



# Genome Sequence of *Clostridium* sp. Strain P21, a CO-Fermenting Acetogen Isolated from Old Hay

D. Annie Doyle,<sup>a</sup> Kathleen E. Duncan,<sup>a</sup> Ralph S. Tanner<sup>a</sup>

<sup>a</sup>Department of Microbiology and Plant Biology, University of Oklahoma, Norman, Oklahoma, USA

**ABSTRACT** Here, we report the genome sequence of *Clostridium* sp. strain P21, isolated from old hay from Stillwater, Oklahoma. This announcement describes the generation and annotation of the 5.6-Mb genomic sequence of strain P21, which will aid in studies targeting genes involved in the enhancement of acid-alcohol production.

*Clostridium* sp. strain P21 (DSM 111390) is a Gram-positive, strictly anaerobic rod-shaped bacterium isolated from old hay from Stillwater, Oklahoma. Strain P21 was isolated in parallel with *Clostridium carboxidivorans* P7<sup>T</sup> (DSM 15243) and “*Clostridium ragsdalei*” P11 (DSM 15248) and enriched with CO at 38°C (1). Axenicity was confirmed via PCR amplification of the regions V1 to V9 of the 16S rRNA gene, as described by Weisburg et al. (2). Strain P21 was selected for whole-genome sequencing to confirm the presence of genes for the Wood-Ljungdahl pathway and acid-alcohol production.

The 16S rRNA gene sequence placed P21 within group 1 *Clostridium* with 97.9% similarity to *C. carboxidivorans* P7<sup>T</sup>, isolated from a settling lagoon (1). Growth characteristics of P21 on CO, on fructose, and on H<sub>2</sub> are in Table 1. Its growth was similar to that of related acetogens (1). Strain P21 was grown anaerobically in 10 ml of basal medium containing fructose, as described by Liou et al. (1) and centrifuged for 20 min at 6,000 × *g*. Cells were washed, and RLA lysis buffer (Maxwell 16; Promega) was added. DNA extraction was performed using the automated Maxwell 16 tissue LEV total RNA purification kit (Promega) purification system v4.90 modified from the manufacturer's instructions as described by Oldham et al. (3). DNA library preparation and genome sequencing were performed at Oklahoma Memorial Research Foundation Next-Generation Sequencing Core using the NEBNext Ultra II library prep kit (New England BioLabs) and MiSeq reagent kit v3 (Illumina) on an Illumina MiSeq instrument as per the manufacturer's instructions. Through the Department of Energy Systems Biology Knowledgebase (KBase) data platform (4), paired-end sequencing reads were trimmed using Trimmomatic v0.36 (5) with TruSeq3-PE, TruSeq2-PE, and NexteraPE-PE adapter files. Further adapters were removed, and low-complexity reads were filtered using Cutadapt v1.18 (6) and PRINSEQ v0.20.4 (7). Quality-filtered paired-end reads were assembled using SPAdes v3.13.0 (8). Contigs with coverage of <120× were removed using Geneious Prime v2020.1.1. Default parameters were used except where noted otherwise. The draft genome consists of 67 scaffolds from 7,670,130 reads (read length, 2 × 300 bp) with a depth of coverage of 405× and an *N*<sub>50</sub> value of 223,637 bp. Gene prediction and annotation was performed using RAST v0.1.1 (9, 10) and NCBI Prokaryotic Genome Annotation Pipeline (11). The assembled draft genome is 5,645,749 bp with a G+C content of 29.45% and contains 109 RNA genes, including 21 rRNA genes (8 5S, 6 16S, and 7 23S), 80 tRNA genes, and 8 noncoding RNA (ncRNA) genes.

Genomic analysis using UniProt (12) and BLAST (13) confirmed the presence of genes in P21 that encode the Wood-Ljungdahl pathway, various acid-alcohol-producing pathways, and an Rnf-complex. We hope that this genome sequence proves useful for finding more information on the genomic potential of this acetogen for acid and alcohol production, particularly for industrial applications.

**Citation** Doyle DA, Duncan KE, Tanner RS. 2021. Genome sequence of *Clostridium* sp. strain P21, a CO-fermenting acetogen isolated from old hay. Microbiol Resour Announc 10: e00864-20. <https://doi.org/10.1128/MRA.00864-20>.

**Editor** J. Cameron Thrash, University of Southern California

**Copyright** © 2021 Doyle et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Ralph S. Tanner, [rtanner@ou.edu](mailto:rtanner@ou.edu).

**Received** 31 July 2020

**Accepted** 22 February 2021

**Published** 18 March 2021

**TABLE 1** Growth of strain P21 on CO<sup>a</sup>

Substrate	Doubling time (h)	Final A <sub>600</sub>	Acetate produced (mM)	Butyrate produced (mM)	Final pH
None <sup>b</sup>		0.05	3.7	0	6.0 <sup>c</sup>
Fructose	5.4	0.95	32.6	3.2	4.6
CO/CO <sub>2</sub>	7.8	0.79	28.1	6.5	4.4
H <sub>2</sub> /CO <sub>2</sub>	21.0	0.30	23.8	1.4	4.3

<sup>a</sup> Methods in Liou et al. (1).

<sup>b</sup> Base medium contained 1 g/liter yeast extract (1).

<sup>c</sup> Initial pH 6.1 (1).

**Data availability.** The 16S rRNA gene sequence, whole-genome sequence (WGS), and raw sequencing reads for strain P21 were deposited in GenBank and the Sequence Read Archive (SRA) under the accession numbers [MT176110](https://doi.org/10.1093/bioinformatics/btr026), [JABBNI000000000](https://doi.org/10.1093/bioinformatics/btr026), and [SRR11451959](https://doi.org/10.1093/bioinformatics/btr026). The WGS and SRA reports can be found with BioProject number [PRJNA613251](https://doi.org/10.1093/bioinformatics/btr026).

## ACKNOWLEDGMENTS

*Clostridium* sp. strain P21 was isolated during support from the USDA-CSREES Special Research Grant (award 01-34447-10302) and the Oklahoma Agricultural Experimental Station.

We thank Rahul Thunguntla and Hasan K. Atiyeh for metabolite measurements.

## REFERENCES

- Liou JS-C, Balkwill DL, Drake GR, Tanner RS. 2005. *Clostridium carboxidivivans* sp. nov., a solvent-producing clostridium isolated from an agricultural settling lagoon, and reclassification of the acetogen *Clostridium scatologenes* strain SL1 as *Clostridium drakei* sp. nov. *Int J Syst Evol Microbiol* 55:2085–2091. <https://doi.org/10.1099/ijs.0.63482-0>.
- Weisburg WG, Barns SM, Pelletier DA, Lane DJ. 1991. 16S ribosomal DNA amplification for phylogenetic study. *J Bacteriol* 173:697–703. <https://doi.org/10.1128/JB.173.2.697-703.1991>.
- Oldham AL, Drilling HS, Stamps BW, Stevenson BS, Duncan KE. 2012. Automated DNA extraction platforms offer solutions to challenges of assessing microbial biofouling in oil production facilities. *AMB Express* 2:60. <https://doi.org/10.1186/2191-0855-2-60>.
- Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J-M, Chia J-M, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, Joachimiak MP, Keegan KP, Kondo S, Kumar V, Land ML, Meyer F, Mills M, Novichkov PS, Oh T, Olsen GJ, Olson R, Parrello B, Pasternak S, Pearson E, Poon SS, Price GA, Ramakrishnan S, Ranjan P, Ronald PC, Schatz MC, Seaver SMD, Shukla M, Sutormin RA, Syed MH, Thomason J, Tintle NL, Wang D, Xia F, Yoo H, Yoo S, Yu D. 2018. KBase: the United States Department of Energy systems biology knowledgebase. *Nat Biotechnol* 36:566–569. <https://doi.org/10.1038/nbt.4163>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet J* 17:10–12. <https://doi.org/10.14806/ej.17.1.200>.
- Schmieder R, Edwards R. 2011. Quality control and preprocessing of metagenomic datasets. *Bioinformatics* 27:863–864. <https://doi.org/10.1093/bioinformatics/btr026>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- The UniProt Consortium. 2019. UniProt: a worldwide hub of protein knowledge. *Nucleic Acids Res* 47:D506–D515. <https://doi.org/10.1093/nar/gky1049>.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).