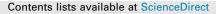


Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Bioorganic & Medicinal Chemistry Letters 27 (2017) 139-142



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Synthesis and *in vitro* antiviral evaluation of 4-substituted 3,4-dihydropyrimidinones



Dhanabal Kumarasamy^{a,*}, Biswajit Gopal Roy^b, Joana Rocha-Pereira^c, Johan Neyts^c, Satheeshkumar Nanjappan^d, Subhasis Maity^a, Musfiqua Mookerjee^a, Lieve Naesens^c

^a Department of Pharmaceutical Chemistry, NSHM College of Pharmaceutical Technology, 124, B.L Saha Road, Kolkata 700053, India

^b Department of Chemistry, School of Physical Sciences, Sikkim University, 6th Mile, Tadong, Gangtok, Sikkim 737102, India

^c Laboratory of Virology and Chemotherapy, Rega Institute for Medical Research, KU Leuven, Minderbroedersstraat 10, Leuven 3000, Belgium

^d Department of Pharmaceutical Analysis, National Institute of Pharmaceutical Education and Research [NIPER-H], Balanagar, Hyderabad 500037, India

ARTICLE INFO

Article history: Received 22 May 2016 Revised 8 November 2016 Accepted 2 December 2016 Available online 5 December 2016

Keywords: Dihydropyrimidinones DHPM Antiviral activity Cytotoxicity Bunyavirus

ABSTRACT

A series of 4-substituted 3,4-dihydropyrimidine-2-ones (DHPM) was synthesized, characterized by IR, ¹H NMR, ¹³C NMR and HRMS spectra. The compounds were evaluated *in vitro* for their antiviral activity against a broad range of DNA and RNA viruses, along with assessment for potential cytotoxicity in diverse mammalian cell lines. Compound **4m**, which possesses a long lipophilic side chain, was found to be a potent and selective inhibitor of Punta Toro virus, a member of the *Bunyaviridae*. For Rift Valley fever virus, which is another Bunyavirus, the activity of **4m** was negligible. DHPMs with a C-4 aryl moiety bearing halogen substitution (**4b**, **4c** and **4d**) were found to be cytotoxic in MT4 cells.

© 2016 Elsevier Ltd. All rights reserved.

Emerging and re-emerging viruses are capable of rapidly overcoming their natural geographical boundaries and spreading among the human population. Most of these viral outbreaks are due to zoonotic viruses, some of which were previously unknown.^{1,2} With an increasing risk for vector-borne outbreaks, the Bunyaviridae present a particular and global threat to public health. Members of this virus family are the cause of Hantavirus cardiopulmonary syndrome in the Americas and Schmallenberg virus-induced congenital malformations in ruminants, which emerged in Europe in 2011. Severe fever with thrombocytopenia syndrome (SFTS) is an emerging infectious disease caused by the SFTS virus. In addition, some well-known Bunyaviruses like Rift Valley fever virus (RVFV) and Crimean-Congo haemorrhagic fever virus continue to emerge in new geographical locations.^{3,4} RVFV is transmitted by a wide variety of mosquitoes and its geographic distribution is expanding, with the potential of reaching Europe in the near future. This virus causes devastating epidemics in Africa and Arabia, killing hundreds of humans and thousands of wild and livestock animals. The transmission of Bunyaviruses, and their cell entry pathways and cell receptors remain poorly characterized,

* Corresponding author. *E-mail address: dhanabalrx@gmail.com* (D. Kumarasamy). partly due to the fact that experiments with wild-type RVFV are limited to enhanced biosafety level-4 (BSL-4) facilities, which are very few in number.⁵ The lack of vaccines and antiviral medications makes the potential spread of these Bunyaviruses a global concern. Recently, in an effort to repurpose FDA-approved drugs for RVFV treatment, the anticancer drug sorafenib was found to be an effective inhibitor of RVFV replication.⁶ Another agent that is able to suppress RVFV replication in cell culture is the polyanionic drug suramin.^{4,7}

Biginelli condensation is a well-known reaction that is more than 120 years old. It is an exceptionally simple and straightforward acid-catalyzed one-pot cyclocondensation reaction of ethylacetoacetate (1), aldehyde (2) and urea (3) that affords 4-substituted 3,4-dihydropyrimidine-2-ones (DHPM) or Biginelli compounds.⁸ A number of variations to give higher yields and expand the scope versus the traditional three-component Biginelli condensation, resulted in many simple derivatives that emerged as promising therapeutic agents. Monastrol was discovered as a prototype anti-cancer drug that inhibits the mitotic kinesin-5 protein.⁹ Methyl 4-(2-chloro-4-fluorophenyl)-6-methyl-2-(pyridin-2-yl)-1,4-dihydropyrimidine-5-carboxylate (HAP-1) and its derivatives were developed as anti-hepatitis B virus (HBV) agents targeting viral capsid assembly.¹⁰ Moreover, marine alkaloids Crambescidins



and Crambescins containing an DHPM core and isolated from the sponges *Crambe crambe* and *Batzella* sp. were demonstrated to possess antiviral and cytotoxic properties.¹¹ Batzelladines A and B with DHPM core were identified as inhibitors of HIV gp-120 binding to CD4 receptors (Fig. 1).¹² However, the majority of studies exclude or limit the aliphatic aldehydes within their scope due to their troublesome behavior and unacceptable yield in this multi-component reaction.^{13,14} It is still desirable to optimize this reaction while employing aliphatic aldehydes.

The present work was aimed to synthesize 4-substituted 3,4-dihydropyrimidin-2(1H)-ones using various aromatic and aliphatic aldehydes and evaluate their antiviral activity against a broad range of DNA and RNA viruses, along with their cytotoxicity assessment in diverse mammalian cell lines.

The synthesis of desired compounds is depicted in Scheme 1. 3,4-dihydropyrimidine-2-one bearing C-4 aryl (**4a**–**g**; **4i**) and styryl (**4h**) moiety were prepared in excellent yields (83–95%) via neat condition¹⁵ without solvent and catalyst at ~98 °C. Whereas, reaction (Scheme 2) employing salicylic aldehyde (**2a**) in neat conditions, a tricyclic compound i.e. oxygen bridged pyrimidinone (**5**) was obtained in good yield (76%). However, utilizing aliphatic aldehydes in neat conditions lead to negligible yield of the desired compounds (data not shown). C-4 unsubstituted DHPM (**4j**) and Dihydropyrimidine-2-one derivatives with C-4 aliphatic group (**4k**–**4m**) were achieved by using excess urea under reflux conditions employing acetic acid as a mild acid catalyst in ethanol. The yields ranged from 53 to 68%. All the synthesized compounds were characterized by IR, ¹H NMR, ¹³C NMR and HR-MS. (Supplementary data)

Antiviral activity and cytotoxicity were determined by previously described methods.^{16–18} The compounds plus viruses were added to 96-well plates containing subconfluent cell cultures. After 3–5 days incubation, the virus-induced cytopathic effect (CPE) was scored which, depending on the virus, consisted of cell lysis, plaque formation, or giant cell formation. Scores of 1, 2, 3 or 4 were given to conditions in which the virus affected 25, 50, 75 or 100% of the

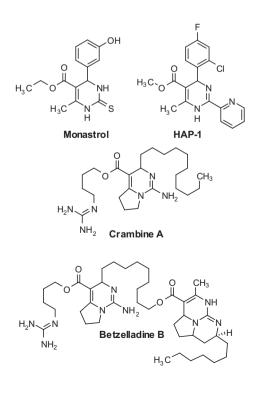
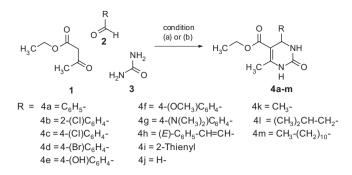
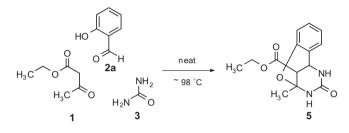


Fig. 1. Biologically active DHPMs.



Scheme 1. Synthesis of DHPMs (a) Compounds **4a–i**, neat reaction condition at ~98 °C; (b) Compounds **4j–m** CH₃COOH, ethanol, reflux.



Scheme 2. Synthesis of oxygen bridged pyrimidinone (5).

cell monolayer, respectively. The virus panel included: (i) evaluated in African green monkey kidney Vero cells: Parainfluenza-3 virus; Reovirus-1; Sindbis virus; Coxsackie virus B4; Punta Toro virus; Rift Valley fever virus (MP-12 strain); and Yellow fever virus; (ii) evaluated in human embryonic lung (HEL) fibroblast cells: herpes simplex virus types 1 and 2; varicella-zoster virus; cytomegalovirus; vaccinia virus; and adenovirus; (iii) evaluated in human cervix carcinoma HeLa cells: vesicular stomatitis virus; Coxsackie virus B4; and respiratory syncytial virus; (iv) evaluated in Crandell-Rees feline kidney (CRFK) cells: feline herpesvirus and feline coronavirus; (v) evaluated in Madin-Darby canine kidney (MDCK) cells: influenza A and B virus; (vi) evaluated in human MT-4 T-lymphoblast cells: HIV types 1 and 2. The antiviral activity was assessed based on CPE reduction, as determined by microscopic scoring of the CPE or virus plaque formation. For HIV-1 and -2, the CPE was measured with the colorimetric formazan-based MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2Htetrazolium] assay.

Cytotoxicity of the compounds was assessed by microscopic examination, yielding the minimal cytotoxic concentration (MCC). Alternatively, the colorimetric formazan-based MTS assay was used to determine the 50% cytotoxic concentration (CC_{50}) value.

The antiviral activities and cytotoxicity of the synthesized compounds (**4a**–**m** and **5**) were determined in CPE reduction assays with a broad and diverse panel of DNA and RNA viruses and using relevant mammalian cell lines. The results of the evaluation in Vero cell cultures are presented in Table 1. Compound **4m** was found to be a selective and potent inhibitor of Punta Toro virus (PTV), a member of the family *Bunyaviridae* and the genus *Phlebovirus*. **4m** was approximately 7 times more potent than the viral entry inhibitor dextran sulphate (DS-10000) and broad-spectrum acting mycophenolic acid (an inhibitor of the cellular IMP dehydrogenase). Compared to the broad antiviral agent ribavirin, **4m** proved 17 times more potent with superior activity (EC₅₀ value of 3 μ M) and a selectivity index (ratio of 50% cytotoxic concentration to 50% effective antiviral concentration) of \geq 33. The anti-PTV effect of **4m** was dose-dependent achieving 81% protection against

Table 1
Antiviral activity and cytotoxicity in Vero cell cultures

Compound	Cytotoxicity MCC $(\mu M)^a$	Antiviral EC ₅₀ $(\mu M)^{\rm b}$							
		Para-influenza-3 virus	Reovirus-1	Sindbis virus	Coxsackie virus B4	Punta Toro virus	Yellow fever virus		
4a	>100	>100	>100	>100	>100	>100	>100		
4b	>100	>100	>100	>100	>100	>100	>100		
4c	>100	>100	>100	>100	>100	>100	>100		
4d	>100	>100	>100	>100	>100	>100	>100		
4e	>100	>100	>100	>100	>100	>100	>100		
4f	>100	>100	>100	>100	>100	>100	>100		
4g	100	>100	>100	>100	>100	>100	>100		
4h	100	>100	>100	>100	>100	>100	>100		
4i	>100	>100	>100	>100	>100	>100	>100		
4j	>100	>100	>100	>100	>100	>100	>100		
4k	>100	>100	>100	>100	>100	>100	>100		
41	>100	>100	>100	>100	>100	>100	>100		
4m	≥100	>100	>100	>100	>100	3.0 ± 1.4^{e}	>100		
5	>100	>100	>100	>100	>100	>100	>100		
DS-10,000 ^c	>100	>100	>100	20	58	20	0.8		
Ribavirin	>250	112	>250	>250	>250	50	112		
MPA ^d	>100	0.8	1.8	4	>250	20	4		

^a MCC: minimal cytotoxic concentration, or compound concentration producing minimal alterations in cell morphology, as detected by microscopical inspection.

^b EC₅₀: 50% effective concentration, or compound concentration giving 50% protection against virus-induced cytopathic effect, as determined by microscopy.

 $^{\rm c}$ DS-10,000: dextran sulphate of MW 10,000; these data are expressed in μg per ml.

^d MPA: mycophenolic acid.

^e Mean ± SD of two independent tests.

Cytotoxicity in diverse mammalian cell lines.

Compound	Cytotoxicity									
	MCC (µM) ^a	$CC_{50} (\mu M)^{b}$								
	Vero	HeLa	MDCK	HEL	CRFK	MT-4				
4a	>100	>100	>100	>100	>100	>125				
4b	>100	>100	>100	>100	>100	96				
4c	>100	>100	>100	>100	>100	76				
4d	>100	>100	>100	>100	>100	64				
4e	>100	>100	>100	>100	>100	>125				
4f	>100	>100	>100	>100	>100	>125				
4g	100	>100	>100	>100	>100	>125				
4h	100	≥100	>100	>100	>100	>125				
4i	>100	>100	>100	>100	>100	>125				
4j	>100	>100	>100	>100	>100	>125				
4k	>100	>100	>100	>100	>100	>125				
41	>100	>100	>100	>100	>100	>125				
4m	≥100	>100	>100	100	>100	50				
5	>100	>100	>100	>100	>100	>125				

The following cell lines were used: Vero, African green monkey kidney cells; HeLa, cervix carcinoma cells; MDCK, Madin-Darby canine kidney cells; HEL, human embryonic lung fibroblast cells; CRFK, Crandell-Rees feline kidney cells; MT-4, human T-lymphoblast cells.

^a MCC: minimal cytotoxic concentration, or compound concentration producing minimal alterations in cell morphology, as detected by microscopical inspection.

^b CC₅₀: 50% cytotoxic concentration, determined by the colorimetric and formazan-based MTS cell viability test.

virus-induced CPE at 20 μ M of compound. Since PTV is related to the most clinically relevant Phlebovirus RVFV,¹⁹ we next evaluated the effect of **4m** against RVFV. This virus is classified as a BSL-4 virus,^{20,21} and we therefore used the low-pathogenic MP-12 strain. Although about 40% reduction in RVFV-induced CPE was seen at a compound concentration of 100 μ M, **4m** was unable to achieve 50% inhibition at this high concentration (data not shown). Also, when evaluated against PTV in HeLa cells, **4m** was found to be inactive, which contrasts to the marked effect seen in Vero cells (see above). Hence, the antiviral effect of **4m** appears to be cell-dependent, alike what was described for the anti-Ebola virus activity of brincidofovir, an antiviral drug with a lipophilic moiety that is structurally similar to that of **4m**.²²

Among the DHPM series that we synthesized, the distinguished structural feature of **4m** is its very long lipophilic chain at position C-4 of the polar dihydropyrimidinone scaffold, leading us to assume that the observed anti-PTV activity mainly depends on

the undecyl moiety which confers amphiphilic properties to **4m**. Some amphiphilic molecules with a large polar head group (inverted-cone-shaped) are known to target viral membranes and display broad-spectrum antiviral activity.²³ However, except for Punta Toro virus, **4m** did not have antiviral activity against all other viruses tested. This molecule displayed only modest cytotoxicity (Table 2), with a minimal cytotoxic concentration (MCC) of 100 μ M in Vero and human embryonic lung (HEL) cells, and no toxicity at 100 μ M compound concentration in the other cell lines tested. Human MT4 T-lymphoblast cells were an exception with a 50% cytotoxic concentration (CC₅₀) of 50 μ M. Of the other DHPM compounds tested, none had noticeable antiviral activity. Compound **4h** had marginal activity against varicella zoster virus (EC₅₀ of 57 μ M and MCC of 100 μ M). Compounds **4b**, **4c** and **4d** displayed no antiviral activity yet some cytotoxicity in MT-4 cells.

In summary, an efficient reaction condition was developed for utilizing aliphatic aldehydes in biginelli condensation. Ethyl 6-methyl-2-oxo-4-undecyl-1,2,3,4-tetrahydro-5-pyrimidinecarboxylate (4m) displayed selective and potent antiviral activity against Punta Toro virus. Based on these first results, further evaluation of **4m** against other Bunyaviruses (such as Crimean-Congo haemorrhagic fever virus) is warranted. Also, mechanistic studies are underway to understand the biochemical basis for its antiviral effect against Punta Toro virus.

Acknowledgements

The authors acknowledge excellent technical assistance from Leentje Persoons, Frieda De Meyer, Lies Van den Heurck, Kristien Erven and Jasper Rymenants. We thank Richard Elliott (University of St Andrews, UK) for providing the RVFV MP-12 strain. This work was supported by the Flemish Fonds voor Wetenschappelijk Onderzoek (FWO) via the Infect-ERA EScential network.

A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.bmcl.2016.12. 010. These data include MOL files and InChiKeys of the most important compounds described in this article.

References

- 1. Jones KE, Patel NG, Levy MA, et al. Nature. 2008;451:990-993.
- 2. Zappa A, Amendola A, Romanò L, Zanetti A. Blood Transfusion. 2009;7:167–171.
- Elliott RM, Brennan B. Curr Opin Virol. 2014;5:50-57. 3.
- Jiao L, Ouyang S, Liang M, et al. J Virol. 2013;87:6829-6839. 4
- Mendenhall M, Wong M-H, Skirpstunas R, Morrey JD, Gowen BB. Virology. 5. 2009;395:143-151.
- 6. Benedict A. Bansal N. Senina S. et al. Front Microbiol. 2015:6:676.
- Ellenbecker M, Lanchy J-M, Lodmell JS. Antimicrob Agents Chemother. 7. 2014;58:7405-7415.
- 8. Oliver Kappe C. Tetrahedron, 1993;49:6937–6963.
- Hague SA, Hasaka TP, Brooks AD, Lobanov PV, Baas PW. Cell Motil Cytoskeleton. 9. 2004.58.10-16
- 10. Zhu X, Zhao G, Zhou X, et al. Bioorg Med Chem Lett. 2010;20:299-301.
- 11. Blunt JW, Munro MHG, eds. Dictionary of Marine Natural Products with CD-ROM. USA: Chapman & Hall/CRC, Taylor & Francis Group; 2008.
- 12. Patil AD, Kumar NV, Kokke WC, et al. J Org Chem. 1995;60:1182–1188.
- 13. Kappe CO. Eur J Med Chem. 2000;35:1043-1052. 14. Kappe CO. Acc Chem Res. 2000;33:879-888.
- 15. Ranu BC, Hajra A, Dey SS. Org Process Res Dev. 2002;6:817-818.
- 16. Naesens L, Vanderlinden E, Rőth E, et al. Antiviral Res. 2009;82:89-94.
- 17. Vanderlinden E, Göktas F, Cesur Z, et al. J Virol. 2010;84:4277–4288.
- 18. Pannecouque C, Daelemans D, De Clercq E. Nat Protoc. 2008;3:427-434.
- 19. Stefan CP, Chase K, Coyne S, Kulesh DA, Minogue TD, Koehler JW. Virol J. 2016:13:54.
- 20. Rolin AI, Berrang-Ford L, Kulkarni MA. Emerg Microbes Infect. 2013;2:e81.
- 21. Dar O, McIntyre S, Hogarth S, Heymann D. Emerg Infect Dis. 2013;19:189–193.
- 22. McMullan LK, Flint M, Dyall J, et al. Antiviral Res. 2016;125:71-78.
- 23. Vigant F, Santos NC, Lee B. Nat Rev Microbiol. 2015;13:426-437.