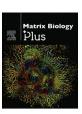
ELSEVIER

Contents lists available at ScienceDirect

Matrix Biology Plus

journal homepage: www.sciencedirect.com/journal/matrix-biology-plus



<u>S</u>mall <u>I</u>ntegrin <u>b</u>inding <u>Ligand N</u>-linked <u>G</u>lycoproteins, prostate-specific antigen and time to prostate cancer diagnosis

Alka Jain ^{a,b}, Ying Ni ^c, Daisy Zhang ^c, Eleanor M. Simonsick ^d, E. Jeffrey Metter ^{d,e}, Kalu U. Ogbureke ^f, Larry W. Fisher ^g, Neal S. Fedarko ^{a,*} ^o

- ^a Department of Medicine, Johns Hopkins University, Baltimore, MD 21224, USA
- ^b Staff Scientist, ThermoFisher Scientific, Middletown, VA 22645, USA
- c Research Laboratory Core, Institute for Clinical and Translational Research, Johns Hopkins University, Baltimore, MD 21224, USA
- ^d Intramural Research Program, National Institute on Aging, National Institutes of Health, Baltimore, MD 21225, USA
- e Department of Neurology, University of Tennessee Health Science Center Memphis, TN 38163, USA
- f Department of Diagnostic & Biomedical Sciences, School of Dentistry, University of Texas Health Science Center at Houston, Houston, TX 77054, USA
- 8 Matrix Biochemistry Section, National Institute of Dental and Craniofacial Research, National Institutes of Health, Bethesda, MD 20892, USA

ARTICLE INFO

Keywords: Osteopontin Bone sialoprotein Dentin sialophosphoprotein Prostate-specific antigen Preclinical Prostate cancer

ABSTRACT

Background: Small Integrin Binding Ligand M-linked Glycoproteins (SIBLINGs1) were associated with cancer in cross-sectional studies. Whether SIBLINGs associate with preclinical disease is unknown. Methods: A retrospective longitudinal case control study was performed to determine the association of SIBLINGs and prostatespecific antigen (PSA) with preclinical disease. Paired serum samples from 109 cancer-free Baltimore Longitudinal Study on Aging participants were divided into those that were either most distal or proximal to diagnosis (cases) or censored (controls). Dentin sialophosphoprotein (DSPP), bone sialoprotein (BSP), osteopontin (OPN), and PSA were measured by immunoassay and dichotomized into low or high based on their respective cut-off values. Associations of time to diagnosis or death, modeled as disease-free survival (DFS) or overall survival (OS), were assessed using Kaplan Meier and Cox proportional hazard survival estimates on individual and aggregated biomarkers in distal or proximal sets separately. Models were adjusted for relevant covariates. A false discovery rate analysis assessed significance of hazard ratios (HRs) in sets. Results: Biomarkers/aggregates identified as true discoveries for DFS included DSPP + PSA, OPN + PSA, DSPP + BSP + PSA, DSPP + OPN + PSA, where unadjusted distal HRs ranged between 11 and 27 and after adjusting for age from 7 to 15, while proximal HRs ranged between 6 and 10 unadjusted and 5 to 12 after adjusting for age. For proximal OS, true discoveries included DSPP + BSP, DSPP + OPN, DSPP + BSP + OPN, and DSPP + OPN + PSA where unadjusted HRs ranged between 6 and 20 while age-adjusted HRs ranged between 5 and 12. Conclusions: These observations support SIBLINGs as biomarkers that associate with DFS and OS in prediagnosis samples.

Based on 2018 to 2022 data, at some point during their lifetime 12.8% of men will be diagnosed with prostate cancer and 2.4% will die from the cancer [1]. Prostate cancer is a heterogeneous disease varying in presentation from localized indolent to a rapidly progressing lethal metastatic disease [2]. Disease progression and metastases are the cause of 90% of human cancer deaths [3]. An underlying assumption to prostate cancer clinical research has been that early detection and treatment will lead to improved quality of life and extended survival. An alternative approach to developing novel biomarkers of symptomatic

cancer is to identify biomarkers of pre-symptomatic disease that can be used to assess risk status for disease development and subsequent progression. Identifying asymptomatic individuals at risk for disease diagnosis and progression would facilitate the targeting of surveillance and/or treatment to those who would benefit the most.

Members of the Small Integrin Binding Ligand N-linked Glycoprotein (SIBLING¹) gene family are intrinsically disordered, secreted glycoproteins that have multiple binding partners, including immunoregulatory proteins (complement factor H, inducible T-cell costimulatory ligand),

E-mail address: nfedark1@jhmi.edu (N.S. Fedarko).

https://doi.org/10.1016/j.mbplus.2025.100171

 $^{^{\}ast}$ Corresponding author.

¹ Abbreviations: SIBLINGs, Small Integrin Binding Ligand N-linked Glycoproteins; DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; Baltimore Longitudinal Study of Aging (BLSA); DFS, disease-free survival; OS, overall survival; FDR, false discovery rate.

 $\begin{tabular}{ll} \textbf{Table 1}\\ \textbf{Characteristics, main predictors and outcomes for pre-diagnosis longitudinal samples.} \end{tabular}$

Model Covariates	Longitudinal Samples Pre diagnosis				
	Distal (n =	Proximal (n =	P value		
	109)	109)			
Age at visit, mean (SD), y	69 (10)	81 (8)			
median (range)	71 (41–86)	82 (53-95)	< 0.0001		
Race, % Caucasian	97	97			
BMI, mean (SD), kg/m ²	24.9 (2.4)	25.0 (3.0)	n.s.		
18 < BMI < 25, number (%)	55 (50)	57 (52)			
25 > BMI < 30, number (%)	51 (47)	46 (42)			
BMI > 30, number (%)	3 (3)	6 (6)			
Systolic BP, mean (SD), mm	138 (25)	145 (24)	≤0.005		
Hg					
Diastolic BP, mean (SD), mm	82 (12)	78 (12)	n.s.		
Hg					
Hypertension, number (%)	75 (69)	84 (77)			
Predictors					
DSPP, mean (SD), ng/ml	175 (94)	223 (129)			
median (range)	162 (33-569)	209 (24-802)	< 0.0001		
BSP, mean (SD), ng/ml	101 (52.0)	142 (64.9)			
median (range)	91.9	153 (16.5-267)	< 0.0001		
	(16.8-216)				
OPN, mean (SD), ng/ml	15.8 (23.6)	30.7 (58.0)			
median (range)	3.08 (0.1-106)	113.9 (0.10-434)	< 0.005		
PSA, mean (SD), ng/ml	2.4 (2.0)	11.3 (38.5)			
median (range)	2.0 (0.2-10.8)	3.4 (0.4-352)	< 0.0001		
Outcomes					
PCA diagnosis %	81 (74 %)	81 (74 %)			
Deaths %	109 (100 %)	109 (100 %)			
Time to diagnosis, mean SD, y	15 ± 6	4 ± 4			
median (range)	16 (1-28)	3 (1-20)	< 0.0001		
Time to death, mean SD, y	18 ± 8	7 ± 5			
median (range)	20 (4–40)	6 (1–31)	< 0.0001		

Wilcoxon matched-pairs signed rank test.

Abbreviations: SD, standard deviation; y, years; BMI, body mass index; BP, blood pressure; DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate specific antigen. PCA, prostate cancer; n.s., not significant; y, years.

Hypertension defined as systolic BP \geq 130 or diastolic BP \geq 80 mm Hg.

receptors (integrins and CD44 variants) as well as specific proteases [4–6]. Normally restricted in expression to skeletal tissue and metabolically active ductal epithelial cells, SIBLINGs are also upregulated in certain cancers [5,7,8]. The association of SIBLING levels with established prostate cancer has been demonstrated using tissue arrays, immunohistochemistry, in situ hybridization of biopsies and ELISAs targeting individual SIBLINGs in serum [9–11]. The above observations were made in cross-sectional studies. The current study was undertaken to determine the association of time to prostate cancer diagnosis with serum SIBLING and PSA levels in longitudinal pre-diagnosis samples. A secondary outcome was to test for association between these biomarkers and overall survival.

Methods

Human subjects. Sets of serum samples were obtained under local approved protocols (Johns Hopkins Medicine Institutional Review Board). A longitudinal set of 218 pre-diagnosis serum samples from 109 men was obtained from the Baltimore Longitudinal Study of Aging (BLSA). The BLSA cohort study, run by the National Institute on Aging, has been previously used to assess the association of PSA velocity with prostate cancer aggressiveness [12]. Participants were disease-free on enrollment. Those under age 60 were assessed every 4 years, between ages 60–79 years were assessed every 2 years and aged 80 and older assessed annually. Assessments included comprehensive health, cognitive, and functional evaluations. Clinical measures associated with each BLSA sample include age, BMI, blood pressure, and PSA levels while for subjects who later developed prostate cancer, additional clinical measures included diagnosis date and disease stage. Ten samples had

missing PSA values and clinical staging was incomplete for 25 subjects. All subjects in this set are currently deceased with known date of death. Hypertension was defined as systolic BP ≥ 130 or diastolic BP ≥ 80 mm Hg.

Immunoassays. Serum samples were stored at -80C until a controlled thaw and analysis by ELISAs. SIBLINGs possess an acidic pI, readily dissolve in aqueous solutions and they are intrinsically disordered. These physical properties and the buffering capacity of serum likely contribute to their ability to undergo at least 6 freeze/thaw cycles with little or no loss of signal. Analytical assays were performed without knowledge of the patients' clinical data or demographics. PSA levels for men were originally measured as part of the BLSA assessment using the Hybritech Tandem-R total PSA immunoradiometric assay (Beckman Coulter) and the clinically validated cut-off of 4.0 ng/ml was used for evaluation [13]. The ten samples with missing PSA values and 10 randomly selected samples were analyzed using the chemiluminescent Access Hybritech total PSA assay that uses the same monoclonal antibodies employed in the Hybritech Tandem total PSA assay and yields PSA concentrations based on calibration to the original Hybritech Tandem-R assay. The average coefficient of variance (%CV) between the original and rerun PSA values was 11 \pm 5 %.

SIBLING levels were quantified using SIBLING ELISAs that had been used to determine biomarker cut-off values of 190 ng/ml for DSPP, 120 ng/ml for BSP and 11 ng/ml for OPN in 310 community-dwelling older adult males (see Supplemental Data: Methods). An underlying assumption to the current study's design was that this training set contains subjects who, at the time of sample collection, are likely to be preclinical and will be diagnosed in their lifetime. The 218 longitudinal samples are from older adult males who also have no symptoms of prostate cancer and contains some who will not develop prostate cancer and others who will be diagnosed with the disease at some future date. The analytical plan was to use SIBLING median values determined in the training group as the cut point for dichotomizing the biomarker data in the longitudinal study. The primary goal of this study is to address whether there is an association between SIBLING levels in preclinical samples and time to diagnosis. The association of dichotomized SIBLING levels and time to event (diagnosis, death) was assessed by Kaplan-Meier and Cox Proportional Hazard survival estimates.

Statistical analysis and scientific rigor. In modeling the relationship between biomarker and time to diagnosis as disease-free survival, censoring occurred for those subjects who were not diagnosed with prostate cancer. In modeling biomarker and time to death as overall survival, no censoring occurred as all subjects are deceased. Paired longitudinal serum samples from each BLSA participant were divided into those that were most distal or proximal to diagnosis (cases) or were censored (controls). No assumptions were made about the distribution of the data and non-parametric tests were employed. Comparisons between cases (those who eventually will be diagnosed with prostate cancer) and controls (those who will not be diagnosed with prostate cancer in their lifetime) used the Mann Whitney U test. Comparisons of measures between subjects' most proximal and distal samples were analyzed by Wilcoxon matched pairs signed rank test. Potential correlations between continuous measures (DSPP, BSP, OPN, PSA, age, BMI, time to events) were analyzed by Spearman correlation. Associations between dichotomized biomarkers and categorical measures (e.g., hypertension) were assessed by Goodman & Kruskal's lambda. The association of biomarkers with survival curves, median disease-free and overall survival times were modeled nonparametrically by Kaplan-Meier survival estimates. Biomarker levels were dichotomized into either low or high levels based on their respective cut-off values and separately analyzing samples grouped as most distal or proximal to diagnosis or censoring. Pairs of biomarkers were analyzed by stratifying samples into three groups: those with low levels of both biomarkers, high levels of one biomarker, and high levels of both biomarkers. Similarly, three biomarkers were stratified into four groups: those with all low biomarker levels, one high biomarker, two high biomarkers, and all

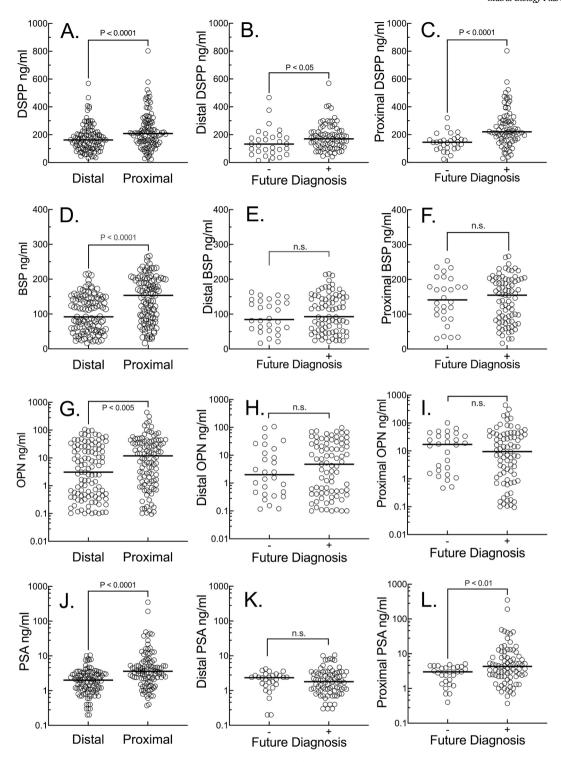


Fig. 1. SIBLING and PSA levels by time point and by future diagnosis. Longitudinal levels of SIBLINGs and PSA in serum from 109 donors were segregated into groups that were either most distal to or most proximal to time of diagnosis. Those groups were further stratified into subgroups that did not receive a diagnosis of prostate cancer (–) and those that did (+). Subgroups were compared for DSPP (A, B, C), BSP (D, E F), OPN, (G, H, I) and PSA (J, K, L). Comparisons were between most distal and proximal times to diagnosis/censoring (A, D, G, J); future diagnosis at the most distal times to diagnosis/censoring (B, E, H, K) and future diagnosis at the most proximal times to diagnosis/censoring (C, F, I, L). Abbreviations: DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; n.s., not significant.

three biomarkers high. A log-rank test was used to assess the p value for individual biomarkers, while a log-rank test for trend in the survival analysis was used to assess whether a linear trend was apparent between three or more survival curves. Hazard ratios were determined by Cox proportional hazard regression modeling using individual, pairs or trios of biomarkers dichotomized into low or high based on each biomarker's

respective cut-off value and analyzing distal or proximal sets separately. There were four sets that were analyzed: distal disease-free survival, distal overall survival, proximal disease-free survival, and proximal overall survival. Comparisons of hazard ratios were performed within each set where there was a total of 14 different biomarker aggregates (4 individual biomarkers, 6 pairs and 4 trios of biomarkers). To assess

Table 2Biomarker associations by Spearman correlation.

		DSPP			BSP			OPN			PSA	
Distal	r _s	95 % CI	P value	rs	95 % CI	P value	r _s	95 % CI	P value	rs	95 % CI	P value
Time to diagnosis	-0.28	$(-0.45 \text{ to} \\ -0.10)$	<0.005	0.04	(-0.16 to 0.23)	n.s.	-0.43	$(-0.58 \text{ to} \\ -0.26)$	<0.0001	-0.24	$(-0.42 \text{ to} \\ -0.05)$	<0.05
Time to death	-0.25	(-0.42 to -0.06)	< 0.01	0.05	(-0.14 to 0.24)	n.s.	-0.44	(-0.58 to -0.26)	< 0.0001	-0.30	(-0.46 to -0.11)	< 0.005
Age	0.27	(0.08 to 0.44)	0.0005	0.02	(-0.18 to 0.21)	n.s.	0.40	(0.23 to 0.55)	< 0.0001	0.42	(0.25 to 0.57)	<0.0001
BMI	0.10	(-0.10 to 0.28)	n.s.	0.02	(-0.18 to 0.21)	n.s.	0.01	(-0.19 to 0.19)	n.s.	-0.10	(-0.29 to 0.09)	n.s.
Hypertension	-0.02	(-0.21 to 0.17)	n.s.	0.20	(0.01 to 0.38)	< 0.05	0.16	(-0.04 to 0.34)	n.s.	-0.03	(-0.23 to 0.16)	n.s.
Proximal												
Time to diagnosis	-0.34	(-0.50 to -0.16)	<0.0005	-0.27	(-0.44 to -0.08)	<0.005	0.10	(-0.09 to 0.29)	n.s.	-0.38	$(-0.53 \text{ to} \\ -0.20)$	< 0.0001
Time to death	-0.20	(-0.38 to -0.01)	< 0.05	-0.24	(-0.41 to -0.04)	< 0.05	-0.36	$(-0.52 \text{ to} \\ -0.18)$	<0.0005	0.08	(-0.11 to 0.27)	n.s.
Age	0.25	(0.05 to 0.42	< 0.01	0.07	(-0.12 to 0.26)	n.s.	0.34	(0.15 to 0.50)	< 0.0005	0.05	(-0.15 to 0.24)	n.s.
BMI	-0.03	(-0.22 to 0.17)	n.s.	0.01	(-0.19 to 0.19)	n.s.	-0.01	(-0.20 to 0.18)	n.s.	-0.01	(-0.21 to 0.18)	n.s.
Hypertension	-0.07	(-0.26 to 0.12)	n.s.	-0.15	(-0.33 to 0.05)	n.s.	0.01	(-0.18 to 0.21)	n.s.	-0.08	(-0.27 to 0.11)	n.s.

Abbreviations: DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate specific antigen;

hazard ratios within a set, a false discovery rate (FDR) analysis using the two-step method of Benjamini, Krieger and Yekutieli with a desired FDR Q of 1 % was employed [14].

Results

Longitudinal set characteristics

The characteristics, main predictors and outcomes associated with these samples obtained through the BLSA (n=218) from subjects who were disease-free on enrollment and longitudinally followed are given in Table 1. These samples were from a broad age range (41–95 years). Almost all subjects were Caucasian and approximately 50 % had BMIs in the normal range while the percentage that were obese was less than 10 %. Systolic blood pressure was significantly different between the proximal and distal groups when compared by Wilcoxon matched pairs signed rank test. Hypertension was present in 69 % of subjects at the most distal time to diagnosis/censoring and the number increased to 77 % by the most proximal time to diagnosis. 74 % of subjects were eventually diagnosed with prostate cancer and 100 % are deceased. There was an average and median difference of at least 10 years between distal and proximal time to diagnosis and time to death for all subjects.

Biomarker distribution

Serum levels of DSPP, BSP, OPN and PSA exhibited a broad range of values and were significantly different between distal and proximal groups (Table 1, Fig. 1). The median value of DSPP at both the most distal and most proximal times to diagnosis/censoring was significantly higher in subjects who received a future diagnosis of prostate cancer when compared with those who remained disease-free (Fig. 1). BSP and OPN did not exhibit a difference in levels between subjects who received a future diagnosis of prostate cancer and those who remained disease-free at both the most distal and most proximal times to diagnosis/censoring. There was also no difference in median PSA levels between subjects who received a future diagnosis of prostate cancer and those who remained disease-free at the most distal times to diagnosis/censoring, while the difference was significant at the most proximal to diagnosis/censoring.

Biomarker associations

At the most distal time point, DSPP, OPN and PSA were negatively correlated with time to diagnosis, time to death, and positively correlated with age at visit, but not correlated with BMI or hypertension (Table 2). Distal BSP was correlated positively with hypertension. In the corresponding most proximal samples, DSPP associated negatively with time to diagnosis, time to death, and positively with age. Proximal BSP correlated negatively with time to diagnosis, time to death but not with age. Proximal OPN correlated negatively with time to death and positively with age. Proximal PSA was negatively correlated with only time to diagnosis (Table 2). Proximal levels of DSPP, BSP and OPN were not correlated with BMI or hypertension. DSPP and OPN were positively correlated at distal times to diagnosis/censoring (Spearman $r=0.25,\,95$ % CI 0.06 to 0.43, and P < 0.01). At proximal times to diagnosis/ censoring, BSP and OPN exhibited a Spearman r of 0.20 with a 95 % CI of 0.007 to 0.38, P < 0.05. Proximal BSP and PSA exhibited a Spearman r of 0.19 with a 95 % CI of -0.0002 to 0.37), P < 0.05.

Biomarkers and time to events

Survival curves and time to diagnosis (disease-free survival) and time to death (overall survival) were profiled by Kaplan-Meier estimates of separately analyzed distal and proximal samples segregated into low versus high biomarker levels (Fig. 2). For the samples drawn most distal to diagnosis, high DSPP levels were associated with a 44 % decrease in median time to diagnosis and a 43 % decrease in time to death (Table 3). The probability of prostate cancer diagnosis (disease-free survival) and death (overall survival) was estimated by the Cox proportional hazard regression model. For the distal group, a subject with high DSPP had twice the probability of being diagnosed with prostate cancer by the next time point compared to subjects with low distal DSPP. A subject with high DSPP levels in the distal group had over twice the probability of having died by the next time point compared to a subject with low DSPP (Table 3). For the samples drawn most proximal to diagnosis, high DSPP levels were associated with an 80 % reduction in median diseasefree survival and a 25 % reduction in median overall survival (Table 3). In the proximal group, a subject with high DSPP who had not been diagnosed with prostate cancer at a given time point had over 4 times the probability of being diagnosed by the next time point compared to a

r_s, Spearman correlation coefficient, 95% CI, 95% Confidence Interval; n.s., not significant.

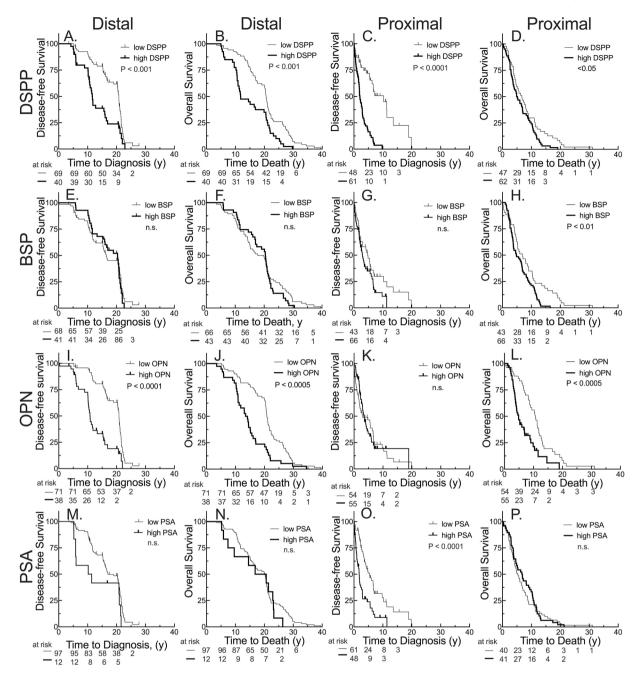


Fig. 2. Survival curves modeling time to diagnosis (disease-free survival) or time to death (overall survival). Longitudinal levels of SIBLINGs or PSA in serum from 109 donors were segregated as either most distal to or most proximal to time of diagnosis. Kaplan-Meier survival estimates were performed comparing high versus low predictor values at the most distal times to diagnosis/censoring (A, B, E, F, I, J, M, N) and most proximal times to diagnosis/censoring (C, D, G, H, K, L, O, P) for DSPP (A, B, C, D), BSP (E, F, G, H), OPN, (I, J, K, L) or PSA (M, N, O, P). The P value reported in each graph is from a Log-rank (Mantel-Cox) test. Abbreviations: DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; n.s., not significant. The number at risk are reported below each graph.

subject in the low DSPP proximal group. High DSPP levels in the proximal group associated with a 3-fold increase in the probability of dying by the next time point.

BSP levels were only modestly associated with time to event at the most proximal times to diagnosis/censoring, where a 37 % decrease in median disease-free and 30 % decrease in overall survival were observed. A subject with high proximal BSP levels had about twice the probability of dying by the next time point. High distal OPN levels were associated with a 2-fold increase in probability of diagnosis and death. There was no significant association between OPN and time to diagnosis at proximal times (Fig. 2, Table 3). However, high proximal OPN levels

did associate with a decrease in disease-free survival and overall survival of 48 % and 34 %, respectively. A subject with a high proximal OPN levels had approximately twice the probability of dying by the next time point compared to a subject in the low OPN group. For the samples drawn most distal to diagnosis, survival curves were not different between low and high PSA levels for disease-free and overall survival (Fig. 2 and Table 3). For the samples drawn most proximal to diagnosis, high PSA levels were associated with a greater than 67 % reduction in median disease-free survival. A subject with elevated PSA levels in the proximal group who had not been diagnosed with prostate cancer at a certain time point had an over 2-fold probability of being diagnosed by

Table 3Survival Estimates for individual biomarkers.

	Distal		Proximal	
	DFS	OS	DFS	OS
DCDD				
DSPP Median				
survival, y				
low DSPP	20.8	20.6	10.3	6.71
high DSPP	11.6	11.8	2.06	5.01
	2.10	2.62	2.06 4.71	3.15
HR, high DSPP				
	(1.33–3.28)	(1.63–4.18)	(2.77–8.40)	(1.90–5.45)
P value	≤0.001	< 0.0001	< 0.0001	< 0.0001
BSP				
Median survival, y				
low BSP	16.5	19.9	4.72	6.97
high BSP	20.4	20.4	2.99	4.88
HR, high BSP	1.06	1.28	1.49	1.95
	(0.66-1.65)	(0.80-2.02)	(0.94-2.42)	(1.20-3.27)
P value	n.s.	n.s.	n.s.	< 0.01
OPN				
Median survival, y				
low OPN	20.8	20.8	2.89	10.6
high OPN	10.9	13.8	3.27	4.38
HR, high	1.91	2.13	0.85	2.27
OPN	(1.22-3.99)	(1.36-3.35)	(0.55-1.33)	(1.41-3.69)
P value	< 0.005	< 0.001	n.s.	< 0.001
PSA				
Median survival, y				
low PSA	17.0	20.1	5.41	5.28
high PSA	10.8	18.7	1.80	6.80
HR, high PSA	1.89	1.8	2.48	1.22
, 0	(0.94-3.46)	(0.89-3.33)	(1.58-3.90)	(0.78-1.90)
P value	n.s.	n.s.	< 0.0001	n.s.

Abbreviations: DFS, disease-free survival; OS, overall survival; DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; n.s., not significant; y, years.

Hazard ratio determined by Cox proportional Hazard modeling of biomarkers dichotomized by their respective cut-off values. Reference is the group with both biomarkers low.

the next time point compared to a subject with low PSA.

SIBLINGs pairs and time to event

Serum levels of paired, dichotomized DSPP and BSP at most distal or proximal times to diagnosis/censoring were segregated into three groups: low levels of both DSPP and BSP, high levels of either DSPP or BSP, or high levels of both DSPP and BSP. Kaplan-Meier estimates of survival between the three groups was performed (Fig. 3, Table 4). A log-rank trend test revealed there was an association with survival. High distal DSPP and BSP associated with a 45 % reduction in median diseasefree survival and a 2-fold increase in probability of a diagnosis event occurring (Table 3). A subject with high proximal DSPP and BSP, compared to the reference group, had an 80 % decrease in disease-free survival, a 39 % decrease in overall survival and a 5-fold increase in the probability of being diagnosed and dying by the next time point. With the pairing of BSP and OPN, there was no difference in median disease-free survival between subjects with high levels relative to subjects with low values (Table 4). Proximal overall survival was reduced 62 % and the probability of dying increased almost 5-fold in subjects with high BSP and OPN.

The pairing of DSPP with OPN yielded a log-rank trend test that was significant for disease-free and overall survival at both distal and proximal times to diagnosis/censoring (Fig. 3). Compared to subjects with low levels of both DSPP and OPN at the most distal time point, median disease-free was reduced 49 % and overall survival 45 % in subjects who had high distal DSPP and OPN (Table 3). Median disease-free survival was reduced 74 % and overall survival by 57 % in subjects with high

proximal DSPP and OPN. A subject with high distal DSPP and OPN, compared to the reference group, had a 3- and 4-fold increase in probability of being diagnosed and dying, respectively, by the next time point. For proximal disease-free survival and overall survival, subjects with high DSPP and OPN had a 4-fold and 9-fold increase in hazard, respectively.

For DSPP paired with PSA, the trend across the three groups (both biomarkers low, one biomarker high, and both biomarkers high) and was significant except for overall survival at the most proximal time point (Fig. 4). Compared to the reference group (subjects with low distal levels of both DSPP and PSA), median disease-free survival was reduced 72 % in subjects who had both high distal DSPP and high PSA (Table 4). Distal median overall survival was reduced 65 % for high DSPP and PSA. Compared to the reference group, proximal median disease-free survival was reduced 86 % in subjects who had both high DSPP and PSA. Using the Cox proportional hazard estimator, a subject with high distal DSPP and PSA had an increase in probability of being diagnosed by the next time point of 20-fold and an increase in probability of dying of 5-fold, compared to the reference group. High proximal DSPP and PSA had a 11-fold increase in probability of being diagnosed by the next time point. Overall survival exhibited a 11-fold and 3-fold increase in probability of dving by the next time point for subjects with high distal and proximal DSPP and PSA, respectively.

When BSP and PSA were combined, the log-rank trend test and survival estimates were only significant at proximal times to diagnosis/ censoring for disease-free survival (Fig. 4). A subject with high BSP and PSA had approximately a 68 % decrease in median disease-free and overall survival, and a 3-fold increase in probability of being diagnosed by the next time point, compared to subjects with low BSP and PSA. Combined OPN and PSA yielded a significant log-rank trend test for both distal and proximal disease-free survival and overall survival. For the group with high distal OPN and PSA, there was a 72 % decrease in median disease-free survival and a 51 % decrease in median overall survival. High proximal OPN and PSA were associated with a 61 % reduction in proximal median disease-free survival, and a 47 % decrease in median overall survival. High OPN and PSA at distal times to diagnosis/censoring was associated with an over 5-fold increase in probability of a prostate cancer diagnosis event occurring by the next time point. For distal overall survival, proximal overall and disease-free survival, a subject with high OPN and PSA had a 3-fold increase in probability of an event occurring by the next time point.

SIBLING and PSA trios and time to events

A composite of DSPP, BSP and OPN was made using dichotomized values and survival assessed for subjects with high levels of one, two or three of the biomarkers compared to the reference group – samples with low levels of all three biomarkers (Fig. 5). The log-rank trend across groups test was significant for disease-free and overall survival at both distal and proximal times to diagnosis/censoring. Subjects with high distal levels of DSPP, BSP and OPN had a 51 % decrease in disease-free survival and a 32 % decrease in overall survival (Table 4). A subject with all three biomarkers high had a 3-fold increase in probability of being diagnosed and 5-fold increase in risk of dying by the next time point, compared to the reference group. Subjects with high proximal DSPP, BSP and OPN had a 79 % decrease in median disease-free survival and a 73 % decrease in overall survival. High proximal DSPP, BSP and OPN was also associated with a 6-fold increase in probability of being diagnosed by the next time point and a 20-fold increase in hazard for overall survival. It should be noted that the confidence interval on this latter estimate is wide (see Table 5). The broad range likely reflects the limited number of subjects in the reference group (only 9 subjects had low levels of DSPP, BSP, and OPN).

For the trio of DSPP + BSP + PSA, log-rank trend test p values were significant at distal and proximal times to diagnosis (Fig. 5). A subject with high distal DSPP, BSP and PSA had a 62 % decrease in median

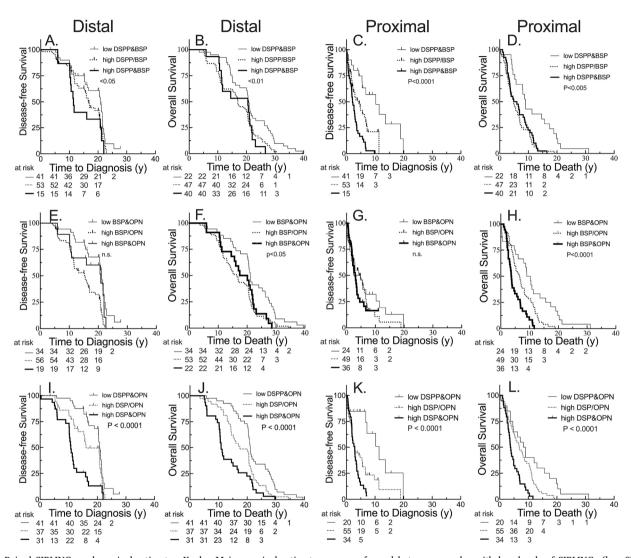


Fig. 3. Paired SIBLINGs and survival estimates. Kaplan-Meier survival estimates were performed between samples with low levels of SIBLINGs (low SIBLING-&SIBLING), high levels of one SIBLING (high SIBLING/SIBLING), or high levels of both SIBLINGs (high SIBLING&SIBLING). The pairing of DSPP with OPN (A, B, C, D), DSPP with BSP (E, F, G, H), and BSP with OPN (I, J, K, L) for disease-free survival (A, C, E, G, I, K) or overall survival (B, D, F, H, J, L) for the groups segregated as most distal to diagnosis (A, B, E, F, I, J) and most proximal to diagnosis (C, D, G, H, K, L). The P value reported for each pairing/group is from a log-rank trend test for three or more survival curves. Abbreviations: DSPP dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin. The number at risk are reported below each graph.

Table 4 Survival Estimates for biomarker pairs.

	Distal DFS	Distal OS	Proximal DFS	Proximal OS
DSPP + BSP				
Median				
survival (y)				
low	20.8	20.8	10.3	9.00
DSPP&BSP	16.6	17.0	4.14	4.84
high DSPP/ BSP	10.0	17.0	4.14	4.04
high	11.4	20.5	2.09	5.44
DSPP&BSP				
HR, high	2.01	2.88	5.22	5.39
DSPP&BSP	(1.00-3.84)	(1.39-5.71)	(2.61-11.5)	(2.66-12.0)
P value	< 0.05	n.s.	< 0.0001	< 0.0001
BSP + OPN				
Median				
survival (y) low BSP&OPN	21.2	21.1	4.72	9.66
high BSP/OPN	15.3	16.5	4.32	6.49
high	20.9	21.6	2.99	3.66
BSP&OPN				
HR, high	1.68	2.26	1.41	4.74
BSP&OPN	(0.86-3.24)	(1.14-4.44)	(0.76-2.69)	(2.35-10.0)
P value	n.s.	< 0.05	n.s.	< 0.0001
DSPP + OPN				
Median				
survival (y) low	20.8	21.1	10.3	8.83
DSPP&OPN	20.6	21.1	10.3	0.03
high DSPP/	16.06	16.5	2.89	6.49
OPN				
high	10.7	11.5	2.62	3.78
DSPP&OPN				
HR, high	3.30	3.77	3.85	8.99
DSPP&OPN	(1.85-5.83)	(2.10-6.75)	(1.95-8.05)	(4.24-20.7)
P value	< 0.0001	< 0.0001	< 0.0005	< 0.0001
DSPP + PSA Median				
survival,				
years				
low	20.6	27.3	11.4	5.69
DSPP&PSA				
high DSPP/	12.8	16.6	3.60	5.99
PSA				
high	5.86	9.60	1.61	6.09
DSPP&PSA	20.8	E 20	11 /	2 17 (1 66 +0
HR, high DSPP&PSA	(6.39–58.2)	5.30 (1.79–12.7)	11.4 (5.52–25.4)	3.17 (1.66 to 6.40)
P value	< 0.0001	< 0.001	< 0.0001	<0.001.
BSP + PSA	(0.0001	(0.001	(0.0001	(0.001.
Median				
survival (y)				
low BSP&PSA	16.7	19.9	5.41	6.97
high BSP/PSA	18.9	20.4	3.60	4.70
high BSP&PSA	20.9	21.4	1.75	6.80
HR, high	1.36	1.44	3.24	2.01
BSP&PSA P value	(0.40–3.41)	(0.43–3.66) n.s.	(1.77–6.15)	(1.10–3.77)
OPN + PSA	n.s.	11.5.	<0.0005	n.s.
Median				
survival (y)				
low OPN&PSA	20.5	20.8	6.64	7.30
high OPN/PSA	15.3	15.8	3.27	6.72
high	5.99	14.5	1.89	3.90
OPN&PSA				
HR, high	5.18	3.25	2.57	3.41
OPN&PSA	(1.74–12.5)	(1.09–7.88)	(1.30–5.10)	(1.70–6.88)
P value	< 0.001	< 0.05	< 0.01	< 0.005

Abbreviations: DFS, disease-free survival; OS, overall survival; DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; HR, hazard ratio; n.s., not significant.

Groups analyzed are: low levels of both biomarkers (e.g., low DSPP&BSP), high levels of one of the biomarkers (e.g., high DSPP/BSP), and high levels of both biomarkers (e.g., high DSPP&BSP).

Hazard ratio determined by Cox proportional Hazard modeling of biomarkers

dichotomized by their respective cut-off values. Reference is the both biomarkers low group.

disease-free survival and a 11-fold increase in hazard. An 84 % reduction in median disease-free survival and 10-fold increase in probability of diagnosis was observed with all three biomarkers high at proximal times to diagnosis/censoring. There was a 4-fold increase in probability of dying by the next time point for subjects with high proximal DSPP, BSP and PSA (Table 5).

The combination of DSPP, OPN and PSA yielded a positive log-rank trend test at distal and proximal time to events (Fig. 5). A subject with high distal DSPP, OPN and PSA had a 72 % decrease in median diseasefree survival and a 16-fold increase in hazard (Table 5). An 84 % reduction in median disease-free survival and 12-fold increase in probability of diagnosis was observed with all three biomarkers high at proximal times to diagnosis/censoring. Median overall survival was decreased 72 % and 56 % at distal and proximal times where there was a 4- and 10-fold increase in probability of dying by the next distal or proximal time point respectively, compared to subjects with low DSPP, OPN and PSA. A subject with high distal BSP, OPN and PSA had a 62 % decrease in median disease-free survival and 17-fold increase in hazard. An 68 % reduction in median disease-free survival and 3-fold increase in probability of diagnosis was observed with all three biomarkers high at proximal times to diagnosis/censoring. Median overall survival and hazard ratios were not significant at distal time points. There was a 57 %reduction in proximal median survival and a 5-fold increase in probability of dying by the next distal time point, compared to subjects with low BSP, OPN and PSA.

Survival model adjustments

Age is the most common risk factor for prostate cancer and age has a strong association with time to prostate cancer diagnosis and death [15]. Cox proportional hazard modeling with age as a covariate was carried out on individual biomarkers and pairs or trios of biomarkers to account for potential differences in age. Adjusting for age had a minimal effect on distal or proximal disease-free survival hazard ratios (Fig. 6). After adjusting for age, the only hazard ratios for distal overall survival that remained significant were with DSPP alone as the biomarker or with paired DSPP and OPN. When models of overall survival with distal levels of individual and grouped biomarkers were adjusted for age, only the paring of high DSPP and OPN remained significant. For both distal and proximal disease-free survival and proximal overall survival, biomarker aggregates that remained significant after adjusting for age include DSPP with OPN, DSPP with PSA, DSPP with BSP and PSA, and DSPP with OPN and PSA. BMI and hypertension have also been associated with prostate cancer risk [16,17]. Adjusting for BMI or hypertension in the Cox proportional hazard models did not change any of the hazard ratios significantly. Finally, because of the multiple comparisons made, an FDR analysis was carried out within each set (Fig. 6). Biomarkers/ aggregates identified as true discoveries (marked by asterisks in the figure) changed little between distal and proximal disease-free survival. For disease-free survival at both distal and proximal time points, the biomarkers/aggregates identified in the age-adjusted models as discoveries were DSPP alone, PSA alone, BSP with OPN, DSPP with PSA, DSPP with OPN and PSA. The hazard ratios for distal overall survival did not qualify as true discoveries. For proximal overall survival, the biomarkers/aggregates identified as discoveries were DSPP alone, DSPP with BSP, BSP with OPN, DSPP with OPN, DSPP with OPN and PSA.

Discussion

Prostate cancer is the most common cancer in men and the second leading cause of cancer death in men [18]. The introduction of PSA into screening has been associated with an increase in the detection of early stage, localized cancers [6]. PSA is an organ-specific, but not cancer-

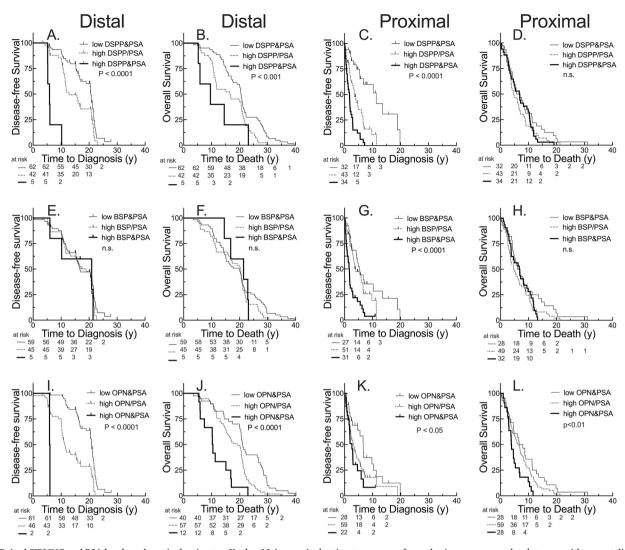


Fig. 4. Paired SIBLING and PSA levels and survival estimates. Kaplan-Meier survival estimates were performed using serum samples that were either most distal to or most proximal to time of diagnosis and comparing survival between samples with low levels of a SIBLING and PSA (low SIBLING&PSA), high levels of a SIBLING or PSA (high SIBLING/PSA), or high levels of both SIBLING and PSA (high SIBLING&PSA). Survival estimates were performed for DSPP with PSA (A, B, C, D), BSP with PSA (E, F, G, H), and OPN with PSA (I, J, K, L), modeling time to diagnosis (A, C, E, G, I, K) and time to death (B, D, F, H, J, L). The P value reported for each pairing is from a log-rank trend test for three or more survival curves. Abbreviations: DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen. The number at risk are reported below each graph.

specific, biomarker. Attempts to improve the detection of prostate cancer have included aggregated tests that incorporate urinary mRNA expression levels, multiplexed protein analysis, and clinical variables, blood biomarkers, genetic markers, and prostate examination [19,20]. While the use of multiple biomarkers in a combination with imaging modalities have also improved diagnostics and treatment prognosis [21,22], these advances do not address risk stratification at the level of neoplastic transformation or other early pre-symptomatic events in cancer development.

SIBLINGs are secreted and intrinsically disordered extracellular matrix proteins. They have multiple binding partners and can tether their respective ligands to the cell surface via interactions with integrins and CD44 variants [4,7,23–29]. Though initially isolated from mineralizing tissue, SIBLING expression by normal non-mineralizing tissue and upregulation in multiple cancer types were also observed [5,30]. The transcription factor RUNX2 has been implicated in regulating the expression of the SIBLINGs BSP [31,32], OPN [33], and DMP1 [34]. RUNX2 is expressed in multipotent mesenchymal cells, osteoblast-lineage cells, and chondrocytes [35]. RUNX2 is also expressed in the prostate where it has been associated with epithelial stem cells during development [36]. RUNX2 has been implicated in regulating the

epithelial to mesenchymal transition, where epithelial cells progressively acquire characteristics of mesenchymal or fibroblasts-like cells [37,38] that contribute to prostate cancer invasiveness [39]. RUNX2 expression levels associated with increased Gleason score and metastasis of prostate tumors [40].

Tissue expression of SIBLINGs in established prostate cancer strongly correlated with the conventional histopathological prognostic indicators such as Gleason score, pathological staging and metastasis [41–45]. DSPP serum levels were elevated in early-stage prostate cancer and combining DSPP and PSA values increased sensitivity and specificity of detection of established prostate cancer [9]. DSPP and its proteolytically processed products — dentin sialoprotein and dentin phosphoprotein have been associated with mesenchymal and epithelial cell differentiation and in homeostatic regeneration [46–48]. DSPP expression and serum levels were elevated in the earliest stages of prostate cancer [9,43]. Suppressing DSPP expression in squamous cell carcinoma *in vitro* models was found to reduce the expression of genes involved in metastasis, angiogenesis, and epithelial to mesenchymal transition [49], downregulate cancer stem cell markers [50], and decrease cancer cell viability, migration and invasion while increasing apoptosis [51,52].

OPN has been proposed as a diagnostic biomarker for prostate cancer

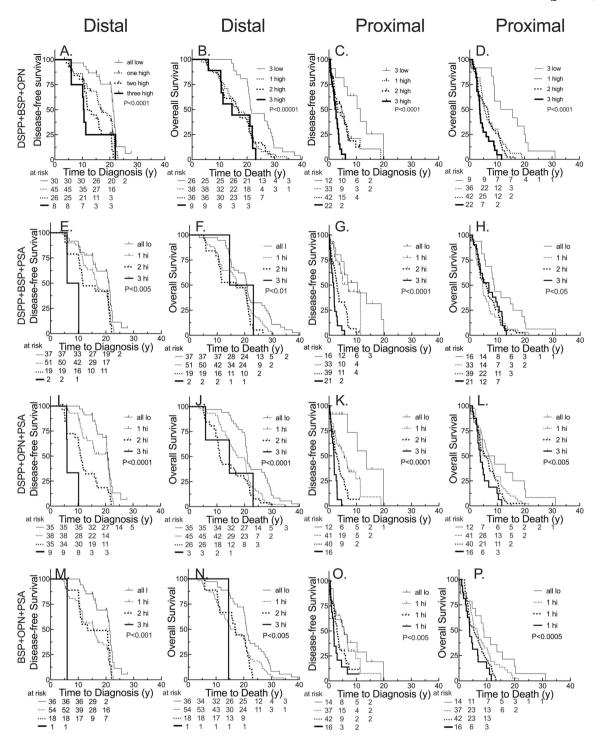


Fig. 5. Trios of SIBLINGs and PSA levels and survival estimates. Kaplan-Meier survival estimates were performed using serum samples that were either most distal to or most proximal to time of diagnosis and comparing survival between subjects with 1 biomarker high, 2 biomarkers high, or all 3 biomarkers high to subjects with low levels of all three biomarkers (reference). Survival estimates were performed for DSPP + BSP + OPN (A, B, C, D), DSPP + BSP + PSA (E, F, G, H), BSP + OPN + PSA (I, J, K, L), and DSPP + OPN + PSA, modeling time to diagnosis (A, C, E, G, I, K, M, O) and time to death (B, D, F, H, J, L, N, P). The P value reported for each pairing is from a log-rank trend test for three or more survival curves. Abbreviations: DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen. The number at risk are reported below each graph.

and a prognostic biomarker of response to treatment [45,53]. Underlying OPN biology are sentinel pathways involving neovascularization and angiogenesis, innate and adaptive immunity, hypoxia, stress response, and apoptosis [54–58]. BSP expression status in patients with moderately differentiated tumors or with pathologically confined tumors associated with higher risk of biochemical progression of prostate cancer [59]. High serum BSP levels were predictive of reduced postoperative

and long-term survival [60]. Serum BSP and PSA doubling time were identified as independent predictors of prostate cancer metastasis to bone [44]. The above observations were focused on SIBLING expression or circulating levels and established cancer detection/prognosis.

The current study is the first to use longitudinal preclinical samples to assess SIBLING associations with time to events (diagnosis, censoring and death). In this study, individual biomarkers exhibited different

Table 5Survival estimates of trios of biomarkers.

	Distal DFS	Distal OS	Proximal DFS	Proximal OS
DSPP + BSP + OPN				
Median survival, y				
All biomarkers low	21.2	21.4	11.4	13.5
1 biomarker high	17.0	16.9	4.72	5.28
2 biomarkers high	15.9	16.7	2.94	6.56
3 biomarkers high	10.4	14.5	2.44	3.66
HR, 3 biomarkers high	3.34 (1.34-7.69)	4.90 (1.91–11.8)	6.48 (2.65–17.8)	20.2 (6.98–74.5)
P value	< 0.01	< 0.0005	< 0.0001	< 0.0001
DSPP + BSP + PSA				
Median survival, y				
All biomarkers low	21.1	20.8	10.3	8.83
1 biomarker high	16.6	17.4	6.96	4.61
2 biomarkers high	11.7	16.8	2.91	5.99
3 biomarkers high	8.10	18.8	1.61	6.88
HR, 3 biomarkers high	10.8 (1.67-40.6)	2.84 (0.45-9.98)	9.95 (4.21–26.0)	3.92 (1.80-9.19)
P value	< 0.005	n.s.	P < 0.0001	P < 0.001
DSPP + OPN + PSA				
Median survival, y				
All biomarkers low	20.8	21.0	11.4	8.68
1 biomarker high	20.8	20.1	5.41	6.56
2 biomarkers high	10.7	11.3	2.61	4.88
3 biomarkers high	5.86	5.86	1.86	3. 68
HR, 3 biomarkers high	16.1 (3.56-53.12)	4.42 (1.03-13.2)	12.1 (4.52–36.6)	9.67 (3.67-28.9)
P value	< 0.0001	< 0.05	< 0.0001	< 0.0001
BSP OPN + PSA				
Median survival, y				
All biomarkers low	21.2	21.2	5.50	8.83
1 biomarker high	15.3	15.2	5.56	6.49
2 biomarkers high	10.7	18.8	2.94	5.44
3 biomarkers high	5.86	18.8	1.75	3.82
HR, 3 biomarkers high	16.9 (2.54–66.2)	3.39 (0.53-12.2)	3.16 (1.36-7.64)	4.96 (2.07–12.4)
P value	< 0.0005	n.s.	< 0.01	< 0.0005

Abbreviations: DFS, disease-free survival; OS, overall survival; DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; HR, hazard ratio; n.s., not significant.

Groups analyzed: low levels of all three biomarkers, high levels of one of the biomarkers, and high levels of two biomarkers, and high levels of all three biomarkers. Hazard ratio determined by Cox proportional Hazard modeling of biomarkers dichotomized by their respective cut-off values. Reference group is the all biomarkers low group.

associations with survival. High DSPP levels at the most distal and most proximal times to diagnosis/censoring were associated with a decreased time to prostate cancer diagnosis (reduction in median disease-free survival), whereas high BSP levels associated with decreased time to death (reduction in median overall survival) only at the most proximal times to diagnosis/censoring. High OPN levels at the most distal and proximal times to diagnosis/censoring were associated with decreased time to death, while high PSA levels only associated with decreased time to diagnosis at more proximal timepoints. For both pairs and trios of biomarkers, if DSPP was in the group, higher distal and proximal hazard ratios were observed. Proximal hazard ratios were higher when groups contained OPN, PSA or BSP. These observations are consistent with DSPP as a biomarker of early events in the development of cancer, while OPN, BSP and PSA are biomarkers of cancer progression.

The limitations to the current study include a modest sample size of 109 subjects with 218 samples, limited diversity in the racial composition of longitudinal study group, missing staging data, and only two longitudinal time points for each subject were analyzed. The BSP assay exhibited the highest %CV values for controls and the widest 95 % confidence intervals. This, in combination with the limited number of samples may have affected detection of associations between BSP levels in preclinical samples and time to events (diagnosis, death). The translation of SIBLING measures to clinical utility has been hindered by their biology. SIBLINGs have splice variants, microheterogeneity in post-translational modifications, and cellular and extracellular sequestration that can complicate their quantitation [61–75]. For example, a specific OPN splice variant (OPNc) has been identified in breast cancer and, depending on the antibodies used in immunohistochemistry or immunoassays, the variant won't be detected [61,62]. The extent of

phosphorylation and N-linked oligosaccharide modifications can also interfere with an antibody binding to its epitope.

The proteolytic processing of SIBLINGs also complicates their analysis. Very rapidly after its synthesis, DSPP is cleaved by bone morphogenetic proteins generating dentin sialoprotein (DSP) and dentin phosphoprotein containing fragments [76]. BSP may be processed by cathepsins and matrix metalloproteinases [77,78]. OPN cleavage by thrombin has been well characterized [79,80]. OPN and DMP1can be processed by matrix metalloproteinases (MMPs) [71,73,80,81]. Thus, ELISAs to measure DSPP, BSP and OPN may be detecting full length or fragments or both, depending on the detection antibody epitopes. Sandwich-based ELISAs can be engineered to be more specific than the competitive assay since the 'sandwich' involves two antibody recognition steps. In the current study, competitive ELISAs that measure the DSP portion of DSPP, total BSP (intact plus fragments) as well a sandwich-based ELISAs that measures intact OPN were used.

Observations from this current study are consistent with SIBLINGs meriting further study as serum biomarkers of events across the life history of prostate cancer development. Future studies of SIBLINGs and PSA with larger retrospective longitudinal repository study samples will help to define the furthest time before diagnosis where the biomarkers retain a predictive capacity. Such studies would also enable analysis of rates of changes in these biomarkers over time to be determined and the exploration of their predictive capability.

CRediT authorship contribution statement

Alka Jain: Writing – review & editing, Writing – original draft, Supervision, Methodology, Formal analysis, Data curation. Ying Ni:

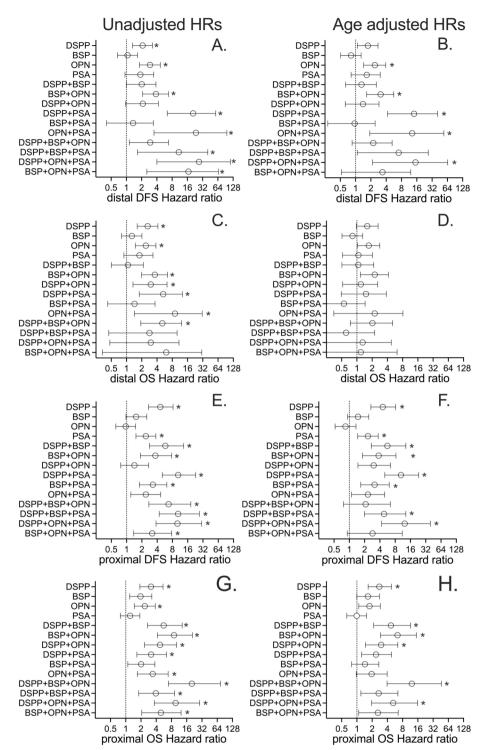


Fig. 6. Cox proportional hazard ratios. Proportional hazard modeling of unadjusted (A, C, E, G) and adjusted for age (B, D, F, H) hazard ratios for disease-free survival (A, B, E, F), overall survival (C, D, G, H) for the groups segregated as most distal to diagnosis (A, B, C, D) and most proximal to diagnosis (E, F, G, H). Asterisks indicate biomarkers that were identified as positive discoveries after applying the adaptive FDR method of Benjamini, Krieger and Yekutieli. Abbreviations: DSPP, dentin sialophosphoprotein; BSP, bone sialoprotein; OPN, osteopontin; PSA, prostate-specific antigen; DFS, disease-free survival; OS, overall survival.

Writing – review & editing, Resources, Methodology, Data curation. Daisy Zhang: Writing – review & editing, Methodology. Eleanor M. Simonsick: Writing – review & editing, Resources, Formal analysis, Conceptualization. E. Jeffrey Metter: Writing – review & editing, Resources, Formal analysis. Kalu U. Ogbureke: Writing – review & editing, Resources, Formal analysis, Conceptualization. Larry W. Fisher: Writing – review & editing, Resources, Conceptualization. Neal S. Fedarko: Writing – review & editing, Resources, Formal analysis,

Conceptualization, Writing – original draft, Supervision, Project administration.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work was supported by a grant from the National Institutes of Health, 1R01CA149273 (N.S.F.). This research was also funded in part by the Intramural Research Program of the National Institute on Aging ZIA AG000015 (E.S., E.J.M.) and by the Intramural Research Program of the National Institute on Dental and Craniofacial Research ZIA DE000074 (L.W.F.) of the National Institutes of Health. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Appendix A. Supplementary data

Supplementary data to this article can be found online at $\frac{https:}{doi.}$ org/10.1016/j.mbplus.2025.100171.

Data availability

The complete datasets for the data presented in the manuscript have been uploaded to the Mendeley Data repository and is available at doi: 10.17632/6x9z7t76hm.1.

References

- [1] SEER*Explorer: An interactive website for SEER cancer statistics [Internet]. Data source(s): SEER Incidence Data, November 2023 Submission (1975-2021), SEER 22 registries. Surveillance Research Program, National Cancer Institute;, 2024 April 17. https://seer.cancer.gov/explorer. (Accessed August 19, 2024).
- [2] M.C. Haffner, W. Zwart, M.P. Roudier, L.D. True, W.G. Nelson, J.I. Epstein, A.M. De Marzo, P.S. Nelson, S. Yegnasubramanian, Genomic and phenotypic heterogeneity in prostate cancer, Nature Reviews Urology 18 (2) (2021) 79–92, https://doi.org/ 10.1038/s41585-020-00400-w.
- [3] P. Mehlen, A. Puisieux, Metastasis: a question of life or death, Nature Reviews Cancer 6 (6) (2006) 449–458, https://doi.org/10.1038/nrc1886.
- [4] A. Jain, A. Karadag, B. Fohr, L.W. Fisher, N.S. Fedarko, Three SIBLINGs (small integrin-binding ligand, N-linked glycoproteins) enhance factor H's cofactor activity enabling MCP-like cellular evasion of complement-mediated attack, The Journal of Biological Chemistry 277 (16) (2002) 13700–13708, https://doi.org/10.1074/jbc.M110757200.
- [5] A. Bellahcene, V. Castronovo, K.U. Ogbureke, L.W. Fisher, N.S. Fedarko, Small integrin-binding ligand N-linked glycoproteins (SIBLINGs): multifunctional proteins in cancer, Nature Reviews Cancer 8 (3) (2008) 212–226, https://doi.org/ 10.1038/nrc2345.
- [6] D. Raineri, C. Dianzani, G. Cappellano, F. Maione, G. Baldanzi, I. Iacobucci, N. Clemente, G. Baldone, E. Boggio, C.L. Gigliotti, R. Boldorini, J.M. Rojo, M. Monti, L. Birolo, U. Dianzani, A. Chiocchetti, Osteopontin binds ICOSL promoting tumor metastasis, Communications Biology 3 (1) (2020) 615, https://doi.org/10.1038/s42003-020-01333-1.
- [7] L.W. Fisher, N.S. Fedarko, Six genes expressed in bones and teeth encode the current members of the SIBLING family of proteins, Connective Tissue Research 44 (Suppl 1) (2003) 33–40.
- [8] K.U. Ogbureke, L.W. Fisher, SIBLING expression patterns in duct epithelia reflect the degree of metabolic activity, The Journal of Histochemistry and Cytochemistry 55 (4) (2007) 403–409, https://doi.org/10.1369/jhc.6A7075.2007.
- [9] A. Jain, D.A. McKnight, L.W. Fisher, E.B. Humphreys, L.A. Mangold, A.W. Partin, N.S. Fedarko, Small integrin-binding proteins as serum markers for prostate cancer detection, Clinical cancer research: an official journal of the American Association for, Cancer Research 15 (16) (2009) 5199–5207, https://doi.org/10.1158/1078-0432.CCR-09-0783.
- [10] C.C. Anunobi, K. Koli, G. Saxena, A.A. Banjo, K.U. Ogbureke, Expression of the SIBLINGs and their MMP partners in human benign and malignant prostate neoplasms, Oncotarget 7 (30) (2016) 48038–48049, https://doi.org/10.18632/ oncotarget.10110.
- [11] J. Aseervatham, S. Geetu, C.C. Anunobi, K. Koli, K.U.E. Ogbureke, Survey of dentin sialophosphoprotein and its cognate matrix metalloproteinase-20 in human cancers, Cancer Medicine 8 (5) (2019) 2167–2178, https://doi.org/10.1002/ cam4.2117.
- [12] S. Loeb, A. Kettermann, L. Ferrucci, P. Landis, E.J. Metter, H.B. Carter, PSA doubling time versus PSA velocity to predict high-risk prostate cancer: data from the Baltimore Longitudinal Study of Aging, European Urology 54 (5) (2008) 1073–1080, https://doi.org/10.1016/j.eururo.2008.06.076.
- [13] H.B. Carter, L. Ferrucci, A. Kettermann, P. Landis, E.J. Wright, J.I. Epstein, B. J. Trock, E.J. Metter, Detection of life-threatening prostate cancer with prostate-specific antigen velocity during a window of curability, Journal of the National Cancer Institute 98 (21) (2006) 1521–1527, https://doi.org/10.1093/jnci/djj410.

- [14] Y. Benjamini, A.M. Krieger, D. Yekutieli, Adaptive linear step-up procedures that control the false discovery rate, Biometrika 93 (2006) 491–507.
- [15] R.A. Godtman, K.S. Kollberg, C.-G. Pihl, M. Månsson, J. Hugosson, The Association between age, prostate cancer risk, and higher gleason score in a long-term screening program: results from the göteborg-1 prostate cancer screening trial, European Urology 82 (3) (2022) 311–317, https://doi.org/10.1016/j. europ. 2022.01.018
- [16] N. Tzenios, M.E. Tazanios, M. Chahine, The impact of body mass index on prostate cancer: An updated systematic review and meta-analysis, Medicine (Baltimore) 101 (45) (2022) e30191, https://doi.org/10.1097/MD.0000000000030191.
- [17] Z. Liang, B. Xie, J. Li, X. Wang, S. Wang, S. Meng, A. Ji, Y. Zhu, X. Xu, X. Zheng, L. Xie, Hypertension and risk of prostate cancer: a systematic review and metaanalysis, Scientific Reports 6 (2016) 31358, https://doi.org/10.1038/srep31358.
- [18] R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2019, CA: a Cancer Journal for Clinicians 69 (1) (2019) 7–34, https://doi.org/10.3322/caac.21551.
- [19] W.C.H. Visser, H. de Jong, W.J.G. Melchers, P.F.A. Mulders, J.A. Schalken, Commercialized blood-, urinary- and tissue-based biomarker tests for prostate cancer diagnosis and prognosis, Cancers (Basel) 12 (12) (2020), https://doi.org/ 10.3300/cancers/12/13700
- [20] H. Gronberg, J. Adolfsson, M. Aly, T. Nordstrom, P. Wiklund, Y. Brandberg, J. Thompson, F. Wiklund, J. Lindberg, M. Clements, L. Egevad, M. Eklund, Prostate cancer screening in men aged 50-69 years (STHLM3): a prospective populationbased diagnostic study, The Lancet Oncology 16 (16) (2015) 1667–1676, https:// doi.org/10.1016/S1470-2045(15)00361-7.
- [21] V.C. Munteanu, R.A. Munteanu, D. Gulei, V.H. Schitcu, B. Petrut, I. Berindan Neagoe, P. Achimas Cadariu, I. Coman, PSA based biomarkers, imagistic techniques and combined tests for a better diagnostic of localized prostate cancer, Diagnostics (basel) 10 (10) (2020), https://doi.org/10.3390/diagnostics10100806.
- [22] S. Wach, O. Al-Janabi, K. Weigelt, K. Fischer, T. Greither, M. Marcou, G. Theil, E. Nolte, H.J. Holzhausen, R. Stohr, V. Huppert, A. Hartmann, P. Fornara, B. Wullich, H. Taubert, The combined serum levels of miR-375 and urokinase plasminogen activator receptor are suggested as diagnostic and prognostic biomarkers in prostate cancer, International Journal of Cancer 137 (6) (2015) 1406–1416, https://doi.org/10.1002/ijc.29505.
- [23] A. Jain, A. Karadag, L.W. Fisher, N.S. Fedarko, Structural requirements for bone sialoprotein binding and modulation of matrix metalloproteinase-2, Biochemistry 47 (38) (2008) 10162–10170, https://doi.org/10.1021/bi801068p.
- [24] A. Jain, L.W. Fisher, N.S. Fedarko, Bone sialoprotein binding to matrix metalloproteinase-2 alters enzyme inhibition kinetics, Biochemistry 47 (22) (2008) 5986–5995, https://doi.org/10.1021/bi800133n.
- [25] N.S. Fedarko, A. Jain, A. Karadag, L.W. Fisher, Three small integrin binding ligand N-linked glycoproteins (SIBLINGs) bind and activate specific matrix metalloproteinases, The FASEB Journal 18 (6) (2004) 734–736, https://doi.org/ 10.1096/fj.03-0966fje.
- [26] L.W. Fisher, D.A. Torchia, B. Fohr, M.F. Young, N.S. Fedarko, Flexible structures of SIBLING proteins, bone sialoprotein, and osteopontin, Biochemical and Biophysical Research Communications 280 (2) (2001) 460–465, https://doi.org/10.1006/ bbr. 2000 4146
- [27] N.S. Fedarko, B. Fohr, P.G. Robey, M.F. Young, L.W. Fisher, Factor H binding to bone sialoprotein and osteopontin enables tumor cell evasion of complementmediated attack, The Journal of Biological Chemistry 275 (22) (2000) 16666–16672, https://doi.org/10.1074/jbc.M001123200.
- [28] K. Koli, G. Saxena, K.U. Ogbureke, Expression of matrix metalloproteinase (MMP)-20 and potential interaction with dentin sialophosphoprotein (DSPP) in human major salivary glands, The Journal of Histochemistry and Cytochemistry 63 (7) (2015) 524–533, https://doi.org/10.1369/0022155415580817.
- [29] G. Saxena, K. Koli, J. de la Garza, K.U. Ogbureke, Matrix metalloproteinase 20-dentin sialophosphoprotein interaction in oral cancer, Journal of Dental Research 94 (4) (2015) 584–593, https://doi.org/10.1177/0022034515570156.
- [30] M. Prasad, Q. Zhu, Y. Sun, X. Wang, A. Kulkarni, A. Boskey, J.Q. Feng, C. Qin, Expression of dentin sialophosphoprotein in non-mineralized tissues, The Journal of Histochemistry and Cytochemistry 59 (11) (2011) 1009–1021, https://doi.org/ 10.1369/0022155411423406.
- [31] M. Riminucci, A. Corsi, K. Peris, L.W. Fisher, S. Chimenti, P. Bianco, Coexpression of bone sialoprotein (BSP) and the pivotal transcriptional regulator of osteogenesis, Cbfa1/Runx2, in malignant melanoma, Calcified Tissue International 73 (3) (2003) 281–289, https://doi.org/10.1007/s00223-002-2134-y.
- [32] G.L. Barnes, A. Javed, S.M. Waller, M.H. Kamal, K.E. Hebert, M.Q. Hassan, A. Bellahcene, A.J. Van Wijnen, M.F. Young, J.B. Lian, G.S. Stein, L.C. Gerstenfeld, Osteoblast-related transcription factors Runx2 (Cbfa1/AML3) and MSX2 mediate the expression of bone sialoprotein in human metastatic breast cancer cells, Cancer Research 63 (10) (2003) 2631–2637.
- [33] M. Sato, E. Morii, T. Komori, H. Kawahata, M. Sugimoto, K. Terai, H. Shimizu, T. Yasui, H. Ogihara, N. Yasui, T. Ochi, Y. Kitamura, Y. Ito, S. Nomura, Transcriptional regulation of osteopontin gene in vivo by PEBP2alphaA/CBFA1 and ETS1 in the skeletal tissues, Oncogene 17 (12) (1998) 1517–1525, https://doi.org/10.1038/sj.onc.1202064.
- [34] J.Q. Fen, J. Zhang, S.L. Dallas, Y. Lu, S. Chen, X. Tan, M. Owen, S.E. Harris, M. MacDougall, Dentin matrix protein 1, a target molecule for Cbfa1 in bone, is a unique bone marker gene, Journal of Bone and Mineral Research: the Official Journal of the American Society for Bone and Mineral Research 17 (10) (2002) 1822–1831, https://doi.org/10.1359/jbmr.2002.17.10.1822.
- [35] T. Komori, Runx2, an inducer of osteoblast and chondrocyte differentiation, Histochemistry and Cell Biology 149 (4) (2018) 313–323, https://doi.org/ 10.1007/s00418-018-1640-6.

- [36] Y. Li, C. Ge, R.T. Franceschi, Role of Runx2 in prostate development and stem cell function, The Prostate 81 (4) (2021) 231–241, https://doi.org/10.1002/ pros. 24099
- [37] D. Ribatti, R. Tamma, T. Annese, Epithelial-Mesenchymal Transition in Cancer: A Historical Overview, Translational Oncology 13 (6) (2020) 100773, https://doi. org/10.1016/j.tranon.2020.100773.
- [38] D.C. Voon, J.P. Thiery, The Emerging Roles of RUNX Transcription Factors in Epithelial-Mesenchymal Transition, Advances in Experimental Medicine and Biology 962 (2017) 471–489, https://doi.org/10.1007/978-981-10-3233-2 28.
- [39] C. Ge, G. Zhao, Y. Li, H. Li, X. Zhao, G. Pannone, P. Bufo, A. Santoro, F. Sanguedolce, S. Tortorella, M. Mattoni, S. Papagerakis, E.T. Keller, R. T. Franceschi, Role of Runx2 phosphorylation in prostate cancer and association with metastatic disease, Oncogene 35 (3) (2016) 366–376, https://doi.org/ 10.1038/onc.2015.91.
- [40] J. Akech, J.J. Wixted, K. Bedard, M. van der Deen, S. Hussain, T.A. Guise, A.J. van Wijnen, J.L. Stein, L.R. Languino, D.C. Altieri, J. Pratap, E. Keller, G.S. Stein, J. B. Lian, Runx2 association with progression of prostate cancer in patients: mechanisms mediating bone osteolysis and osteoblastic metastatic lesions, Oncogene 29 (6) (2010) 811–821, https://doi.org/10.1038/onc.2009.389.
- [41] G. De Pinieux, T. Flam, M. Zerbib, P. Taupin, A. Bellahcene, D. Waltregny, A. Vieillefond, M.F. Poupon, Bone sialoprotein, bone morphogenetic protein 6 and thymidine phosphorylase expression in localized human prostatic adenocarcinoma as predictors of clinical outcome: a clinicopathological and immunohistochemical study of 43 cases, The Journal of Urology 166 (5) (2001) 1924–1930, https://doi. org/10.1097/00005392-200111000-00085.
- [42] G. Carlinfante, D. Vassiliou, O. Svensson, M. Wendel, D. Heinegard, G. Andersson, Differential expression of osteopontin and bone sialoprotein in bone metastasis of breast and prostate carcinoma, Clinical & Experimental Metastasis 20 (5) (2003) 437–444, https://doi.org/10.1023/a:1025419708343.
- [43] M. Chaplet, D. Waltregny, C. Detry, L.W. Fisher, V. Castronovo, A. Bellahcene, Expression of dentin sialophosphoprotein in human prostate cancer and its correlation with tumor aggressiveness, International Journal of Cancer 118 (4) (2006) 850–856, https://doi.org/10.1002/ijc.21442.
- [44] Y. Wang, X.F. Zhang, J. Dai, Y.C. Zheng, M.G. Zhang, J.J. He, Predictive value of serum bone sialoprotein and prostate-specific antigen doubling time in patients with bone metastasis of prostate cancer, Journal of Huazhong University of Science and Technology Medical Sciences 33 (4) (2013) 559–562, https://doi.org/ 10.1007/s11596-013-1158-z.
- [45] A. Yu, K. Guo, Q. Qin, C. Xing, X. Zu, Clinicopathological and prognostic significance of osteopontin expression in patients with prostate cancer: a systematic review and meta-analysis, Bioscience Reports 41 (8) (2021), https:// doi.org/10.1042/BSR20203531.
- [46] K. Gu, S. Chang, H.H. Ritchie, B.H. Clarkson, R.B. Rutherford, Molecular cloning of a human dentin sialophosphoprotein gene, European Journal of Oral Sciences 108 (1) (2000) 35–42, https://doi.org/10.1034/j.1600-0722.2000.00765.x.
- [47] S. Chen, S. Rani, Y. Wu, A. Unterbrink, T.T. Gu, J. Gluhak-Heinrich, H.H. Chuang, M. Macdougall, Differential regulation of dentin sialophosphoprotein expression by Runx2 during odontoblast cytodifferentiation, The Journal of Biological Chemistry 280 (33) (2005) 29717–29727, https://doi.org/10.1074/jbc.M502929200.
 [48] M. Nakatomi, H. Ida-Yonemochi, H. Ohshima, Lymphoid enhancer-binding factor 1
- [48] M. Nakatomi, H. Ida-Yonemochi, H. Ohshima, Lymphoid enhancer-binding factor 1 expression precedes dentin sialophosphoprotein expression during rat odontoblast differentiation and regeneration, Journal of Endodontia 39 (5) (2013) 612–618, https://doi.org/10.1016/j.joen.2012.12.016.
 [49] J. Aseervatham, K.U.E. Ogbureke, Effects of DSPP and MMP20 Silencing on
- [49] J. Aseervatham, K.U.E. Ogbureke, Effects of DSPP and MMP20 Silencing on Adhesion, Metastasis, Angiogenesis, and Epithelial-Mesenchymal Transition Proteins in Oral Squamous Cell Carcinoma Cells, International Journal of Molecular Sciences 21 (13) (2020), https://doi.org/10.3390/ijms21134734.
- [50] N.G. Nikitakis, I. Gkouveris, J. Aseervatham, K. Barahona, K.U.E. Ogbureke, DSPP-MMP20 gene silencing downregulates cancer stem cell markers in human oral cancer cells, Cellular & Molecular Biology Letters 23 (2018) 30, https://doi.org/10.1186/s11658-018-0096-y.
- [51] I. Gkouveris, N.G. Nikitakis, J. Aseervatham, K.U.E. Ogbureke, Interferon gamma suppresses dentin sialophosphoprotein in oral squamous cell carcinoma cells resulting in antitumor effects, via modulation of the endoplasmic reticulum response, International Journal of Oncology 53 (6) (2018) 2423–2432, https://doi. org/10.3892/ijo.2018.4590.
- [52] R. Joshi, A. Tawfik, N. Edeh, V. McCloud, S. Looney, J. Lewis, S. Hsu, K. U. Ogbureke, Dentin sialophosphoprotein (DSPP) gene-silencing inhibits key tumorigenic activities in human oral cancer cell line, OSC2, PLoS One1 5 (11) (2010) e13974, https://doi.org/10.1371/journal.pone.0013974.
- [53] B. Smolska-Ciszewska, J. Mrochem-Kwarciak, K. Karczmarczyk, R. Deja, R. Suwinski, Pretreatment plasma osteopontin level as a marker of response to curative radiotherapy and hormonal treatment for prostate cancer, Radiotherapy and Oncology 200 (2024) 110518, https://doi.org/10.1016/j. radonc.2024.110518.
- [54] M.A. Icer, M. Gezmen-Karadag, The multiple functions and mechanisms of osteopontin, Clinical Biochemistry 59 (2018) 17–24, https://doi.org/10.1016/j. clinbiochem.2018.07.003.
- [55] E.Y. Lin, W. Xi, N. Aggarwal, M.L. Shinohara, Osteopontin (OPN)/SPP1: from its biochemistry to biological functions in the innate immune system and the central nervous system (CNS), International Immunology 35 (4) (2023) 171–180, https:// doi.org/10.1093/intimm/dxac060.
- [56] H.R. Moorman, D. Poschel, J.D. Klement, C. Lu, P.S. Redd, K. Liu, Osteopontin: a key regulator of tumor progression and immunomodulation, Cancers (Basel) 12 (11) (2020), https://doi.org/10.3390/cancers12113379.

- [57] L. Wang, X. Niu, Immunoregulatory roles of osteopontin in diseases, Nutrients 16 (2) (2024), https://doi.org/10.3390/nu16020312.
- [58] A.S. Lamort, I. Giopanou, I. Psallidas, G.T. Stathopoulos, Osteopontin as a link between inflammation and cancer: the thorax in the spotlight, Cells 8 (8) (2019), https://doi.org/10.3390/cells8080815.
- [59] D. Waltregny, A. Bellahcene, I. Van Riet, L.W. Fisher, M. Young, P. Fernandez, W. Dewe, J. de Leval, V. Castronovo, Prognostic value of bone sialoprotein expression in clinically localized human prostate cancer, Journal of the National Cancer Institute 90 (13) (1998) 1000–1008, https://doi.org/10.1093/jnci/90.13.1000.
- [60] S.H. Loosen, P. Hoening, N. Puethe, M. Luedde, M. Spehlmann, T.F. Ulmer, D. V. Cardenas, S. Roy, F. Tacke, C. Trautwein, U.P. Neumann, T. Luedde, C. Roderburg, Elevated serum levels of bone sialoprotein (BSP) predict long-term mortality in patients with pancreatic adenocarcinoma, Scientific Reports 9 (1) (2019) 1489, https://doi.org/10.1038/s41598-018-38352-2.
- [61] C. Hao, Z. Wang, Y. Gu, W.G. Jiang, S. Cheng, Prognostic value of osteopontin splice variant-c expression in breast cancers: a meta-analysis, Biomed Research International 2016 (2016) 7310694, https://doi.org/10.1155/2016/7310694.
- [62] B. He, M. Mirza, G.F. Weber, An osteopontin splice variant induces anchorage independence in human breast cancer cells, Oncogene 25 (15) (2006) 2192–2202, https://doi.org/10.1038/sj.onc.1209248.
- [63] S. Suzuki, H. Hoshino, K. Yoshida, J. Nakanishi, S. Tsuchiya-Hirata, S. Kobuke, N. Haruyama, F. Nishimura, H. Shiba, Genome-wide identification of chromatinenriched RNA reveals that unspliced dentin matrix protein-1 mRNA regulates cell proliferation in squamous cell carcinoma, Biochemical and Biophysical Research Communications 495 (3) (2018) 2303–2309, https://doi.org/10.1016/j. bbrc.2017.12.136.
- [64] Y. Yamakoshi, J.P. Simmer, Structural features, processing mechanism and gene splice variants of dentin sialophosphoprotein, Jpn Dent Sci Rev 54 (4) (2018) 183–196, https://doi.org/10.1016/j.jdsr.2018.03.006.
- [65] C. Chent, P.D. Delmas, Platelets contribute to circulating levels of bone sialoprotein in human, Journal of Bone and Mineral Research: the Official Journal of the American Society for Bone and Mineral Research 7 (1) (1992) 47–54, https://doi.org/10.1002/jbmr.5650070108.
- [66] C.I. Irion, J.C. Dunkley, K. John-Williams, J.M. Condor Capcha, S.A. Shehadeh, A. Pinto, M. Loebe, K.A. Webster, N.A. Brozzi, L.A. Shehadeh, Nuclear osteopontin is a marker of advanced heart failure and cardiac allograft vasculopathy: evidence from transplant and retransplant hearts, Frontiers in Physiology 11 (2020) 928.
- [67] K. Narayanan, A. Ramachandran, J. Hao, G. He, K.W. Park, M. Cho, A. George, Dual functional roles of dentin matrix protein 1: implications in biomineralization and gene transcription by activation of intracellular Ca2+ store, The Journal of Biological Chemistry 278 (19) (2003) 17500–17508, https://doi.org/10.1074/jbc. M212700200.
- [68] S. Ravindran, K. Narayanan, A.S. Eapen, J. Hao, A. Ramachandran, S. Blond, A. George, Endoplasmic reticulum chaperone protein GRP-78 mediates endocytosis of dentin matrix protein 1, The Journal of Biological Chemistry 283 (44) (2008) 29658–29670, https://doi.org/10.1074/jbc.M800786200.
- [69] A. Siyam, S. Wang, C. Qin, G. Mues, R. Stevens, R.N. D'Souza, Y. Lu, Nuclear localization of DMP1 proteins suggests a role in intracellular signaling, Biochemical and Biophysical Research Communications 424 (3) (2012) 641–646, https://doi.org/10.1016/j.bbrc.2012.07.037.
- [70] K. Zduniak, P. Ziolkowski, C. Ahlin, A. Agrawal, S. Agrawal, C. Blomqvist, M. L. Fjallskog, G.F. Weber, Nuclear osteopontin-c is a prognostic breast cancer marker, British Journal of Cancer 112 (4) (2015) 729–738, https://doi.org/10.1038/bjc.2014.664.
- [71] C. Chaussain, A.S. Eapen, E. Huet, C. Floris, S. Ravindran, J. Hao, S. Menashi, A. George, MMP2-cleavage of DMP1 generates a bioactive peptide promoting differentiation of dental pulp stem/progenitor cell, European Cells & Materials 18 (2009) 84–95, https://doi.org/10.22203/ecm.v018a08.
- [72] S. Peraramelli, Q. Zhou, Q. Zhou, B. Wanko, L. Zhao, T. Nishimura, T.H. Leung, S. Mizuno, M. Ito, T. Myles, T.M. Stulnig, J. Morser, L.L.K. Leung, Thrombin cleavage of osteopontin initiates osteopontin's tumor-promoting activity, Journal of Thrombosis and Haemostasis 20 (5) (2022) 1256–1270, https://doi.org/10.1111/jth.15663.
- [73] C. Qin, J.C. Brunn, R.G. Cook, R.S. Orkiszewski, J.P. Malone, A. Veis, W.T. Butler, Evidence for the proteolytic processing of dentin matrix protein 1. Identification and characterization of processed fragments and cleavage sites, The Journal of Biological Chemistry 278 (36) (2003) 34700–34708, https://doi.org/10.1074/jbc. M305315200
- [74] S.A. Sharif, X. Du, T. Myles, J.J. Song, E. Price, D.M. Lee, S.B. Goodman, M. Nagashima, J. Morser, W.H. Robinson, L.L. Leung, Thrombin-activatable carboxypeptidase B cleavage of osteopontin regulates neutrophil survival and synoviocyte binding in rheumatoid arthritis, Arthritis and Rheumatism 60 (10) (2009) 2902–2912, https://doi.org/10.1002/art.24814.
- [75] Q. Zhu, M.P. Gibson, Q. Liu, Y. Liu, Y. Lu, X. Wang, J.Q. Feng, C. Qin, Proteolytic processing of dentin sialophosphoprotein (DSPP) is essential to dentinogenesis, The Journal of Biological Chemistry 287 (36) (2012) 30426–30435, https://doi.org/ 10.1074/jbc.M112.388587.
- [76] Y. Yamakoshi, Dentin Sialophophoprotein (DSPP) and Dentin, Journal of Oral Biosciences 50 (1) (2008) 33–44, https://doi.org/10.2330/joralbiosci.50.33.
- [77] D. Brömme, S. Wilson, Role of Cysteine Cathepsins in Extracellular Proteolysis, in: W.C. Parks, R.P. Mecham (Eds.), Extracellular Matrix Degradation, Springer Berlin Heidelberg, Berlin, Heidelberg, 2011, pp. 23–51.
- [78] P. Hou, T. Troen, M.C. Ovejero, T. Kirkegaard, T.L. Andersen, I. Byrjalsen, M. Ferreras, T. Sato, S.D. Shapiro, N.T. Foged, J.M. Delaisse, Matrix metalloproteinase-12 (MMP-12) in osteoclasts: new lesson on the involvement of

- MMPs in bone resorption, Bone 34 (1) (2004) 37–47, https://doi.org/10.1016/j.
- [79] D.R. Senger, C.A. Perruzzi, A. Papadopoulos-Sergiou, L. Van de Water, Adhesive properties of osteopontin: regulation by a naturally occurring thrombin-cleavage in close proximity to the GRGDS cell-binding domain, Molecular Biology of the Cell 5 (5) (1994) 565–574, https://doi.org/10.1091/mbc.5.5.565.
- [80] T. Myles, L.L. Leung, Thrombin hydrolysis of human osteopontin is dependent on thrombin anion-binding exosites, The Journal of Biological Chemistry 283 (26) (2008) 17789–17796, https://doi.org/10.1074/jbc.M708629200.
 [81] B.M. Steiglitz, M. Ayala, K. Narayanan, A. George, D.S. Greenspan, Bone
- [81] B.M. Steiglitz, M. Ayala, K. Narayanan, A. George, D.S. Greenspan, Bone morphogenetic protein-1/Tolloid-like proteinases process dentin matrix protein-1, The Journal of Biological Chemistry 279 (2) (2004) 980–986, https://doi.org/ 10.1074/jbc.M310179200.