



# Hypoxia equally reduces the respiratory compensation point and the NIRS-derived [HHb] breakpoint during a ramp-incremental test in young active males

Rafael D. A. Azevedo  | Béjar Saona J. E. | Erin Calaine Inglis | Danilo Iannetta |  
 Juan M. Murias 

Faculty of Kinesiology, University of  
 Calgary, Calgary, AB, Canada

## Correspondence

Juan M. Murias, Faculty of Kinesiology,  
 University of Calgary, KNB 434, 2500  
 University Drive NW, Calgary, AB, Canada  
 T2N 1N4.  
 Email: jmmurias@ucalgary.ca

## Funding information

Funding for this study was provided by the  
 National Science and Engineering Research  
 Council of Canada (RGPIN-2016-03698)  
 and the Heart & Stroke Foundation of  
 Canada (1047725). Rafael A. Azevedo  
 was supported by Eyes High Doctoral  
 Recruitment Scholarship.

## Abstract

This study investigated the effect of reduced inspired fraction of O<sub>2</sub> (FiO<sub>2</sub>) in the correspondence between the respiratory compensation point (RCP) and the breakpoint in the near-infrared spectroscopy-derived deoxygenated hemoglobin signal ([HHb]<sub>bp</sub>) during a ramp-incremental (RI) test to exhaustion. Eleven young males performed, on two separated occasions, a RI test either in normoxia (NORM, FiO<sub>2</sub> = 20.9%) or hypoxia (HYPO, FiO<sub>2</sub> = 16%). Oxygen uptake ( $\dot{V}O_2$ ), and [HHb] signal from the vastus lateralis muscle were continuously measured. Peak  $\dot{V}O_2$  (2.98 ± 0.36 vs. 3.39 ± 0.26 L min<sup>-1</sup>) and PO (282 ± 29 vs. 310 ± 19 W) were lower in HYPO compared to NORM condition, respectively. The  $\dot{V}O_2$  and PO associated with RCP and [HHb]<sub>bp</sub> were lower in HYPO (2.35 ± 0.24 and 2.34 ± 0.26 L min<sup>-1</sup>; 198 ± 37 and 197 ± 30 W, respectively) when compared to NORM (2.75 ± 0.26 and 2.75 ± 0.28 L min<sup>-1</sup>; 244 ± 29 and 241 ± 28 W, respectively) (*p* < .05). Within the same condition, the  $\dot{V}O_2$  and PO associated with RCP and [HHb]<sub>bp</sub> were not different (*p* > .05). Bland–Altman plots mean average errors between RCP and [HHb]<sub>bp</sub> were not different from zero in HYPO (0.01 L min<sup>-1</sup> and 1.1 W) and NORM (0.00 L min<sup>-1</sup> and 3.6 W) conditions. The intra-individual changes between thresholds associated with  $\dot{V}O_2$  and PO in HYPO from NORM were strongly correlated (*r* = .626 and 0.752, *p* < .05). Therefore, breathing a lower FiO<sub>2</sub> during a RI test resulted in proportional reduction in the RCP and the [HHb]<sub>bp</sub> in terms of  $\dot{V}O_2$  and PO, which further supports the notion that these physiological responses may arise from similar metabolic changes reflecting a common phenomenon.

## KEYWORDS

cycling, deoxygenation breakpoint, exercise intensity, RCP, thresholds

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. *Physiological Reports* published by Wiley Periodicals LLC on behalf of The Physiological Society and the American Physiological Society.

## 1 | INTRODUCTION

During a ramp-incremental (RI) test to exhaustion, the rate of  $O_2$  uptake ( $\dot{V}O_2$ ) increases rather linearly from the exercise onset until maximal  $\dot{V}O_2$  ( $\dot{V}O_{2max}$ ) or task failure ensue. At approximately 80% of  $\dot{V}O_{2max}$ , two physiological responses become discernible: at the pulmonary level the respiratory compensation point (RCP) (Whipp, Davis, & Wasserman, 1989) is expressed, whereas at the level of the muscle (i.e. the *vastus lateralis* muscle), a breakpoint in the near-infrared spectroscopy (NIRS)-derived deoxygenated hemoglobin signal ( $[HHb]_{bp}$ ) is observed (Spencer, Murias, & Paterson, 2012).

At the pulmonary level, the RCP is identified as the onset of a more rapid increase in minute ventilation that is disproportional from the rate of carbon dioxide production ( $VCO_2$ ), which causes the arterial tension of  $CO_2$  to fall (Whipp et al., 1989). This hyperventilatory response offers partial compensation to the blood accumulation of hydrogen ions ( $[H^+]$ ) arising from the accelerated glycolytic rate within the active musculature (Whipp et al., 1989). At the muscle level (within the *vastus lateralis* of the quadriceps muscles), the  $[HHb]_{bp}$  demarcates the beginning of a plateau-like response in the  $[HHb]$  signal (Spencer et al., 2012), with the adjustment of the  $[HHb]$  being a proxy for local  $O_2$  extraction, as it represents the balance between delivery and off-loading of  $O_2$  to and out of the capillary microvasculature, respectively. (Grassi & Quaresima, 2016). Although still a subject of debate, our group has suggested that the  $[HHb]_{bp}$  and the subsequent plateau might be related to improved blood flow redistribution toward the active tissues of the of the *vastus lateralis* (Azevedo, Béjar Saona, Inglis, Iannetta, & Murias, 2020; Inglis, Iannetta, & Murias, 2017; Murias, Spencer, Keir, & Paterson, 2013), whereby the plateau in the  $[HHb]$  signal would result from an increased local  $O_2$  delivery in the presence of a continuous increase in  $\dot{V}O_2$  (Murias, Spencer, et al., 2013). However, others have proposed that the occurrence of the plateau in the  $[HHb]$  signal might be due to other physiological factors, such as the achievement of the upper limit for  $O_2$  extraction in the superficial portions of muscle (Okushima et al., 2015), and/or limitations to perfusive or diffusive provision of  $O_2$  during high-intensity exercise (Okushima et al., 2020).

Representative of whole-body and local physiological responses, the RCP and the  $[HHb]_{bp}$ , respectively, have been suggested to indicate the metabolic rate associated with the transition from the heavy into the severe exercise intensity domains (Keir et al., 2015; Keir, Pogliaghi, & Murias, 2019). This suggestion has been based on studies that have shown their correspondence on the basis of  $\dot{V}O_2$  and power output (PO) during RI tests (Fontana et al., 2015; Iannetta, Passfield, Murias, Calaine, & Christopher, 2018;

Iannetta, Qahtani, Millet, & Murias, 2017; Inglis, Iannetta, Keir, & Murias, 2019), and their occurrence at a  $\dot{V}O_2$  similar to that associated with the maximal lactate steady state (MLSS) and critical power (CP) (Bellotti, Calabria, Capelli, & Pogliaghi, 2013; Iannetta, Passfield, et al., 2018; Keir et al., 2015). In this context, a recent report has demonstrated that the RCP and the  $[HHb]_{bp}$ , (as well as MLSS) of trained cyclists changed uniformly during the course of a competitive season (Inglis et al., 2019). Despite these lines of evidence, however, the idea of an equivalence between the RCP and the  $[HHb]_{bp}$  is still under debate and other research groups have suggested plausible alternative interpretations (Boone, Barstow, Celie, Prieur, & Bourgois, 2016; Broxterman, Craig, & Richardson, 2018; Caen, Vermeire, Bourgois, & Boone, 2018). Therefore, interventions that can alter the occurrence of both the RCP and the  $[HHb]_{bp}$  are needed to further test whether these two indices represent a similar physiological event during RI exercise and, thus, can possibly be seen as equivalent.

From this perspective, breathing hypoxic air is an intervention that can be used to potentially alter the occurrence of both the RCP and  $[HHb]_{bp}$  during a RI test, and thus provide insights into the relationship between these two physiological indices of exercise intensity (Leo, Sabapathy, Simmonds, & Cross, 2017). Indeed, hypoxia has been shown to limit  $O_2$  provision to the peripheral tissues as indicated by a reduction in arterial  $O_2$  saturation and capillary partial pressure of  $O_2$  (Richardson et al., 2006). These impairments typically result in lower  $\dot{V}O_{2max}$  and peak power output (PPO) during incremental exercise (Knight et al., 1992), and a smaller PO associated with the heavy-to-severe boundary of exercise intensity, as demonstrated by reduced CP in hypoxia (La Monica et al., 2018; Parker Simpson, Jones, Skiba, Vanhatalo, & Wilkerson, 2014; Townsend, Nichols, Skiba, Racinais, & Périard, 2017). Thus, it could be expected that breathing a lower fraction of  $O_2$  ( $FiO_2$ ) during a RI test would also affect the RCP and the  $[HHb]_{bp}$  by reducing the  $\dot{V}O_2$  and PO at which they occur with respect to normoxia. Indeed, previous findings have shown a reduction in the PO associated with the  $[HHb]_{bp}$  in hypoxia compared to normoxia (Azevedo et al., 2020; Osawa, Kime, Hamaoka, Katsumura, & Yamamoto, 2011). Although suggested by a previous study (Osawa et al., 2011), no investigation has quantified if the changes in the RCP and the  $[HHb]_{bp}$  in hypoxia compared to normoxia would be of similar magnitude, and, thus, be linked to one another.

Therefore, this study explored the changes in the  $\dot{V}O_2$  and PO associated with the RCP and the  $[HHb]_{bp}$  in response to a diminished  $FiO_2$  (16%) during a RI test to exhaustion. It was hypothesized that a reduction in both the  $\dot{V}O_2$  and PO at the RCP and the  $[HHb]_{bp}$  would occur as a consequence of the decreased  $FiO_2$ , and that these changes would be proportional on an individual basis.

## 2 | METHODS

### 2.1 | Participants

Eleven healthy males were recruited (age:  $29 \pm 6$  years; height:  $179 \pm 6$  cm; weight:  $79 \pm 9$  kg). Participants gave informed written consent to participate in this study after completing the physical activity readiness questionnaire (PAR-Q+) and being cleared for exercise. All participants were free of any medical condition that could have altered their normal cardiovascular responses to exercise. The study was approved by the Conjoint Health Research Ethics Board at the University of Calgary. This study was part of a larger project designed to answer different research questions (Azevedo et al., 2020), whereby, out of 11 participants, 10 were included in our previous publication.

### 2.2 | Experimental design

Participants reported to the laboratory on two separate occasions at similar time of the day ( $\pm 1$  hr). The visits were separated by at least 48 hr, during which participants performed a RI test to exhaustion either in normoxia (NORM,  $FiO_2 = 20.9\%$ ) or hypoxia (HYPO,  $FiO_2 = 16\%$ ). The  $FiO_2$  for hypoxia was chosen with reference of previous investigations demonstrating that this specific  $FiO_2$  was effective in altering arterial and muscle  $O_2$  saturation during exercise (Amann, Romer, Subudhi, Pegelow, & Dempsey, 2007). Before each RI test, a moderate intensity step transition, consisting of cycling at 20 W for 6 min and at 80 W for another 6 min, was performed to subsequently compute the mean response time (MRT) of  $\dot{V}O_2$  (Iannetta, Murias, & Keir, 2019) (see data analyses for details). Thereafter, the participants were given 2 min of rest followed by a 4 min baseline cycling at 20 W prior to the RI test. The RI tests to volitional exhaustion consisted of  $30 \text{ W min}^{-1}$  continuous increments (i.e. 1 W every 2 s). Each exercise session was preceded by a 5 min wash-in period with the predetermined  $FiO_2$  (either hypoxia or normoxia), during which participants rested seated on the bike. All exercise testing was performed on an electromagnetically braked cycle ergometer (Velotron Dynafit Pro; Racer Mate).

### 2.3 | Measurements

#### 2.3.1 | Pulmonary $O_2$ uptake

A breath-by-breath metabolic cart system (Quark CPET, Cosmed), which was calibrated before each test as per the manufacturer's recommendations, was used to measure ventilatory and gas exchange variables. The expiratory and

inspiratory air volume rates and the concentrations of inspired and expired  $O_2$  and  $CO_2$  were measured through a low-resistance flowmeter and high precision gas analyzers, respectively. During all sessions, participants breathed through a mask connected to the flowmeter which was attached to a two-way low-resistance T-valve (Hans Rudolph In.; 2600 series, medium two-way NRBV). To control the  $FiO_2$  during HYPO, inspired air was delivered by a gas-mixing device (Altitrainer NP, SMTEC) connected with a tube to the T-valve to provide the desired concentrations of  $O_2$ . During NORM, the same system was used to deliver room air in order to blind participants from the experimental condition.

#### 2.3.2 | NIRS-derived signals

[HHb] ( $\mu\text{M}$ ) and tissue  $O_2$  saturation index ( $StO_2$ ) (%) signals of the VL muscle were measured using a two-channel frequency-domain NIRS device (Oxiplex TS; ISS) at a sampling rate of 2 Hz, as described elsewhere (Iannetta, Qahtani, Millet, et al., 2017). The probe was placed on the belly of VL muscle, midway between the greater trochanter and the proximal border of the patella and secured in place by double-sided tape as well as an elastic strap to prevent movement. Additionally, the NIRS probe was covered with an optically dense, black vinyl sheet to minimize possible intrusion of extraneous light. Before removing the probe, the area was marked to ensure the consistency of the placement for the following visit. The NIRS system was calibrated before each test, as per the manufacturer's recommendations.

#### 2.3.3 | Pulse $O_2$ saturation

The pulse oxygen saturation ( $SpO_2$ ) was measured via a pulse oximetry pod (ADInstruments) from the forefinger of the right hand at a 1 Hz sampling rate and at wavelengths of 660 and 910 nm for red and infrared lights, respectively. The oximeter was connected to the acquisition apparatus (Power Lab, ADInstruments) linked to a computer software (LabChart 8, ADInstruments).

#### 2.3.4 | Blood lactate concentration

Within 1 min after the RI task failure, capillary blood lactate concentrations ( $[\text{Lac}^-]$ ) were measured with a portable lactate analyzer (Lactate Scout, SensLab GmbH). A lancet was used to perform a pinprick after wiping the finger with an alcohol swab, and a  $2 \mu\text{l}$  capillary sample of whole blood was collected and immediately analyzed.

## 2.4 | Data analyses

### 2.4.1 | $\dot{V}O_2$

$\dot{V}O_2$  data were individually analyzed as previously described (Lamarra, Whipp, Ward, & Wasserman, 2017). Briefly, aberrant data points that were three standard deviations (*SD*) from the local mean were removed. Data were then linearly interpolated to 1 s intervals.  $\dot{V}O_{2\max}$  was defined as the highest  $\dot{V}O_2$  computed from a 20-s rolling average. Gas exchange threshold (GET) and RCP were identified by three independent investigators by examining raw respiratory data. Briefly, GET corresponded to the breakpoint in the  $\dot{V}O_2$ -to- $\dot{V}CO_2$  relationship (i.e. V-slope method) concomitant with an increase in the ventilatory equivalent of  $O_2$  and a leveling off of end-tidal pressure of  $CO_2$  ( $PCO_2$ ) (Beaver, Wasserman, & Whipp, 1986). The RCP corresponded to the second disproportional increase (i.e. second breakpoint) in the  $V_E/\dot{V}O_2$  relation, where end-tidal  $PCO_2$  began to fall after a period of isocapnic buffering (Whipp et al., 1989). In case of disagreement of more than  $100 \text{ ml min}^{-1}$ , investigators would reevaluate together the profiles until consensus was reached. To account for the circulatory transit time delay of deoxygenated hemoglobin from the active musculature to reach the lungs and the kinetics of  $\dot{V}O_2$ , the MRT was calculated on an individual basis, as previously described (Iannetta, Murias, et al., 2019), using a customized function of a computer software (Origin, Origin Lab). A linear regression of the  $\dot{V}O_2$  versus PO relationship was fitted from the onset of the systemic rise in  $\dot{V}O_2$  until the previously established GET. The steady-state  $\dot{V}O_2$  from the moderate step transition (i.e. 80 W for 6 min) was superimposed on the  $\dot{V}O_2$  versus PO relationship. Thereafter, the MRT corresponded to the difference between (a) the PO equivalent to the abscissa of the intersection between the  $\dot{V}O_2$  and the linear fit versus PO, and (b) the steady-state  $\dot{V}O_2$  corresponding to 80 W (i.e. measured during the step transition) (Iannetta, Murias, et al., 2019).

### 2.4.2 | NIRS-derived signals

The [HHb]-time relationship for VL was modeled with the following piecewise equation that included two linear segments (e.g. “double linear”), as previously described (Spencer et al., 2012):

$$f = \text{if}(x < \text{TD}, g(x), h(x))$$

$$g(x) = i1 + (s1 \cdot x)$$

$$i2 = i1 + (s1 \cdot \text{TD})$$

$$h(x) = i2 + [s2 \cdot (x - \text{TD})]$$

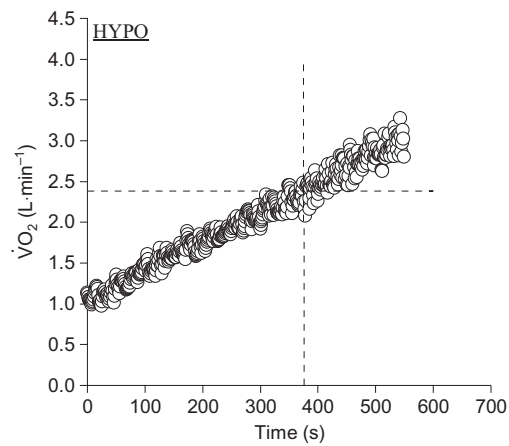
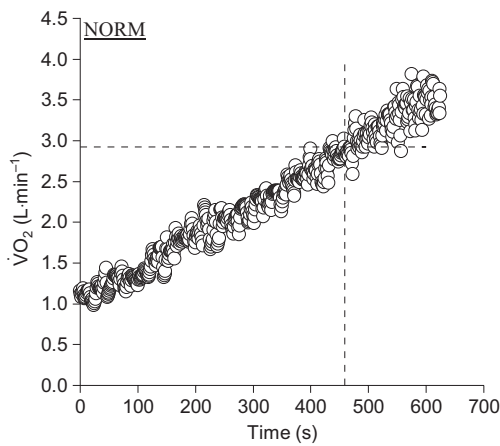
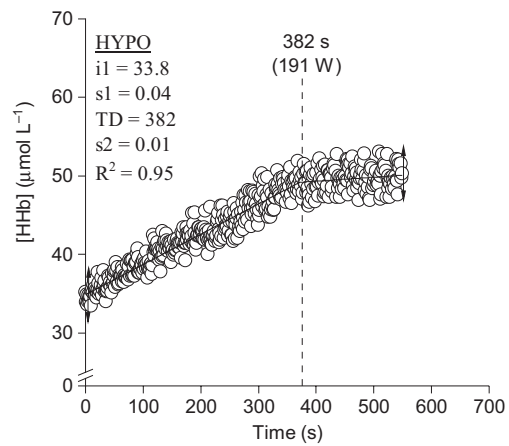
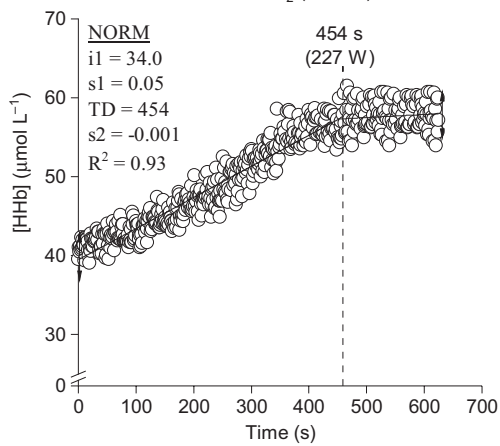
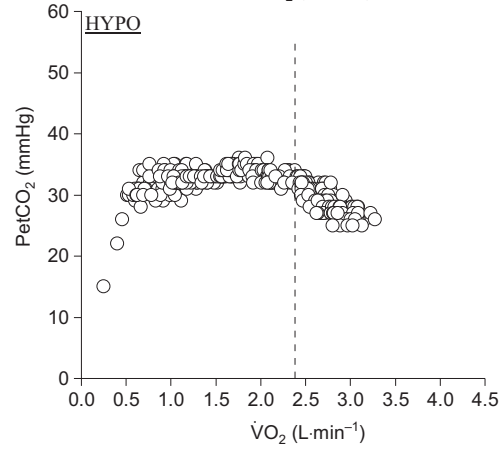
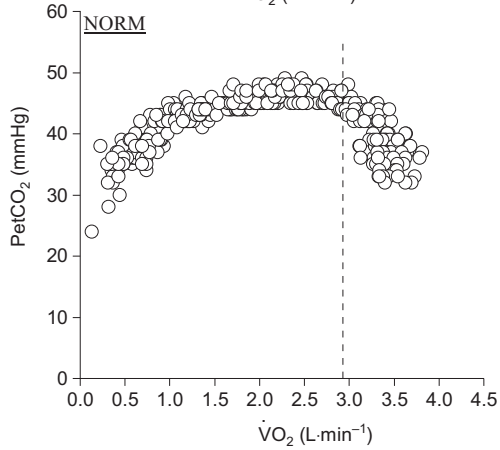
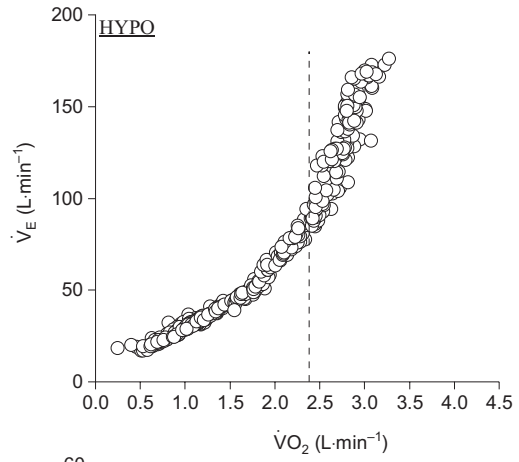
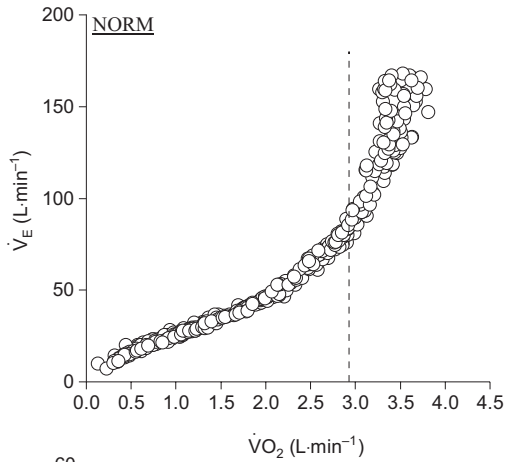
fit  $f$  to  $y$ ,

where  $f$  is the “double-linear” function,  $x$  is time and  $y$  is [HHb], TD (i.e. time delay) is the time coordinate corresponding to the interception of the two regression lines (i.e. [HHb]<sub>bp</sub>),  $i1$  and  $i2$  are the intercepts of the first and second linear function, respectively and  $s1$  and  $s2$  are the slopes (i.e. SL1 and SL2, respectively). Model parameter estimates were determined by linear least-square regression analysis in which the best fit was defined by minimization of the residual sum of squares (RSS) and highest coefficient of determination ( $R^2$ ). The “double-linear” fit was performed plotting the [HHb] data against PO with the fit starting at the onset of the systematic increase in the [HHb] signal until the last data point corresponding to the end of the RI test. Aberrant data that were  $3 \pm SD$  from the local mean were removed. The piecewise equation was selected to fit this specific range of data where the equation parameters (i.e.  $i1$ ,  $s1$ ,  $s2$ , and TD) were not previously fixed and were automatically selected by the mathematical software. Thereafter, the  $\dot{V}O_2$  associated with the [HHb]<sub>bp</sub> was identified after having “left-shifted” the  $\dot{V}O_2$  data (Fontana et al., 2015). Finally, the end-exercise  $SpO_2$  and  $StO_2$  (i.e. last 10% of the RI test) were utilized to compare the effectiveness of distinct  $FiO_2$  in altering the  $O_2$  saturation at the arterial and muscle level, respectively. Figure 1 shows a representative participant in whom the RCP and the [HHb]<sub>bp</sub> were identified.

## 2.5 | Statistical analyses

Descriptive data are presented as mean  $\pm$  standard deviation (*SD*). Data normal distribution was tested by Shapiro–Wilk test and Q-Q plots. End-exercise PPO,  $\dot{V}O_{2\max}$ ,  $SpO_2$ ,  $StO_2$ , and [Lac<sup>-</sup>] were compared using paired samples  $t$  tests. The  $\dot{V}O_2$  and the PO associated with the RCP and the [HHb]<sub>bp</sub> variables (i.e. absolute and relative values) and the [HHb] NIRS-derived slopes before and after the [HHb]<sub>bp</sub> (i.e. SL1 and SL2, respectively) were compared utilizing a two-way repeated measures ANOVA. When  $F$  values were significant, Tukey *post hoc* were used to determine the loci of significant differences. The effect size was computed as partial eta squared ( $\eta_p^2$ ) for ANOVA comparisons [i.e. assuming the small (<0.02), medium (0.02–0.26)

**FIGURE 1** Identification of the RCP and the [HHb]<sub>bp</sub> with associated  $\dot{V}O_2$  during the RI tests performed in normoxia and hypoxia. NORM, normoxia condition ( $FiO_2 = 20.9\%$ ); HYPO, hypoxia condition ( $FiO_2 = 16.0\%$ ); [HHb], deoxygenated hemoglobin;  $\dot{V}O_2$ , oxygen uptake;  $V_E$ , minute ventilation;  $PetCO_2$ , end-tidal partial pressure of  $CO_2$ . Dashed vertical line indicates deoxygenated hemoglobin and respiratory compensation point occurrence among the different variables for the same participant in distinct  $FiO_2$  condition



and large ( $>0.26$ ) effect sizes] (Bakeman, 2005). For two group comparisons as Cohen  $D$  ( $d$ ) [i.e. ranked as trivial (0–0.19), small (0.20–0.49), medium (0.50–0.79), and large (0.80 and greater) effect sizes] (Cumming, 2014). Bland–Altman plot analyses were used to compare the  $\dot{V}O_2$  and PO average errors and limits of agreement (LoA) between the RCP and  $[HHb]_{bp}$  at each  $FiO_2$  condition (Bland & Altman, 1986; Inglis et al., 2019). In addition, the association between values of  $\dot{V}O_2$  and PO at the RCP and the  $[HHb]_{bp}$  were tested by linear regression and Pearson's product moment correlation. Finally, the  $\dot{V}O_2$  and the PO associated with the RCP and the  $[HHb]_{bp}$  in HYPO condition were compared based on the delta change from NORM condition with a Pearson's product moment correlation. The significance level was set at  $p < .05$ . All statistical analyses were performed using a statistical software package (Statistica, version 10.0).

### 3 | RESULTS

#### 3.1 | Ramp-incremental physiological responses and $[HHb]$ signal adjustment in NORM and HYPO conditions

$\dot{V}O_{2max}$  was lower in HYPO ( $2.98 \pm 0.36$  L  $min^{-1}$ ) compared to NORM ( $3.39 \pm 0.26$  L  $min^{-1}$ ) ( $p < .001$ ,  $d = -1.32$ ). Similarly, PPO was lower in HYPO ( $282 \pm 29$  W) than in NORM ( $310 \pm 19$  W) ( $p = .002$ ,  $d = -1.12$ ).  $[Lac^-]$  was not different between conditions (HYPO,  $9.6 \pm 2.6$  mM  $L^{-1}$ ; NORM:  $10.0 \pm 2.3$  mM  $L^{-1}$ ;  $p = .522$ ,  $d = -0.16$ ).  $SpO_2$  and  $StO_2$  in HYPO ( $81\% \pm 4\%$  and  $58\% \pm 11\%$ , respectively) were lower than in NORM ( $90\% \pm 5\%$  vs.  $65\% \pm 10\%$ , respectively) ( $p = .001$  and  $0.007$ ,  $d = -1.85$  and  $-0.63$ ) at the last 10% of the RI test. The slope of the  $[HHb]$  NIRS signal before the  $[HHb]_{bp}$  (i.e. SL1; HYPO,  $0.039 \pm 0.022$   $\mu M s^{-1}$ ; NORM,  $0.044 \pm 0.024$   $\mu M s^{-1}$ ) was greater than after the  $[HHb]_{bp}$  (i.e. SL2; HYPO,  $0.019 \pm 0.022$   $\mu M s^{-1}$ ; NORM,  $0.019 \pm 0.020$   $\mu M s^{-1}$ ) ( $p = .003$ ,  $\eta_p^2 = 0.583$ ) but there was no difference between conditions ( $p = .562$ ,  $\eta_p^2 = 0.034$ ). The “double-linear” fitting parameters for  $[HHb]_{bp}$  identification

showed high values for goodness-of-fit within the data set, in accordance with a previous study (Spencer et al., 2012), as given by the RSS and  $R^2$  for HYPO (RSS =  $709 \pm 269$ ;  $R^2 = 0.92 \pm 0.06$ ) and NORM (RSS =  $658 \pm 247$ ;  $R^2 = 0.93 \pm 0.03$ ) conditions.

#### 3.2 | The $\dot{V}O_2$ and PO associated with the RCP and the $[HHb]_{bp}$ variables between the different $FiO_2$ conditions

The RCP and the  $[HHb]_{bp}$  associated with absolute and relative  $\dot{V}O_2$  and PO values are shown in Table 1. The absolute values for  $\dot{V}O_2$  associated with the RCP and the  $[HHb]_{bp}$  were lower in HYPO compared to NORM condition ( $p < .001$ ,  $\eta_p^2 = 0.785$ ) but there was no difference between variables within the same condition ( $p = .964$ ,  $\eta_p^2 < 0.001$ ). The % $\dot{V}O_{2max}$  associated with the RCP and the  $[HHb]_{bp}$  were neither different between conditions ( $p = .235$ ,  $\eta_p^2 = 0.069$ ) nor variables within the same condition ( $p = .924$ ,  $\eta_p^2 < 0.001$ ). The absolute values for the PO associated with the RCP and the  $[HHb]_{bp}$  were lower in HYPO when compared to NORM ( $p < .001$ ,  $\eta_p^2 = 0.826$ ), but there was no difference between the RCP and the  $[HHb]_{bp}$  within the same condition ( $p = .848$ ,  $\eta_p^2 = 0.001$ ). The %PPO associated with RCP and  $[HHb]_{bp}$  in HYPO were lower compared to NORM ( $p < .001$ ,  $\eta_p^2 = 0.575$ ) but there was no difference between the RCP and the  $[HHb]_{bp}$  within the same condition ( $p = .756$ ,  $\eta_p^2 = 0.004$ ). The MRT obtained for HYPO ( $37 \pm 10$  s) and NORM ( $39 \pm 13$  s) conditions were not different ( $p = .363$ ,  $d = -.329$ ).

#### 3.3 | Correspondence between the RCP and the $[HHb]_{bp}$ variables within the same $FiO_2$ condition

The comparison between the  $\dot{V}O_2$  and the PO associated with the RCP and the  $[HHb]_{bp}$  within the HYPO and NORM conditions are shown in the Bland–Altman plots (Figure 2). The  $\dot{V}O_2$  mean average error for HYPO ( $0.01$  L  $min^{-1}$ , LoA, lower:

**TABLE 1** Respiratory compensation point (RCP) and deoxyhemoglobin breakpoint ( $[HHb]_{bp}$ ) variables in each  $FiO_2$  condition

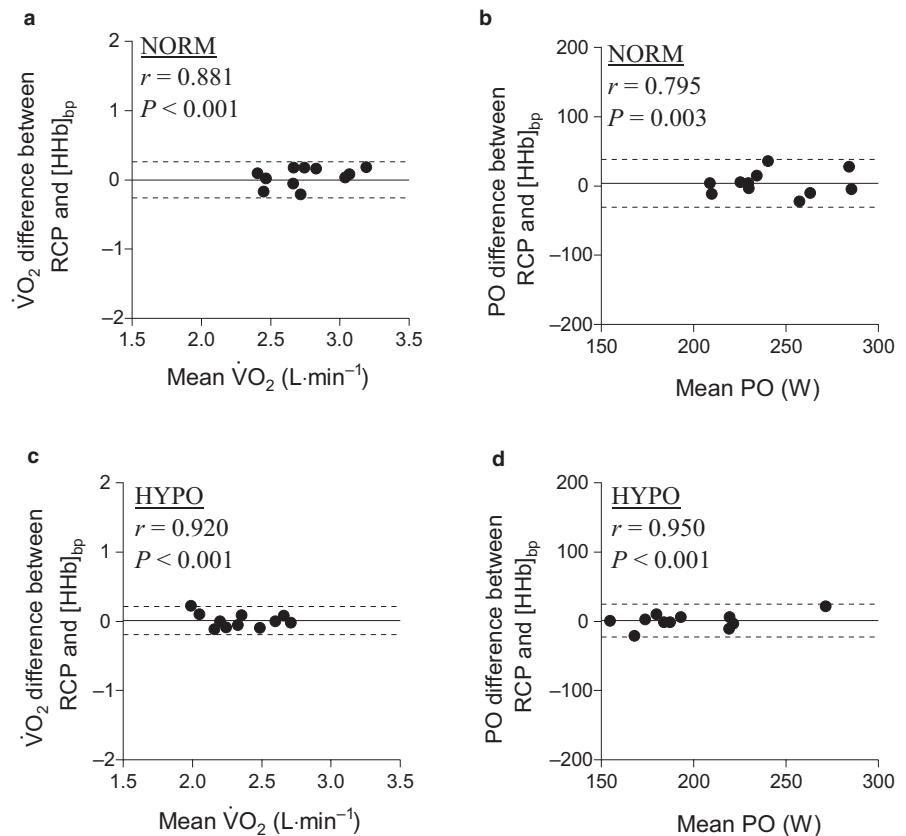
	NORM		HYPO	
	RCP	$[HHb]_{bp}$	RCP	$[HHb]_{bp}$
$\dot{V}O_2$ (L $min^{-1}$ )	$2.75 \pm 0.26^*$ (2.24–3.26)	$2.75 \pm 0.28^*$ (2.19–3.31)	$2.35 \pm 0.24$ (1.89–2.81)	$2.34 \pm 0.26$ (1.82–2.86)
$\dot{V}O_2$ (%)	$81 \pm 5$	$81 \pm 6$	$79 \pm 6$	$78 \pm 5$
PO (W)	$244 \pm 29^*$ (188–300)	$241 \pm 28^*$ (186–294)	$198 \pm 37$ (126–270)	$197 \pm 30$ (138–255)
PO (%)	$79 \pm 6^*$	$78 \pm 7^*$	$69 \pm 7$	$69 \pm 5$

Note: Data are presented as mean  $\pm$  SD and 95% confidence intervals in brackets.

Abbreviations: %, percentage of peak power output and  $\dot{V}O_{2max}$ ;  $[HHb]_{bp}$ , deoxygenated hemoglobin breakpoint; HYPO, hypoxia condition ( $FiO_2 = 16.0\%$ ); NORM, normoxia condition ( $FiO_2 = 20.9\%$ );  $\dot{V}O_2$ , oxygen uptake; PO, power output; RCP, respiratory compensation point.

\*Statistically different from HYPO ( $p < .05$ ).

**FIGURE 2** Bland–Altman plots comparison between the  $\dot{V}O_2$  and the PO associated with the respiratory compensation point (RCP) and the deoxyhemoglobin breakpoint ( $[HHb]_{bp}$ ) for each  $FiO_2$  condition. Black lines indicate mean difference error between the oxygen uptake ( $\dot{V}O_2$ ) associated with RCP and  $[HHb]_{bp}$ ; Dashed black lines indicate lower and upper limits of agreement ( $\pm 2 SD$ ). (a)  $\dot{V}O_2$  associated with RCP and  $[HHb]_{bp}$  in normoxia condition ( $FiO_2 = 20.9\%$ ); (b) Power output associated with RCP and  $[HHb]_{bp}$  in normoxia condition; (c)  $\dot{V}O_2$  associated with RCP and  $[HHb]_{bp}$  in hypoxia condition ( $FiO_2 = 16.0\%$ ); (d) PO associated with RCP and  $[HHb]_{bp}$  in hypoxia condition



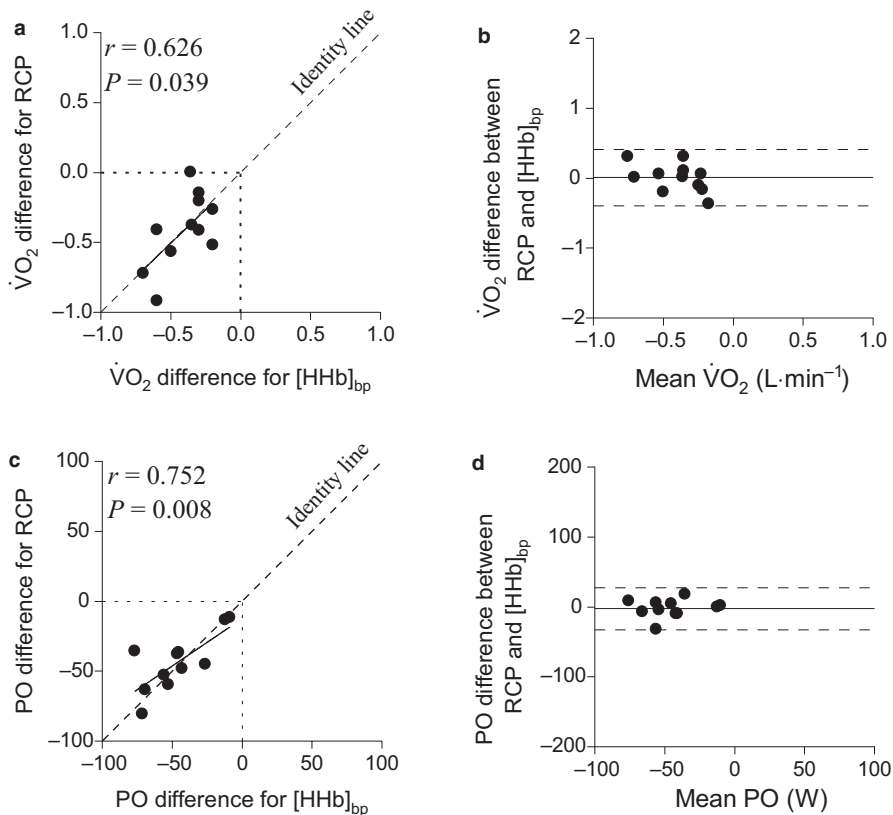
$-0.19 \text{ L}\cdot\text{min}^{-1}$ ; upper,  $0.21 \text{ L}\cdot\text{min}^{-1}$ ;  $p = .761$ ) (Figure 2a) and NORM ( $0.00 \text{ L}\cdot\text{min}^{-1}$ , LoA: lower,  $-0.26 \text{ L}\cdot\text{min}^{-1}$ ; upper,  $0.26 \text{ L}\cdot\text{min}^{-1}$ ;  $p = .997$ ) (Figure 2c) were not different from zero and strongly correlated. The PO mean average error bias for HYPO ( $1.1 \text{ W}$ , LoA, lower:  $-23 \text{ W}$ ; upper,  $25 \text{ W}$ ;  $p = .749$ ) (Figure 2b) and NORM ( $3.6 \text{ W}$ , LoA, lower:  $-31 \text{ W}$ ; upper,  $38 \text{ W}$ ;  $p = .481$ ) (Figure 2d) was also not different from zero and strongly correlated. The change in  $\dot{V}O_2$  and PO associated with the RCP and the  $[HHb]_{bp}$  in HYPO compared to NORM condition are shown in Figure 3. The  $\dot{V}O_2$  and PO associated with RCP and  $[HHb]_{bp}$  were equally decreased in HYPO compared to NORM, as demonstrated by a significant correlation for each variable (Figure 3a and c). Moreover, the mean average error bias for  $\dot{V}O_2$  ( $0.00 \text{ L}\cdot\text{min}^{-1}$ , LoA, lower:  $-0.39 \text{ L}\cdot\text{min}^{-1}$ ; upper,  $0.41 \text{ L}\cdot\text{min}^{-1}$ ;  $p = .877$ ) (Figure 3b) and PO ( $-2 \text{ W}$ , LoA, lower:  $-32 \text{ W}$ ; upper,  $27 \text{ W}$ ;  $p = .572$ ) (Figure 3d) associated with the RCP and the  $[HHb]_{bp}$  in HYPO compared to NORM condition was not statistically different from zero.

## 4 | DISCUSSION

This study investigated whether the strong relationship between the RCP and the  $[HHb]_{bp}$  that is typically observed in normoxia during a RI test (Fontana et al., 2015; Iannetta, Qahtani, Maturana, & Murias, 2017; Iannetta, Qahtani,

Millet, et al., 2017; Inglis et al., 2019; Keir et al., 2015; Murias, Keir, Spencer, & Paterson, 2013) was affected by breathing hypoxic air ( $FiO_2 16\%$ ). The main and novel findings were that: (a) the RCP and  $[HHb]_{bp}$  were equally reduced during hypoxia in terms of both  $\dot{V}O_2$  and PO; (b) these changes were highly correlated, showing small and nonsignificant biases within each individual in each experimental condition (i.e. NORM and HYPO). Taken together, these results indicate that breathing a lower  $FiO_2$  during a RI test results in a proportional reduction in both the  $\dot{V}O_2$  and PO associated with the RCP and the  $[HHb]_{bp}$ . These data further support the notion that these indices of intensity reflect a similar physiological response that is linked to metabolic changes within the active muscles.

In line with previous studies (Knight et al., 1992; Osawa et al., 2011; Richardson et al., 2006), the detrimental effects of hypoxia in the present study are evidenced by the reduced  $SpO_2$  and  $StO_2$ , and lower  $\dot{V}O_{2max}$  and PPO throughout and at the end of RI test, respectively, compared to NORM. Most importantly, the earlier development of metabolic perturbations associated with exercise in hypoxia is evident in the attenuated  $\dot{V}O_2$  and PO at which RCP and  $[HHb]_{bp}$  occurred. Acute changes in  $FiO_2$  have detrimental effects on exercise performance and maximal aerobic capacity due to reduced  $O_2$  delivery and diffusive driving pressure of  $O_2$  at the muscular level (Knight et al., 1992; Osawa et al., 2011; Richardson et al., 2006). Thus, in order to maintain the adequate balance



**FIGURE 3** Correlation and Bland–Altman plots for the  $\dot{V}O_2$  and PO delta difference between hypoxia (HYPO) and normoxia (NORM) conditions for the respiratory compensation point (RCP) and deoxyhemoglobin breakpoint ( $[HHb]_{bp}$ ). (a) correlation for oxygen uptake ( $\dot{V}O_2$ ) delta change for associated with the RCP and the  $[HHb]_{bp}$  between hypoxia and normoxia; (b)  $\dot{V}O_2$  delta change for the RCP and the  $[HHb]_{bp}$  between hypoxia and normoxia conditions; (c) correlation for power output (PO) delta change for associated with the RCP and the  $[HHb]_{bp}$  between hypoxia and normoxia; (d) PO delta change for the RCP and the  $[HHb]_{bp}$  between hypoxia and normoxia conditions. Black lines indicate mean difference error and dashed black lines indicate lower and upper limits of agreement

between  $O_2$  delivery and  $O_2$  utilization, several physiological adjustments need to occur (Calbet, Rådegran, Boushel, & Saltin, 2009). For example, for a given absolute submaximal exercise intensity, there is a greater cardiac output and increased local vasodilatory responses when compared to normoxia (Calbet et al., 2009). In fact, it has been suggested that the hypoxic-induced hyperemia response is proportional to the hypoxia-induced fall in arterial  $O_2$  content (Casey & Joyner, 2011). However, as the exercise intensity increases, these adjustments cannot counteract the reduced driving pressure of  $O_2$  at the capillary-to-muscle interface, limiting mitochondrial ATP resynthesis rate (Richardson, Leigh, Wagner, & Noyszewski, 1999). Thus, during a RI test to exhaustion in hypoxic conditions, there is an earlier reliance on anaerobic resources to sustain the ATP requirement when compared to normoxia (Connett, Honig, Gayeski, & Brooks, 1990; Linnarsson, Karlsson, Fagraeus, & Saltin, 1974). This is also due to increased circulation of catecholamines (i.e. epinephrine and noradrenaline) and an earlier recruitment of type II fibers which increase the glycolytic contribution to the ATP resynthesis in hypoxia compared to normoxia (Moritani, Sherman, Shibata, Matsumoto, & Shinohara, 1992; Osawa et al., 2011). This is evidenced by previous studies showing greater lactate efflux from the muscle to blood and greater depletion of intramuscular phosphagen resources at a given submaximal absolute work rate in hypoxia compared to normoxia (Hogan, Richardson, & Haseler, 1999; Linnarsson et al., 1974). Additionally, the present results and previous

findings (Azevedo et al., 2020; Osawa et al., 2011) have consistently shown that the occurrence of  $[HHb]_{bp}$  is associated with lower absolute PO but the rate of change on  $[HHb]$  signal (i.e. SL1 and SL2) is not different either before or after the  $[HHb]_{bp}$  (Azevedo et al., 2020). Thus, based on those abovementioned mechanisms, it would be plausible to suggest that breathing hypoxic air promotes a “left-shift” in the physiological responses throughout a RI test when compared to normoxia.

In relation to the NORM condition, the present results are in line with previous studies which have shown a correspondence between the RCP and the  $[HHb]_{bp}$  both in terms of  $\dot{V}O_2$  and PO in normoxia condition (Fontana et al., 2015; Iannetta, Qahtani, Millet, et al., 2017; Keir et al., 2015). Murias et al. (2013) were the first to demonstrate that the RCP and the  $[HHb]_{bp}$  occurred at the same  $\dot{V}O_2$  throughout a RI test, which was later confirmed by several other studies in large and heterogenous samples of individuals and in a test–retest conditions (Fontana et al., 2015; Iannetta, Passfield, et al., 2018; Iannetta, Qahtani, Millet, et al., 2017; Inglis et al., 2019; Keir et al., 2015). In direct connection with the goal of this investigation, the present data further confirm the correspondence between the RCP and  $[HHb]_{bp}$  when exercising under distinct  $FiO_2$  conditions. Based on the Bland–Altman plots analysis, the bias obtained for the  $\dot{V}O_2$  associated with the RCP and  $[HHb]_{bp}$  was not different from zero neither in NORM ( $0.00 \text{ L min}^{-1}$ ) nor in HYPO conditions ( $0.01 \text{ L min}^{-1}$ ) (Figure 2). These biases are similar to those



previously reported in other studies (Fontana et al., 2015; Iannetta, Qahtani, Maturana, et al., 2017). Similarly, the PO associated with the RCP and  $[\text{HHb}]_{\text{bp}}$ , displayed biases that were small in NORM (i.e. 3.6 W) and HYPO (i.e. 1.1 W), and similar to previously reported findings (Inglis et al., 2019). In addition to this, the  $\dot{V}\text{O}_2$  and the PO change in HYPO from NORM condition for RCP and  $[\text{HHb}]_{\text{bp}}$  were significantly correlated (Figure 3a and c), showing a bias that was not different from zero (Figure 3b and d). This is an important finding, as it indicates that both, the  $\dot{V}\text{O}_2$  and the PO associated with the RCP and the  $[\text{HHb}]_{\text{bp}}$ , were negatively affected by the same degree in hypoxia.

The fact that the RCP and  $[\text{HHb}]_{\text{bp}}$  continued to occur at the same metabolic rate, independently of the  $\text{FiO}_2$ , provides further support to the idea that these events might be triggered by common physiological responses occurring at the systemic and muscular level (Fontana et al., 2015; Keir et al., 2015). In other words, both the RCP and the  $[\text{HHb}]_{\text{bp}}$  are linked to a metabolic rate corresponding to the transition from the heavy to the severe exercise intensity domain (Keir et al., 2015), which elicits important metabolic changes as a consequence of metabolites accumulation within the muscle and bloodstream (Whipp et al., 1989). For example, up to a critical metabolic rate, increases in ventilation are sufficient to constraint the increases in arterial  $[\text{H}^+]$  linked to  $\text{CO}_2$  production (Wasserman, Beaver, Sun, & Stringer, 2011). However, the increase in rate of  $[\text{H}^+]$  accumulation, when progressively increasing the intensities above the critical metabolic rate, exceeds the capacity of this buffering system, which results in a more rapid accumulation of  $[\text{H}^+]$ , triggering a reflex increase in ventilation (i.e. the RCP) as a result of its uncoupling in relation to  $\text{VCO}_2$ . Even though the exact mechanisms underpinning the hyperventilatory response throughout a RI test, and more specifically above the RCP, are still under debate (Nicolò, Marcora, & Sacchetti, 2020), there are several relevant systemic and local muscular changes that may play a direct effect (Wasserman et al., 2011; Whipp et al., 1989). Furthermore, the transition into the severe intensity domain is characterized not only by an increase in  $[\text{H}^+]$ , with a subsequent reduction in pH, but also by a decrease in the partial pressure of  $\text{O}_2$  within the active muscles, which are known to favor local vasodilation (Casey & Joyner, 2011) and, consequently, redistribution of blood flow to the active tissues. Accordingly, it has been suggested that increased blood flow distribution to the vastus lateralis muscle might be the main mechanism responsible for the observed plateau in the  $[\text{HHb}]$  signal during ramp-incremental exercise (Azevedo et al., 2020; Iannetta, Okushima, et al., 2018; Inglis et al., 2017; Murias, Spencer, et al., 2013), as the improved local blood flow would reduce the need to further increase  $\text{O}_2$  extraction to support the continued increase in muscle  $\dot{V}\text{O}_2$ . This is in line with findings from animal models, whereby the greater activation of type

II fibers when surpassing the heavy-to-severe boundary of exercise intensity triggers disproportionate increases in blood flow in these fibers (Copp, Hirai, Musch, & Poole, 2010). However, although in humans the superficial portions of the vastus lateralis (which are the target of the NIRS light) are characterized by a greater proportion of type II muscle fibers (Johnson, Polgar, Weightman, & Appleton, 1973), this interpretation must be taken with caution because, since the human skeletal muscle is characterized by a mosaic of different fiber types, which may present disparate vascular dynamics and not necessary demonstrate such abrupt changes. In this regard, the leveling off of the  $[\text{HHb}]$  signal in the vastus lateralis muscle has been demonstrated not to be depth dependent (Iannetta, Okushima, et al., 2018) or affected by differential muscle recruitment patterns (Okushima et al., 2020) and exercise modes (Iannetta, Passfield, Qahtani, MacInnis, & Murias, 2019), which would suggest that this phenomenon may not be ascribed to a unique fiber type population.

Alternatively, it has been proposed that even though the RCP and the  $[\text{HHb}]_{\text{bp}}$  might share somewhat similar underpinning mechanisms, they manifest an order of occurrence (Boone, Barstow, et al., 2016; Boone, Vandekerckhove, Coomans, Prieur, & Bourgois, 2016). Accordingly, as intensity increases there is an increasing recruitment of type II fibers leading to the development of metabolic acidosis that triggers the hyperventilatory response (i.e. RCP) and, subsequently, the occurrence of the  $[\text{HHb}]_{\text{bp}}$  as a consequence of the attainment of a supposedly maximal  $\text{O}_2$  extraction (Boone, Barstow, et al., 2016; Boone, Vandekerckhove, et al., 2016). It should be noted, however, that one of these studies has actually shown that the RCP and the  $[\text{HHb}]_{\text{bp}}$  occurred at the same metabolic rate (Boone, Barstow, et al., 2016). Furthermore, it has been recently discussed that discrepancies in views as to whether or not the RCP and the  $[\text{HHb}]_{\text{bp}}$  are expressed simultaneously or in sequential order might be explained by the approach used to account for the MRT, as detailed elsewhere (Boone, Caen, Vermeire, Bourgois, & Bourgois, 2019; Keir et al., 2019).

## 4.1 | Experimental considerations

It should be noted that the goal of this study was to determine whether changes (or the lack thereof) in the  $\dot{V}\text{O}_2$  and PO associated with the RCP and the  $[\text{HHb}]_{\text{bp}}$  when breathing hypoxic air were proportional. In this context, it could be argued that including a hyperoxia condition would have provided additional insights. In fact, this study originally included such condition and, when the RCP and the  $[\text{HHb}]_{\text{bp}}$  in hyperoxia were analyzed (data presented elsewhere (Azevedo et al., 2020)), they occurred at the same  $\% \dot{V}\text{O}_{2\text{max}}$  and  $\% \text{PPO}$  compared to normoxia condition. However, as stated in our previous manuscript (Azevedo et al., 2020) and also reported

by other studies (Amann et al., 2007; Oussaidene et al., 2013; Rausch, Whipp, Wasserman, & Huszczuk, 1991; Ulrich et al., 2017), measurements of  $\dot{V}O_2$  using current systems are often not valid during hyperoxic conditions, (even after attempting corrections as proposed elsewhere (Lang, Herold, Kraft, Harth, & Preisser, 2018)). Therefore, the inclusion of the hyperoxia condition is not appropriate since the  $\dot{V}O_2$  data are not physiologically justifiable.

In conclusion, the present study demonstrated that the RCP and the  $[HHb]_{bp}$  during a RI test occur at the same metabolic rate and PO, even under distinct  $FiO_2$  conditions that alter the absolute values at which these intensity indices occur during the test. Furthermore, and most importantly, the magnitudes of the changes in both the RCP and the  $[HHb]_{bp}$  from normoxic to hypoxic conditions were proportional on an individual basis. Taken together, data from the present study reinforce the idea that the RCP and the  $[HHb]_{bp}$  parameters represent a similar physiological response and that the mechanisms underpinning their occurrence are likely linked to metabolic changes that take place when surpassing the heavy-to-severe boundary of exercise intensity.

## 5 | PERSPECTIVES AND SIGNIFICANCE

This study is the first to evaluate the correspondence between the RCP and the  $[HHb]_{bp}$  during a RI test under distinct  $FiO_2$  conditions (i.e. normoxia and hypoxia). It was found that the  $\dot{V}O_2$  and PO associated with the RCP and the  $[HHb]_{bp}$  were not different within conditions but both reduced under hypoxia condition. Most importantly, the changes in hypoxia, in terms of the  $\dot{V}O_2$  and PO, negatively affected the RCP and the  $[HHb]_{bp}$  in a similar magnitude compared to normoxia. Thus, the present data reinforce the idea that both markers of intensity represent a similar physiological response that delimitates the metabolic rate associated with the transition from the heavy to the severe intensity domain. These findings have important implications to further our understanding on the physiological underpinnings of these markers.

### ACKNOWLEDGMENTS

We express our gratitude to the participants in this study.

### CONFLICT OF INTEREST

The authors declare no conflict of interest.

### ORCID

Rafael D. A. Azevedo  <https://orcid.org/0000-0002-0346-7007>

[org/0000-0002-0346-7007](https://orcid.org/0000-0002-0346-7007)

Juan M. Murias  <https://orcid.org/0000-0002-6460-6453>

## REFERENCES

- Amann, M., Romer, L. M., Subudhi, A. W., Pegelow, D. F., & Dempsey, J. A. (2007). Severity of arterial hypoxaemia affects the relative contributions of peripheral muscle fatigue to exercise performance in healthy humans. *The Journal of Physiology*, *581*(1), 389–403. <https://doi.org/10.1113/jphysiol.2007.129700>
- Azevedo, R. D. A., Béjar Saona, J. E., Inglis, E. C., Iannetta, D., & Murias, J. M. (2020). The effect of the fraction of inspired oxygen on the NIRS-derived deoxygenated hemoglobin “breakpoint” during ramp-incremental test. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, *318*(2), R399–R409. <https://doi.org/10.1152/ajpregu.00291.2019>
- Bakeman, R. (2005). Recommended effect size statistics for repeated measures designs. *Behavior Research Methods*, *37*(3), 379–384. <https://doi.org/10.3758/BF03192707>
- Beaver, W. L., Wasserman, K., & Whipp, B. J. (1986). A new method for detecting anaerobic threshold by gas exchange. *Journal of Applied Physiology*, *60*, 2020–2027. <https://doi.org/10.1152/jappl.1986.60.6.2020>
- Bellotti, C., Calabria, E., Capelli, C., & Pogliaghi, S. (2013). Determination of maximal lactate steady state in healthy adults: Can NIRS help? *Medicine & Science in Sports & Exercise*, *45*(6), 1208–1216. <https://doi.org/10.1249/MSS.0b013e3182828ab2>
- Bland, J. M., & Altman, D. G. (1986). Statistical methods for assessing agreement between two methods of clinical measurement. *Lancet*, *327*, 307–310. [https://doi.org/10.1016/S0140-6736\(86\)90837-8](https://doi.org/10.1016/S0140-6736(86)90837-8)
- Boone, J., Barstow, T. J., Celie, B., Prieur, F., & Bourgois, J. (2016). The interrelationship between muscle oxygenation, muscle activation, and pulmonary oxygen uptake to incremental ramp exercise: Influence of aerobic fitness. *Applied Physiology, Nutrition, and Metabolism*, *41*, 55–62.
- Boone, J., Caen, K., Vermeire, K., Bourgois, G., & Bourgois, J. (2019). The question should be: Is HHb<sub>bp</sub> equivalent to RCP? *Medicine & Science in Sports & Exercise*, *51*(4), 829–<https://doi.org/10.1249/MSS.0000000000001850>
- Boone, J., Vandekerckhove, K., Coomans, I., Prieur, F., & Bourgois, J. G. (2016). An integrated view on the oxygenation responses to incremental exercise at the brain, the locomotor and respiratory muscles. *European Journal of Applied Physiology*, *116*, 2085–2102. <https://doi.org/10.1007/s00421-016-3468-x>
- Broxterman, R. M., Craig, J. C., & Richardson, R. S. (2018). The respiratory compensation point and the deoxygenation break point are not valid surrogates for critical power and maximum lactate steady state. *Medicine & Science in Sports & Exercise*, *50*(11), 2379–2382. <https://doi.org/10.1249/MSS.0000000000001699>
- Caen, K., Vermeire, K., Bourgois, J. G., & Boone, J. (2018). Exercise thresholds on trial: Are they really equivalent? *Medicine & Science in Sports & Exercise*, *50*(6), 1277–1284. <https://doi.org/10.1249/MSS.0000000000001547>
- Calbet, J. A. L., Rådegran, G., Boushel, R., & Saltin, B. (2009). On the mechanisms that limit oxygen uptake during exercise in acute and chronic hypoxia: Role of muscle mass. *The Journal of Physiology*, *587*, 477–490.
- Casey, D. P., & Joyner, M. J. (2011). Local control of skeletal muscle blood flow during exercise: Influence of available oxygen. *Journal of Applied Physiology*, *111*, 1527–1538.
- Connett, R. J., Honig, C. R., Gayeski, T. E. J., & Brooks, G. A. (1990). Defining hypoxia: A systems view of  $VO_2$ , glycolysis, energetics,

- and intracellular PO<sub>2</sub>. *Journal of Applied Physiology*, 68(3), 833–842. <https://doi.org/10.1152/jappl.1990.68.3.833>.
- Copp, S. W., Hirai, D. M., Musch, T. I., & Poole, D. C. (2010). Critical speed in the rat: Implications for hindlimb muscle blood flow distribution and fibre recruitment. *The Journal of Physiology*, 588, 5077–5087.
- Cumming, G. (2014). The new statistics: Why and how. *Psychological Science*, 25, 7–29.
- Fontana, F. Y., Keir, D. A., Bellotti, C., De Roia, G. F., Murias, J. M., & Pogliaghi, S. (2015). Determination of respiratory point compensation in healthy adults: Can non-invasive near-infrared spectroscopy help? *Journal of Science and Medicine in Sport*, 18(5), 590–595. <https://doi.org/10.1016/j.jsams.2014.07.016>
- Grassi, B., & Quaresima, V. (2016). Near-infrared spectroscopy and skeletal muscle oxidative function in vivo in health and disease: a review from an exercise physiology perspective. *J. Biomed. Opt.*, 21(9). <https://doi.org/10.1117/1.JBO.21.9.091313>.
- Hogan, M. C., Richardson, R. S., & Haseler, L. J. (1999). Human muscle performance and PCr hydrolysis with varied inspired oxygen fractions: A31P-MRS study. *Journal of Applied Physiology*, 86, 1367–1373.
- Iannetta, D., Murias, J. M., & Keir, D. A. (2019). A simple method to quantify the VO<sub>2</sub> mean response time of ramp-incremental exercise. *Medicine & Science in Sports & Exercise*, 51(5), 1080–1086. <https://doi.org/10.1249/MSS.0000000000001880>
- Iannetta, D., Okushima, D., Inglis, E. C., Kondo, N., Murias, J. M., & Koga, S. (2018). Blood flow occlusion-related O<sub>2</sub> extraction “reserve” is present in different muscles of the quadriceps but greater in deeper regions after ramp-incremental test. *Journal of Applied Physiology*, 125, 313–319.
- Iannetta, D., Passfield, L., Murias, J. M., Calaine, E., & Christopher, I. (2018). Metabolic and performance-related consequences of exercising at and slightly above MLSS. *Scandinavian Journal of Medicine & Science in Sports*, 28(12), 2481–2493. <https://doi.org/10.1111/sms.13280>
- Iannetta, D., Passfield, L., Qahtani, A., MacInnis, M. J., & Murias, J. M. (2019). Interlimb differences in parameters of aerobic function and local profiles of deoxygenation during double-leg and counterweighted single-leg cycling. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 317(6), R840–R851. <https://doi.org/10.1152/ajpregu.00164.2019>
- Iannetta, D., Qahtani, A., Maturana, F. M., & Murias, J. M. (2017). The near-infrared spectroscopy-derived deoxygenated haemoglobin breaking-point is a repeatable measure that demarcates exercise intensity domains. *Journal of Science and Medicine in Sport*, 20(9), 873–877. <https://doi.org/10.1016/j.jsams.2017.01.237>
- Iannetta, D., Qahtani, A., Millet, G. Y., & Murias, J. M. (2017). Quadriceps muscles O<sub>2</sub> extraction and EMG breakpoints during a ramp incremental test. *Frontiers in Physiology*, 8, 1–9. <https://doi.org/10.3389/fphys.2017.00686>
- Inglis, E. C., Iannetta, D., Keir, D. A., & Murias, J. M. (2019). Training-induced changes in the RCP, [HHb]BP and MLSS: Evidence of equivalence. *International Journal of Sports Physiology and Performance*, 1, 1–2. <https://doi.org/10.1123/ijssp.2019-0046>.
- Inglis, E. C., Iannetta, D., & Murias, X. J. M. (2017). The plateau in the NIRS-derived [HHb] signal near the end of a ramp incremental test does not indicate the upper limit of O<sub>2</sub> extraction in the vastus lateralis. *The American Journal of Physiology-Regulatory, Integrative and Comparative*, 313, 723–729.
- Johnson, M. A., Polgar, J., Weightman, D., & Appleton, D. (1973). Data on the distribution of fibre types in thirty-six human muscles. *Journal of the Neurological Sciences*, 18(1), 111–129. [https://doi.org/10.1016/0022-510X\(73\)90023-3](https://doi.org/10.1016/0022-510X(73)90023-3)
- Keir, D. A., Fontana, F. Y., Robertson, T. C., Murias, J. M., Paterson, D. H., Kowalchuk, J. M., & Pogliaghi, S. (2015). Exercise intensity thresholds: Identifying the boundaries of sustainable performance. *Medicine and Science in Sports and Exercise*, 47, 1932–1940.
- Keir, D. A., Pogliaghi, S., & Murias, J. M. (2019). Response. *Medicine & Science in Sports & Exercise*, 51(4), 830–<https://doi.org/10.1249/MSS.0000000000001851>
- Knight, D. R., Poole, D. C., Schaffartzik, W., Guy, H. J., Prediletto, R., Hogan, M. C., & Wagner, P. D. (1992). Relationship between body and leg VO<sub>2</sub> during maximal cycle ergometry. *Journal of Applied Physiology*, 73(3), 1114–1121. <https://doi.org/10.1152/jappl.1992.73.3.1114>
- La Monica, M. B., Fukuda, D. H., Starling-Smith, T. M., Wang, R., Hoffman, J. R., & Stout, J. R. (2018). Effects of normobaric hypoxia on upper body critical power and anaerobic working capacity. *Respiratory Physiology & Neurobiology*, 249, 1–6. <https://doi.org/10.1016/j.resp.2017.12.002>
- Lamarra, N., Whipp, B. J., Ward, S. A., & Wasserman, K. (2017). Effect of interbreath fluctuations on characterizing exercise gas exchange kinetics. *Journal of Applied Physiology*, 62(5), 2003–2012. <https://doi.org/10.1152/jappl.1987.62.5.2003>
- Lang, S., Herold, R., Kraft, A., Harth, V., & Preisser, A. M. (2018). Spiroergometric measurements under increased inspiratory oxygen concentration (FIO<sub>2</sub>)—Putting the Haldane transformation to the test. *PLoS One*, 13, e0207648. <https://doi.org/10.1371/journal.pone.0207648>
- Leo, J. A., Sabapathy, S., Simmonds, M. J., & Cross, T. J. (2017). Response to editor-in-chief. *Medicine & Science in Sports & Exercise*, 49(12), 2609. <https://doi.org/10.1249/MSS.0000000000001376>
- Linnarsson, D., Karlsson, J., Fagraeus, L., & Saltin, B. (1974). Muscle metabolites and oxygen deficit with exercise in hypoxia and hyperoxia. *Journal of Applied Physiology*, 36(4), 399–402. <https://doi.org/10.1152/jappl.1974.36.4.399>
- Moritani, T., Sherman, W. M., Shibata, M., Matsumoto, T., & Shinohara, M. (1992). Oxygen availability and motor unit activity in humans. *European Journal of Applied Physiology and Occupational Physiology*, 64(6), 552–556. <https://doi.org/10.1007/BF00843767>
- Murias, J. M., Keir, D. A., Spencer, M. D., & Paterson, D. H. (2013). Sex-related differences in muscle deoxygenation during ramp incremental exercise. *Respiratory Physiology & Neurobiology*, 189(0), 530–536. <https://doi.org/10.1016/j.resp.2013.08.011>
- Murias, J. M., Spencer, M. D., Keir, D. A., & Paterson, D. H. (2013). Systemic and vastus lateralis muscle blood flow and O<sub>2</sub> extraction during ramp incremental cycle exercise. *The American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 304, R720–R725.
- Nicolò, A., Marcora, S. M., & Sacchetti, M. (2020). Time to reconsider how ventilation is regulated above the respiratory compensation point during incremental exercise. *Journal of Applied Physiology*, 128(5), 1447–1449. <https://doi.org/10.1152/japphysiol.00814.2019>
- Okushima, D., Poole, D. C., Barstow, T. J., Kondo, N., Chin, L. M. K., & Koga, S. (2020). Effect of differential muscle activation patterns on muscle deoxygenation and microvascular haemoglobin

- regulation. *Experimental Physiology*, 105(3), 531–541. <https://doi.org/10.1113/EP088322>
- Okushima, D., Poole, D. C., Rossiter, H. B., Barstow, T. J., Kondo, N., Ohmae, E., & Koga, S. (2015). Muscle deoxygenation in the quadriceps during ramp incremental cycling: Deep vs. superficial heterogeneity. *Journal of Applied Physiology*, 119, 1313–1319.
- Osawa, T., Kime, R., Hamaoka, T., Katsumura, T., & Yamamoto, M. (2011). Attenuation of muscle deoxygenation precedes emg threshold in normoxia and hypoxia. *Medicine & Science in Sports & Exercise*, 43(8), 1406–1413. <https://doi.org/10.1249/MSS.0b013e3182100261>
- Oussaidene, K., Prieur, F., Bougault, V., Borel, B., Matran, R., & Mucci, P. (2013). Cerebral oxygenation during hyperoxia-induced increase in exercise tolerance for untrained men. *European Journal of Applied Physiology*, 113(8), 2047–2056. <https://doi.org/10.1007/s00421-013-2637-4>
- Parker Simpson, L., Jones, A. M., Skiba, P. F., Vanhatalo, A., & Wilkerson, D. (2014). Influence of hypoxia on the power-duration relationship during high-intensity exercise. *International Journal of Sports Medicine*, 36(02), 113–119. <https://doi.org/10.1055/s-0034-1389943>
- Rausch, S. M., Whipp, B. J., Wasserman, K., & Huszczuk, A. (1991). Role of the carotid bodies in the respiratory compensation for the metabolic acidosis of exercise in humans. *The Journal of Physiology*, 444, 567–578. <https://doi.org/10.1113/jphysiol.1991.sp018894>
- Richardson, R. S., Duteil, S., Wary, C., Wray, D. W., Hoff, J., & Carlier, P. G. (2006). Human skeletal muscle intracellular oxygenation: The impact of ambient oxygen availability. *The Journal of Physiology*, 571, 415–424.
- Richardson, R. S., Leigh, J. S., Wagner, P. D., & Noyszewski, E. A. (1999). Cellular PO<sub>2</sub> as a determinant of maximal mitochondrial O<sub>2</sub> consumption in trained human skeletal muscle. *The Journal of Applied Physiology*, 87, 325–331.
- Spencer, M. D., Murias, J. M., & Paterson, D. H. (2012). Characterizing the profile of muscle deoxygenation during ramp incremental exercise in young men. *European Journal of Applied Physiology*, 112(9), 3349–3360. <https://doi.org/10.1007/s00421-012-2323-y>
- Townsend, N. E., Nichols, D. S., Skiba, P. F., Racinais, S., & Périard, J. D. (2017). Prediction of critical power and w' in hypoxia: Application to work-balance modelling. *Frontiers in Physiology*, 8, 1–9.
- Ulrich, S., Hasler, E. D., Müller-Mottet, S., Keusch, S., Furian, M., Latshang, T. D., ... Bloch, K. E. (2017). Mechanisms of improved exercise performance under hyperoxia. *Respiration*, 93, 90–98. <https://doi.org/10.1159/000453620>
- Wasserman, K., Beaver, W. L., Sun, X.-G., & Stringer, W. W. (2011). Arterial H<sup>+</sup> regulation during exercise in humans. *Respiratory Physiology & Neurobiology*, 178(2), 191–195. <https://doi.org/10.1016/j.resp.2011.05.018>
- Whipp, B. J., Davis, J. A., & Wasserman, K. (1989). Ventilatory control of the 'isocapnic buffering' region in rapidly-incremental exercise. *Respiration Physiology*, 76(3), 357–367. [https://doi.org/10.1016/0034-5687\(89\)90076-5](https://doi.org/10.1016/0034-5687(89)90076-5)

**How to cite this article:** Azevedo RDA, Saona JEB, Inglis EC, Iannetta D, Murias JM. Hypoxia equally reduces the respiratory compensation point and the NIRS-derived [HHb] breakpoint during a ramp-incremental test in young active males. *Physiol Rep.* 2020;8:e14478. <https://doi.org/10.14814/phy2.14478>