor in oesophagogastric cancer: a review of progress to date and immunotherapy combination strategies. Front Oncol 2019;9:618–18.
- 34 Terme M, Colussi O, Marcheteau E, et al. Modulation of immunity by antiangiogenic molecules in cancer. Clin Dev Immunol 2012;2012:1–8.
- 35 Herbst RS, Arkenau H-T, Santana-Davila R, et al. Ramucirumab plus pembrolizumab in patients with previously treated advanced non-small-cell lung cancer, gastro-oesophageal cancer, or urothelial carcinomas (JVDF): a multicohort, non-randomised, open-label, phase 1a/b trial. Lancet Oncol 2019;20:1109–23.
- 36 Fukuoka S, Hara H, Takahashi N, et al. Regorafenib plus nivolumab in patients with advanced gastric (GC) or colorectal cancer (CRC): an open-label, dose-finding, and doseexpansion phase 1B trial (REGONIVO, EPOC1603). J Clin Oncol 2020:JCO1903296.
- 37 Janjigian YY, Chou JF, Simmons M, *et al.* First-Line pembrolizumab (P), trastuzumab (T), capecitabine (C) and oxaliplatin (o) in HER2positive metastatic esophagogastric adenocarcinoma (mEGA). *JCO* 2019;37:62.
- 38 Janjigian YY, Bang Y-J, Fuchs CS, et al. KEYNOTE-811 pembrolizumab plus trastuzumab and chemotherapy for HER2+ metastatic gastric or gastroesophageal junction cancer (mG/GEJC): a double-blind, randomized, placebo-controlled phase 3 study. JCO 2019;37:TPS4146–TPS46.
- 39 Hirsch FR, McElhinny A, Stanforth D, et al. Pd-L1 immunohistochemistry assays for lung cancer: results from phase 1 of the blueprint PD-L1 IHC assay comparison project. J Thorac Oncol 2017;12:208–22.
- 40 Wainberg ZA, Yoon HH, Catenacci DVT, et al. Efficacy and safety of pembrolizumab (pembro) alone or in combination with chemotherapy (chemo) in patients (PTS) with advanced gastric or gastroesophageal (G/GEJ) cancer: long-term follow up from KEYNOTE-059. JCO 2019;37:4009–09.
- 41 Bodor JN, Boumber Y, Borghaei H. Biomarkers for immune checkpoint inhibition in non-small cell lung cancer (NSCLC). *Cancer* 2020;126:260–70.
- 42 Coutzac C, Pernot S, Chaput N, *et al.* Immunotherapy in advanced gastric cancer, is it the future? *Crit Rev Oncol Hematol* 2019;133:25–32.
- 43 Lantuejoul S, Sound-Tsao M, Cooper WA, et al. Pd-L1 testing for lung cancer in 2019: perspective from the IASLC pathology Committee. J Thorac Oncol 2020;15:499–519.
- 44 Lei M, Siemers N, Pandya D, et al. Abstract 2673: association of PD-L1 combined positive score and immune gene signatures with efficacy of nivolumab (NIVO) ± ipilimumab (IPI) in patients with metastatic gastroesophageal cancer (mGEC). *Cancer Res* 2019;79:2673–73.

- 45 Naseem M, Barzi A, Brezden-Masley C, et al. Outlooks on Epstein-Barr virus associated gastric cancer. Cancer Treat Rev 2018;66:15–22.
- 46 Tan GW, Visser L, Tan LP, et al. The microenvironment in Epstein-Barr virus-associated malignancies. *Pathogens* 2018;7:40.
- 47 Kang BW, Seo AN, Yoon S, et al. Prognostic value of tumorinfiltrating lymphocytes in Epstein-Barr virus-associated gastric cancer. Ann Oncol 2016;27:494–501.
- 48 Panda A, Mehnert JM, Hirshfield KM, et al. Immune activation and benefit from Avelumab in EBV-positive gastric cancer. J Natl Cancer Inst 2018;110:316–20.
- 49 Fukayama M, Abe H, Kunita A, et al. Thirty years of Epstein-Barr virus-associated gastric carcinoma. *Virchows Arch* 2020;476:353–65.
- 50 Wang F, Wei XL, Wang FH, et al. Safety, efficacy and tumor mutational burden as a biomarker of overall survival benefit in chemo-refractory gastric cancer treated with toripalimab, a PD-1 antibody in phase Ib/II clinical trial NCT02915432. Ann Oncol 2019;30:1479–86.
- 51 Richman S. Deficient mismatch repair: read all about it (review). Int J Oncol 2015;47:1189–202.
- 52 Ratti M, Lampis A, Hahne JC, et al. Microsatellite instability in gastric cancer: molecular bases, clinical perspectives, and new treatment approaches. *Cell Mol Life Sci* 2018;75:4151–62.
- 53 Vasen HFA, Blanco I, Aktan-Collan K, et al. Revised guidelines for the clinical management of Lynch syndrome (HNPCC): recommendations by a group of European experts. Gut 2013;62:812–23.
- 54 Polom K, Marano L, Marrelli D, et al. Meta-Analysis of microsatellite instability in relation to clinicopathological characteristics and overall survival in gastric cancer. Br J Surg 2018;105:159–67.
- 55 Li K, Luo H, Huang L, et al. Microsatellite instability: a review of what the oncologist should know. Cancer Cell Int 2020;20:16.
- 56 Smyth EC, Wotherspoon A, Peckitt C, et al. Mismatch repair deficiency, microsatellite instability, and survival: an exploratory analysis of the medical Research Council adjuvant gastric infusional chemotherapy (magic) trial. JAMA Oncol 2017;3:1197–203.
- 57 Mandal R, Samstein RM, Lee K-W, et al. Genetic diversity of tumors with mismatch repair deficiency influences anti-PD-1 immunotherapy response. Science 2019;364:485–91.
- 58 Meléndez B, Van Campenhout C, Rorive S, et al. Methods of measurement for tumor mutational burden in tumor tissue. *Transl Lung Cancer Res* 2018;7:661–7.
- 59 Otoshi T, Nagano T, Tachihara M, et al. Possible biomarkers for cancer immunotherapy. Cancers 2019;11:935.
- 60 Yarchoan M, Hopkins A, Jaffee EM. Tumor mutational burden and response rate to PD-1 inhibition. N Engl J Med 2017;377:2500–1.
- 61 Rizvi H, Sanchez-Vega F, La K, et al. Molecular Determinants of Response to Anti-Programmed Cell Death (PD)-1 and Anti-Programmed Death-Ligand 1 (PD-L1) Blockade in Patients With Non-Small-Cell Lung Cancer Profiled With Targeted Next-Generation Sequencing. J Clin Oncol 2018;36:633–41.
- 62 Hellmann MD, Ciuleanu T-E, Pluzanski A, et al. Nivolumab plus ipilimumab in lung cancer with a high tumor mutational burden. N Engl J Med 2018;378:2093–104.
- 63 Samstein RM, Lee C-H, Shoushtari AN, et al. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. *Nat Genet* 2019;51:202–6.
- 64 Morrison C, Pabla S, Conroy JM, *et al*. Predicting response to checkpoint inhibitors in melanoma beyond PD-L1 and mutational burden. *J Immunother Cancer* 2018;6:32.
- 65 Mishima S, Kawazoe A, Nakamura Y, *et al.* Clinicopathological and molecular features of responders to nivolumab for patients with advanced gastric cancer. *J Immunother Cancer* 2019;7:24.
- 66 Goodman AM, Kato S, Bazhenova L, et al. Tumor mutational burden as an independent predictor of response to immunotherapy in diverse cancers. *Mol Cancer Ther* 2017;16:2598–608.
- 67 Cristescu R, Mogg R, Ayers M, et al. Pan-tumor genomic biomarkers for PD-1 checkpoint blockade-based immunotherapy. Science 2018;362:eaar3593.
- 68 Gandara DR, Paul SM, Kowanetz M, et al. Blood-Based tumor mutational burden as a predictor of clinical benefit in non-smallcell lung cancer patients treated with atezolizumab. *Nat Med* 2018;24:1441–8.
- 69 Wu T, Dai Y. Tumor microenvironment and therapeutic response. *Cancer Lett* 2017;387:61–8.
- 70 Hendry S, Salgado R, Gevaert T, et al. Assessing tumor-infiltrating lymphocytes in solid tumors: a practical review for pathologists and proposal for a standardized method from the International Immuno-Oncology biomarkers Working group: Part 2: TILs in melanoma, gastrointestinal tract carcinomas, non-small cell lung carcinoma

and mesothelioma, endometrial and ovarian carcinomas, squamous cell carcinoma of the head and neck, genitourinary carcinomas, and primary brain tumors. *Adv Anat Pathol* 2017;24:311–35.

- 71 Kang BW, Kim JG, Lee IH, *et al.* Clinical significance of tumorinfiltrating lymphocytes for gastric cancer in the era of immunology. *World J Gastrointest Oncol* 2017;9:293–9.
- 72 Chang W-J, Du Y, Zhao X, et al. Inflammation-Related factors predicting prognosis of gastric cancer. World J Gastroenterol 2014;20:4586–96.
- 73 Miller BC, Sen DR, Al Abosy R, et al. Subsets of exhausted CD8⁺ T cells differentially mediate tumor control and respond to checkpoint blockade. Nat Immunol 2019;20:326–36.
- 74 Force J, Leal JHS, McArthur HL. Checkpoint blockade strategies in the treatment of breast cancer: where we are and where we are heading. *Curr Treat Options Oncol* 2019;20:35.
- 75 Adams S, Schmid P, Rugo HS, et al. Pembrolizumab monotherapy for previously treated metastatic triple-negative breast cancer: cohort a of the phase II KEYNOTE-086 study. Ann Oncol 2019;30:397–404.
- 76 Basile D, Pelizzari G, Vitale MG, et al. Atezolizumab for the treatment of breast cancer. Expert Opin Biol Ther 2018;18:595–603.
- 77 Loupakis F, Depetris I, Biason P, et al. Prediction of benefit from checkpoint inhibitors in mismatch repair deficient metastatic colorectal cancer: role of tumor infiltrating lymphocytes. Oncologist.
- 78 Yu Y, Zeng D, Ou Q, et al. Association of survival and immunerelated biomarkers with immunotherapy in patients with non-small cell lung cancer: a meta-analysis and individual patient-level analysis. JAMA Netw Open 2019;2:e196879–e79.
- 79 Esensten JH, Muller YD, Bluestone JA, et al. Regulatory T-cell therapy for autoimmune and autoinflammatory diseases: the next frontier. J Allergy Clin Immunol 2018;142:1710–8.
- 80 Tumeh PC, Harview CL, Yearley JH, et al. Pd-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* 2014;515:568–71.
- 81 Pagès F, Mlecnik B, Marliot F, et al. International validation of the consensus immunoscore for the classification of colon cancer: a prognostic and accuracy study. *Lancet* 2018;391:2128–39.
- 82 Tong M, Wang J, He W, et al. Predictive biomarkers for tumor immune checkpoint blockade. Cancer Manag Res 2018;10:4501–7.
- 83 Ayers M, Lunceford J, Nebozhyn M, et al. IFŇ-γ–related mRNA profile predicts clinical response to PD-1 blockade. J Clin Invest 2017;127:2930–40.
- 84 Auslander N, Zhang G, Lee JS, *et al.* Robust prediction of response to immune checkpoint blockade therapy in metastatic melanoma. *Nat Med* 2018;24:1545–9.
- 85 Fehrenbacher L, Spira A, Ballinger M, et al. Atezolizumab versus docetaxel for patients with previously treated non-small-cell lung cancer (poplar): a multicentre, open-label, phase 2 randomised controlled trial. *Lancet* 2016;387:1837–46.
- 86 Socinski MA, Jotte RM, Cappuzzo F, et al. Atezolizumab for firstline treatment of metastatic Nonsquamous NSCLC. N Engl J Med 2018;378:2288–301.
- 87 Li W, Deng Y, Chu Q, *et al*. Gut microbiome and cancer immunotherapy. *Cancer Lett* 2019;447:41–7.
- 88 Figueroa-Protti L, Soto-Molinari R, Calderón-Osorno M, et al. Gastric cancer in the era of immune checkpoint blockade. J Oncol 2019;2019:1–11.
- 89 Silva R, Gullo I, Carneiro F. The PD-1:PD-L1 immune inhibitory checkpoint in Helicobacter pylori infection and gastric cancer: a comprehensive review and future perspectives. *Porto Biomed J* 2016;1:4–11.
- 90 YY W, Lin CW, Cheng KS, et al. Increased programmed deathligand-1 expression in human gastric epithelial cells in Helicobacter pylori infection. *Clin Exp Immunol* 2010;161:551–9.
- 91 Knipp U, Birkholz S, Kaup W, et al. Suppression of human mononuclear cell response by *Helicobacter pylori* : Effects on isolated monocytes and lymphocytes. *FEMS Immunol Med Microbiol* 1994;8:157–66.
- 92 Frankel AE, Deshmukh S, Reddy A, et al. Cancer immune checkpoint inhibitor therapy and the gut microbiota. *Integr Cancer Ther* 2019;18:153473541984637–79.
- 93 Vétizou M, Pitt JM, Daillère R, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. Science 2015;350:1079–84.
- 94 McQuade JL, Daniel CR, Helmink BA, *et al.* Modulating the microbiome to improve therapeutic response in cancer. *Lancet* Oncol 2019;20:e77–91.
- 95 Gopalakrishnan V, Spencer CN, Nezi L, et al. Gut microbiome modulates response to anti–PD-1 immunotherapy in melanoma patients. Science 2018;359:97–103.

Open access

- 96 Matson V, Fessler J, Bao R, et al. The commensal microbiome is associated with anti–PD-1 efficacy in metastatic melanoma patients. Science 2018;359:104–8.
- 97 Pinato DJ, Howlett S, Ottaviani D, et al. Association of prior antibiotic treatment with survival and response to immune checkpoint inhibitor therapy in patients with cancer. JAMA Oncol 2019;5:1774–8.
- 98 Greally M, Chou JF, Chatila WK, et al. Clinical and molecular predictors of response to immune checkpoint inhibitors in patients with advanced esophagogastric cancer. *Clin Cancer Res* 2019;25:6160–9.
- 99 Inamura K. Roles of microbiota in response to cancer immunotherapy. Semin Cancer Biol 2020.
- 100 Dubin K, Callahan MK, Ren B, et al. Intestinal microbiome analyses identify melanoma patients at risk for checkpoint-blockade-induced colitis. Nat Commun 2016;7:10391–91.
- 101 Guner A, Kim H-I. Biomarkers for evaluating the inflammation status in patients with cancer. *J Gastric Cancer* 2019;19:254–77.
- 102 Jiang T, Qiao M, Zhao C, *et al.* Pretreatment neutrophil-tolymphocyte ratio is associated with outcome of advanced-stage cancer patients treated with immunotherapy: a meta-analysis. *Cancer Immunol Immunother* 2018;67:713–27.
- 103 Li M, Spakowicz D, Burkart J, et al. Change in neutrophil to lymphocyte ratio during immunotherapy treatment is a non-linear predictor of patient outcomes in advanced cancers. J Cancer Res Clin Oncol 2019;145:2541–6.
- 104 Granot Z. Neutrophils as a therapeutic target in cancer. *Front Immunol* 2019;10:1710.
- 105 Ocana A, Nieto-Jiménez C, Pandiella A, et al. Neutrophils in cancer: prognostic role and therapeutic strategies. *Mol Cancer* 2017;16:137–37.

- 106 Bagley SJ, Kothari S, Aggarwal C, et al. Pretreatment neutrophilto-lymphocyte ratio as a marker of outcomes in nivolumab-treated patients with advanced non-small-cell lung cancer. Lung Cancer 2017;106:1–7.
- 107 Cassidy MR, Wolchok RE, Zheng J, *et al.* Neutrophil to lymphocyte ratio is associated with outcome during ipilimumab treatment. *EBioMedicine* 2017;18:56–61.
- 108 Jeyakumar G, Kim S, Bumma N, *et al.* Neutrophil lymphocyte ratio and duration of prior anti-angiogenic therapy as biomarkers in metastatic RCC receiving immune checkpoint inhibitor therapy. *J Immunother Cancer* 2017;5:82.
- 109 Sacdalan DB, Lucero JA, Sacdalan DL. Prognostic utility of baseline neutrophil-to-lymphocyte ratio in patients receiving immune checkpoint inhibitors: a review and meta-analysis. *Onco Targets Ther* 2018;11:955–65.
- 110 Li S, Zou J, Liu C, et al. Baseline derived neutrophil-to-lymphocyte ratio as a prognostic biomarker for non-colorectal gastrointestinal cancer patients treated with immune checkpoint blockade. *Clin Immunol* 2020;212:108345.
- 111 Ota Y, Takahari D, Suzuki T, et al. Changes in the neutrophil-tolymphocyte ratio during nivolumab monotherapy are associated with gastric cancer survival. Cancer Chemother Pharmacol 2020;85:265–72.
- 112 Kiriu T, Yamamoto M, Nagano T, *et al.* Pseudo-Progression and the neutrophil-to-lymphocyte ratio in non-small cell lung cancer treated with immune checkpoint inhibitors: a case-control study. *Onco Targets Ther* 2019;12:10559–68.
- 113 Kim CG, Kim KH, Pyo K-H, et al. Hyperprogressive disease during PD-1/PD-L1 blockade in patients with non-small-cell lung cancer. Ann Oncol 2019;30:1104–13.