



## Heat Shock Protein 60 in Cardiovascular Physiology and Diseases

Yaoyun Duan<sup>1†</sup>, Huayuan Tang<sup>1†</sup>, Kali Mitchell-silbaugh<sup>2</sup>, Xi Fang<sup>2</sup>, Zhen Han<sup>3\*</sup> and Kunfu Ouyang<sup>1\*</sup>

<sup>1</sup> State Key Laboratory of Chemical Oncogenomics, School of Chemical Biology and Biotechnology, Peking University Shenzhen Graduate School, Shenzhen, China, <sup>2</sup> Department of Medicine, School of Medicine, University of California, San Diego, La Jolla, CA, United States, <sup>3</sup> Department of Cardiovascular Surgery, Peking University Shenzhen Hospital, Shenzhen, China

### **OPEN ACCESS**

### Edited by:

Antonella Marino Gammazza, University of Palermo, Italy

#### Reviewed by:

Paula Fernandez-Guerra, Aarhus University Hospital, Denmark Magdalena Gorska Ponikowska, Medical University of Gdańsk, Poland

### \*Correspondence:

Zhen Han 2547671020@qq.com Kunfu Ouyang ouyang\_kunfu@pku.edu.cn <sup>†</sup>These authors have contributed equally to this work

#### Specialty section:

This article was submitted to Protein Folding, Misfolding and Degradation, a section of the journal Frontiers in Molecular Biosciences

> Received: 10 January 2020 Accepted: 31 March 2020 Published: 30 April 2020

### Citation:

Duan Y, Tang H, Mitchell-silbaugh K, Fang X, Han Z and Ouyang K (2020) Heat Shock Protein 60 in Cardiovascular Physiology and Diseases. Front. Mol. Biosci. 7:73. doi: 10.3389/fmolb.2020.00073 Heat shock protein 60 (HSP60) is a highly conserved protein abundantly expressed in both prokaryotic and eukaryotic cells. In mammals, HSP60 has been primarily considered to reside in the mitochondria, where HSP60 and HSP10 form a complex and facilitate mitochondrial protein folding. However, HSP60 is also observed in the cytoplasm, the plasma membrane, and the extracellular space. HSP60 regulates a broad spectrum of cellular events including protein trafficking, peptide hormone signaling, cell survival, cell proliferation, inflammation, and immunization. In the cardiovascular system, growing evidence indicates that HSP60 could not only play an important role under physiological conditions, but also regulate the initiation and progression of heart failure and atherosclerosis. In this review, we focus on recent progress in understanding the function of HSP60 in cardiomyocytes, endothelial cells, and vascular smooth muscle cells (VSMCs), respectively, and discuss the related signaling pathways that have been found in these cells, so as to illustrate the role of HSP60 in the development of cardiovascular disease.

### Keywords: heat shock protein, HSP60, heart failure, atherosclerosis, cardiomyocyte

### INTRODUCTION

The human heat shock protein 60 (HSP60), which is also known as 60 kDa chaperonin, belongs to a family of the most ancient and conserved proteins in both prokaryotic and eukaryotic cells. They have a high homology between species and are ubiquitously expressed in most cells. HSP60 was initially found as a mitochondrial protein that plays a critical role in regulating mitochondrial protein homeostasis (Cheng et al., 1989; Ostermann et al., 1989). It has been shown that human HSP60 and HSP10 form a symmetrical football complex (Nisemblat et al., 2015), whereas the bacteria homolog of HSP60, GroEL, is organized in two rings, producing a barrel-like structure (Ostermann et al., 1989; Horwich et al., 2006). A survey of interactors of the human HSP60 suggested that most HSP60-interacting proteins are localized to the mitochondrial matrix space and

involved in various mitochondrial functions and metabolic pathways (Bie et al., 2020). Interestingly, accumulating studies have demonstrated that HSP60 is also localized in extramitochondrial compartments including the cytosol, plasma membrane, and extracellular space, as well as in blood circulation (Meng et al., 2018). Depending on protein localization, HSP60 not only regulates the mitochondrial chaperoning activity, but also plays a functional role in multiple cellular processes including cell proliferation, apoptosis, migration, and immune responses (Henderson et al., 2013).

The cardiovascular system is comprised of the heart and the network of arteries, veins, and capillaries that transport blood throughout the body. In humans, cardiovascular disease is a leading cause of mortality throughout the world (Go et al., 2014). It has been shown that many risk factors for cardiovascular disease, including smoking, lipopolysaccharide, chlamydia pneumoniae, shear stress, and ischemia, can promote the expression of HSP60 (Jakic et al., 2017). In the cardiovascular system, HSP60 has been supposed to play a regulatory role in cardiomyocytes, endothelial cells (ECs), vascular smooth muscle cells (VSMCs), and immune cells under both physiological and pathological conditions such as heart failure and atherosclerosis. The functions of HSP60 in immune cells and immune regulation have been well addressed and discussed elsewhere (Grundtman et al., 2011; Quintana and Cohen, 2011; Wick et al., 2014; Zininga et al., 2018). This review focuses on the functional roles that HSP60 performs in cardiomyocytes, ECs, and VSMCs, respectively, and the involvement of HSP60 in the pathogenesis of heart failure and atherosclerosis.

### **CARDIAC HSP60 AND HEART FAILURE**

HSP60 is highly expressed in cardiac tissues, and has been found in different subcellular locations inside cardiomyocytes, including on the membrane and in the mitochondria, cytoplasm, and extracellular space (Gupta and Knowlton, 2007; Lin et al., 2007). A large number of studies using *in vitro* cell culture models and *in vivo* animal models have revealed that HSP60 plays an important role in regulating cardiac physiology and pathophysiology. Here we describe several major roles of HSP60 in cardiomyocytes and the involvement of HSP60 in the progression of heart failure (**Figure 1**).

## Intracellular HSP60 May Play Protective Roles in Cardiac Cells

In cardiac cells, HSP60 is mainly located inside the mitochondria, while a small portion of HSP60 (approximately 20–40%) can be observed in the cytoplasm (Lin et al., 2007). In cultured neonatal rat cardiomyocytes, overexpression of HSP60 alone or together with its co-chaperone HSP10 protected myocytes against apoptosis induced by simulated ischemia and reoxygenation (Lau et al., 1997; Lin et al., 2001). The protective function of overexpressed HSP60 and HSP10 is associated with reduced mitochondrial Cytochrome c release and suppressed Caspase-3 activity, as well as an increase of ATP recovery and elevated activities of mitochondrial complexes III and IV. These results

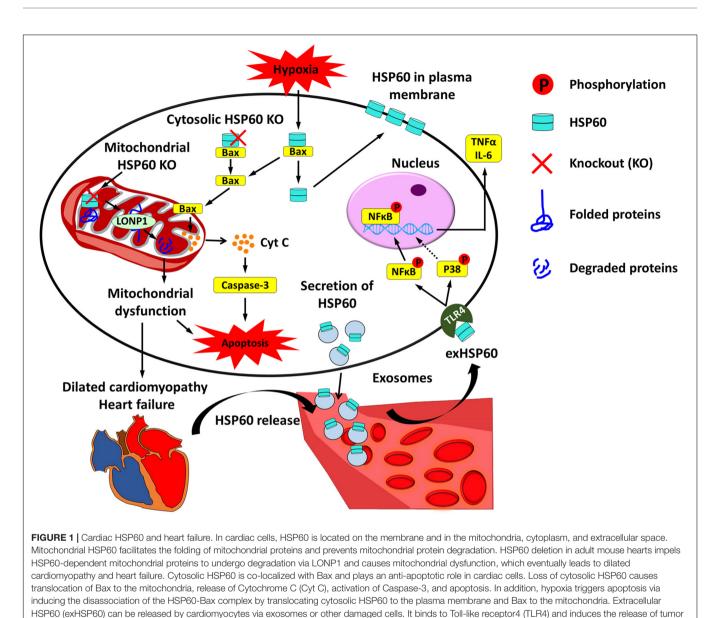
suggest that mitochondrial chaperonin HSP60 plays a critical role in regulating mitochondrial integrity and capacity for ATP production, which are essential for determining the survival of cardiomyocytes undergoing ischemia and reperfusion injury.

Human mitochondrial HSP60 and its cochaperonin HSP10 form a symmetrical football complex (Nisemblat et al., 2015), which facilitates the folding of mitochondrial proteins and therefore confers their stability. This consequently prevents mitochondrial protein degradation and the induction of mitochondrial unfolded protein responses. Furthermore, HSP60 may protect mitochondrial proteins from aggregation, especially under stressful conditions. Myrtucommulone, a natural product that can inhibit the refolding activity of the HSP60/HSP10 complex, prevents the reactivation of denatured malate dehydrogenase in a protein refolding assay. Under heat shock, the interference of myrtucommulone with HSP60 is accompanied by aggregation of the Lon protease-like protein (LONP) and the leucine-rich PPR motif-containing protein (LRP130) (Wiechmann et al., 2017; Meng et al., 2018). In future studies, it will be very interesting to examine whether HSP60 deletion in cardiac cells could also result in the aggregation of certain mitochondrial proteins under stress.

HSP60 is also observed in the cytosol, where it may exist in monomeric or heptametric forms (Taguchi et al., 1994; Levy-Rimler et al., 2001). HSP60 is synthesized in the cytosol with a mitochondrial transport signal (Singh et al., 1990). After translocation to the mitochondria, the mitochondrial transport signal is then cleaved and a certain amount of HSP60 protein may return to the cytosol. On the other hand, HSP60 with the mitochondrial target peptide may also be observed in the cytosol as the newly synthesized protein and accumulation of such proteins could be found in certain circumstance without apparent mitochondrial release (Chandra et al., 2007). However, the mechanism underlying the distribution of HSP60 between the mitochondria and the cytosol remains unclear. HSP60 in the cytosol has also been considered to play an antiapoptotic role in cardiac cells. Immuno-electron microscopy has demonstrated that HSP60 is co-localized with Bax in the cytosol of normal rat hearts (Gupta and Knowlton, 2005). The decrease of cytosolic HSP60 induced by an antisense phosphorothioate oligonucleotide facilitates the translocation of Bax to the mitochondria and induces apoptosis, evidenced by the release of mitochondrial Cytochrome c, activation of Caspase 3, and induction of DNA fragmentation (Kirchhoff et al., 2002). Moreover, the hypoxia can trigger the disassociation of the HSP60-Bax complex, accompanied with the translocation of Bax to the mitochondria and cytosolic HSP60 to the plasma membrane, which may be sufficient to induce cell apoptosis in adult rat cardiomyocytes (Gupta and Knowlton, 2002; Knowlton and Gupta, 2003).

## HSP60 Is Essential for Maintaining Mitochondrial Function and Cardiac Physiology

The importance of HSP60 has been characterized in *Escherichia coli* and yeast, in which the deficiency of HSP60 leads to



necrosis factor  $\alpha$  (TNF $\alpha$ ) and Interleukin 6 (IL-6) via activation of NF $\kappa$ B and JNK. In heart failure, HSP60 is released from cardiomyocytes. Increased serum levels of

a lethal phenotype (Cheng et al., 1989; Fayet et al., 1989). HSP60 is also essential for survival of Drosophila and mice (Perezgasga et al., 1999; Christensen et al., 2010). Inactivation of the *Hspd1* gene in mice results in early embryonic lethality while *Hspd1* haploinsufficiency can also cause a late onset and slowly progressive deficit in motor functions (Christensen et al., 2010; Magnoni et al., 2013). Moreover, it has been shown that missense mutations in human HSPD1 gene are associated with two extremely rare monogenic disorders-hereditary spastic paraplegia and a recessively inherited white matter disorder (Hansen et al., 2002; Magen et al., 2008). Recently, we generated a mouse model with inducible cardiac-specific HSP60 deletion and investigated the role of HSP60 in regulating mitochondrial function and cardiac physiology (Fan et al., 2019). We found that deletion of HSP60 in adult cardiomyocytes dramatically altered the activities of mitochondrial complexes, reduced mitochondrial membrane potential, increased ROS production, and eventually resulted in dilated cardiomyopathy, heart failure, and death of mice. Proteomic analysis in purified HSP60-deficient mitochondria suggested that about 20% of mitochondrial-localized proteins are HSP60-dependent, meaning they rely on HSP60 to mediate correct protein folding in the mitochondria. A survey of HSP60-interacting proteins was recently performed in HEK293 cells using coimmunoprecipitation and mass spectrometry and identified more than 300 proteins (Bie et al., 2020), 46 of which including ALDH2, CPT2, IDH3A, and SUCLG2 were downregulated in mitochondria of HSP60-deficient cardiomyocytes (Fan et al., 2019). Furthermore, an assessment of the mitochondrial protein import and stability found that deletion of HSP60 has no effect

HSP60 are related to the severity of heart failure.

on mitochondrial protein import. However, HSP60 deletion impels HSP60-dependent mitochondrial proteins to undergo degradation after import, which suggests that the protein exhibits low stability in HSP60-deficient mitochondria. Moreover, deletion of HSP60 activates the mitochondrial unfolded protein response (Fan et al., 2019) and is also accompanied with increased cell apoptosis (Fan et al., 2019). All these results together demonstrate that HSP60 is required for maintaining normal mitochondrial function and cardiac physiology.

# Extracellular HSP60 May Have an Injurious Effect on Cardiomyocytes

It is now clear that HSP60 also exists in the plasma, as well as in the extracellular space of cardiomyocytes (Pockley et al., 2000; Lewthwaite et al., 2002; Giannessi et al., 2007; Gupta and Knowlton, 2007; Lin et al., 2007; Kim et al., 2009; Li et al., 2011; Blasi et al., 2012), even though the exact mechanism underlying how HSP60 is secreted from cardiomyocytes is still under debate. First, HSP60 can be passively released as the intact or fragmented protein from damaged or dead cells, respectively (Basu et al., 2000). Secondly, HSP60 in certain non-cardiomyocytes can be secreted through the conventional endoplasmic reticulum-Golgi secretory pathway (Hayoun et al., 2012; Campanella et al., 2016). However, a nonconventional secretion mechanism, the lipid raftexosome pathway, is a more widely accepted way by which HSP60 in cardiomyocytes is secreted (Gupta and Knowlton, 2007; Lin et al., 2007). HSP60 in the exosomes is found to bind with the exosome membrane, and is released via exosomes in both the basal state and subsequent mild stress (Gupta and Knowlton, 2007; Lin et al., 2007). HSP60 can be used as a marker to indicate the number of extracellular vesicles released from the heart (Giricz et al., 2014). Alternatively, another point of view suggests that HSP60 may be stabilized within the exosome under multiple certain physiological conditions and is not released to prevent its toxicity to cardiomyocytes (Malik et al., 2013).

It is worthy to note that HSP60 in the plasma/serum reflects the total amount of the protein released by all types of organs and tissues, and extracellular HSP60 (exHSP60) in the heart tissue can be released by cardiomyocytes via exosomes as well as by necrosis or other routes, as mentioned above. In any case, exHSP60 is generally considered as an injurious signal in cardiomyocytes (Kim et al., 2009; Tian et al., 2013), even though exHSP60 has also been shown to play a beneficial effect on certain non-cardiomyocytes like B cells (Cohen-Sfady et al., 2009). exHSP60 may function as a ligand for the Toll-like receptors (TLRs) in many cell types including cardiomyocytes. TLR4 is the most highly expressed subtype of TLRs in cardiomyocytes (Frantz et al., 1999; Heiserman et al., 2015), and has been considered as the receptor of exHSP60 (Ohashi et al., 2000; Kim et al., 2009; Tian et al., 2013). exHSP60 can induce the release of tumor necrosis factor a (TNF $\alpha$ ) and Interleukin 6 from cardiomyocytes, which can be impeded by the inhibitors of P38 and NFkB (Tian et al., 2013). Meanwhile, exHSP60 also increases the expression of TLR2 and TLR4 in cardiomyocytes, which can be abolished by the inhibitors of JNK and NFkB (Tian et al., 2013). Activation of NFκB eventually leads to the release of Cytochrome C and AIF from the mitochondria, activation of Caspase-3/7, and cell apoptosis in cardiomyocytes treated with HSP60 (Kim et al., 2009; Knowlton, 2017). Consistently, treatment with an anti-TLR4 blocking antibody or deletion of TLR4 is able to totally abolish the exHSP60-induced cell apoptosis in cardiomyocytes (Kim et al., 2009; Heiserman et al., 2015). In addition, exHSP60 released from cardiomyocytes via exosomes or in other ways may also play a role in regulating cardiac fibroblast and endothelial cell functions, and thus affects the adaptive responses of hearts under stress (Cervio et al., 2015).

## HSP60 May Act as a Biomarker of Heart Failure

Heart failure, a process of chronic inflammation and progressive injury of cardiac muscle, is one of the most common complications of cardiovascular disease (Rizzo et al., 2011). Heart failure can result from many forms of heart disease, including dilated and ischemic cardiomyopathy. It has been shown that the expression of endogenous HSP60 is significantly elevated in the myocardium of patients with dilated and ischemic cardiomyopathy (Knowlton et al., 1998; Latif et al., 1999). During the progression of heart failure, NFkB is chronically activated, resulting in increased binding to the two NFkB binding elements in the HSP60 gene. This may account for increased expression of HSP60 in cardiomyocytes (Wong et al., 1998; Wang et al., 2010). In addition to the overall change of the HSP60 protein, HSP60 may redistribute between different subcellular locations in the cardiomyocyte under stress. As mentioned above, HSP60 in the cytosol can be translocated to the plasma membrane, which may cause the movement of Bax to the mitochondria as well as the activation of Caspases and apoptosis in the cardiomyocytes during heart failure (Gupta and Knowlton, 2007). This is consistent with a finding that HSP60 levels in the cytosol is reduced in dilated cardiomyopathy (DCM) hearts (Sidorik et al., 2005).

More importantly, serum HSP60 (sHSP60) may act as a biomarker for heart failure. Detectable sHSP60 levels have been observed in both healthy control human patients (Lewthwaite et al., 2002; Halcox et al., 2005) and patients with cardiovascular diseases (Xu et al., 2000; Shamaei-Tousi et al., 2006, 2007; Zhang et al., 2008). Acute myocardial infarction is able to induce HSP60 release, as manifested by a rise of sHSP60 levels soon after the onset of acute myocardial infarction and a positive correlation of these levels. Acute myocardial infarction also leads to adverse cardiovascular events and increased levels of creatine phosphokinase and troponin (Zhang et al., 2008; Novo et al., 2011). In patients with advanced chronic heart failure secondary to ischemic or idiopathic dilated cardiomyopathy, sHSP60 is correlated to the severity of the disease and is associated with a high risk of adverse cardiac events (Niizeki et al., 2008). In patients with acute heart failure, increased sHSP60 is also correlated with a higher risk for subsequent death/readmission for acute heart failure (Bonanad et al., 2013). Thus, sHSP60 levels could emerge as promising independent predictors of adverse cardiac events. Alternatively, exosomal HSP60 can serve as a

biomarker for diagnostics, assessing prognosis, and monitoring disease progression (Bavisotto et al., 2017).

### VASCULAR HSP60 AND ATHEROSCLEROSIS

Atherosclerosis progression is a complicated process that involves the participation of endothelial cells (ECs), VSMCs, macrophages, and other lymphocytes (Bennett et al., 2016; Gimbrone and Garcia-Cardena, 2016; Tabas and Bornfeldt, 2016). In atherosclerosis, the endothelial cell layer is disrupted by oxidized low-density lipoproteins (oxLDLs) (Negre-Salvayre et al., 2017). Macrophage foam cell formation occurs, and smooth muscle cells undergo migration and proliferation (Bennett et al., 2016). These mechanisms contribute to atherosclerotic plaque formation. It has been generally supposed that HSP60 is atherogenic (Grundtman et al., 2011). HSP60 can activate both the innate immune system via TLR4 and the adaptive immune system (Quintana and Cohen, 2011). However, the role of HSP60 in ECs and VSMCs has only been investigated in a limited number of studies. Here we describe the current understanding of how HSP60 functions on ECs and VSMCs, respectively, and discuss how vascular HSP60 is involved in the development of atherosclerosis (Figure 2).

### HSP60 May Regulate Cell Survival of ECs

Various atherogenic risk factors including physical insult (shear stress and heat), chemical stress (smoking, oxygen radicals, drugs, high sodium, and high glucose), infection (e.g., Chlamydia pneumoniae), and inflammation (LPS, inflammatory cytokines, and oxLDLs) possess the ability to regulate the expression or localization of HSP60 in ECs (Amberger et al., 1997; Hochleitner et al., 2000; Hirono et al., 2003; Wick et al., 2008, 2014; Kreutmayer et al., 2011, 2013; Mohammad and Kowluru, 2011; Zhao et al., 2015; Jakic et al., 2017). Shear stress plays a major role in the generation, progression, and destabilization of atherosclerotic plaques (Souilhol et al., 2019). Shear stress is able to induce HSP60 expression in ECs in vitro and in vivo (Hochleitner et al., 2000). ECs in atherosclerotic lesions of rabbits and humans also exhibit higher HSP60 levels than those in other regions of normal arterial intima (Kleindienst et al., 1993; Xu et al., 1994). Cigarette smoking is another significant risk factor for atherosclerosis (Knoflach et al., 2003). It has been shown that prolonged exposure to cigarette smoke alters mitochondrial structures. As a result, HSP60 is released from the mitochondria, transported to the cell surface, and then dispensed into the cell culture supernatant in ECs. Thus, cigarette smoke exposure likely accounts for the increased levels of HSP60 found in the serum of healthy young individuals exposed to second hand smoke (Bernhard et al., 2004; Kreutmayer et al., 2011). exHSP60 has been shown to induce TNFa in human umbilical vein ECs (Martinus and Goldsbury, 2018). However, there is one exceptional condition in which exHSP60 is beneficial to ECs. exHSP60 may bind to membrane ATP synthase and serve a protective role against EC acidification and cell apoptosis in the presence of anti-ATP

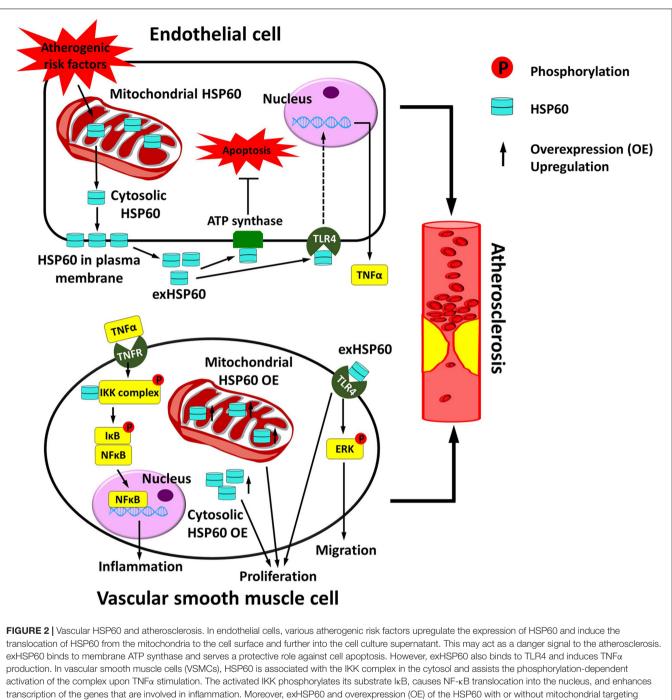
synthase antibodies (Champagne et al., 2006; Alard et al., 2011). On the other hand, downregulation of mitochondrial HSP60 in ECs is associated with accelerated apoptosis (Mohammad and Kowluru, 2011), whereas overexpression of HSP60 in ECs may exert a protective role in digoxin-induced apoptosis (Qiu et al., 2008). A recent study using Tie2-Cre to delete HSP60 in both hematopoietic cells and ECs suggests that HSP60 may be required for embryonic erythropoiesis and vascular development (Duan et al., 2019). However, it remains to be investigated in more detail whether and how HSP60 regulates EC physiology *in vivo*.

### HSP60 May Regulate Proliferation, Survival, and Migration of VSMCs

It has been shown that the overexpression of the HSP60 with or without mitochondrial targeting peptide can induce an increase in VSMC proliferation (Hirono et al., 2003; Deniset et al., 2018). exHSP60 may also play a very important role in regulating cell proliferation of VSMCs. Recombinant human HSP60 promotes cell proliferation in venous VSMCs, which can be inhibited by the application of TLR2 and TLR4 antibodies (de Graaf et al., 2006). Interestingly, exHSP60 can also promote cell migration of VSMCs. VSMCs exposed to HSP60 exhibit increased cell migration, which is accompanied with increased expression of TLR4 and ERK activity. Knockdown of TLR4 or the use of ERK inhibitors can significantly reduce HSP60induced VSMC migration (Zhao et al., 2015). On the other hand, cytosolic HSP60 plays an anti-apoptotic role in VSMCs. Deletion of cytosolic HSP60 in VSMCs is found to reduce the IKB kinase activation, repress the induction of NFKB-dependent survival genes, enhance apoptotic death in response to  $TNF\alpha$ , and markedly inhibit the neointimal thickening in the ballooninjured arterial vessels (Choi et al., 2015).

## **HSP60 Is Atherogenic**

Under stressful states, HSP60 is translocated to the cytosol and appears on the plasma membrane and in the extracellular space. HSP60 can also be directly released from damaged and dying cells (Xu et al., 1994; Wick et al., 2014). Higher expression of HSP60 can be found in ECs in atherosclerotic lesions of rabbits and humans compared with those in other parts of normal arterial intima (Kleindienst et al., 1993; Xu et al., 1994). Consistently, elevated levels of soluble or circulating HSP60 have been shown to correlate with an increased risk of atherosclerosis (Pockley et al., 2000; Xu et al., 2000; Shamaei-Tousi et al., 2007; Grundtman et al., 2011). HSP60 on the membrane and in the extracellular space has been widely considered to act as a danger signal to the atherosclerosis in several aspects. First, HSP60 has a high degree of homology in both protein and DNA levels between different bacterial species, and from prokaryotic to eukaryotic cells (Craig et al., 1993; Grundtman et al., 2011). All humans develop cellular and humoral immunity against bacterial HSP60 either by infection or vaccination. This protective defense may increase the risk of cross-reactivity with autologous HSP60, which causes the adhesion of HSP60reactive T cells to the endothelial cells and the initiation of the



peptide can induce cell proliferation. In addition, exHSP60 can bind to TLR4 to promote cell migration via activation of ERK. In short, HSP60 regulates the inflammation, proliferation and migration of VSMCs, which may further accelerate atherosclerosis.

earliest inflammatory responses of atherosclerosis. In addition, antibodies against HSP60 are able to accelerate and perpetuate the disease (Schett et al., 1997; Stocker and Keaney, 2004; Grundtman et al., 2011; Wick et al., 2014). If the reversible early inflammatory stage of atherosclerosis is not interfered or risk factors remain present, irreversible late stages with atherosclerotic plaques will develop and lead to clinical consequences. Second, exHSP60 can react to TLR4 expressed in endothelial cells, smooth

muscle cells, and macrophages (Xu et al., 2012), as well as activate NF $\kappa$ B-dependent signaling pathways. It can also promote the production of various proteolytic enzymes and cytokine such as TNF, IL10, adhesion molecules, and growth factors (Moghimpour Bijani et al., 2012). Third, exHSP60 may induce the proliferation and migration of VSMCs (Hirono et al., 2003; de Graaf et al., 2006; Zhao et al., 2015; Deniset et al., 2018), which may further accelerate atherosclerosis.

## CONCLUSION

It is now clear that HSP60 plays multiple regulatory roles in cardiovascular physiology. Mitochondrial HSP60 together with its chaperonin HSP10 regulates mitochondrial protein folding. Deletion of HSP60 results in mitochondrial dysfunction, chamber dilation, and heart failure. HSP60 undergoes subcellular translocation and secretion in response to stress and cell / tissue injury, and is considered as a pathogenic signal in heart failure, atherosclerosis, and other cardiovascular diseases. exHSP60 may induce cell apoptosis in cardiomyocytes and thus exacerbates the disease state of heart failure. exHSP60 may also participate in the initial step of atherosclerosis by inducing multiple inflammatory responses, and accelerates the progression of atherosclerosis by promoting VSMC cell proliferation and migration. All of these results have provided us with a better understanding of HSP60 functions in the cardiovascular system and may contribute to the future development of novel therapies targeting HSP60 for the treatment of cardiovascular disease.

### REFERENCES

- Alard, J. E., Hillion, S., Guillevin, L., Saraux, A., Pers, J. O., Youinou, P., et al. (2011). Autoantibodies to endothelial cell surface ATP synthase, the endogenous receptor for hsp60, might play a pathogenic role in vasculatides. *Plos One* 6:e14654. doi: 10.1371/journal.pone.0014654
- Amberger, A., Maczek, C., Jurgens, G., Michaelis, D., Schett, G., Trieb, K., et al. (1997). Co-expression of ICAM-1, VCAM-1, ELAM-1 and Hsp60 in human arterial and venous endothelial cells in response to cytokines and oxidized low-density lipoproteins. *Cell Stress Chaperones* 2, 94–103.
- Basu, S., Binder, R. J., Suto, R., Anderson, K. M., and Srivastava, P. K. (2000). Necrotic but not apoptotic cell death releases heat shock proteins, which deliver a partial maturation signal to dendritic cells and activate the NF-kappa B pathway. *Int. Immunol.* 12, 1539–1546. doi: 10.1093/intimm/12.11.1539
- Bavisotto, C. C., Cappello, F., Macario, A. J. L., de Macario, E. C., Logozzi, M., Fais, S., et al. (2017). Exosomal HSP60: a potentially useful biomarker for diagnosis, assessing prognosis, and monitoring response to treatment. *Expert Rev. Mol. Diagn.* 17, 815–822. doi: 10.1080/14737159.2017.1356230
- Bennett, M. R., Sinha, S., and Owens, G. K. (2016). Vascular smooth muscle cells in atherosclerosis. *Circ. Res.* 118, 692–702. doi: 10.1161/CIRCRESAHA.115. 306361
- Bernhard, D., Huck, C. W., Jakschitz, T., Pfister, G., Henderson, B., Bonn, G. K., et al. (2004). Development and evaluation of an in vitro model for the analysis of cigarette smoke effects on cultured cells and tissues. *J. Pharmacol. Toxicol. Methods* 50, 45–51. doi: 10.1016/j.vascn.2004.01.003
- Bie, A. S., Comert, C., Korner, R., Corydon, T. J., Palmfeldt, J., Hipp, M. S., et al. (2020). An inventory of interactors of the human HSP60/HSP10 chaperonin in the mitochondrial matrix space. *Cell Stress Chaperones* [Epub ahead of print]. doi: 10.1007/s12192-020-01080-6
- Blasi, C., Kim, E., and Knowlton, A. A. (2012). Improved metabolic control in diabetes, HSP60, and proinflammatory mediators. *Autoimmune Dis.* 2012:346501. doi: 10.1155/2012/346501
- Bonanad, C., Nunez, J., Sanchis, J., Bodi, V., Chaustre, F., Chillet, M., et al. (2013). Serum heat shock protein 60 in acute heart failure: a new biomarker? *Congest Heart Fail* 19, 6–10. doi: 10.1111/j.1751-7133.2012.00299.x
- Campanella, C., D'Anneo, A., Marino Gammazza, A., Caruso Bavisotto, C., Barone, R., Emanuele, S., et al. (2016). The histone deacetylase inhibitor SAHA induces HSP60 nitration and its extracellular release by exosomal vesicles in human lung-derived carcinoma cells. *Oncotarget* 7, 28849–28867. doi: 10. 18632/oncotarget.6680
- Cervio, E., Barile, L., Moccetti, T., and Vassalli, G. (2015). Exosomes for intramyocardial intercellular communication. *Stem Cells Int.* 2015, 482171. doi: 10.1155/2015/482171

## **AUTHOR CONTRIBUTIONS**

YD, HT, KM, XF, ZH, and KO wrote the manuscript.

## FUNDING

This work was supported by the Shenzhen Basic Research Foundation (JCYJ20160428154108239, KQJSCX20170330155020 267, JCYJ20170818090044949), the Guangdong Province Basic Research Foundation (2018A030310012), the National Key Basic Research Program of China (2013CB531200), and the National Science Foundation of China (81700289, 31800767, 81970421).

## ACKNOWLEDGMENTS

We would like to thank Dr. Ju Chen at University of California, San Diego for critical reading of the manuscript.

- Champagne, E., Martinez, L. O., Collet, X., and Barbaras, R. (2006). Ecto-F1Fo ATP synthase/F1 ATPase: metabolic and immunological functions. *Curr. Opin. Lipidol.* 17, 279–284. doi: 10.1097/01.mol.0000226120.27931.76
- Chandra, D., Choy, G., and Tang, D. G. (2007). Cytosolic accumulation of HSP60 during apoptosis with or without apparent mitochondrial release: evidence that its pro-apoptotic or pro-survival functions involve differential interactions with caspase-3. J. Biol. Chem. 282, 31289–31301. doi: 10.1074/jbc.M702777200
- Cheng, M. Y., Hartl, F. U., Martin, J., Pollock, R. A., Kalousek, F., Neupert, W., et al. (1989). Mitochondrial heat-shock protein hsp60 is essential for assembly of proteins imported into yeast mitochondria. *Nature* 337, 620–625. doi: 10. 1038/337620a0
- Choi, B., Choi, M., Park, C., Lee, E. K., Kang, D. H., Lee, D. J., et al. (2015). Cytosolic Hsp60 orchestrates the survival and inflammatory responses of vascular smooth muscle cells in injured aortic vessels. *Cardiovasc Res.* 106, 498–508. doi: 10. 1093/cvr/cvv130
- Christensen, J. H., Nielsen, M. N., Hansen, J., Fuchtbauer, A., Fuchtbauer, E. M., West, M., et al. (2010). Inactivation of the hereditary spastic paraplegiaassociated Hspd1 gene encoding the Hsp60 chaperone results in early embryonic lethality in mice. *Cell Stress Chaperones* 15, 851–863. doi: 10.1007/ s12192-010-0194-x
- Cohen-Sfady, M., Pevsner-Fischer, M., Margalit, R., and Cohen, I. R. (2009). Heat shock protein 60, via MyD88 innate signaling, protects B cells from apoptosis, spontaneous and induced. *J. Immunol.* 183, 890–896. doi: 10.4049/jimmunol. 0804238
- Craig, E. A., Gambill, B. D., and Nelson, R. J. (1993). Heat shock proteins: molecular chaperones of protein biogenesis. *Microbiol. Rev.* 57, 402–414.
- de Graaf, R., Kloppenburg, G., Peter, J. H. M. K., Bruggeman, C. A., and Stassen, F. (2006). Human heat shock protein 60 stimulates vascular smooth muscle cell proliferation through Toll-like receptors 2 and 4. *Microbes Infect.* 8, 1859–1865. doi: 10.1016/j.micinf.2006.02.024
- Deniset, J. F., Hedley, T. E., Hlavackova, M., Chahine, M. N., Dibrov, E., O'Hara, K., et al. (2018). Heat shock protein 60 involvement in vascular smooth muscle cell proliferation. *Cell Signal.* 47, 44–51.
- Duan, Y., Wang, H., Mitchell-Silbaugh, K., Cai, S., Fan, F., Li, Y., et al. (2019). Heat shock protein 60 regulates yolk sac erythropoiesis in mice. *Cell Death Dis.* 10:766. doi: 10.1038/s41419-019-2014-2
- Fan, F., Duan, Y., Yang, F., Trexler, C., Wang, H., Huang, L., et al. (2019). Deletion of heat shock protein 60 in adult mouse cardiomyocytes perturbs mitochondrial protein homeostasis and causes heart failure. *Cell Death Differ*. 27, 587–600. doi: 10.1038/s41418-019-0374-x
- Fayet, O., Ziegelhoffer, T., and Georgopoulos, C. (1989). The groES and groEL heat shock gene products of *Escherichia coli* are essential for bacterial growth at

all temperatures. J. Bacteriol. 171, 1379–1385. doi: 10.1128/jb.171.3.1379-1385. 1989

- Frantz, S., Kobzik, L., Kim, Y. D., Fukazawa, R., Medzhitov, R., Lee, R. T., et al. (1999). Toll4 (TLR4) expression in cardiac myocytes in normal and failing myocardium. J. Clin. Invest. 104, 271–280. doi: 10.1172/JCI6709
- Giannessi, D., Colotti, C., Maltinti, M., Del Ry, S., Prontera, C., Turchi, S., et al. (2007). Circulating heat shock proteins and inflammatory markers in patients with idiopathic left ventricular dysfunction: their relationships with myocardial and microvascular impairment. *Cell Stress Chaperones* 12, 265–274. doi: 10. 1379/csc-272.1
- Gimbrone, M. A. Jr., and Garcia-Cardena, G. (2016). endothelial cell dysfunction and the pathobiology of atherosclerosis. *Circ. Res.* 118, 620–636. doi: 10.1161/ CIRCRESAHA.115.306301
- Giricz, Z., Varga, Z. V., Baranyai, T., Sipos, P., Paloczi, K., Kittel, A., et al. (2014). Cardioprotection by remote ischemic preconditioning of the rat heart is mediated by extracellular vesicles. *J. Mol. Cell Cardiol.* 68, 75–78. doi: 10.1016/ j.yjmcc.2014.01.004
- Go, A. S., Mozaffarian, D., Roger, V. L., Benjamin, E. J., Berry, J. D., Blaha, M. J., et al. (2014). Heart disease and stroke statistics–2014 update: a report from the American heart association. *Circulation* 129, e28–e292. doi: 10.1161/01.cir. 0000441139.02102.80
- Grundtman, C., Kreutmayer, S. B., Almanzar, G., Wick, M. C., and Wick, G. (2011). Heat shock protein 60 and immune inflammatory responses in atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 31, 960–968. doi: 10.1161/ATVBAHA.110. 217877
- Gupta, S., and Knowlton, A. A. (2002). Cytosolic heat shock protein 60, hypoxia, and apoptosis. *Circulation* 106, 2727–2733. doi: 10.1161/01.cir.0000038112. 64503.6e
- Gupta, S., and Knowlton, A. A. (2005). HSP60, Bax, apoptosis and the heart. J. Cell Mol. Med. 9, 51–58.
- Gupta, S., and Knowlton, A. A. (2007). HSP60 trafficking in adult cardiac myocytes: role of the exosomal pathway. Am. J. Physiol. Heart Circ. Physiol. 292, H3052– H3056. doi: 10.1152/ajpheart.01355.2006
- Halcox, J. P., Deanfield, J., Shamaei-Tousi, A., Henderson, B., Steptoe, A., Coates, A. R., et al. (2005). Circulating human heat shock protein 60 in the blood of healthy teenagers: a novel determinant of endothelial dysfunction and early vascular injury? *Arterioscler Thromb. Vasc. Biol.* 25:e141. doi: 10.1161/01.ATV. 0000185832.34992.ff
- Hansen, J. J., Durr, A., Cournu-Rebeix, I., Georgopoulos, C., Ang, D., Nielsen, M. N., et al. (2002). Hereditary spastic paraplegia SPG13 is associated with a mutation in the gene encoding the mitochondrial chaperonin Hsp60. Am. J. Hum. Genet. 70, 1328–1332. doi: 10.1086/339935
- Hayoun, D., Kapp, T., Edri-Brami, M., Ventura, T., Cohen, M., Avidan, A., et al. (2012). HSP60 is transported through the secretory pathway of 3-MCAinduced fibrosarcoma tumour cells and undergoes N-glycosylation. *Febs. J.* 279, 2083–2095. doi: 10.1111/j.1742-4658.2012.08594.x
- Heiserman, J. P., Chen, L., Kim, B. S., Kim, S. C., Tran, A. L., Siebenborn, N., et al. (2015). TLR4 mutation and HSP60-induced cell death in adult mouse cardiac myocytes. *Cell Stress Chaperones* 20, 527–535. doi: 10.1007/s12192-015-0577-0
- Henderson, B., Fares, M. A., and Lund, P. A. (2013). Chaperonin 60: a paradoxical, evolutionarily conserved protein family with multiple moonlighting functions. *Biol. Rev.* 88, 955–987. doi: 10.1111/brv.12037
- Hirono, S., Dibrov, E., Hurtado, C., Kostenuk, A., Ducas, R., and Pierce, G. N. (2003). Chlamydia pneumoniae stimulates proliferation of vascular smooth muscle cells through induction of endogenous heat shock protein 60. *Circ. Res.* 93, 710–716. doi: 10.1161/01.RES.0000095720.46043.F2
- Hochleitner, B. W., Hochleitner, E. O., Obrist, P., Eberl, T., Amberger, A., Xu, Q., et al. (2000). Fluid shear stress induces heat shock protein 60 expression in endothelial cells in vitro and in vivo. *Arterioscler. Thromb. Vasc. Biol.* 20, 617–623.
- Horwich, A. L., Farr, G. W., and Fenton, W. A. (2006). GroEL-GroES-mediated protein folding. *Chem. Rev.* 106, 1917–1930. doi: 10.1021/cr040435v
- Jakic, B., Buszko, M., Cappellano, G., and Wick, G. (2017). Elevated sodium leads to the increased expression of HSP60 and induces apoptosis in HUVECs. *Plos One* 12:e0179383. doi: 10.1371/journal.pone.0179383
- Kim, S. C., Stice, J. P., Chen, L., Jung, J. S., Gupta, S., Wang, Y., et al. (2009). Extracellular heat shock protein 60, cardiac myocytes, and apoptosis. *Circ. Res.* 105, 1186–1195. doi: 10.1161/CIRCRESAHA.109.209643

- Kirchhoff, S. R., Gupta, S., and Knowlton, A. A. (2002). Cytosolic heat shock protein 60, apoptosis, and myocardial injury. *Circulation* 105, 2899–2904. doi: 10.1161/01.cir.0000019403.35847.23
- Kleindienst, R., Xu, Q., Willeit, J., Waldenberger, F. R., Weimann, S., and Wick, G. (1993). Immunology of atherosclerosis. demonstration of heat shock protein 60 expression and T lymphocytes bearing alpha/beta or gamma/delta receptor in human atherosclerotic lesions. *Am. J. Pathol.* 142, 1927–1937.
- Knoflach, M., Kiechl, S., Kind, M., Said, M., Sief, R., Gisinger, M., et al. (2003). Cardiovascular risk factors and atherosclerosis in young males: ARMY study (Atherosclerosis Risk-Factors in Male Youngsters). *Circulation* 108, 1064–1069. doi: 10.1161/01.CIR.0000085996.95532.FF
- Knowlton, A. A. (2017). Paying for the tolls: the high cost of the innate immune system for the cardiac myocyte. *Adv. Exp. Med. Biol.* 1003, 17–34. doi: 10.1007/ 978-3-319-57613-8\_2
- Knowlton, A. A., and Gupta, S. (2003). HSP60, Bax, and cardiac apoptosis. *Cardiovasc. Toxicol.* 3, 263–268.
- Knowlton, A. A., Kapadia, S., Torre-Amione, G., Durand, J. B., Bies, R., Young, J., et al. (1998). Differential expression of heat shock proteins in normal and failing human hearts. J. Mol. Cell Cardiol. 30, 811–818. doi: 10.1006/jmcc.1998.0646
- Kreutmayer, S., Csordas, A., Kern, J., Maass, V., Almanzar, G., Offterdinger, M., et al. (2013). Chlamydia pneumoniae infection acts as an endothelial stressor with the potential to initiate the earliest heat shock protein 60-dependent inflammatory stage of atherosclerosis. *Cell Stress Chaperones* 18, 259–268.
- Kreutmayer, S. B., Messner, B., Knoflach, M., Henderson, B., Niederegger, H., Boeck, G., et al. (2011). Dynamics of heat shock protein 60 in endothelial cells exposed to cigarette smoke extract. *J. Mol. Cell Cardiol.* 51, 777–780.
- Latif, N., Taylor, P. M., Khan, M. A., Yacoub, M. H., and Dunn, M. J. (1999). The expression of heat shock protein 60 in patients with dilated cardiomyopathy. *Basic Res. Cardiol.* 94, 112–119.
- Lau, S., Patnaik, N., Sayen, M. R., and Mestril, R. (1997). Simultaneous overexpression of two stress proteins in rat cardiomyocytes and myogenic cells confers protection against ischemia-induced injury. *Circulation* 96, 2287–2294. doi: 10.1161/01.cir.96.7.2287
- Levy-Rimler, G., Viitanen, P., Weiss, C., Sharkia, R., Greenberg, A., Niv, A., et al. (2001). The effect of nucleotides and mitochondrial chaperonin 10 on the structure and chaperone activity of mitochondrial chaperonin 60. *Eur. J. Biochem.* 268, 3465–3472. doi: 10.1046/j.1432-1327.2001.02243.x
- Lewthwaite, J., Owen, N., Coates, A., Henderson, B., and Steptoe, A. (2002). Circulating human heat shock protein 60 in the plasma of British civil servants: relationship to physiological and psychosocial stress. *Circulation* 106, 196–201. doi: 10.1161/01.cir.0000021121.26290.2c
- Li, Y., Si, R., Feng, Y., Chen, H. H., Zou, L., Wang, E., et al. (2011). Myocardial ischemia activates an injurious innate immune signaling via cardiac heat shock protein 60 and Toll-like receptor 4. *J. Biol. Chem.* 286, 31308–31319. doi: 10. 1074/jbc.M111.246124
- Lin, K. M., Lin, B., Lian, I. Y., Mestril, R., Scheffler, I. E., and Dillmann, W. H. (2001). Combined and individual mitochondrial HSP60 and HSP10 expression in cardiac myocytes protects mitochondrial function and prevents apoptotic cell deaths induced by simulated ischemia-reoxygenation. *Circulation* 103, 1787–1792.
- Lin, L., Kim, S. C., Wang, Y., Gupta, S., Davis, B., Simon, S. I., et al. (2007). HSP60 in heart failure: abnormal distribution and role in cardiac myocyte apoptosis. *Am. J. Physiol. Heart C* 293, H2238–H2247.
- Magen, D., Georgopoulos, C., Bross, P., Ang, D., Segev, Y., Goldsher, D., et al. (2008). Mitochondrial hsp60 chaperonopathy causes an autosomalrecessive neurodegenerative disorder linked to brain hypomyelination and leukodystrophy. Am. J. Hum. Genet. 83, 30–42. doi: 10.1016/j.ajhg.2008.0 5.016
- Magnoni, R., Palmfeldt, J., Christensen, J. H., Sand, M., Maltecca, F., Corydon, T. J., et al. (2013). Late onset motoneuron disorder caused by mitochondrial Hsp60 chaperone deficiency in mice. *Neurobiol. Dis.* 54, 12–23. doi: 10.1016/j. nbd.2013.02.012
- Malik, Z. A., Kott, K. S., Poe, A. J., Kuo, T., Chen, L., Ferrara, K. W., et al. (2013). Cardiac myocyte exosomes: stability, HSP60, and proteomics. *Am. J. Physiol. Heart C* 304, H954–H965. doi: 10.1152/ajpheart.00835.2012
- Martinus, R. D., and Goldsbury, J. (2018). Endothelial TNF-alpha induction by Hsp60 secreted from THP-1 monocytes exposed to hyperglycaemic conditions. *Cell Stress Chaperones* 23, 519–525. doi: 10.1007/s12192-017-0858-x

- Meng, Q. L., Li, B. X., and Xiao, X. S. (2018). Toward developing chemical modulators of Hsp60 as potential therapeutics. *Front. Mol. Biosci.* 5:35. doi: 10.3389/fmolb.2018.00035
- Moghimpour Bijani, F., Vallejo, J. G., and Rezaei, N. (2012). Toll-like receptor signaling pathways in cardiovascular diseases: challenges and opportunities. *Int. Rev. Immunol.* 31, 379–395. doi: 10.3109/08830185.2012.706761
- Mohammad, G., and Kowluru, R. A. (2011). Novel role of mitochondrial matrix metalloproteinase-2 in the development of diabetic retinopathy. *Invest Ophthalmol. Vis. Sci.* 52, 3832–3841. doi: 10.1167/iovs.10-6368
- Negre-Salvayre, A., Auge, N., Camare, C., Bacchetti, T., Ferretti, G., and Salvayre, R. (2017). Dual signaling evoked by oxidized LDLs in vascular cells. *Free Radic. Biol. Med.* 106, 118–133. doi: 10.1016/j.freeradbiomed.2017.02.006
- Niizeki, T., Takeishi, Y., Watanabe, T., Nitobe, J., Miyashita, T., Miyamoto, T., et al. (2008). Relation of serum heat shock protein 60 level to severity and prognosis in chronic heart failure secondary to ischemic or idiopathic dilated cardiomyopathy. *Am. J. Cardiol.* 102, 606–610. doi: 10.1016/j.amjcard.2008.0 4.030
- Nisemblat, S., Yaniv, O., Parnas, A., Frolow, F., and Azem, A. (2015). Crystal structure of the human mitochondrial chaperonin symmetrical football complex. *Proc. Natl. Acad. Sci. U. S. A.* 112, 6044–6049. doi: 10.1073/pnas. 1411718112
- Novo, G., Cappello, F., Rizzo, M., Fazio, G., Zambuto, S., Tortorici, E., et al. (2011). Hsp60 and heme oxygenase-1 (Hsp32) in acute myocardial infarction. *Transl. Res.* 157, 285–292. doi: 10.1016/j.trsl.2011.01.003
- Ohashi, K., Burkart, V., Flohe, S., and Kolb, H. (2000). Cutting edge: heat shock protein 60 is a putative endogenous ligand of the toll-like receptor-4 complex. *J. Immunol.* 164, 558–561. doi: 10.4049/jimmunol.164.2.558
- Ostermann, J., Horwich, A. L., Neupert, W., and Hartl, F. U. (1989). Protein folding in mitochondria requires complex formation with hsp60 and ATP hydrolysis. *Nature* 341, 125–130. doi: 10.1038/341125a0
- Perezgasga, L., Segovia, L., and Zurita, M. (1999). Molecular characterization of the 5' control region and of two lethal alleles affecting the hsp60 gene in Drosophila melanogaster. *Febs Lett.* 456, 269–273. doi: 10.1016/s0014-5793(99)00963-1
- Pockley, A. G., Wu, R., Lemne, C., Kiessling, R., de Faire, U., and Frostegard, J. (2000). Circulating heat shock protein 60 is associated with early cardiovascular disease. *Hypertension* 36, 303–307. doi: 10.1161/01.hyp.36.2.303
- Qiu, J., Gao, H. Q., Liang, Y., Yu, H., and Zhou, R. H. (2008). Comparative proteomics analysis reveals role of heat shock protein 60 in digoxin-induced toxicity in human endothelial cells. *Biochim. Biophys. Acta* 1784, 1857–1864. doi: 10.1016/j.bbapap.2008.07.006
- Quintana, F. J., and Cohen, I. R. (2011). The HSP60 immune system network. *Trends Immunol.* 32, 89–95. doi: 10.1016/j.it.2010.11.001
- Rizzo, M., Macario, A. J. L., de Macario, E. C., Gouni-Berthold, I., Berthold, H. K., Rini, G. B., et al. (2011). Heat shock protein-60 and risk for cardiovascular disease. *Curr. Pharm. Design.* 17, 3662–3668.
- Schett, G., Metzler, B., Kleindienst, R., Moschen, I., Hattmannsdorfer, R., Wolf, H., et al. (1997). Salivary anti-hsp65 antibodies as a diagnostic marker for gingivitis and a possible link to atherosclerosis. *Int. Arch. Allergy Immunol.* 114, 246–250. doi: 10.1159/000237675
- Shamaei-Tousi, A., Stephens, J. W., Bin, R., Cooper, J. A., Steptoe, A., Coates, A. R., et al. (2006). Association between plasma levels of heat shock protein 60 and cardiovascular disease in patients with diabetes mellitus. *Eur. Heart J.* 27, 1565–1570. doi: 10.1093/eurheartj/ehl081
- Shamaei-Tousi, A., Steptoe, A., O'Donnell, K., Palmen, J., Stephens, J. W., Hurel, S. J., et al. (2007). Plasma heat shock protein 60 and cardiovascular disease risk: the role of psychosocial, genetic, and biological factors. *Cell Stress Chaperones* 12, 384–392. doi: 10.1379/csc-300.1
- Sidorik, L., Kyyamova, R., Bobyk, V., Kapustian, L., Rozhko, O., Vigontina, O., et al. (2005). Molecular chaperone, HSP60, and cytochrome P450 2E1 co-expression in dilated cardiomyopathy. *Cell Biol Int.* 29, 51–55. doi: 10.1016/j.cellbi.2004.1 1.011
- Singh, B., Patel, H. V., Ridley, R. G., Freeman, K. B., and Gupta, R. S. (1990). Mitochondrial import of the human chaperonin (HSP60) protein. *Biochem. Biophys. Res. Commun.* 169, 391–396. doi: 10.1016/0006-291x(90)90344-m

- Souilhol, C., Serbanovic-Canic, J., Fragiadaki, M., Chico, T. J., Ridger, V., Roddie, H., et al. (2019). Endothelial responses to shear stress in atherosclerosis: a novel role for developmental genes. *Nat. Rev. Cardiol.* 17, 52–63. doi: 10.1038/s41569-019-0239-5
- Stocker, R., and Keaney, J. F. Jr. (2004). Role of oxidative modifications in atherosclerosis. *Physiol. Rev.* 84, 1381–1478. doi: 10.1152/physrev.00047.2003
- Tabas, I., and Bornfeldt, K. E. (2016). Macrophage phenotype and function in different stages of atherosclerosis. *Circ. Res.* 118, 653–667. doi: 10.1161/ CIRCRESAHA.115.306256
- Taguchi, H., Makino, Y., and Yoshida, M. (1994). Monomeric chaperonin-60 and its 50-kDa fragment possess the ability to interact with non-native proteins, to suppress aggregation, and to promote protein folding. *J. Biol. Chem.* 269, 8529–8534.
- Tian, J., Guo, X., Liu, X. M., Liu, L., Weng, Q. F., Dong, S. J., et al. (2013). Extracellular HSP60 induces inflammation through activating and up-regulating TLRs in cardiomyocytes. *Cardiovasc. Res.* 98, 391–401.
- Wang, Y., Chen, L., Hagiwara, N., and Knowlton, A. A. (2010). Regulation of heat shock protein 60 and 72 expression in the failing heart. J. Mol. Cell Cardiol. 48, 360–366. doi: 10.1016/j.yjmcc.2009.11.009
- Wick, G., Jakic, B., Buszko, M., Wick, M. C., and Grundtman, C. (2014). The role of heat shock proteins in atherosclerosis. *Nat. Rev. Cardiol.* 11, 516–529.
- Wick, M. C., Mayerl, C., Backovic, A., van der Zee, R., Jaschke, W., Dietrich, H., et al. (2008). In vivo imaging of the effect of LPS on arterial endothelial cells: molecular imaging of heat shock protein 60 expression. *Cell Stress Chaperones* 13, 275–285. doi: 10.1007/s12192-008-0044-2
- Wiechmann, K., Muller, H., Konig, S., Wielsch, N., Svatos, A., Jauch, J., et al. (2017). Mitochondrial chaperonin HSP60 is the apoptosis-related target for myrtucommulone. *Cell Chem. Biol.* 24, 614.e6–623.e6. doi: 10.1016/j.chembiol. 2017.04.008
- Wong, S. C., Fukuchi, M., Melnyk, P., Rodger, I., and Giaid, A. (1998). Induction of cyclooxygenase-2 and activation of nuclear factor-kappaB in myocardium of patients with congestive heart failure. *Circulation*98, 100–103. doi: 10.1161/01. cir.98.2.100
- Xu, Q., Schett, G., Perschinka, H., Mayr, M., Egger, G., Oberhollenzer, F., et al. (2000). Serum soluble heat shock protein 60 is elevated in subjects with atherosclerosis in a general population. *Circulation* 102, 14–20. doi: 10.1161/ 01.cir.102.1.14
- Xu, Q., Schett, G., Seitz, C. S., Hu, Y., Gupta, R. S., and Wick, G. (1994). Surface staining and cytotoxic activity of heat-shock protein 60 antibody in stressed aortic endothelial cells. *Circ Res.* 75, 1078–1085. doi: 10.1161/01.res.75.6.1078
- Xu, Q. B., Metzler, B., Jahangiri, M., and Mandal, K. (2012). Molecular chaperones and heat shock proteins in atherosclerosis. *Am. J. Physiol. Heart. C* 302, H506–H514.
- Zhang, X., He, M., Cheng, L., Chen, Y., Zhou, L., Zeng, H., et al. (2008). Elevated heat shock protein 60 levels are associated with higher risk of coronary heart disease in Chinese. *Circulation* 118, 2687–2693. doi: 10.1161/ CIRCULATIONAHA.108.781856
- Zhao, Y., Zhang, C. X., Wei, X. G., Li, P., Cui, Y., Qin, Y. H., et al. (2015). Heat shock protein 60 stimulates the migration of vascular smooth muscle cells via Toll-like receptor 4 and ERK MAPK activation. *Sci. Rep.* 5:15352. doi: 10.1038/srep15352
- Zininga, T., Ramatsui, L., and Shonhai, A. (2018). Heat shock proteins as immunomodulants. *Molecules* 23:2846. doi: 10.3390/molecules23112846

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Duan, Tang, Mitchell-silbaugh, Fang, Han and Ouyang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.