

An Augmented Negative Force-Frequency Relationship and Slowed Mechanical Restitution Are Associated With Increased Susceptibility to Drug-Induced Torsade de Pointes Arrhythmias in the Chronic Atrioventricular Block Dog

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*Correspondence: David J. Sprenkeler D.J.Sprenkeler@umcutrecht.nl

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Department of Medical Physiology, Division of Heart and Lungs, University Medical Center Utrecht, Utrecht, Netherlands

Introduction: In the chronic AV-block (CAVB) dog model, structural, contractile, and electrical remodeling occur, which predispose the heart to dofetilide-induced Torsade de Pointes (TdP) arrhythmias. Previous studies found a relation between electrical remodeling and inducibility of TdP, while structural remodeling is not a prerequisite for arrhythmogenesis. In this study, we prospectively assessed the relation between *in vivo* markers of contractile remodeling and TdP inducibility.

Methods: In 18 anesthetized dogs, the maximal first derivative of left ventricular pressure (LV dP/dt_{max}) was assessed at acute AV-block (AAVB) and after 2 weeks of chronic AV-block (CAVB2). Using pacing protocols, three markers of contractile remodeling, i.e., force-frequency relationship (FFR), mechanical restitution (MR), and post-extrasystolic potentiation (PESP) were determined. Infusion of dofetilide (0.025 mg/kg in 5 min) was used to test for TdP inducibility.

Results: After infusion of dofetilide, 1/18 dogs and 12/18 were susceptible to TdParrhythmias at AAVB and CAVB2, respectively (p = 0.001). The inducible dogs at CAVB2 showed augmented contractility at a CL of 1200 ms (2354 ± 168 mmHg/s in inducible dogs versus 1091 ± 59 mmHg/s in non-inducible dogs, p < 0.001) with a negative FFR, while the non-inducible dogs retained their positive FFR. The time constant (TC) of the MR curve was significantly higher in the inducible dogs (158 ± 7 ms versus 97 ± 8 ms, p < 0.0001). Furthermore, a linear correlation was found between a weighted score of the number and severity of arrhythmias and contractile parameters, i.e., contractility at CL of 1200 ms (r = 0.73, p = 0.002), the slope of the FFR (r = -0.58, p = 0.01) and the TC of MR (r = 0.66, p = 0.003). Thus, more severe arrhythmias were seen in dogs with the most pronounced contractile remodeling.

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Conclusion: Contractile remodeling is concomitantly observed with susceptibility to dofetilide-induced TdP-arrhythmias. The inducible dogs show augmented contractile remodeling compared to non-inducible dogs, as seen by a negative FFR, higher maximal response of MR and PESP and slowed MR kinetics. These altered contractility parameters could reflect disrupted Ca^{2+} handling and Ca^{2+} -overload, which predispose the heart to delayed- and early afterdepolarizations that could trigger TdP-arrhythmias.

Keywords: force-frequency relationship, mechanical restitution, post-extrasystolic potentiation, Torsade de Pointes, contractile remodeling, chronic AV-block dog

INTRODUCTION

Despite advances in treatment and prevention strategies, sudden cardiac death caused by ventricular arrhythmias remains a common cause of death in patients with heart failure or compensated hypertrophy (Tomaselli and Zipes, 2004). In response to certain stressors, these patients exhibit ventricular remodeling, an adaptive process that initially helps to maintain normal cardiac function, but eventually becomes maladaptive, causing electrical instability and an increased risk of lifethreatening ventricular arrhythmias (Konstam et al., 2011).

The chronic AV-block (CAVB) dog model is a widely used animal model to study ventricular remodeling and its relation with ventricular arrhythmias (Oros et al., 2008; Zhou et al., 2008; Wada et al., 2016). In this model, creation of third degree AV-block results in bradycardia and volume overload. To compensate for the resulting drop in cardiac output, ventricular remodeling occurs (Volders et al., 1998; Vos et al., 1998). This remodeling process reduces the so called "repolarization reserve," the ability of the heart to cope with stressors on repolarization (Roden, 1998). When repolarization is then further challenged by anesthesia and administration of a pro-arrhythmic drug such as dofetilide, early afterdepolarizations (EADs), ectopic beats, and TdP arrhythmias start to occur (Dunnink et al., 2010).

Ventricular remodeling is a complex process and can be divided into multiple components, such as structural, electrical, and contractile remodeling. However, the contribution of each of the three different components to arrhythmogenesis is not fully elucidated. Previous studies have shown that electrical remodeling is an important contributor to the susceptibility of TdP, (Vos et al., 1998) while structural remodeling is not a prerequisite (Schoenmakers et al., 2003). Electrical remodeling, which is reflected by prolongation of the action potential duration (APD) and increased spatial and temporal dispersion of repolarization, (Verduyn et al., 1997a,b; Thomsen et al., 2006, 2007) develops in synchrony with TdP-inducibility: both are present after 2 weeks of CAVB (Vos et al., 1998; Schoenmakers et al., 2003). Structural remodeling, on the other hand, follows a much slower path and is fully present after 16 weeks of CAVB (Schoenmakers et al., 2003; Peschar et al., 2004; Donker et al., 2005).

Yet, the time course of contractile remodeling and its relation to TdP-inducibility is less well described. Contractile adaptations, as a result of changes in Ca^{2+} handling, can be measured in vivo by three physiological mechanisms of the heart: forcefrequency relationship (FFR), mechanical restitution (MR) and post-extrasystolic potentiation (PESP). The FFR accounts for potentiation of contractility when heart rate is increased. MR and PESP relate to changes in contractile force of the extrasystolic beat and post-extrasystolic beat, respectively, when a basic stimulation train is interrupted with extrastimuli. MR represents the restoration of contractile force of the extrasystolic beat when the CI of the extrastimulus is lengthened. PESP displays the opposite behavior: when CI is shortened, the contractility of the post-extrasystolic beat is enhanced. Thus, the earlier an extrasystole occurs, the lower the contractility of the extrasystolic beat and the higher the contractility of the post-extraystolic beat.

de Groot et al. (2000) showed that FFR and PESP are altered in the CAVB-dogs after 6 weeks of remodeling and that these contractile parameters are associated with delayed afterdepolarizations (DADs) in vivo. Furthermore, in isolated cardiomyocytes from CAVB-dogs, Ca²⁺overload of the sarcoplasmic reticulum (SR) can result in spontaneous Ca²⁺ release, which triggers EADs/DADs (Sipido, 2006). However, it is unknown whether these macroscopic measures of Ca²⁺-handling (FFR, MR, PESP) also reflect the propensity for sustained TdP-arrhythmias. If contractility and arrhythmogenesis are intertwined, contractile parameters could function as markers of pro-arrhythmia and might eventually help to identify the patient at risk for life-threatening arrhythmias. Therefore, we aimed to investigate the relation between in vivo contractility measures and susceptibility to TdP-arrhythmias in the CAVB dog. We assessed these measures after 2 weeks of remodeling, since CAVB-dogs are susceptible to TdP-arrhythmias from that moment onward.

MATERIALS AND METHODS

Animal handling was in accordance with the "Directive 2010/63/EU of the European Parliament and of the Council of 22 September 2010 on the protection of animals used for scientific purposes" and the Dutch law, laid down in the Experiments

Abbreviations: AAVB, acute AV-block; CAVB, chronic AV-block; CI, coupling interval; CL, cycle length; DAD, delayed afterdepolarization; EAD, early afterdepolarization; FFR, force-frequency relationship; MAPD, monophasic action potential duration; MR, mechanical restitution; NCX, Na⁺-Ca²⁺ exchanger; PESP, post-extrasystolic potentiation; RyR2, ryanodine receptor 2; SERCA2a, sarcoplasmic reticulum Ca²⁺-ATPase 2a; SR, sinus rhythm; STV, short-term variability; TdP, Torsade de Pointes.

on Animals Act. All experiments were performed with approval of the Central Authority for Scientific Procedures on Animals (CCD), registered as project AVD115002016531.

Animal Preparation

Eighteen adult purpose-bred mongrel dogs of either sex (Marshal, United States; 5 females, 13 males; bodyweight 26 \pm 0.63 kg) were included. Experiments were performed under general anesthesia with mechanical ventilation at 12 breaths/min. The dogs were premedicated with a mixture of 0.5 mg/kg methadone, 0.5 mg/kg vetranquil and 0.02 mg/kg atropine i.m.. General anesthesia was induced with 25 mg/kg pentobarbital i.v. and maintained by isoflurane (1.5%) in a mixture of O₂ and N₂O (1:2). Periprocedurally antibiotics (ampicillin 1000 mg i.v. preoperatively and ampicillin 1000 mg i.m. postoperatively) and analgesics (metacam 0.2 mg/kg s.c. preoperatively and buprenorphine 0.3 mg i.m. postoperatively) were administered.

Ten surface-ECG leads (six limb leads, four precordial leads) were recorded throughout the experiment and stored on hard disk. Under aseptic conditions, the femoral artery and vein and carotid artery were dissected and sheaths were placed by Seldinger technique. Right and left ventricular monophasic action potential (RV and LV MAP) catheters (Hugo Sachs Elektronik GmbH, March, Germany) and a left ventricular 7F pressure catheter (CD Leycom Inc., Zoetermeer, Netherlands) were positioned under fluoroscopic guidance.

Experimental Protocol

Two serial experiments were performed. In the first experiment, baseline surface ECG, LV, and RV MAPD and left ventricular pressure during SR were recorded. Subsequently, His bundle ablation was done as described previously (Schoenmakers et al., 2003). When the idioventricular rhythm (IVR) was too low, a pacemaker was implanted subcutaneously with a transvenous lead in the RV apex. At AAVB, a pacing protocol (see below) was performed and the effects on LV pressure were recorded. Next, susceptibility to TdP was tested by a pro-arrhythmic challenge with the I_{Kr} blocker dofetilide (0.025 mg/kg i.v, infused during 5 min or until the first TdP occurred). TdP was defined as a run of five or more ectopic beats, with twisting morphology of the QRS-complexes. When \geq 3 TdP occurred in the first 10 min after start of dofetilide administration, the dog was considered inducible.

During the second experiment, after 2 weeks of remodeling at CAVB2, baseline ECG, LV and RV MAPD and left ventricular pressure were recorded and the pacing protocol and susceptibility test with dofetilide were repeated.

Pacing Protocol

FFR, MR, and PESP were measured during a pacing protocol from the LV MAP catheter. The FFR protocol consisted of 3 min of steady-state pacing at three different CLs of 300 ms, 750 ms, and 1200 ms. For the MR and PESP protocol, the LV was paced with a basic CL of 600 ms, which was interrupted every 20th beat by an extrastimulus with an incremental CI ranging from 250 ms up to 800 ms, with steps of 50 ms.

Data Analysis

Contractility Measures

As a measure of contractility, the maximal slope of left ventricular pressure rise (LV dP/dt_{max}) was calculated offline with computer software (Conduct NT, CD Leycom). For the FFR, the mean LV dP/dtmax of five consecutive beats was used at the three stimulation frequencies. A straight line was fitted through these points and its slope was calculated to quantify the orientation of the FFR. MR and PESP were defined as the LV dP/dt_{max} of the extrasystolic and post-extrasystolic beat, respectively (Figure 1). In addition, normalized MR and PESP were calculated by taking the ratio of LV dP/dt_{max} of the extrasystolic and post-extrasystolic beat, respectively, to the mean LV dP/dtmax of the five beats immediately preceding the extrasystole. Both MR and PESP were fitted to monoexponential functions using non-linear least squares regression, with the equation: $y = a-b^*e^{-\frac{x}{TC}}$ for MR and $v = a + b^* e^{-\frac{x}{TC}}$ for PESP to calculate the time constant (TC) of MR and PESP.

Electrophysiological Measures

Measurement of RR-interval and QT-interval was performed with calipers on lead II of the surface ECG by use of a software program (EPTracer, Cardiotek, Heilbronn, Germany). QT-interval was corrected for heart rate (QT_c) with the van der Water formula (Van de Water et al., 1989), which gives a better estimate than Bazett formula in anesthetized dogs.





LV and RV MAPD were measured from the initial peak to 80% of repolarization in a custom-made MATLAB software program. We defined interventricular dispersion (Δ MAPD) as the difference between LV MAPD and RV MAPD. STV of LV MAPD (STV) was calculated over 31 consecutive beats using the formula: STV = $\sum |D_{n+1} - D_n|/30 \times \sqrt{2}$, where *D* represents LV MAPD. STV after dofetilide challenge was measured just prior to the first ectopic beat.

Statistical Analysis

Data are expressed as mean \pm standard error of the mean (SEM). Serial data were analyzed with a paired Student's *t*-test or a repeated measure analysis of variance (ANOVA) with *post hoc* Bonferroni correction. Group analysis was performed with an unpaired Student's *t*-test. Correlation analysis was done with Pearson's correlation coefficient. All statistical analyses were performed with GraphPad Prism 6.0 (GraphPad Software Inc., La Jolla, CA, United States). A *p*-value < 0.05 was considered as statistically significant.

RESULTS

Contractile Remodeling

Contractile parameters at AAVB and CAVB are shown in **Figure 2**. Contractile remodeling was present after 2 weeks of CAVB, as seen by a significant increase in LV dP/dt_{max} during

IVR compared to AAVB (**Figure 2A**). At AAVB, a positive FFR was observed with augmentation of LV dP/dt_{max} at higher frequencies (**Figure 2B**). This relationship was blunted at CAVB2. At AAVB, the MR curve demonstrated a monoexponential rise in LV dP/dt_{max} toward a plateau with lengthening of the CI. The PESP curve displayed the opposite behavior. (**Figures 2C,D**). Both these parameters were increased at CAVB2.

Electrical Remodeling

The course of electrical remodeling is depicted in **Table 1**. At AAVB, all electrophysiological parameters have adapted to the sudden drop in heart rate. When challenged with dofetilide, a further significant increase was seen in QT, LV and RV MAPD, and STV. After 2 weeks of CAVB, electrical remodeling was present, as seen by a significant increase in QT, QTc, and LV MAPD under baseline conditions compared to AAVB. RV MAPD did not increase to the same extent as LV MAPD, thus creating a high interventricular dispersion (Δ MAPD). Finally, also STV showed a significant increase, which was further augmented after infusion of dofetilide.

Contractile and Electrical Remodeling and Susceptibility to TdP

Of the 18 dogs tested, one was susceptible to dofetilide-induced TdP at AAVB. After 2 weeks of CAVB, 12/18 dogs (67%, p = 0.002 compared to AAVB) were inducible after dofetilide infusion. Separate measurement of contractile parameters of the



CAVB2 + dof
$1452 \pm 62^{\$}$
$593 \pm 19^{\circ}$
$553 \pm 19^{\circ}$
$470 \pm 30^{\circ}$
$341\pm22^{\$}$
$130 \pm 25^{\circ}$
$2.66\pm0.34^{\$}$

TABLE 1 | Electrophysiological parameters.

 $^{*}p < 0.05$ versus SR. $^{\dagger}p < 0.05$ versus AAVB. $^{\$}p < 0.05$ versus CAVB2.

inducible and non-inducible dogs are depicted in Figure 3. At AAVB, no differences were found in contractile measures between the two groups. However, at CAVB2, the inducible dogs demonstrated a completely different pattern of contractile remodeling compared to the non-inducible dogs. At low stimulation frequency with a CL of 1200 ms, the inducible subjects had augmentation of contractility compared to the noninducible dogs ($2354 \pm 168 \text{ mmHg/s}$ versus $1091 \pm 59 \text{ mmHg/s}$, p < 0.001; Figure 3A). Furthermore, the slope of the FFR-curve was inverted in the inducible dogs, while the non-inducible dogs retained their positively oriented FFR (slope of -0.51 ± 0.19 in inducible dogs versus 0.89 \pm 0.06 in non-inducible dogs, p < 0.001). As seen from Table 2, these differences in LV dP/dt_{max} between inducible and non-inducible dogs are not due to differences in end-diastolic pressure (EDP). A significantly higher end-systolic pressure (ESP) is seen in the inducible dogs at low stimulation rate.

At small CIs, MR was similar in both groups, but diverged when the CI of the extrasystole rised above 450 ms, after which the susceptible dogs showed a significantly higher LV dP/dt_{max} (1483 ± 96 mmHg/s versus 1117 ± 31 mmHg/s, p = 0.01, **Figure 3B**). PESP, on the other hand, was increased in the inducible dogs at whole range of CIs (**Figure 3C**). When MR was normalized to the LV dP/dt_{max} of the preceding beats, the TC of MR appeared significantly higher in the inducible dogs (TC = 158 ± 7 ms versus TC = 97 ± 8 ms in non-inducible dogs, p < 0.001, **Figure 4A**). Normalized PESP was similar between the groups and had equal TC (143 ± 9 ms versus 153 ± 4 ms, p = 0.46, **Figure 4B**).

Electrically, no differences were found between the two groups at AAVB. At CAVB, the RR-interval, QT- and QTc-interval, LV MAPD, RV MAPD, and Δ MAPD were similar between the inducible and non-inducible dogs (**Table 3**). Only STV could distinguish between inducible and non-inducible dogs both at baseline and when challenged with dofetilide. Dogs that were inducible to drug-induced TdP had significantly higher STV at baseline (1.98 \pm 0.41 ms versus 0.61 \pm 0.08 ms, p = 0.009) which increased even more when challenged with dofetilide (3.18 \pm 0.36 ms versus 1.35 \pm 0.08 ms, p = 0.001).

Relation of Contractile and Electrical Remodeling With Severity of Arrhythmias

In order to quantify the severity of arrhythmias, we developed a new weighted scoring system, which takes into account both the number of arrhythmias in 10 min and whether the arrhythmias were sustained or defibrillations were needed (**Table 4**). This weighted score was correlated with both contractile and electrical remodeling. As illustrated in **Figure 5**, a linear correlation was found between this weighted score and LV dP/dt_{max} at CL of 1200 ms (r = 0.71, p = 0.002), the slope of FFR (r = -0.58, p = 0.01) and TC of MR (r = 0.66, p = 0.003). Of the electrical parameters, STV at baseline was also correlated with the weighted score of arrhythmic severity (r = 0.74, p = 0.0006).

The Effect of Ouabain on Contractility and TdP-Inducibility

As a proof of principle of the importance of contractile remodeling and Ca²⁺-overload for the inducibility of TdParrhythmias in the CAVB dog model, we attempted to pharmacologically increase Ca2+-load to alter FFR and potentially convert an initial non-inducible dog into an inducible one. Four non-inducible dogs were given a single dose of the cardiac glycoside ouabain (0.045 mg/kg i.v. in 1 min) prior to administration of dofetilide. In Figure 6, the effects of ouabain administration on FFR are depicted. In all dogs, ouabain resulted in a rise in contractility. Ouabain itself did not induce any arrhythmias. Unfortunately, in three out of four dogs, contractility at a CL of 1200 ms could not reach a level comparable to that of the inducible dogs and retained a positive FFR. As expected, these three dogs remained non-inducible after additional dofetilide infusion. Nevertheless, in one dog, contractility after ouabain administration increased to a level similar to that of the inducible dogs, with a more blunted FFR. Interestingly, this dog did become inducible to TdP-arrhythmias after infusion of dofetilide (Figure 7).

DISCUSSION

The results of this study demonstrate that contractile remodeling is already present after 2 weeks of CAVB and that this remodeling process differs substantially between TdP-susceptible and TdP-resistant dogs. Moreover, the more contractile or electrical remodeling have occurred, the higher the number and severity of arrhythmias after dofetilide challenge. Alterations in Ca²⁺-homeostasis may be the common denominator that explains the observed relation between contractile remodeling and arrhythmogenesis in the CAVB-dog model.





TABLE 2 | Hemodynamic parameters of non-inducible and inducible dogs.

		AAVB		CAV	В
		non-inducible	inducible	non-inducible	Inducible
LV EDP (mmHg)	CL 1200 ms	14 ± 2	16 ± 1	12 ± 2	11 ± 1
	CL 750 ms	14 ± 2	16 ± 1	10 ± 2	10 ± 1
	CL 300 ms	14 ± 1	15 ± 1	11 ± 3	9 ± 1
LV ESP (mmHg)	CL 1200 ms	60 ± 4	55 ± 4	73 ± 3	$88 \pm 2^{*}$
	CL 750 ms	63 ± 9	62 ± 2	85 ± 3	85 ± 3
	CL 300 ms	69 ± 9	69 ± 4	87 ± 6	77 ± 4
LV dP/dtmax (mmHg/s)	CL 1200 ms	547 ± 10	597 ± 42	1091 ± 59	$2354 \pm 168^{*}$
	CL 750 ms	763 ± 69	692 ± 39	1280 ± 68	$1860 \pm 160^{*}$
	CL 300 ms	1304 ± 170	1368 ± 81	1865 ± 94	1745 ± 86

p < 0.05 versus non-inducible during dofetilide infusion.



${\rm Ca}^{2+}$ Overload, DADs, and EADs in the CAVB-Dog

The present study is, to our knowledge, the first to show a direct relation between *in vivo* measurements of contractility

and inducibility of TdP-arrhythmias. Nevertheless, the relation between disturbances of Ca^{2+} -homeostasis and arrhythmogenesis on a cellular level is well established. Previous *in vitro* studies have demonstrated altered Ca^{2+} handling in isolated cardiomyocytes of CAVB-dogs with increased amplitude

TABLE 3	Electrophysiological	parameters of	of non-inducible	and inducible dogs.
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	Baseline		Dofeti	lide
	Non-inducible	Inducible	Non-inducible	Inducible
RR (ms)	1272 ± 33	1257 ± 69	1507 ± 107	1373 ± 72
QT (ms)	383 ± 19	439 ± 20	596 ± 50	587 ± 25
LV MAPD (ms)	292 ± 23	308 ± 14	494 ± 48	460 ± 39
RV MAPD (ms)	236 ± 9	249 ± 8	378 ± 53	328 ± 23
∆MAPD (ms)	57 ± 10	56 ± 9	116 ± 14	135 ± 35
STV (ms)	0.61 ± 0.08	$1.98 \pm 0.41^{*}$	1.35 ± 0.08	$3.18\pm0.36^{\dagger}$

*p < 0.05 versus non-inducible in baseline. $^{\dagger}p < 0.05$ versus non-inducible during dofetilide infusion.

 TABLE 4 | Custom-made weighted score of the number and severity of arrhythmias.

Arrhythmia	Points
Single ectopic beat	1
Double ectopic beats	2
Triple ectopic beats	3
Four ectopic beats	4
nsTdP (<50 complexes)	10
Sustained TdP (>50 complexes) or defibrillation	100
>1 consecutive defibrillations	200

Score is the total number of points during 10 min after start of dofetilide infusion.

and duration of Ca²⁺-transients along with a higher Ca²⁺content of the SR (Sipido et al., 2002). Furthermore, NCX is upregulated, which contributes to further loading of the SR via "reverse mode" Ca²⁺-influx (Sipido et al., 2000). The resulting SR Ca²⁺ overload is associated with spontaneous SR Ca²⁺ release, which can generate a DAD via "forward mode" function of NCX (exchange of 1 Ca²⁺-ion for 3 Na⁺-ions, creating an inward current) (Pogwizd and Bers, 2002). Furthermore, it has become more and more evident that altered Ca²⁺ cycling and upregulation of NCX also play a role in the formation of EADs, which has been regarded as the most important trigger of TdP arrhythmias (Volders et al., 2000; Němec et al., 2010; Horváth et al., 2015).

Our results are consistent with the study by de Groot et al. (2000) which showed that the highest LV dP/dt_{max} postpacing, which reflects an increased SR-load, was associated with the occurrence of DADs on MAP recordings. Since DADs and EADs share a common molecular mechanism, we expected to find the same relation between enhanced contractility and susceptibility of TdP-arrhythmias.

Force-Frequency Relationship and TdP-Arrhythmias

A blunted or inverted FFR has been observed in isolated muscle preparations of failing myocardium (Pieske et al., 1995, 1999, 2002) as well as in animal models (Eising et al., 1994; Neumann et al., 1998) or patients with cardiac dysfunction (Feldman et al., 1988; Hasenfuss et al., 1994; Schotten et al., 1999). In the

CAVB-dog, FFR has also shown to be inverted, with increased contractility at low heart rates (de Groot et al., 2000; Peschar et al., 2004).

In the present study we observed the inverted FFR only in the inducible dogs, while the FFR of the non-inducible dogs remained positive (Figure 3A). There is strong evidence that increased [Na]_i during diastole plays a central role in the inversion of the FFR (Mubagwa et al., 1997; Mills et al., 2007). A study on rabbit papillary muscle strips examined the effect of increased [Na]; on FFR and showed that treatment with the Na⁺ ionophore monensin, which increases diastolic [Na]_i, could convert a positive FFR into a negative FFR, mainly by increasing contractility at lower heart rates (Mubagwa et al., 1997). The high diastolic [Na]; will favor Ca²⁺ influx via NCX and since the ratio of diastole to systole is highest at low heart rates, Ca²⁺ loading of the SR will be enhanced, resulting in increased contractility. Dogs with CAVB have approximately 4 mM higher [Na]_i, compared to controls, probably due to reduced [Na]_i-affinity of the Na⁺-K⁺-ATPase and increased Na⁺-influx via the Na⁺-H⁺-exchanger type 1 (NHE-1) (Verdonck et al., 2003; van Borren et al., 2013). However, in these studies, no dofetilide challenge was performed, therefore it is unknown whether the [Na]_i is higher in inducible dogs compared to non-inducible dogs.

Nevertheless, we could hypothesize that in the inducible dogs a high $[Na]_i$ in combination with enhanced NCX activity results in Ca²⁺-overload, spontaneous Ca²⁺ release, and subsequent EADs and TdP-arrhythmias. Supporting this hypothesis, it has been shown that lowering $[Na]_i$ by administration of the selective late Na⁺-current blocker ranolazine could reduce the number of EADs *in vitro* and TdP-arrhythmias *in vivo* (Antoons et al., 2010). In addition, blockade of NCX by SEA-0400 is effective in prevention of TdP-arrhythmias in the CAVB dog (Bourgonje et al., 2013).

As a proof of concept, we tried to increase $[Na]_i$ in the non-inducible dogs by blockade of the Na⁺-K⁺-ATPase with ouabain. In contrast to the cardiac glycoside digitalis, ouabain has a fast onset of action, reaching his maximum after 5 min of administration (Fuerstenwerth, 2014). The increased $[Na]_i$ caused by inhibition of the Na-K-ATPase will be exchanged for Ca²⁺, causing Ca²⁺-overload of the cardiomyocyte that is responsible for both the inotropic and potential arrhythmogenic effects. As we have shown, only the dog that could reach the contractility level of the inducible dogs and developed a more



blunted FFR, became susceptible to dofetilide-induced TdP, which further illustrates the importance of $[Na]_i$ and $[Ca^{2+}]_i$ for the initiation of TdP-arrhythmias.

Mechanical Restitution/Post-extrasystolic Potentiation and TdP-Arrhythmias

We have demonstrated that inducible dogs have a higher MR and PESP and slower MR kinetics compared to the noninducible dogs. In previous studies altered MR and PESP have been found in patients with hypertrophy and heart failure. (Beck et al., 1971; Merillon et al., 1979; Seed et al., 1984; Prabhu and Freeman, 1995). Both phenomena are explained by time-dependent availability of releasable Ca²⁺ due to gradual translocation of releasable Ca²⁺ from an uptake compartment to a release compartment or because of a time-dependent recovery of the SR Ca²⁺-release channel, the RyR2.

Discussion on the mechanism of slower MR in the CAVB dog remains speculative, since the kinetics of RyR2 have never been investigated. Furthermore, the precise molecular mechanism behind inactivation and recovery of RyR2 remains controversial (Bers, 2008). First, it can be hypothesized that the intrinsic gating properties of the RyR2 have been altered by the remodeling process. Secondly, changes in cytosolic [Ca²⁺] have been proposed to influence the inactivation of RyR2, referred to as Ca²⁺-dependent inactivation. It is hypothesized that increased dyadic [Ca]_i, which is responsible for RyR2 activation, might also play a role in RyR2 inactivation (Laver, 2005). Thirdly, RyR refractoriness might be caused by "functional depletion" of Ca²⁺ of the SR after global Ca²⁺-release. Refilling of the SR during Ca^{2+} -reuptake increases the sensitivity of RyR2 to cytosolic Ca^{2+} and accelerates recovery from inactivation (Szentesi et al., 2004). Since Ca²⁺ homeostasis is an interplay between RyR2, SERCA2a, and NCX, we can assume that alterations in expression and function of one of these proteins, will subsequently affect normal function of the others. Increased NCX function could result in "relatively" diminished activity of SERCA2a, by changing the ratio of Ca²⁺ extrusion versus reuptake. In this regard, a study in a transgenic mouse model showed that overexpression of phospholamban, the main inhibitor of SERCA2a, caused a reduced reuptake of Ca²⁺ in the SR, but also resulted in a



after ouabain (green line) in four non-inducible dogs. As comparison, the mean of the inducible dogs is shown (blue line). Only dog 4 became inducible after ouabain in combination with dofetilide. TCs of the different curves are given.

higher TC of MR and increased PESP compared to control, just as we found in the inducible dogs (Hoit et al., 1999, 2000). Furthermore, in isolated cardiomyocytes, inhibition of SERCA2a could significantly slow down recovery from inactivation of RyR2 (Szentesi et al., 2004). Thus, increased NCX function, which contributes to susceptibility to triggered arrhythmias in the CAVB-dog, might indirectly influence SERCA2a and RyR2 function, and thus alter MR and PESP *in vivo*.

Electrical Remodeling and TdP-Arrhythmias

Electrically, we found that only STV can distinguish between inducible and non-inducible subjects. This is in line with previous studies on STV in the CAVB dog model, which show that STV both at baseline and after dofetilide infusion is a more powerful predictor of drug-induced TdP arrhythmias compared to the QT-interval or LV MAPD itself (Thomsen et al., 2004, 2006, 2007). The molecular basis of STV is not fully elucidated, but have been attributed to alterations in Ca²⁺ handling.

In isolated cardiomyocytes, β -adrenergic stimulation during reduced repolarization reserve (blockade of the repolarization current I_{Ks}) resulted in increased cellular Ca²⁺ load and spontaneous Ca²⁺ release, which was associated with increased STV and the occurrence of DADs and EADs (Johnson et al., 2010, 2013). Buffering of Ca²⁺ by BAPTA-AM, blockade of SR Ca²⁺ release with ryanodine or inhibition of NCX by SEA0400 led to a drastic reduction of STV and eliminated all DADs and EADs (Johnson et al., 2010, 2013). A study by Antoons et al. (2015) showed that STV is highly dependent on SR Ca²⁺ release, which can modulate Ca²⁺-dependent currents making the heart more prone to EADs. These data support the hypothesis that increased STV reflects disrupted Ca²⁺-homeostasis as the underlying mechanism of EADs and TdP-arrhythmias in the CAVB-dog.

Clinical Implications

Since the CAVB dog model is a specific model of compensated hypertrophy caused by volume overload, extrapolation to a population of patients with heart failure with reduced



ejection fraction should be done with caution. Nevertheless, since contractile remodeling and arrhythmogenesis are related, contractile parameters might be used in risk stratification of ventricular arrhythmias and sudden cardiac death. In addition, patients with heart failure with preserved ejection fraction (HFpEF) have also been shown to exhibit disrupted Ca²⁺-handling, therefore the contractile parameters investigated in this study could also be used in these patients. Recently, a study of a non-invasive measurement of PESP of blood pressure, measured via a photoplethysmographic device, showed that higher PESP was correlated with increased mortality in myocardial infarction survivors (Sinnecker et al., 2014). The cause of death was not stratified in this study, however, increased PESP may be associated with increased arrhythmia risk. The currently ongoing "EUropean Comparative Effectiveness Research to assess the use of primary prophylacTic Implantable Cardioverter Defibrillators" (EU-CERT-ICD) study evaluates the relationship between non-invasive measured PESP and the incidence of ICD-shocks in a primary prevention ICD-population. This study will give further insight if non-invasive measures of contractility could function as predictors of life-threatening arrhythmias.

Limitations

No direct measurements of Ca^{2+} transients or SR function have been done, therefore hypotheses on mechanism of contractile remodeling and the relation with TdP-arrhythmias are based on assumptions derived from previous molecular work. Explanations other than related to Ca^{2+} -handling may also be possible, such as mechano-electrical feedback, i.e., direct effects of altered loading conditions on repolarization. Secondly, LV dP/dt_{max} has important limitations as a measure of contractility, since it is also dependent on ventricular loading. While we did not measure ventricular volumes, we have shown that EDP are not different between inducible and non-inducible dogs. Therefore, we can assume the differences found in LV dP/dt_{max} are predominantly caused by changes in contractility. Finally, the relation between contractile remodeling and TdP was only measured at CAVB2, therefore no conclusion can be made on this association later in the remodeling process.

CONCLUSION

In the CAVB dog model, contractile and electrical remodeling are already present after 2 weeks of AV-block and develop concomitantly with susceptibility to dofetilide-induced TdP. Furthermore, contractile parameters are altered to a far larger extent in inducible dogs, as seen by development of an augmented negative FFR, higher maximal response of MR and PESP and slowed MR kinetics.

AUTHOR CONTRIBUTIONS

DS did experimental work, analysis of data, and wrote the manuscript. JB delivered technical support during experiments. MS contributed to experimental concepts and design of the study. MV was responsible for organization of experiments and revised the manuscript thoroughly. All authors approved the submitted version.

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REFERENCES

- Antoons, G., Johnson, D. M., Dries, E., Santiago, D. J., Ozdemir, S., Lenaerts, I., et al. (2015). Calcium release near L-type calcium channels promotes beat-tobeat variability in ventricular myocytes from the chronic AV block dog. J. Mol. Cell. Cardiol. 89, 326–334. doi: 10.1016/j.yjmcc.2015.10.008
- Antoons, G., Oros, A., Beekman, J. D. M., Engelen, M. A., Houtman, M. J. C., Belardinelli, L., et al. (2010). Late Na+ current inhibition by ranolazine reduces torsades de pointes in the chronic atrioventricular block dog model. *J. Am. Coll. Cardiol.* 55, 801–809. doi: 10.1016/j.jacc.2009.10.033
- Beck, W., Chesler, E., and Schrire, V. (1971). Postextrasystolic ventricular pressure responses. *Circulation* 44, 523–533. doi: 10.1161/01.CIR.44.4.523
- Bers, D. M. (2008). Calcium cycling and signaling in cardiac myocytes. Annu. Rev. Physiol. 70, 23–49. doi: 10.1146/annurev.physiol.70.113006.100455
- Bourgonje, V. J. A., Vos, M. A., Ozdemir, S., Doisne, N., Acsai, K., Varro, A., et al. (2013). Combined Na+/Ca2+ exchanger and l-type calcium channel block as a potential strategy to suppress arrhythmias and maintain ventricular function. *Circ. Arrhythm. Electrophysiol.* 6, 371–379. doi: 10.1161/CIRCEP.113.000322
- de Groot, S. H., Schoenmakers, M., Molenschot, M. M., Leunissen, J. D., Wellens, H. J., and Vos, M. A. (2000). Contractile adaptations preserving cardiac output predispose the hypertrophied canine heart to delayed afterdepolarizationdependent ventricular arrhythmias. *Circulation* 102, 2145–2151. doi: 10.1161/ 01.CIR.102.17.2145
- Donker, D. W., Volders, P. G., Arts, T., Bekkers, B. C., Hofstra, L., Spätjens, R. L., et al. (2005). End-diastolic myofiber stress and ejection strain increase with ventricular volume overload–Serial in-vivo analyses in dogs with complete atrioventricular block. *Basic Res. Cardiol.* 100, 372–382. doi: 10.1007/s00395-005-0525-8
- Dunnink, A., Sharif, S., Oosterhoff, P., Winckels, S., Montagne, D., Beekman, J., et al. (2010). Anesthesia and arrhythmogenesis in the chronic atrioventricular block dog model. *J. Cardiovasc. Pharmacol.* 55, 601–608. doi: 10.1097/FJC. 0b013e3181da7768
- Eising, G. P., Hammond, H. K., Helmer, G. A., Gilpin, E., and Ross, J. (1994). Force-frequency relations during heart failure in pigs. Am. J. Physiol. 267, H2516–H2522. doi: 10.1152/ajpheart.1994.267.6.H2516
- Feldman, M. D., Alderman, J. D., Aroesty, J. M., Royal, H. D., Ferguson, J. J., Owen, R. M., et al. (1988). Depression of systolic and diastolic myocardial reserve during atrial pacing tachycardia in patients with dilated cardiomyopathy. *J. Clin. Invest.* 82, 1661–1669. doi: 10.1172/JCI113778
- Fuerstenwerth, H. (2014). On the differences between ouabain and digitalis glycosides. Am. J. Ther. 21, 35–42. doi: 10.1097/MJT.0b013e318217a609
- Hasenfuss, G., Holubarsch, C., Hermann, H. P., Astheimer, K., Pieske, B., and Just, H. (1994). Influence of the force-frequency relationship on haemodynamics and left ventricular function in patients with non-failing hearts and in patients with dilated cardiomyopathy. *Eur. Heart J.* 15, 164–170. doi: 10.1093/oxfordjournals.eurheartj.a060471
- Hoit, B. D., Kadambi, V. J., Tramuta, D. A., Ball, N., Kranias, E. G., and Walsh, R. A. (2000). Influence of sarcoplasmic reticulum calcium loading on mechanical and relaxation restitution. *Am. J. Physiol. Heart Circ. Physiol.* 278, H958–H963. doi: 10.1152/ajpheart.2000.278.3.H958
- Hoit, B. D., Tramuta, D. A., Kadambi, V. J., Dash, R., Ball, N., Kranias, E. G., et al. (1999). Influence of transgenic overexpression of phospholamban on postextrasystolic potentiation. *J. Mol. Cell. Cardiol.* 31, 2007–2015. doi: 10.1006/ jmcc.1999.1031
- Horváth, B., Hegyi, B., Kistamás, K., Váczi, K., Bányász, T., Magyar, J., et al. (2015). Cytosolic calcium changes affect the incidence of early afterdepolarizations in canine ventricular myocytes 1. *Can. J. Physiol. Pharmacol.* 93, 527–534. doi: 10.1139/cjpp-2014-0511
- Johnson, D. M., Heijman, J., Bode, E. F., Greensmith, D. J., van der Linde, H., Abi-Gerges, N., et al. (2013). Diastolic spontaneous calcium release from the sarcoplasmic reticulum increases beat-to-beat variability of repolarization in

FP7/2007-2013 under grant agreement no. 602299, EU-CERT-ICD. The results have been presented as an abstract at the European Heart Rhythm Association (EHRA) congress 2018 and Heart Rhythm annual scientific sessions 2018.

canine ventricular myocytes after β -adrenergic stimulation. Circ. Res. 112, 246–256. doi: 10.1161/CIRCRESAHA.112.275735

- Johnson, D. M., Heijman, J., Pollard, C. E., Valentin, J.-P., Crijns, H. J. G. M., Abi-Gerges, N., et al. (2010). IKs restricts excessive beat-to-beat variability of repolarization during beta-adrenergic receptor stimulation. *J. Mol. Cell. Cardiol.* 48, 122–130. doi: 10.1016/j.yjmcc.2009.08.033
- Konstam, M. A., Kramer, D. G., Patel, A. R., Maron, M. S., and Udelson, J. E. (2011). Left ventricular remodeling in heart failure: current concepts in clinical significance and assessment. *JACC Cardiovasc. Imaging* 4, 98–108. doi: 10.1016/ j.jcmg.2010.10.008
- Laver, D. R. (2005). Coupled calcium release channels and their regulation by luminal and cytosolic ions. *Eur. Biophys. J.* 34, 359–368. doi: 10.1007/s00249-005-0483-y
- Merillon, J. P., Motte, G., Aumont, M. C., Masquet, C., Lecarpentier, Y., and Gourgon, R. (1979). Post-extrasystolic left ventricular peak pressure with and without left ventricular failure. *Cardiovasc. Res.* 13, 338–344. doi: 10.1093/cvr/ 13.6.338
- Mills, G. D., Harris, D. M., Chen, X., and Houser, S. R. (2007). Intracellular sodium determines frequency-dependent alterations in contractility in hypertrophied feline ventricular myocytes. Am. J. Physiol. Heart Circ. Physiol. 292, H1129–H1138. doi: 10.1152/ajpheart.00375.2006
- Mubagwa, K., Lin, W., Sipido, K., Bosteels, S., and Flameng, W. (1997). Monensininduced reversal of positive force–frequency relationship in cardiac muscle: role of intracellular sodium in rest-dependent potentiation of contraction. J. Mol. Cell. Cardiol. 29, 977–989. doi: 10.1006/jmcc.1996.0342
- Němec, J., Kim, J. J., Gabris, B., and Salama, G. (2010). Calcium oscillations and T-wave lability precede ventricular arrhythmias in acquired long QT type 2. *Heart Rhythm* 7, 1686–1694. doi: 10.1016/j.hrthm.2010.06.032
- Neumann, T., Ravens, U., and Heusch, G. (1998). Characterization of excitation-contraction coupling in conscious dogs with pacing-induced heart failure. *Cardiovasc. Res.* 37, 456–466. doi: 10.1016/S0008-6363(97) 00246-0
- Oros, A., Beekman, J. D. M., and Vos, M. A. (2008). The canine model with chronic, complete atrio-ventricular block. *Pharmacol. Ther.* 119, 168–178. doi: 10.1016/j.pharmthera.2008.03.006
- Peschar, M., Vernooy, K., Cornelussen, R. N., Verbeek, X. A. A., Reneman, R. S., Vos, M. A., et al. (2004). Structural, electrical and mechanical remodeling of the canine heart in AV-block and LBBB. *Eur. Heart J. Suppl.* 6, D61–D65. doi: 10.1016/j.ehjsup.2004.05.017
- Pieske, B., Kretschmann, B., Meyer, M., Holubarsch, C., Weirich, J., Posival, H., et al. (1995). Alterations in intracellular calcium handling associated with the inverse force-frequency relation in human dilated cardiomyopathy. *Circulation* 92, 1169–1178. doi: 10.1161/01.CIR.92.5.1169
- Pieske, B., Maier, L. S., Bers, D. M., and Hasenfuss, G. (1999). Ca2+ handling and sarcoplasmic reticulum Ca2+ content in isolated failing and nonfailing human myocardium. *Circ. Res.* 85, 38–46. doi: 10.1161/01.RES.85.1.38
- Pieske, B., Maier, L. S., Piacentino, V., Weisser, J., Hasenfuss, G., and Houser, S. (2002). Rate dependence of [Na+]i and contractility in nonfailing and failing human myocardium. *Circulation* 106, 447–453. doi: 10.1161/01.CIR. 0000023042.50192.F4
- Pogwizd, S. M., and Bers, D. M. (2002). Calcium cycling in heart failure: the arrhythmia connection. J. Cardiovasc. Electrophysiol. 13, 88–91. doi: 10.1046/ j.1540-8167.2002.00088.x
- Prabhu, S. D., and Freeman, G. L. (1995). Effect of tachycardia heart failure on the restitution of left ventricular function in closed-chest dogs. *Circulation* 91, 176–185. doi: 10.1161/01.CIR.91.1.176
- Roden, D. M. (1998). Taking the "idio" out of "idiosyncratic": predicting torsades de pointes. *Pacing Clin. Electrophysiol.* 21, 1029–1034. doi: 10.1111/j.1540-8159. 1998.tb00148.x
- Schoenmakers, M., Ramakers, C., van Opstal, J. M., Leunissen, J. D. M., Londoño, C., and Vos, M. A. (2003). Asynchronous development of electrical

remodeling and cardiac hypertrophy in the complete AV block dog. *Cardiovasc. Res.* 59, 351–359. doi: 10.1016/S0008-6363(03)00430-9

- Schotten, U., Voss, S., Wiederin, T. B., Voss, M., Schoendube, F., Hanrath, P., et al. (1999). Altered force-frequency relation in hypertrophic obstructive cardiomyopathy. *Basic Res. Cardiol.* 94, 120–127. doi: 10.1007/s003950050134
- Seed, W. A., Noble, M. I., Walker, J. M., Miller, G. A., Pidgeon, J., Redwood, D., et al. (1984). Relationships between beat-to-beat interval and the strength of contraction in the healthy and diseased human heart. *Circulation* 70, 799–805. doi: 10.1161/01.CIR.70.5.799
- Sinnecker, D., Dirschinger, R. J., Barthel, P., Muller, A., Morley-Davies, A., Hapfelmeier, A., et al. (2014). Postextrasystolic blood pressure potentiation predicts poor outcome of cardiac patients. J. Am. Heart Assoc. 3:e000857. doi: 10.1161/JAHA.114.000857
- Sipido, K. R. (2006). Calcium overload, spontaneous calcium release, and ventricular arrhythmias. *Heart Rhythm* 3, 977–979. doi: 10.1016/j.hrthm.2006. 01.013
- Sipido, K. R., Volders, P. G., de Groot, S. H., Verdonck, F., Van de Werf, F., Wellens, H. J., et al. (2000). Enhanced Ca(2+) release and Na/Ca exchange activity in hypertrophied canine ventricular myocytes: potential link between contractile adaptation and arrhythmogenesis. *Circulation* 102, 2137–2144. doi: 10.1161/01.CIR.102.17.2137
- Sipido, K. R., Volders, P. G. A., Schoenmakers, M., De Groot, S. H. M., Verdonck, F., and Vos, M. A. (2002). Role of the Na/Ca exchanger in arrhythmias in compensated hypertrophy. *Ann. N. Y. Acad. Sci.* 976, 438–445. doi: 10.1111/j.1749-6632.2002.tb04773.x
- Szentesi, P., Pignier, C., Egger, M., Kranias, E. G., and Niggli, E. (2004). Sarcoplasmic reticulum Ca2+ refilling controls recovery from Ca2+-induced Ca2+ release refractoriness in heart muscle. *Circ. Res.* 95, 807–813. doi: 10.1161/01.RES.0000146029.80463.7d
- Thomsen, M., Oros, A., Schoenmakers, M., van Opstal, J., Maas, J., Beekman, J., et al. (2007). Proarrhythmic electrical remodelling is associated with increased beat-to-beat variability of repolarisation. *Cardiovasc. Res.* 73, 521–530. doi: 10.1016/j.cardiores.2006.11.025
- Thomsen, M. B., Verduyn, S. C., Stengl, M., Beekman, J. D. M., de Pater, G., van Opstal, J., et al. (2004). Increased short-term variability of repolarization predicts d-sotalol-induced torsades de pointes in dogs. *Circulation* 110, 2453–2459. doi: 10.1161/01.CIR.0000145162.64183.C8
- Thomsen, M. B., Volders, P. G. A., Beekman, J. D. M., Matz, J., and Vos, M. A. (2006). Beat-to-Beat variability of repolarization determines proarrhythmic outcome in dogs susceptible to drug-induced torsades de pointes. J. Am. Coll. Cardiol. 48, 1268–1276. doi: 10.1016/j.jacc.2006. 05.048
- Tomaselli, G. F., and Zipes, D. P. (2004). What causes sudden death in heart failure? *Circ. Res.* 95, 754–763. doi: 10.1161/01.RES.0000145047.14691.db
- van Borren, M. M. G. J., Vos, M. A., Houtman, M. J. C., Antoons, G., and Ravesloot, J. H. (2013). Increased sarcolemmal Na+/H+ exchange activity in hypertrophied myocytes from dogs with chronic atrioventricular block. *Front. Physiol.* 4:322. doi: 10.3389/fphys.2013.00322
- Van de Water, A., Verheyen, J., Xhonneux, R., and Reneman, R. S. (1989). An improved method to correct the QT interval of the electrocardiogram for

changes in heart rate. J. Pharmacol. Methods 22, 207-217. doi: 10.1016/0160-5402(89)90015-6

- Verdonck, F., Volders, P. G. A., Vos, M. A., and Sipido, K. R. (2003). Increased Na+ concentration and altered Na/K pump activity in hypertrophied canine ventricular cells. *Cardiovasc. Res.* 57, 1035–1043. doi: 10.1016/S0008-6363(02) 00734-4
- Verduyn, S. C., Vos, M. A., van der Zande, J., Kulcsàr, A., and Wellens, H. J. (1997a). Further observations to elucidate the role of interventricular dispersion of repolarization and early afterdepolarizations in the genesis of acquired torsade de pointes arrhythmias: a comparison between almokalant and *d*-sotalol using the dog as its own control. *J. Am. Coll. Cardiol.* 30, 1575–1584. doi: 10.1016/S0735-1097(97)00333-1
- Verduyn, S. C., Vos, M. A., van der Zande, J., van der Hulst, F. F., and Wellens, H. J. (1997b). Role of interventricular dispersion of repolarization in acquired torsade-de-pointes arrhythmias: reversal by magnesium. *Cardiovasc. Res.* 34, 453–463.
- Volders, P. G., Sipido, K. R., Vos, M. A., Kulcsár, A., Verduyn, S. C., and Wellens, H. J. (1998). Cellular basis of biventricular hypertrophy and arrhythmogenesis in dogs with chronic complete atrioventricular block and acquired torsade de pointes. *Circulation* 98, 1136–1147. doi: 10.1161/01.CIR.98.11.1136
- Volders, P. G., Vos, M. A., Szabo, B., Sipido, K. R., de Groot, S. H., Gorgels, A. P., et al. (2000). Progress in the understanding of cardiac early afterdepolarizations and torsades de pointes: time to revise current concepts. *Cardiovasc. Res.* 46, 376–392. doi: 10.1016/S0008-6363(00)00022-5
- Vos, M. A., de Groot, S. H., Verduyn, S. C., van der Zande, J., Leunissen, H. D., Cleutjens, J. P., et al. (1998). Enhanced susceptibility for acquired torsade de pointes arrhythmias in the dog with chronic, complete AV block is related to cardiac hypertrophy and electrical remodeling. *Circulation* 98, 1125–1135. doi: 10.1161/01.CIR.98.11.1125
- Wada, T., Ohara, H., Nakamura, Y., Yokoyama, H., Cao, X., Izumi-Nakaseko, H., et al. (2016). Impacts of surgically performed renal denervation on the cardiovascular and electrophysiological variables in the chronic atrioventricular block dogs– comparison with those of amiodarone treatment. *Circ. J.* 80, 1556–1563. doi: 10.1253/circj.CJ-16-0198
- Zhou, S., Jung, B.-C., Tan, A. Y., Trang, V. Q., Gholmieh, G., Han, S.-W., et al. (2008). Spontaneous stellate ganglion nerve activity and ventricular arrhythmia in a canine model of sudden death. *Heart Rhythm* 5, 131–139. doi: 10.1016/j. hrthm.2007.09.007

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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