

A Phase 2, Randomized, Double-blind, Placebo-Controlled Trial of Presatovir for the Treatment of Respiratory Syncytial Virus Upper Respiratory Tract Infection in Hematopoietic-Cell Transplant Recipients

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(See the Major Article by Marty et al on pages 2787–95 and the Editorial Commentary by Löwensteyn and Bont on pages 2796–8.)

Background. Hematopoietic-cell transplant (HCT) recipients are at risk for severe respiratory syncytial virus (RSV) infection. We evaluated the RSV fusion inhibitor presatovir in a randomized, double-blind, Phase II trial in HCT recipients with RSV upper respiratory tract infections.

Methods. Patients were stratified by lymphopenia ($<200/\mu$ L) and ribavirin use; were randomized, stratified by lymphopenia ($<200/\mu$ L) and ribavirin use, to receive oral presatovir at 200 mg or a placebo on Days 1, 5, 9, 13, and 17, and were followed through Day 28. The coprimary efficacy endpoints were the time-weighted average change in the nasal RSV viral load between Days 1 and 9 and the proportion of patients developing lower respiratory tract complications (LRTCs) through Day 28.

Results. From 23 January 2015 to 16 June 2017, 189 patients were randomly assigned to treatment (96 to presatovir and 93 to the placebo). Presatovir treatment, compared with the placebo treatment, did not significantly affect (prespecified $\alpha = 0.01$) a time-weighted average decline in the RSV viral load from Day 1 to 9 (treatment difference, $-0.33 \log_{10}$ copies/mL; 95% confidence interval [CI] -.64 to $-.02 \log_{10}$ copies/mL; P = .040) or the progression to LRTC (11.2% vs 19.5%, respectively; odds ratio, 0.50; 95% CI, .22–1.18; P = .11). In a post hoc analysis among patients with lymphopenia, presatovir decreased LRTC development by Day 28 (2/15 [13.3%] vs 9/14 [64.3%], respectively; P = .008), compared with the placebo. Adverse events were similar for patients receiving presatovir and the placebo.

Conclusions. Presatovir had a favorable safety profile in adult HCT recipients with RSV but did not achieve the coprimary endpoints. Exploratory analyses suggest an antiviral effect among patients with lymphopenia.

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Adult recipients of autologous or allogeneic hematopoietic-cell transplants (HCT) are at high risk for respiratory syncytial virus (RSV) infection and associated morbidity and mortality. Up to 17% of HCT recipients may develop an RSV infection [1–7], of whom 17% to 84% progress from an upper respiratory tract infection (URTI) to a lower respiratory tract infection (LRTI) [2, 3, 5, 7–14]. Progression to a LRTI often requires hospitalization, during which oxygen supplementation and intensive care may be required; RSV LRTIs are associated with increased mortality rates, ranging from 6% to 35% [2, 4, 8, 9, 15–20]. Survivors of a

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respiratory viral infection after HCT may have a long-term air-flow decline [15, 21].

Currently, there are no effective vaccines or approved antiviral agents for RSV infection in HCT recipients. Aerosolized ribavirin (Virazole) is approved for the treatment of RSV infections in young children but is not used in general pediatric practice because of efficacy and tolerability concerns and the complexity of the required specialized aerosol delivery system [22-24]. A randomized, placebo-controlled trial of aerosolized ribavirin attempted in HCT recipients recruited only 14 subjects in 5 years due to slow accrual [25]. Epidemiologic studies and a single-center, retrospective analysis suggest that ribavirin-based therapy has some efficacy for preventing RSV-associated morbidity or mortality in high-risk HCT recipients [6, 14, 18]. However, these results are from uncontrolled, retrospective studies and the ribavirin benefit remains unconfirmed. Thus, there remains a significant, unmet medical need for safe, convenient, and effective treatments for RSV infection.

Presatovir (formerly GS-5806) is an oral RSV fusion inhibitor with potent and selective anti-RSV activity in vitro and a terminal half-life of ~34 hours [26]. When tested in a human challenge study of healthy volunteers, presatovir reduced the RSV viral load and the severity of clinical disease [26]. In the current study, we evaluated presatovir's safety, tolerability, and efficacy among HCT recipients with RSV URTIS.

PATIENTS AND METHODS

Patients and Study Design

This Phase II, randomized, double-blind, placebo-controlled, 2-group, parallel study recruited allogeneic or autologous HCT recipients with positive local RSV test results who were 18 to 75 years of age from 43 centers in 9 countries (Supplementary Appendix). Patients with new or worsening respiratory symptoms for \leq 7 days, who had been diagnosed with an RSV infection of the upper respiratory tract for \leq 6 days, and who were without new abnormalities on a chest X-ray obtained <48 hours from the start of study treatment were eligible to participate. Patients with a specified, documented respiratory virus coinfection within 7 days from the start of study treatment or with another significant respiratory or systemic infection were excluded. The full eligibility criteria are provided in the Supplementary Methods.

This study followed International Conference on Harmonization requirements and the principles of the Declaration of Helsinki and was approved by local ethics committees. Written informed consent was obtained from patients or their legally responsible representatives. The protocol amendments and Data Monitoring Committee activities are described in the Supplementary Methods. The trial was registered at ClinicalTrials.gov (NCT02254408) and EudraCT (2014-002474-36) before enrollment began.

Randomization and Masking

Patients were randomly assigned (1:1) to receive presatovir or a placebo, were stratified centrally by lymphopenia (lymphocyte count <200 cells/mm³ within 6 days of screening), and were prescribed the use of ribavirin by any route of administration at randomization. The study treatment assignments were provided by an interactive web response system (Bracket Global, Wayne, PA, USA). Patients, all study staff, and the study sponsor were blinded to study treatment. Allocation was concealed by the use of presatovir and placebo tablets that were identical in appearance.

Procedures

The patients received presatovir at 200 mg (4 × 50 mg tablets) or a placebo orally or by a nasogastric tube during study visits on Days 1, 5, 9, 13, and 17 (\pm 24 hours), and were followed through study Day 28. Based on human pharmacokinetic and pharmacodynamic studies [26], this regimen was predicted to provide plasma concentrations >4-fold over requirements to inhibit the replication of >95% of tested RSV isolates. Patients with detectable RSV by reverse transcription quantitative polymerase chain reaction (RT-qPCR) on Day 22 could participate in an optional extended, weekly follow-up through Day 56. A detailed schedule of the study assessments and procedures is provided in Supplementary Table 1.

Plasma pharmacokinetic methods are described in the Supplementary Methods. For virology assessments, bilateral intranasal swabs were obtained using mid-turbinate, adult, flocked swabs (Copan Diagnostics, Murrieta, CA) at each study visit. Samples were analyzed using RT-qPCR to measure the RSV viral load; RSV F gene sequencing, to detect the development of resistance; and a multiplex assay to identify respiratory viral coinfections. All nasal samples were analyzed at central laboratories; further methodological details are provided in the Supplementary Methods. Chest X-rays or computed tomography scans were performed per standard care in patients with suspected lower respiratory tract complications (LRTC). Imaging studies and results of local microbiology tests were collected for review by the endpoint adjudication committee.

Clinical safety assessments included vital signs, body weight, and oxygen saturation by pulse oximetry; laboratory safety assessments included complete blood cell counts and liver enzyme measurements. Cardiac safety was assessed via electrocardiograms and troponin testing (per US Food and Drug Administration cardiac monitoring requirements) on Days 1, 17, and 28. Additional safety assessments included the evaluation of adverse events (AEs) and the documentation of concomitant medications.

Outcomes

The coprimary endpoints were the time-weighted, average change in the nasal RSV viral load, measured by RT-qPCR (log₁₀ copies/mL) between Day 1 and Day 9, and the proportion

of patients who developed LRTCs—defined as a primary RSV LRTI, a secondary bacterial LRTI, a lower respiratory tract infection due to unusual pathogens, or an LRTC of unknown etiology—from Day 1 through Day 28. The development of an LRTC was determined by an independent, blinded endpoint adjudication committee (details are in the Supplementary Methods). The secondary efficacy endpoint was the proportion of patients who died or developed respiratory failure requiring invasive mechanical ventilation from Day 1 to Day 28. Safety was assessed from AEs, vital signs, electrocardiograms, and clinical laboratory test results.

Statistical Analysis

Assuming a time-weighted average change in the RSV viral load from Day 1 to Day 9 of $-1 \log_{10}$ copies/mL with a standard deviation (SD) of 2 \log_{10} and an LRTC event rate of 30% in patients receiving the placebo, 100 patients per treatment group were planned to provide >80% power to detect a $\ge 1-\log_{10}$ decrease in the first coprimary endpoint with a 2-sided α of 0.01 and >90% power to detect a $\ge 20\%$ reduction in the second coprimary endpoint with a 2-sided α of 0.04 in patients receiving presatovir, relative to the placebo.

The efficacy population included patients who received ≥ 1 dose of presatovir with a quantifiable nasal RSV viral load on Day 1. The coprimary and secondary endpoints were analyzed in the efficacy population and in prespecified subgroups defined by the randomization stratification factors (lymphopenia and ribavirin use on Day 1), and were also analyzed post hoc in subgroups defined by the duration of RSV symptoms, hospitalization status, time after HCT, and graft-vs-host disease (GVHD) status on Day 1. The safety population included patients who received ≥ 1 dose of presatovir.

The first coprimary analysis was performed by a parametric analysis of covariance with the baseline viral load and randomization stratification factors as covariates. The second coprimary analysis and secondary efficacy analysis were performed using 2-sided Cochran-Mantel-Haenszel tests stratified by lymphopenia (<200 cells/mm³) and the intent to use ribavirin at baseline. If the number of events was small, the Fisher exact method was applied. A fallback approach was employed to control the Type I error rate at 0.05 across the coprimary and secondary endpoints (details are in the Supplementary Methods). Subgroup analyses were performed using the corresponding analysis of covariance model for the first coprimary endpoint and the Fisher exact test, with a 95% confidence interval (CI) based on the Clopper-Pearson method, for the second coprimary and secondary endpoints.

RESULTS

Patients

From 23 January 2015 to 16 June 2017, 213 patients were screened for eligibility; 24 patients were excluded, the majority

(n = 14) of whom did not have a documented RSV infection of the upper respiratory tract. A total of 189 patients were randomly assigned to a study treatment (96 to presatovir and 93 to the placebo), and 185 received ≥ 1 dose of a study drug (95 received presatovir and 90 received the placebo; Figure 1). The sponsor halted the study on 20 September 2017, before achieving the planned 200-subject enrollment, because an unplanned interim analysis before a database lock by an unblinded team indicated that results were unlikely to differ if enrollment was extended through another RSV season. Important protocol deviations are described in the Supplementary Results and Supplementary Table 2. Overall, 168 (90.8%) patients (88 assigned to presatovir and 80 to the placebo) completed treatment with a study drug through Day 17 (Figure 1).

Patient demographic and baseline clinical characteristics were generally well balanced between treatment groups, except for hospitalization of a larger number of patients receiving presatovir, compared with the placebo, at the beginning of study treatment (43.2% vs 26.7%, respectively; Table 1). The majority of treated patients (146/185, 78.9%) underwent allogeneic HCT, and 69/185 (37.3%) had GVHD at baseline. Lymphopenia was noted in 29 (15.7%) patients, and 44 (23.8%) patients were treated with aerosolized or oral ribavirin at baseline (Table 1).

Efficacy

Figure 2A-B shows the absolute RSV viral load and change from baseline at each study visit. Despite adequate plasma concentrations (Supplementary Results; Supplementary Table 3), presatovir did not significantly (prespecified $\alpha = 0.01$) reduce the time-weighted average change in the RSV viral load from Day 1 to Day 9, compared with the placebo (mean, -1.26 [SD, 0.964] log₁₀ copies/mL vs -0.91 [SD, 1.145] log₁₀ copies/mL, respectively; treatment difference, -0.33 log₁₀ copies/mL; 95% CI, -.64 to $-.02 \log_{10}$ copies/mL; P = .040). The development of LRTCs through Day 28 is shown in Figure 3. Compared with the placebo, presatovir did not significantly reduce the proportion of patients in the efficacy population who developed an LRTC from Day 1 through Day 28 (10/89 [11.2%] on presatovir vs 17/87 [19.5%] on placebo; P = .11; $\alpha = 0.04$). The majority of LRTC events were adjudicated as having an unknown etiology (presatovir, 7/10 [70%]; placebo, 15/17 [88%]). There were 2 events in each treatment arm that were attributed to a primary RSV LRTI, and 1 event in the presatovir arm was adjudicated as a secondary bacterial infection. Sensitivity analyses are reported in the Supplementary Results. Death or respiratory failure requiring mechanical ventilation through Day 28 occurred in 5/89 (5.6%) patients receiving presatovir and 5/87 (5.7%) patients receiving the placebo (P = .98; Figure 4).

In prespecified subgroup analyses, presatovir numerically decreased the proportion of patients who developed an LRTC from Day 1 through Day 28, relative to the placebo, among patients with baseline lymphopenia (2/15 [13.3%] vs 9/14 [64.3%],

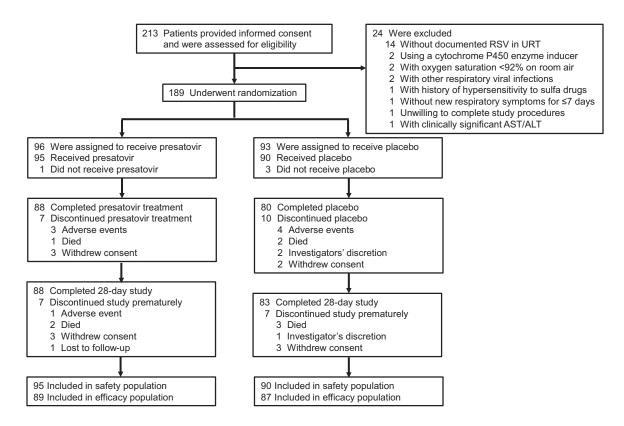


Figure 1. Patient disposition from enrollment through analysis. Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase; RSV, respiratory syncytial virus; URT, upper respiratory tract.

respectively; P = .008) and those not receiving ribavirin (4/64 [6.3%] vs [12/68] 17.6%, respectively; P = .061; Table 2; Supplementary Tables 4 and 5). The proportions of patients receiving presatovir who developed LRTC were similar among patients without baseline lymphopenia and in patients without

ribavirin use at baseline, as compared to those receiving the placebo (Supplementary Tables 4 and 5). Overall, ribavirin use was higher among patients who developed an LRTC (37.0%) versus those who did not (23.5%). Patients hospitalized at baseline had a numerically higher rate of LRTCs, relative to those who

Table 1. Baseline Characteristics and Demographics: Safety Population

	Patients Given Presatovir, n = 95	Patients Given Placebo, n = 90	Total, N = 185
Age, years, median (min, max)	54 (22, 70)	53 (20, 75)	54 (20, 75)
Male sex at birth	55 (57.9)	55 (61.1)	110 (59.5)
Ethnic origin			
White	66 (69.5)	70 (77.8)	136 (73.5)
Asian	13 (13.7)	9 (10.0)	22 (11.9)
African American or African	6 (6.3)	3 (3.3)	9 (4.9)
Other	2 (2.1)	0	2 (1.1)
Not documented	8 (8.4)	8 (8.9)	16 (8.6)
Hispanic or Latino	8 (8.4)	6 (6.7)	14 (7.6)
Body mass index, kg/m², median (min, max)ª	25.0 (13.6, 49.8)	24.3 (16.8, 46.0)	24.6 (13.6, 49.8
Lymphopenia, <200 cells/µL, at randomization	15 (15.8)	14 (15.6)	29 (15.7)
Ribavirin use at randomization	25 (26.3)	19 (21.1)	44 (23.8)
Route of administration ^b			
Aerosolized	4/25 (16.0)	5/19 (26.3)	9/44 (20.5)
Oral	21/25 (84.0)	14/19 (73.7)	35/44 (79.5)
RSV type			
RSV A	44 (46.3)	43 (47.8)	87 (47.0)
RSV B	44 (46.3)	43 (47.8)	87 (47.0)
Both RSV A and RSV B	2 (2.1)	1 (1.1)	3 (1.6)

Table 1. Continued

	Patients Given Presatovir, n = 95	Patients Given Placebo, n = 90	Total, N = 185
Undetectable	5 (5.3)	1 (1.1)	6 (3.2)
Missing	0	2 (2.2)	2 (1.1)
Nasal RSV RNA, log ₁₀ copies/mL, ^c median (min, max)	7.00 (0, 8.51)	7.10 (0, 8.94)	7.00 (0, 8.94)
Respiratory symptom duration before Day 1, days, median (min, max)	4 (1, 7)	4 (1, 10) ^d	4 (1, 10)
Oxygen saturation, %, median (min, max)	96 (87, 100)	96 (90, 100)	96 (87, 100)
Smoking history			
Never	52 (54.7)	52 (57.8)	104 (56.2)
Former	40 (42.1)	35 (38.9)	75 (40.5)
Current	3 (3.2)	3 (3.3)	6 (3.2)
Other respiratory viruses detected			
Rhinovirus or enterovirus	2 (2.1)	3 (3.3)	5 (2.7)
Adenovirus	1 (1.1)	1 (1.1)	2 (1.1)
Coronavirus 229E	0	3 (3.3)	3 (1.6)
Coronavirus HKU1	1 (1.1)	1 (1.1)	2 (1.1)
Coronavirus NL63	0	1 (1.1)	1 (0.5)
Coronavirus OC43	1 (1.1)	0	1 (0.5)
Parainfluenza 1	1 (1.1)	0	1 (0.5)
Parainfluenza 2	1 (1.1)	0	1 (0.5)
Hospitalized on Day 1	41 (43.2)	24 (26.7)	65 (35.1)
Unplanned hospitalization	27 (65.9)	11 (45.8)	38 (58.5)
Planned hospitalization	14 (34.1)	13 (54.2)	27 (41.5)
Hospitalization related to RSV infection	24 (58.5)	8 (33.3)	32 (49.2)
Hospitalization days before Day 1, median (min, max)	0 (0, 48)	0 (0, 75)	0 (0, 75)
Hematopoietic-cell transplant type			
Allogeneic HCT	72 (75.8)	74 (82.2)	146 (78.9)
Autologous HCT	23 (24.2)	16 (17.8)	39 (21.1)
Time from HCT to Day 1, days, median (min, max) ^e	278 (2, 4000)	275 (1, 7538)	278 (1, 7538
Underlying hematologic disease			
Acute leukemia	44 (46.3)	49 (54.4)	93 (50.3)
Myeloma	24 (25.3)	13 (14.4)	37 (20.0)
Lymphoma	11 (11.6)	14 (15.6)	25 (13.5)
Refractory anemia	1 (1.1)	0	1 (0.5)
Chronic lymphocytic leukemia	4 (4.2)	1 (1.1)	5 (2.7)
Other	15 (15.8)	13 (14.4)	28 (15.1)
Acute or chronic graft-vs-host disease	10 (1010)		20 (1011)
Yes	33 (34.7)	36 (40.0)	69 (37.3)
No	37 (38.9)	37 (41.1)	74 (40.0)
Not applicable, autologous HCT	23 (24.2)	16 (17.8)	39 (21.1)
Unknown	2 (2.1)	1 (1.1)	3 (1.6)
HCT donor type	2 (2.1)	1 (1.17	0 (1.0)
Unrelated	44 (46.3)	35 (38.9)	79 (42.7)
Matched-related	24 (25.3)	32 (35.6)	56 (30.3)
Mismatched-related	3 (3.2)	6 (6.7)	9 (4.9)
Autologous	23 (24.2)	17 (18.9)	40 (21.6)
Unknown	1 (1.1)	0	1 (0.5)
Stem-cell source	1 \1.17	U	1 (0.0)
Peripheral blood	72 (75 9)	75 (83.3)	1/7 /70 5
•	72 (75.8)		147 (79.5)
Bone marrow	11 (11.6)	8 (8.9)	19 (10.3)
Cord blood	7 (7.4)	5 (5.6)	12 (6.5)
Other Unknown	2 (2.1) 3 (3.2)	1 (1.1) 1 (1.1)	3 (1.6) 4 (2.2)

Data are presented as n (%) unless otherwise noted.

Abbreviations: CMV, cytomegalovirus; HCT, hematopoietic cell transplant; RSV, respiratory syncytial virus.

^aFor this value, n = 94 for presatovir and n = 184 total.

 $^{\rm b}\mbox{For this value, }n$ = 10 for presatovir, n = 11 for placebo, and n = 21 total.

 $^{\rm c}\text{For this value, }n$ = 88 for placebo and n = 183 total.

^dProtocol deviation related to onset of respiratory symptoms was recorded for 1 placebo-treated patient.

 $^{\rm e}\textsc{For}$ this value, n = 94 for presatovir and n = 184 total.

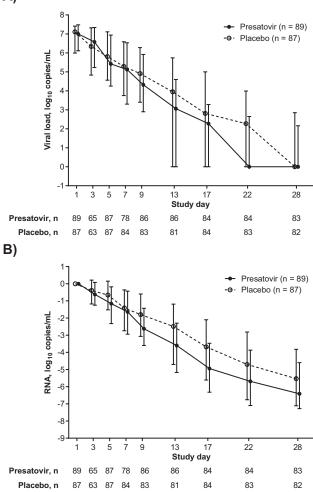


Figure 2. Nasal respiratory syncytial virus (RSV) RNA at each study visit in the efficacy population. *A*, The median nasal RSV RNA. *B*, The median change from baseline in nasal RSV RNA at each study visit in patients treated with presatovir (closed circles, solid line) versus placebo (open circles, dashed line). Error bars represent the interquartile range.

started treatment as outpatients (18/63 [28.6%] vs 9/113 [8.0%], respectively), and the hospitalization status was imbalanced between the presatovir and placebo arms at baseline. The effects of presatovir versus placebo treatment on the time-weighted average change in the viral load from Day 1 to Day 9 and the occurrence of death or respiratory failure requiring mechanical ventilation through Day 28 were similar between patients hospitalized and not hospitalized on Day 1 (Supplementary Table 6). However, treatment with presatovir, relative to the placebo, was associated with a 28% lower LRTC event rate among patients hospitalized on Day 1 (Table 2; Supplementary Table 6). In other post hoc analyses, the proportion of patients who developed LRTCs was numerically lower following presatovir treatment, as compared to placebo treatment, among patients with shorter than median symptom durations (≤ 4 days) and \leq 365 days since HCT (Table 2; Supplementary Tables 6–9). A post hoc multivariate Cox proportional hazard model for the time to an LRTC through Day 28 in patients receiving presatovir, adjusted for lymphopenia and ribavirin use on Day 1, enrollment site, and hospitalization status on Day 1, yielded an adjusted hazard ratio of 0.44 (95% CI, .19–.99; P = .091), as compared to those receiving the placebo. Optional extended RSV monitoring and serologic responses are presented in the Supplementary Results. Patients with treatment-emergent substitutions in RSV F that were associated with presatovir resistance had a numerically smaller change in the time-weighted average RSV load, but not worse clinical outcomes, relative to those with wild-type F sequences; such substitutions occurred at a significantly higher frequency in patients with, versus without, lymphopenia (Supplementary Results; Supplementary Tables 10–11).

Safety

Overall, AEs were reported in 76 (80%) of the patients receiving presatovir and 78 (86.7%) of the patients receiving the placebo, while 18 (18.9%) of the patients receiving presatovir and 23 (25.6%) of the patients receiving the placebo had serious adverse events (SAEs). The most common AEs were diarrhea (15.8%), nausea (13.7%), and pyrexia (12.6%) in the patients receiving presatovir; and diarrhea (15.6%), vomiting (13.3%), and nausea (11.1%) in the patients receiving the placebo (Table 3). Most Grade 3 or 4 AEs and SAEs occurred less frequently in patients receiving presatovir, except for pyrexia as an SAE in 4 (4.2%) patients and GVHD in the gastrointestinal tract as an SAE, Grade 3 pyrexia, and Grade 4 pneumonia in 2 (2.1%) patients each (Supplementary Tables 12-13). There were no imbalances in new electrocardiogram findings or troponin abnormalities between the 2 groups. Overall, 6 patients died during the study; 2 (2.1%) were treated with presatovir and 4 (4.4%) were treated with the placebo. There were 2 patients receiving presatovir who died from gastrointestinal hemorrhage and pneumonia (1 each), and 4 patients receiving the placebo died from an LRTI, pneumonia, recurrent acute myeloid leukemia, and an intracranial hemorrhage (1 each).

DISCUSSION

This is the largest randomized, double blind, placebo-controlled clinical trial to date for the treatment of allogeneic and autologous HCT recipients with RSV URTIs. Presatovir treatment did not meet the coprimary endpoints of a greater time-weighted average change in the RSV viral load from Day 1 to 9 and the reduced development of LRTCs through Day 28, but was well tolerated, with a comparable safety profile relative to the placebo. In a post hoc analysis of patients with lymphopenia, the proportion who developed an LRTC through Day 28 was 51% lower following treatment with presatovir, as compared to the placebo; other post hoc analyses also indicated trends toward a

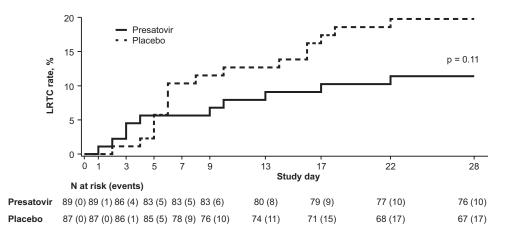


Figure 3. Development of lower respiratory tract complications (LRTC) in the efficacy population. The LRTC rate at each study visit in patients treated with presatovir (solid line) versus placebo (dashed line) is shown.

treatment effect on LRTCs. The results suggest lessons for the design of future clinical trials of drugs for RSV or other respiratory viruses in transplant recipients or other immunocompromised patients.

Among healthy adults with established experimental RSV infections, presatovir treatment, as compared to the placebo treatment, significantly reduced the RSV load and clinical severity [26]. The current study did not reproduce these findings, most likely because the challenge study participants received presatovir at or before symptom onset, whereas the current study patients were treated after a median of 4 days of symptoms. An exploratory analysis revealed trends toward reduced LRTC rates following presatovir treatment, versus placebo treatment, of patients with median or shorter symptom durations (Table 2). Future studies of anti-RSV drugs, particularly fusion inhibitors, should explore whether earlier therapy improves treatment outcomes.

Some transplant centers treat RSV infections in immunocompromised patients with oral or aerosolized ribavirin, despite lacking randomized clinical trial evidence [1]. Ribavirin use in RSV-infected HCT recipients, especially those with URTIs, has been associated with more favorable outcomes in retrospective studies [6, 8, 27]. In the current study, placebo-treated patients who received ribavirin had a higher LRTC progression rate, compared with those who did not (26% vs 18%, respectively), and all patients who developed an LRTC used ribavirin more frequently (37.0%), relative to those without progression (23.5%). As this was not a randomized, controlled study of ribavirin treatment, these observations require confirmation.

The observed rate of LRTCs was lower than the expected 30% used for the sample size calculation, and the Day 28 mortality was very low (~3%) relative to previous retrospective studies [2, 7, 10], possibly due to the recruitment of less severely ill patients who would not typically undergo RSV testing. Lymphopenia is a well-described risk factor for LRTCs in RSV-infected HCT recipients [9, 12, 14, 28], as observed in the current study (64% in placebo-treated patients with lymphopenia vs 11% in those without). Treatment with presatovir reduced the development

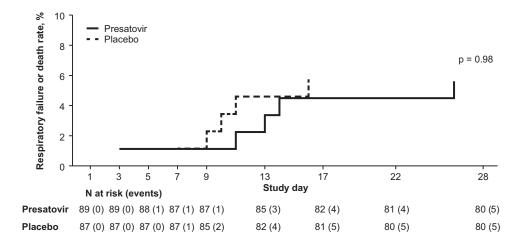


Figure 4. Occurrence of death or respiratory failure requiring mechanical ventilation in the efficacy population. The event rate at each study visit in patients receiving presatovir (solid line) versus placebo (dashed line) is shown.

Table 2. Post Hoc Analyses

Patients developing LRTC, n/N (%)	Presatovir	Placebo	Treatment Difference (95% CI), %	Nominal <i>P</i> Value ^a
Lymphopenia, <200 cells/µL	2/15 (13.3)	9/14 (64.3)	-51.0 (-77.8 to -13.1)	.008
No ribavirin use	4/64 (6.3)	12/68 (17.6)	-11.4 (-28.1 to 5.9)	.061
Symptom duration \leq median, 4 days ^b	5/48 (10.4)	13/49 (26.5)	-16.1 (-35.4 to 3.4)	.066
Hospitalized on Day 1	7/39 (17.9)	11/24 (45.8)	-27.9 (-50.9 to -2.4)	.023
≤365 days after HCT	5/50 (10.0)	12/47 (25.5)	-15.5 (-34.8 to 4.7)	.061

Data are for LRTC development through Day 28 by presence of lymphopenia, duration of symptoms, hospitalization status, and time after HCT at Day 1. Data for other efficacy endpoints and subgroups are provided in Supplementary Tables 4–9.

Abbreviations: CI, confidence interval; HCT, hematopoietic cell transplant; LRTC, lower respiratory tract complications.

^a*P* values were calculated using the Fisher exact test.

^bThe median duration of respiratory symptoms on Day 1 in the efficacy population was 4 days.

of LRTCs in patients with lymphopenia—a surrogate marker of impaired T-cell or humoral immunity—possibly because robust immune responses masked any treatment effect by improving outcomes regardless of the treatment. Furthermore,

Table 3.	Adverse	Events	and	Laboratory	Abnormalities	Reported	in	≥4
Patients in	n a Treatn	nent Gro	up ir	n the Safety	Population			

Adverse Event	Presatovir, n = 95	Placebo, n = 90
Any adverse event	76 (80.0)	78 (86.7)
Serious adverse events	18 (18.9)	23 (25.6)
Grade ≥3 adverse events	22 (23.2)	21 (23.3)
Diarrhea	15 (15.8)	14 (15.6)
Nausea	13 (13.7)	10 (11.1)
Vomiting	11 (11.6)	12 (13.3)
Pyrexia	12 (12.6)	9 (10.0)
Decreased appetite	7 (7.4)	6 (6.7)
Epistaxis	9 (9.5)	3 (3.3)
Headache	5 (5.3)	7 (7.8)
Pneumonia	4 (4.2)	7 (7.8)
Acute kidney injury	3 (3.2)	7 (7.8)
Asthenia	3 (3.2)	7 (7.8)
Cough	6 (6.3)	4 (4.4)
Dizziness	7 (7.4)	3 (3.3)
Rash	4 (4.2)	5 (5.6)
Fatigue	4 (4.2)	4 (4.4)
Neutropenia	3 (3.2)	5 (5.6)
Abdominal pain	3 (3.2)	4 (4.4)
Dyspnea	3 (3.2)	4 (4.4)
Febrile neutropenia	2 (2.1)	5 (5.6)
Hypokalemia	4 (4.2)	3 (3.3)
Anemia	5 (5.3)	1 (1.1)
Insomnia	4 (4.2)	2 (2.2)
Edema peripheral	2 (2.1)	4 (4.4)
Dysgeusia	1 (1.1)	4 (4.4)
Fall	1 (1.1)	4 (4.4)
Fluid overload	4 (4.2)	1 (1.1)
Hypertension	4 (4.2)	1 (1.1)
Pain in extremity	4 (4.2)	1 (1.1)
Dysuria	4 (4.2)	0
Sinusitis	4 (4.2)	0

Data are shown as n (%).

lymphopenia could influence respiratory immunopathology, providing better evidence of presatovir's antiviral efficacy.

Perhaps the most important question is whether an all-cause LRTC rate is a clinically relevant endpoint and, if so, whether the observed trends are clinically meaningful. Respiratory failure and mortality are more clinically significant, but their rates in this study suggest that the sample size required would be prohibitive, especially for HCT recipients. The current study endpoint of LRTCs included multiple etiologies, because RSV URTIs may predispose patients to secondary infections-for example, by disrupting mucociliary function [29, 30]-so treatment could prevent a secondary LRTI, as well as a primary LRTI. Furthermore, any LRTC is a clinically significant event that may prolong hospitalization, necessitate intensive clinical care (including empiric antimicrobial treatment), and potentially result in death. Only a minority of LRTCs in this study were adjudicated as primary RSV LRTIs-likely due to other etiologies, as well as a lack of lower respiratory tract samples for the confirmation of RSV-underscoring the potential importance of nonviral pulmonary events in HCT recipients with RSV infections. Determining the cause of each LRTC event in a clinical trial, while ideal, requires invasive procedures (eg, bronchoscopy or lung biopsy) that could pose significant patient risks and are not globally mandated by the current clinical standard of care. Thus, radiographic confirmation, corroborated by clinical data with central, blinded adjudication, as used here, may be the best approach to classify LRTIs. Whether the near-50% relative reduction in LRTC events is clinically meaningful, despite lacking statistical significance, is left to interpretation. The consistent trends toward a treatment effect in exploratory analyses need confirmation in future studies.

In summary, this study provided important lessons for the design of future clinical trials of drugs for RSV and other respiratory virus infections in HCT recipients. Although the coprimary endpoints were not achieved, presatovir treatment was associated with trends toward an antiviral effect and clinical benefit. Similar future trials should judiciously select suitable at-risk patients (ie, patients with lymphopenia, neutropenia, GVHD, or receiving corticosteroids) to maximize the potential benefits. Because having an LRTC increases the mortality risk, prompt diagnoses, early intervention for RSV URTIs in highrisk patients, and effective antiviral agents are imperative to improve clinical outcomes.

Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes

Author contributions. R. F. C wrote the first draft of the manuscript and contributed to the study design. S. S. D., F. M. M., J. W. C., and M. B. gave input into the manuscript draft and contributed to the study design. Y. G. performed the statistical analysis. G.-S. C., S. N. P., and A. P. L. were members of the endpoint adjudication committee. P. G. performed the pharmacokinetics analysis. D. P. P. performed the virology analysis. All authors reviewed the manuscript and approved the final version for submission.

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References

- Khanna N, Widmer AF, Decker M, et al. Respiratory syncytial virus infection in patients with hematological diseases: single-center study and review of the literature. Clin Infect Dis 2008; 46:402–12.
- Martino R, Porras RP, Rabella N, et al. Prospective study of the incidence, clinical features, and outcome of symptomatic upper and lower respiratory tract infections by respiratory viruses in adult recipients of hematopoietic stem cell transplants for hematologic malignancies. Biol Blood Marrow Transplant 2005; 11:781–96.
- Nichols WG, Gooley T, Boeckh M. Community-acquired respiratory syncytial virus and parainfluenza virus infections after hematopoietic stem cell transplantation: the Fred Hutchinson Cancer Research Center experience. Biol Blood Marrow Transplant 2001; 7(Suppl):11–5S.
- Raboni SM, Nogueira MB, Tsuchiya LR, et al. Respiratory tract viral infections in bone marrow transplant patients. Transplantation 2003; 76:142–6.
- Schiffer JT, Kirby K, Sandmaier B, Storb R, Corey L, Boeckh M. Timing and severity of community acquired respiratory virus infections after myeloablative versus non-myeloablative hematopoietic stem cell transplantation. Haematologica 2009; 94:1101–8.
- Shah JN, Chemaly RF. Management of RSV infections in adult recipients of hematopoietic stem cell transplantation. Blood 2011; 117:2755–63.
- Whimbey E, Champlin RE, Couch RB, et al. Community respiratory virus infections among hospitalized adult bone marrow transplant recipients. Clin Infect Dis 1996; 22:778–82.
- Avetisyan G, Mattsson J, Sparrelid E, Ljungman P. Respiratory syncytial virus infection in recipients of allogeneic stem-cell transplantation: a retrospective study of the incidence, clinical features, and outcome. Transplantation 2009; 88:1222–6.
- Chemaly RF, Ghosh S, Bodey GP, et al. Respiratory viral infections in adults with hematologic malignancies and human stem cell transplantation recipients: a retrospective study at a major cancer center. Medicine (Baltimore) 2006; 85:278–87.
- Chemaly RF, Shah DP, Boeckh MJ. Management of respiratory viral infections in hematopoietic cell transplant recipients and patients with hematologic malignancies. Clin Infect Dis 2014; 59(Suppl 5):S344–51.
- Ghosh S, Champlin RE, Englund J, et al. Respiratory syncytial virus upper respiratory tract illnesses in adult blood and marrow transplant recipients: combination therapy with aerosolized ribavirin and intravenous immunoglobulin. Bone Marrow Transplant 2000; 25:751–5.
- Kim YJ, Guthrie KA, Waghmare A, et al. Respiratory syncytial virus in hematopoietic cell transplant recipients: factors determining progression to lower respiratory tract disease. J Infect Dis 2014; 209:1195–204.

- Seo S, Campbell AP, Xie H, et al. Outcome of respiratory syncytial virus lower respiratory tract disease in hematopoietic cell transplant recipients receiving aerosolized ribavirin: significance of stem cell source and oxygen requirement. Biol Blood Marrow Transplant 2013; 19:589–96.
- Shah DP, Ghantoji SS, Shah JN, et al. Impact of aerosolized ribavirin on mortality in 280 allogeneic haematopoietic stem cell transplant recipients with respiratory syncytial virus infections. J Antimicrob Chemother 2013; 68:1872–80.
- Erard V, Chien JW, Kim HW, et al. Airflow decline after myeloablative allogeneic hematopoietic cell transplantation: the role of community respiratory viruses. J Infect Dis 2006; 193:1619–25.
- Harrington RD, Hooton TM, Hackman RC, et al. An outbreak of respiratory syncytial virus in a bone marrow transplant center. J Infect Dis 1992; 165:987–93.
- Kim YJ, Boeckh M, Englund JA. Community respiratory virus infections in immunocompromised patients: hematopoietic stem cell and solid organ transplant recipients, and individuals with human immunodeficiency virus infection. Semin Respir Crit Care Med 2007; 28:222–42.
- Shah DP, Ghantoji SS, Mulanovich VE, Ariza-Heredia EJ, Chemaly RF. Management of respiratory viral infections in hematopoietic cell transplant recipients. Am J Blood Res 2012; 2:203–18.
- Small TN, Casson A, Malak SF, et al. Respiratory syncytial virus infection following hematopoietic stem cell transplantation. Bone Marrow Transplant 2002; 29:321–7.
- Whimbey E, Champlin RE, Englund JA, et al. Combination therapy with aerosolized ribavirin and intravenous immunoglobulin for respiratory syncytial virus disease in adult bone marrow transplant recipients. Bone Marrow Transplant 1995; 16:393–9.
- de Seynes C, Fleury H, Milpied N, et al. Influence of respiratory viruses on airflow decline after allogenic hematopoietic stem cell transplantation. Eur Respir J 2016; 48(Suppl 60):PA2599.

- 22. US Department of Health and Human Services: Food and Drug Administration: Center for Drug Evaluation and Research. Respiratory syncytial virus infection: developing antiviral drugs for prophylaxis and treatment. Guidance for industry. Draft guidance. October 6, 2017. Available at: https://www.fda.gov/downloads/ Drugs/GuidanceComplianceRegulatoryInformation/Guidances/UCM579756. pdf. Accessed 23 August 2018.
- VIRAZOLE^{*} (Ribavirin for Inhalation Solution, USP). Full prescribing information. Bridgewater, NJ: Valeant Pharmaceuticals North America LLC, 2016.
- American Academy of Pediatrics Committee on Infectious Diseases. Use of ribavirin in the treatment of respiratory syncytial virus infection. Pediatrics 1993; 92(3):501–4.
- 25. Boeckh M, Englund J, Li Y, et al; National Institute of Allergy and Infectious Diseases Collaborative Antiviral Study Group. Randomized controlled multicenter trial of aerosolized ribavirin for respiratory syncytial virus upper respiratory tract infection in hematopoietic cell transplant recipients. Clin Infect Dis 2007; 44:245–9.
- DeVincenzo JP, Whitley RJ, Mackman RL, et al. Oral GS-5806 activity in a respiratory syncytial virus challenge study. N Engl J Med 2014; 371:711–22.
- Waghmare A, Campbell AP, Xie H, et al. Respiratory syncytial virus lower respiratory disease in hematopoietic cell transplant recipients: viral RNA detection in blood, antiviral treatment, and clinical outcomes. Clin Infect Dis 2013; 57:1731-41.
- Ljungman P. Respiratory virus infections in stem cell transplant patients: the European experience. Biol Blood Marrow Transplant 2001; 7(Suppl):5S–7S.
- Smith CM, Kulkarni H, Radhakrishnan P, et al. Ciliary dyskinesia is an early feature of respiratory syncytial virus infection. Eur Respir J 2014; 43:485–96.
- Tristram DA, Hicks W Jr, Hard R. Respiratory syncytial virus and human bronchial epithelium. Arch Otolaryngol Head Neck Surg 1998; 124:777–83.