

## Building Osteogenic Microenvironments With Strontium-Substituted Calcium Phosphate Ceramics

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Wan B, Wang R, Sun Y, Cao J, Wang H, Guo J and Chen D (2020) Building Osteogenic Microenvironments With Strontium-Substituted Calcium Phosphate Ceramics. Front. Bioeng. Biotechnol. 8:591467. doi: 10.3389/fbioe.2020.591467 Bioceramics have experienced great development over the past 50 years. Modern bioceramics are designed to integrate bioactive ions within ceramic granules to trigger living tissue regeneration. Preclinical and clinical studies have shown that strontium is a safe and effective divalent metal ion for preventing osteoporosis, which has led to its incorporation in calcium phosphate-based ceramics. The local release of strontium ions during degradation results in moderate concentrations that trigger osteogenesis with few systemic side effects. Moreover, strontium has been proven to generate a favorable immune environment and promote early angiogenesis at the implantation site. Herein, the important aspects of strontium-enriched calcium phosphate bioceramics (Sr-CaPs), and how Sr-CaPs affect the osteogenic microenvironment, are described.

Keywords: calcium phosphate ceramics, strontium substitution, microenvironment, bone regeneration, biomaterials

## INTRODUCTION

Bone is a metabolically active specialized connective tissue with the capacity for continuous resorption and reformation, but the reconstruction process requires extra support in large bone defect cases, for example, injury, tumor excision, and age-related diseases (Loi et al., 2016; Ginebra et al., 2018). Clinically, autografts still remain the most common strategy to support and stimulate bone growth. While biologically desirable, the supply of autografts from a patient's own body cannot meet the amount required for large-sized bone defects (Wang and Yeung, 2017). Grafts from bone banks or other animal species can provide a vast range of grafting forms by virtue of their relatively easy availability (Wang and Yeung, 2017). Nevertheless, they are still subject to attenuated osteointegration and disease transmission.

Synthetic bioactive ceramics can overcome these limitations. They are obtained by chemical reagent deposition through a controlled process that allows their properties to be adapted to the specific requirements of different clinical situations. Calcium phosphate ceramics (CaPs) are basic bone repair materials with excellent osteoinductive and osteoconductive features; examples include hydroxyapatite (HAP),  $\beta$ -tricalcium phosphate ( $\beta$ -TCP), calcium polyphosphate (CPP), and biphasic calcium phosphate (BCP). They have been widely used in clinical applications in the form of implant surface coatings, the composition of cements, and scaffolds (Jeong et al., 2019). Moreover, CaPs encapsulated with pharmacologics and biologics in scaffolds have shown

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multiple abilities, such as enhanced osteointegration, angiogenesis, and antibacterial properties (Bose and Tarafder, 2012; Marques et al., 2016). However, these biologically active molecules are only surface-loaded and show explosive release effects with little long-term benefit (Wu et al., 2014). This raises some concerns about their safety. For example, recombinant human bone morphogenetic protein-2 (rhBMP-2) is the first commercial synthetic osteoinductive molecule, and the combined use of rhBMP-2 and calcium phosphate ceramics has achieved many clinical successes. But it is worth noting that serious side effects related to ectopic or unwanted bone formation in certain situations have made the FDA increasingly resistant to approving the use of such materials (Vavken et al., 2016).

An alternative long-lasting and potentially safer strategy is the incorporation of trace metallic elements into CaPs. The mineral composition of our bone itself is not a homogenous and bioinert calcium phosphate-based material (Von Euw et al., 2019). Bone incorporates and releases various trace elements (such as  $Mg^{2+}$ ,  $Zn^{2+}$ , and  $Sr^{2+}$ ) into the microenvironment during bone metabolism (Bose et al., 2013). Previous studies have confirmed that CaPs modified with these bivalent trace metallic ions can lead to controlled degradation, increase the mechanical strength of the materials, and enhance bioactive properties. But among these bioactive ions, strontium (Sr) has gained great attention since strontium ranelate (SrRan) has been approved as an antiosteoporotic drug for post-menopausal osteoporosis since it increases bone strength (Neves et al., 2017; Marx et al., 2020). Recent findings suggest that oral administration of SrRan may have no anabolic action on bone formation in humans, and even inhibits osteogenic differentiation under the in vitro experiment conditions (Wornham et al., 2014; Marx et al., 2020). However, the comprehensively described SrCaPs do exert positively influence on new bone formation and accelerate the healing process (Neves et al., 2017). These discrepancies may be partially due to the implant microenvironments where biomaterials interact with various cells. The local delivery of Sr ions from SrCaPs implanted in bone changes local microenvironment, which is involved in several biological processes such as osteogenesis, angiogenesis, osteoimmunomodulation.

Consequently, in this review, we present recent developments in strontium-substituted calcium phosphate ceramics (SrCaPs) and offer insights into how Sr ions released from implants influence the immune response, angiogenesis, and osteoblastic differentiation of bone marrow stromal cells (bMSCs).

## Sr-CaPs: CURRENT STATE OF THE ART

In the 1960s, surgeons valued the inertness of surgical materials in the human tissue environment, and first-generation bioceramics (alumina, zirconia, diverse forms of carbon) were developed as bone substitutes, mainly for femoral head fabrication (Smith, 1963). However, they elicited a foreign body reaction, forming fibrous capsules that isolated them from the body. In the 1980s, Larry Hench invented a bioactive glass with the ability to bond to living tissue, and Heughebaert reported that CaPs could induce bone formation (Hench and Wilson, 1984; Heughebaert et al., 1988). Since then, osteoinductive and tissue-inducing materials have become the mainstream of biomaterial science and engineering (Hench and Polak, 2002). CaPs are the most studied and implanted bioactive ceramics in the clinic because of their cost-effective preparation from chemical regents or natural resources. Various calcium phosphate compounds are in different crystalline phases and often classified on the basis of the Ca/P ratio. Of great interest is the subgroup of apatite compounds, which exhibit similarities to vertebrate hard tissues. Indeed, both bone and teeth are well-crystallized apatites (hydroxyapatite), and this structure can easily accommodate many cations and anions for substitution, thereby achieving adaptive biophysical functions (Drouet et al., 2017).

Strontium (Sr) is naturally deposited in the mineral phase of bones via consuming a normal diet, and it replaces approximately 0.035% of Ca content (International Programme on Chemical and Inter-Organization Programme for the Sound Management, 2010). The majority of in vitro experiments support a dualacting mechanism in which Sr stimulates bone formation and hinders bone resorption (Saidak and Marie, 2012). The exact mechanism of the role of Sr in bone remains unclear, but it has been proposed that Sr acts on similar cellular targets as  $Ca^{2+}$  by activating the calcium sensing receptor (CaSR), thus interacting with Ca-driven signaling pathways related to bone metabolism regulation (Saidak and Marie, 2012). Furthermore, small animal trials and clinical trials have produced overwhelming evidence that Sr benefits bone remodeling and increases bone-mineral density, especially in the treatment of osteoporosis (Meunier et al., 2004; Reginster et al., 2012; Marx et al., 2020). Taking these beneficial effects into consideration, it is likely that local Sr ion delivery will enhance osteoinduction and osseointegration at the bone-implant interface, contributing to a faster healing microenvironment.

Recently, Sr has been widely incorporated into calcium phosphate-based materials for biomedical applications. Krishnan reported using Sr doped hydroxyapatite (SrHAP) particles to repair dental enamel with the potential to treat white spot lesions and incipient carious lesions (Krishnan et al., 2016). It is also an effective additive for toothpaste in the prevention of cariogenesis (Surdacka et al., 2007). These particles cannot only be used as a raw material, but also associated with other biomaterial compounds to facilitate new functions. Sr-CaPs are fabricated as coatings on the surfaces of metallic implants in order to accelerate bone healing at early implantation times (Li et al., 2010; Arcos and Vallet-Regí, 2020). Titanium and titanium-based alloys are commonly used for load-bearing applications (such as total joint replacement and fracture fixation elements), but carry a risk of loosening, especially when implanted in osteoporotic bones. Tao et al. (2016a) conducted an in vivo study on rats with ovariectomy-induced osteoporosis and reported that a strontium-containing HAP coating on shaped titanium implants was better than an Sr-free HAP coating in terms of new bone formation and push-out force. Furthermore, they conducted an comparative study that found that under the same the molar ratio [Zn, Mg, Sr/(Zn, Mg, Sr+Ca) = 10%], the SrHAP coating exhibited better osseointegration than zinc- and magnesiumsubstituted HAP coatings (Tao et al., 2016b).

SrCaP cements (SrCPC) consist of a combination of a precursor powder and liquid phase, allowing it to fill and heal bone defects during minimally invasive procedures. Kuang et al. (2015) prepared pre-set SrCPC for a metaphyseal defect in Sprague-Dawley rats and found early endochondral ossification at 2 weeks post-operation. Thormann et al. (2013) further demonstrated that Sr was deposited at newly formed tissues around the implanted SrCPC as visualized by Time-of-flight secondary ion mass spectrometry (ToF-SIMS) imaging and found a significant increase of mineralized extracellular matrix in the SrCPC group compared to the Sr-free calcium phosphate cement (CPC) and empty defect groups. Sr-CaP scaffolds can be further modified with drugs, especially antibiotics and growth factors. The chemical composition of SrHAP provides strong surface adsorption capacity and increases surface area, allowing drug release for a longer period of time (Xu et al., 2018). Lin et al. (2013) reported that SrHAP microspheres with 3D hierarchically mesoporous structures not only promoted an osteogenic response, but exhibited sustained vancomycin release. Yan et al. (2018) developed an Sr-nHAP/SF-Hep-BMP-2 scaffold system with controlled BMP-2 release that supported critical size calvarial defect healing in Sprague-Dawley rats. Tao et al. (2018) revealed synergistic bioactive properties of BMP-2 and Sr released from CPC composites, which achieved rapid bone healing in osseous defects in an ovariectomized rat model. These new applications for strontium-containing calcium phosphate extends their biomedical potential.

## FAVORABLE POST-IMPLANTATION ENVIRONMENTS FOR Sr-CaPs

Sr is chemically and physically close to calcium but has a larger ionic radius (112 vs. 99 pm). The partial substitution

of Ca by Sr results in higher solubility when compared with Sr-free CaPs owning to the enlargement of the unit cell (Zhu et al., 2018). The surfaces of SrCaPs are reactive and biodegradable, and bone-like apatite forms on them through continuous dissolution and precipitation. The newly formed apatite creates a strong bond between the living tissues and implants. However, it degrades over time when in contact with bodily fluids and it elevates the local concentrations of  $Sr^{2+}$ ,  $Ca^{2+}$ , and  $PO_4^{3-}$  ions. These released ions affect important components of the bone microenvironment, including the mineral deposited layer phase, the cellular phase (osteoblasts, macrophages, and vascular endothelial cells), and the soluble factor phase (growth factors and/or cytokines) and provide an extracellular osteogenic signaling network. The incorporation of an appropriate proportion of Sr into CaPs allows them to build an osteogenic microenvironment via modulating the inflammatory response, stimulating osteogenic differentiation of bone marrow mesenchymal stem cells, and promoting early angiogenesis (Figure 1).

## **EFFECTS ON OSTEOGENESIS**

SrCaPs is a promising biomaterial to help bone reconstruction. Mounting evidence shows that Sr replacement of Ca in CaPs leads to increased solubility due to the expansion of the crystal structure, and the high local  $\text{Sr}^{2+}$ ,  $\text{Ca}^{2+}$ , and  $\text{PO}_4{}^{3-}$  ionic concentrations are believed to be of great importance for osteoinduction and osteoconduction (Bose et al., 2013; Neves et al., 2017). Current research shows that both Sr and Ca mediate key cellular functions in osteogenesis-related cells via Ca-sensing receptor (CaSR)-dependent mechanisms (Marx et al., 2020). In the extracellular microenvironment, a high Ca concentration is



a potent chemical signal for bMSCs osteogenic differentiation (Dvorak and Riccardi, 2004; Barradas et al., 2012). A lack of calcium significantly retards cell growth and differentiation (Dvorak et al., 2004; Barradas et al., 2012). Interestingly, CaPs with Sr modification tend to have decreased Ca concentrations around the material due to induced apatite precipitation, though lower concentrations of Ca do not impair the proliferation or osteogenic differentiation of bMSCs (Schumacher et al., 2013; Kruppke et al., 2019b). In an in vitro study, Kruppke et al. (2019a) confirmed that Sr may compensate for the adverse effects of lowered Ca ion concentrations to a certain degree and potentially stimulate bone regeneration. They investigated the effect of varied Ca and Sr ion concentrations, respectively, on proliferation and differentiation of bMSCs and concluded that under low calcium concentrations culture conditions (0.3-0.4 mM due to the addition of fetal calf serum), Sr at 0.4-0.9 mM promoted differentiation of bMSCs while 0.9-1.8 mM stimulated the highest proliferation. And in a second study exploring the relationship between Sr and Ca on bone formation, they conducted in vitro and in vivo experiments to evaluate the osteogenesis effects of Sr together with two concentration gradients of Ca (normal calcium of 1.8 mM or high calcium of 9 mM) and concluded that Sr (1 mM) enhanced mineralization and ALP expression of MC3T3-E1 pre-osteoblast cells under a high dose of calcium (9 mM) (Xie et al., 2018). This may explain the results by Wornham et al. (2014) in which concentrations of Sr ranging from 10 µM to 1 mM inhibit the mineralization ability of primary rat osteoblasts under 1.8 mM Ca present in the culture system. These experiments illustrate the importance of the implant microenvironment because bone has higher Ca levels than plasma. It also suggests that both Sr and Ca from ion-leaching SrCaPs may interact to promote bone regeneration at the

TABLE 1 | The osteogenic differentiation function of various Sr concentrations.

bone-implant interface. In addition, the phosphate ion is a basic component in the human body and accumulates in bone minerals in the form of calcium phosphates along with calcium ions (Khoshniat et al., 2011). When phosphate ions are not present in high quantities, the generation of new bone will be blocked due to insufficient formation of hydroxyapatite (Penido and Alon, 2012).

Sr-rich CaPs consistently outperform Sr-free CaPs in *in vitro* and *in vivo* studies (Neves et al., 2017; Marx et al., 2020). In the absence of doping with other osteogenic ions, the local release of Sr is sufficient to promote bone formation, but it is difficult to ignore the effects that surface morphology, porosity, hydrophilicity, dissolution process, and local pH changes may have on osteogenesis regulation at the implantation site.

#### **EFFECTS ON ANGIOGENESIS**

Increased angiogenesis can relieve the symptoms of ischemia and facilitate the transport nutrients, which is essential for bone repair/regeneration. Sr-CaPs have been found to increase neovascularization in many convincing experiments (Liu et al., 2011; Gu et al., 2013; Wang et al., 2014). Endothelial cells are the major seed cells that drive angiogenesis. Chen et al. (2008) observed that degradation products of Sr doped calcium polyphosphate (SCPPs) promote the proliferation, migration, and tube-like structure formation of endothelial cells (ECV304). It has been reported that *in situ* production of angiogenic growth factors, such as vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF), can promote more cell infiltration, extracellular matrix synthesis, and faster angiogenesis (Saberianpour et al., 2018).

Cell lines	Strontium salt	Sr concentration	Function	References
C3H10T1/2 and mice primary bone marrow mesenchymal stem cells	Strontium chloride	1.0 mM, 3.0 mM	Promote osteogenic differentiation	Peng et al., 2009
Ovariectomy bone marrow mesenchymal stem cells	Strontium ranelate	0.25–0.5 mM	Promote osteogenic differentiation	Guo et al., 2016
Primary fetal mouse calvaria cells	Strontium ranelate	0.1–1 mM	Promote osteogenic differentiation	Bonnelye et al., 2008
Human bone marrow mesenchymal stem cells	Strontium ranelate	2.4–240 μM	Promote osteogenic differentiation	Sila-Asna et al., 2007
Human adipose-derived stem cells	Strontium ranelate	25–500 μΜ	Promote osteogenic differentiation	Aimaiti et al., 2017
	Strontium ranelate	1–3 mM	Apoptosis and inhibit osteogenic differentiation	
MC3T3-E1 pre-osteoblast cells	Strontium chloride	1 mM	Inhibit osteogenic differentiation	Xie et al., 2018
Rat bone marrow mesenchymal stem cells	Strontium ranelate	0.1 mM, 1 mM	Inhibit proliferation and promote osteogenic differentiation	Li et al., 2012
Primary rat osteoblastic cells	Strontium ranelate; strontium chloride	0.1–1 mM	Inhibit mineral deposition	Wornham et al., 2014

Liu et al. (2011) directly seeded osteoblasts (ROS17/2.8) on SCPP scaffolds with various amounts of Sr and found increased secretion of VEGF and bFGF in the culture medium. Wang et al. (2014) observed the stimulating effects of SCPP on protein secretion and mRNA expression of VEGF, bFGF, and MMP-2 from endothelial cells in vitro. Gu et al. (2014) co-cultured umbilical vein endothelial cells and osteoblasts on SCPP scaffolds in vitro and demonstrated higher VEGF and bFGF protein production compared with CPP and HAP. Furthermore, Ye et al. (2019) fabricated Sr-doped calcium phosphate/polycaprolactone/chitosan (Sr-CaP/PCL/CS) nanohybrid fibrous membranes and demonstrated high VEGF secretion from bMSCs. In the future, more experiments are needed to test the ability of SrCaPs to promote blood vessel formation in vivo, and more attention should be paid to the impact of the immune response.

## EFFECTS ON OSTEOIMMUNOMODULATION

Every implantable material must be biocompatible, meaning that only a limited inflammatory response occurs in the host body. Huang et al. (2020) reported the incorporation of fishderived nano CaPs (Ca/P molar ratio was 2.35) into gelatin methacrylate generated a suitable immune microenvironment by inducing the secretion of cytokines and promoting macrophage phenotype conversion. The controllable inflammation may promote the osteogenic differentiation and angiogenesis (Zhao et al., 2018; Huang et al., 2020). The local release of Sr from implants to the adjacent host bone can reduce unfavorable inflammatory responses (Renaudin et al., 2008), and Sr has been proven to be an anti-inflammatory agent (Wang et al., 2020). Buache et al. (2012) first verified that Sr-BCP can decrease the production of pro-inflammatory cytokines (TNF- $\alpha$  and IL-6) and the chemokine interleukin 8 in human monocytes. Braux et al. (2016) demonstrated the downregulating effect of Sr-BCP on inflammatory mediator production (MCP-1 and Gro- $\alpha$ ) by human primary osteoblasts. Sr has also been shown to suppress the expression of proinflammatory cytokines (IL-6 and TNF- $\alpha$ ) in periodontal ligament cells and macrophages (Römer et al., 2012; Gu et al., 2014). Among various immune-related cells, macrophages play a significant role in regulating bone healing after trauma or the implantation of biomaterials (Alexander et al., 2011; Batoon et al., 2019). At the early time of bone repair, resident or infiltrating monocytederived macrophages mostly present the pro-inflammatory M1 phenotype, and a timely switch from M1 to M2 (antiinflammatory phenotype) leads to osteogenic cytokine release, including that of IL-10, TGFB, and VEGF. Interestingly, prolonged M1 polarization leads to an increased release of fiber-enhancing cytokines from M2 macrophages, which leads to the formation of fibrous capsules (Chen et al., 2016). Previous evidence suggests that Sr-integrated implants elicit more pro-regenerative M2 macrophages; examples include Srdoped calcium polyphosphate particles, Sr-substituted bioactive glass, and Ca- and Sr-modified titanium implant surfaces (Gu et al., 2014; Lee et al., 2016; Zhao et al., 2018). The incorporation of Sr into CaPs endows traditional CaPs with osteoimmunomodulation properties and positively regulates cytokine production and immune cell functions for better bone remodeling.



# CONTROVERSY OVER THE EFFECTIVE STRONTIUM CONCENTRATION

Local Sr release is a critical property of SrCaPs, but the optimal concentration remains to be determined. Evidence from in vitro studies shows that Sr promotes osteogenic differentiation in a dose-dependent manner, and different cell lineages respond differently to Sr at similar concentrations (Table 1). Peng et al. (2009) showed that 1 mM can promote osteogenic differentiation of C3H10T1/2 or mice primary bMSCs, while Aimaiti et al. (2017) reported that 1 mM induces apoptosis and inhibits osteogenic differentiation in human primary adiposederived stem cells. Moreover, inhibition at this concentration also occurs for primary rat osteoblastic cells and MC3T3-E1 cells (Wornham et al., 2014; Xie et al., 2018). However, the maximum circulating concentrations of Sr in the serum of patients treated with strontium ranelate (2 g per day for 3 years) is about 0.1 mM (Meunier et al., 2004). Although several in vitro studies have shown that Sr concentration above 1 mM can stimulate osteogenesis, it is preferable to take clinical trial results into consideration, and the effective range (below 500 mM) is recommended here (Bonnelye et al., 2008; Peng et al., 2009; Guo et al., 2016). To the best of our knowledge, no in vivo study on the local administration of Sr with the use of modified CaPs has reported adverse effects on bone formation and osseointegration. The reasons may be that, first, the implantation microenvironment in the body is dynamically changing, and a high dose of Sr does not last for a long time. Second, the degradation product is not only Sr, but also Ca, which matters because of the equilibrium effect of Ca to Sr (Xie et al., 2018). Third, Sr positively modulates the microenvironment via regulating immune cells and endothelial cells in the process of bone formation.

## CONCLUSION

In summary, integrating Sr ions into CaPs offers an alternative to biologics in the design of bioactive materials. It is of low

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cost, has a longer shelf life, and presents low systemic risk compared to growth factors. These added benefits make Sr as therapeutic agent attractive in tissue engineering and regenerative medicine applications (Jiménez et al., 2019; Prabha et al., 2019; Mao et al., 2020). However, the mechanism of osteoinduction of SrCaPs remains complicated because inflammation and angiogenesis accompany the entire process of bone healing. In light of the previous studies, we propose a hypothesis for the osteoinduction mechanism of SrCaPs: SrCaPs build the osteoinductive microenvironment of the host bone-implant interface (Figure 2). Functional ion release and apatite laver formation on the surfaces of SrCaP ceramics allow them to interact with cells and extracellular matrices in the host system, providing bioactive bonding to bone. Specifically, a higher local concentration of Sr stimulates the osteoblastic differentiation of MSCs (1 in Figure 1), induces macrophage polarization toward a pro-regenerative M2 phenotype (2 in Figure 1), and contributes to angiogenesis (3 in Figure 1). Collectively, understanding the biological microenvironment of implant-to-tissue interactions at the bone site is important for the development of highperformance bioceramics.

## **AUTHOR CONTRIBUTIONS**

BW was the first author and drafted the manuscript. DC and RW designed the work. YS, JG, JC, and HW summarized related publications. All authors approved the final version to be published.

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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