OPEN

Prognostic Significance of Hypoxia-Inducible Factor Expression in Renal Cell Carcinoma

A PRISMA-compliant Systematic Review and Meta-Analysis

Yang Fan, MD, Hongzhao Li, MD, Xin Ma, MD, Yu Gao, MD, Luyao Chen, MD, Xintao Li, MD, Xu Bao, MD, Qingshan Du, MD, Yu Zhang, MD, and Xu Zhang, MD

Abstract: The prognostic value of hypoxia-inducible factor (HIF) in renal cell carcinoma (RCC) has been evaluated in a large number of studies, but the reports were inconsistent and remained inconclusive. Therefore, we conducted a systematic review and meta-analysis to clarify the significance of HIF expression in RCC prognosis.

PubMed, Embase, Web of Science, Cochrane Library, EBSCO, Cumulative Index to Nursing and Allied Health Literature (CINAHL), and Biological Abstracts were searched for eligible studies. Hazard ratio (HR) data for overall survival (OS), cancer-specific survival (CSS), and progression-free survival (PFS) with 95% confidence interval (CI) related to the expression status of HIF-1 α or HIF-2 α detected by immunohistochemistry were all extracted. Data were combined using a random- or fixed-effects model based on the corresponding inter-study heterogeneity. Subgroup analyses were also performed.

A total of 14 studies composed of 1258 patients for HIF-1 α evaluation and 619 patients for HIF-2 α evaluation were included for further analysis. When initially analyzed as a whole, the HIF-1 α expression was not significantly correlated with OS (HR 1.637, 95% CI 0.898–2.985, P = 0.108), CSS (HR 1.110, 95% CI 0.595–2.069, P = 0.744), and PFS (HR 1.113, 95% CI 0.675–1.836, P = 0.674). Similarly, HIF-2 α expression was not significantly correlated with CSS (HR 1.597, 95% CI 0.667–3.824, P = 0.293) and PFS (HR 0.847, 95% CI 0.566–1.266, P = 0.417). However, subgroup analyses concerning subcellular localization of HIFs revealed that the high nuclear expression of HIF-1 α was significantly associated with poor OS (HR 2.014, 95% CI 1.206–3.363, P = 0.007) and the high cytoplasmic expression of HIF -2 α was significantly associated with poor CSS (HR 2.356, 95% CI 1.629–3.407, P = 0.000).

The increased nuclear expression of HIF-1 α and cytoplasmic expression of HIF-2 α indicate unfavorable prognosis in RCC patients, which may serve as biomarkers for disease management.

Editor: Giuseppe Lucarelli.

Correspondence: Xu Zhang, Department of Urology, Chinese PLA General Hospital, Beijing 100853, PR China (e-mail: xzhang@tjh.tjmu.edu.cn). YF and HL equally contributed to this study.

Funding: This work was financially supported by the National High Technology Research and Development Program ("863" Program) of China

(2012AA02101 and 2014AA020607). The authors have no conflicts of interest to disclose.

DOI: 10.1097/MD.00000000001646

(Medicine 94(38):e1646)

Abbreviations: ccRCC = clear cell renal cell carcinoma, CI = confidence interval, CSS = cancer-specific survival, DFS = disease-free survival, eIF4E = eukaryotic translation initiation factor 4E, HIF = hypoxia-inducible factor, HR = hazard ratio, IHC = immunohistochemistry, OS = overall survival, PDGF = platelet-derived growth factor, PFS = progression-free survival, PRISMA = Preferred Reporting Items for Systematic Reviews and Meta-Analyses, RCC = renal cell carcinoma, VEGF = vascular endothelial growth factor, VHL = von Hippel–Lindau.

INTRODUCTION

R enal cell carcinoma (RCC), which accounts for 2% to 3% of all adult malignancies, is one of the most prevalent urologic cancers and the second leading cause of death among its cancer type.¹ RCC is highly aggressive; ~30% of RCC patients present metastasis at initial diagnosis, and another 20% to 30% of RCC patients with clinically localized disease eventually develop metastasis even after curative nephrectomy.^{2,3} Although surgery remains the gold standard among treatment strategies for localized RCC, this method provides limited benefits to RCC patients with locally advanced or metastatic disease; in this regard, early systematic therapy is required.⁴ Considering that the current surveillance of RCC mostly relies on imaging tests,⁴ identifying novel biomarkers to stratify patients with poor prognosis in the early stage of RCC is significantly needed.

Given that clear-cell RCC (ccRCC) represents $\sim 80\%$ of RCC subtypes⁵ and loss of von Hippel-Lindau (VHL) tumor suppressor gene is found in the majority (75-85%) of ccRCC,⁶ VHL may play a central role in RCC biology. In the absence of a functional VHL protein, VHL-associated proteolysis of hypoxia-inducible factor (HIF) occurring in normoxia is lost. This behavior leads to an accumulation of HIF-1 α and HIF-2 α , as well as subsequent transcription of HIF target genes involved in angiogenesis, such as vascular endothelial growth factor (VEGF) and platelet-derived growth factor (PDGF).⁷ Although HIF-1 α and HIF-2 α exhibit 48% amino acid sequence identity and similar protein structures, they contain distinct target genes and regulatory mechanisms.^{8,9} With recent advancement in the understanding of molecular basis of RCC tumorigenesis and metastasis, many studies concerning HIF-1 α and HIF-2 α were conducted in terms of outcome prediction and potential therapeutic targets. Several studies such as Klatte et al¹⁰ and Minardi et al¹¹ directly implicated that overexpression of HIF-1 α was a critical factor in RCC development, which was associated with poor prognosis. However, Biswas et al¹² reported that HIF-2 α was more tumorigenic in RCC and others even implicated HIF-1 α as a tumor suppressor gene.^{13,14} HIF was considered an

Received: July 8, 2015; revised: August 22, 2015; accepted: August 27, 2015.

From the State Key Laboratory of Kidney Diseases, Department of Urology, Military Postgraduate Medical College, Chinese People's Liberation Army General Hospital, Beijing, People's Republic of China (YF, HL, XM, LC, XL, QD, YZ, XZ); and Medical School, Nankai University, Tianjin, People's Republic of China (XB).

Copyright © 2015 Wolters Kluwer Health, Inc. All rights reserved.

Attribution License 4.0, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. ISSN: 0025-7974

unfavorable prognostic marker in other types of tumors such as colorectal cancer¹⁵ and gynecological cancer¹⁶ using metaanalysis, but its prognosis remained inconclusive in RCC patients. Hence, we conducted a systematic review and metaanalysis of eligible studies to quantitatively evaluate the prognostic values and explore the exact roles of different HIF isoforms in RCC.

MATERIALS AND METHODS

Search Strategy

This meta-analysis was conducted following the guidelines of Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA),¹⁷ which is available in the supplementary materials (PRISMA Checklist). A literature search was performed until August 15, 2015, in PubMed, Embase, Web of Science, Cochrane Library, EBSCO, Cumulative Index to Nursing and Allied Health Literature (CINAHL), and Biological Abstracts by using the following terms with different combinations: ("HIF" or "hypoxia-inducible factor 1" or "HIF-1 α " or "endothelial PAS domain-containing protein 1" or "hypoxia-inducible factor 2" or "EPAS1" or "HIF-2 α ") and ("carcinoma" or "neoplasm" or "tumor" or "cancer" or "malignancy") and ("kidney" or "renal") and ("survival" or "prognosis"). A manual search through reference lists of relevant literature was additionally performed. The approval by an institutional review board is not required because this study was based on published studies.

Criteria for Inclusion and Exclusion

Inclusion criteria were as follows: (1) HIF expression (HIF-1 α or HIF-2 α) should be evaluated by immunohistochemistry (IHC) in RCC tissues; (2) the association between the HIF expression and prognosis of RCC should be described; and (3) study design should be clearly defined (observational study, including retrospective or prospective). Studies were excluded if they met the following criteria: (1) not written in English; (2) reviews, abstracts, case reports, meeting records, animal, or cell line studies; (3) not providing sufficient data for estimating hazard ratios (HRs) and their 95% confidence intervals (CI) of HIF expression on the prognosis of overall survival (OS), cancer-specific survival (CSS), or progression-free survival (PFS). For overlapping articles, we only retrieved the most recent and most complete studies for further analyses.

Data Extraction and Quality Assessment

Data from all the studies that met the inclusion criteria were extracted independently by 2 investigators (Y Fan and H Li), and any discrepancy was resolved by consulting a third investigator (X Ma). The basic characteristics of each study were recorded as follows: first author's last name, year of publication, country of origin, study design, pathological type, number of patients, patients' age, HIF isoforms and detecting method, source and dilution of antibodies, location of protein expression, cutoff value, adjuvant therapy, follow-up period, and survival outcomes (Table 1). The primary survival outcomes associated with HIF expression were focused on OS, CSS, and PFS. HRs with corresponding 95% CIs for each outcome were also extracted. If the HR and 95% CI were not directly available, the values can be estimated as reported by Tierney et al.¹⁸

The Newcastle–Ottawa scale was used to assess the quality of the included studies.¹⁹ Eight methodology items in each

study can be assessed with a total score of 0 to 9. Studies with scores of ≥ 6 or more were graded as high quality. The total scores of each eligible study for this meta-analysis are listed in Table 1 and Supplementary Table 1.

Statistical Analysis

For the pooled analysis of the association between HIF expression and prognosis, HR and 95% CI concerning OS, CSS, and PFS were used. An observed HR > 1 implied unfavorable prognosis for the group of high HIF expression. A heterogeneity test of pooled HRs was conducted using Cochran's Q test and Higgins I-squared statistic. An I^2 value >50% indicated significant heterogeneity among studies. In this case, either a random- $(I^2 > 50\%)$ or fixed-effects $(I^2 \le 50\%)$ model was used. HIF-1 α and HIF-2 α were analyzed independently, and subgroup analyses were conducted in terms of expression location (nuclear or cytoplasmic), targeted therapy (received or not received), and HR availability (directly or indirectly). To investigate the heterogeneous studies, a sensitivity analysis was conducted to evaluate the influence of individual studies on the stability of pooled results. Publication bias was visually assessed by funnel plot and statistically evaluated by Begg's test and Egger's test. All statistical analyses were performed using Stata 12.0 software (StatCorp, College Station, TX), and a Pvalue < 0.05 was considered significant.

RESULTS

Eligible Studies

A total of 2637 records were retrieved from the primary literature search (through electronic databases and reference lists of relevant literature), and 1398 records were excluded for duplication. After screening the titles and abstracts, 40 articles were assessed for full-text review. To avoid the heterogeneity caused by the detection method, only the studies with IHC-based evaluation were included, and articles with no relevant outcome or insufficient data to estimate HR were further excluded. Finally, a total of 14 eligible studies composed of 1258 patients for HIF-1 α evaluation and 619 patients for HIF-2 α evaluation were included in this meta-analysis.^{10–12,20–30} A flowchart of the study selection process is shown in Figure 1, and the characteristics of eligible studies are presented in Table 1.

Effects of Different HIF Isoform Expression on Survival Outcomes of RCC and Subgroup Analyses

Given that HIF-1 α and HIF-2 α exhibit distinct target genes and may exert different effects on RCC patients, we performed a meta-analysis of different HIF isoforms independently. The primary outcomes considered for pooled HRs were OS, CSS, and PFS. We did not evaluate the prognostic value of HIF-2 α on OS because only 1 study was available in this concern.

The effects of different HIF isoform expressions on RCC prognosis were initially analyzed as a whole. The HIF-1 α expression was not significantly correlated with OS (HR 1.637, 95% CI 0.898–2.985, P = 0.108; random-effects model, $I^2 = 76.5\%$, P = 0.002; Figure 2A), CSS (HR 1.110, 95% CI 0.595–2.069, P = 0.744; random-effects model, $I^2 = 85.1\%$, P = 0.000; Figure 2B), and PFS (HR 1.113, 95% CI 0.675–1.836, P = 0.674; random-effects model, $I^2 = 68.3\%$, P = 0.024; Figure 2C). Similarly, the HIF-2 α expression was not significantly correlated with CSS (HR 1.597, 95% CI

TABLE 1.	Chara	cteristic	s of Eli	igible Stud	ies in Meta-Ané	alysis											
Study	Year	Country	Study Design	Pathological Type	Number of Patients	Mean or Median Age(years)	HIF Isoform	Detecting Method	Antibody Source	Antibody Dilution	Expression Location	Definition of HIF Positive	Adjuvant Therapy	Median Follow-up (Months)	Survival Outcome	HR Q Availability 5	uality Score
Lidgren	2006 Sv	veden	К	Clear cell	176	65.3 (25–87)	HIF-1α	IHC	Novus	1:2000	C	Medium or strong	NA	152 (59–272)	I SO	ndirectly	٢
Di Cristofano	2007 Ita	ıly	К	Clear cell	105	NA	HIF-1α	IHC	Zymed	1:50	C and N	staining 50% for cytoplasmic staining and 10% for nuclear	YN	74 (5–154)	CSS I	ndirectly	٢
Klatte Minardi	2007 Ur 2008 Ita	nited States Jv	R R	Clear cell Clear cell	141 50	61 (27–88) 62.06 (38–83)	HIF-1α HIF-1α	IHC	Novus Santa Cruz	6 ug/ml 1:100	N C and N	staining 35% 25%	NA NA 15	102 (1.2–202.8) 91.66 (5–264)	DSS I DSS I	Directly	8
Minervini	2008 Ita	lly	Ч	Clear cell	174	NA	HIF-1 α	IHC	Zymed	NA 1.2000 E-1	C and N	Median level	NA	66 (5-154)	CSS 1 DEG CGG 1	ndirectly	∞ 1
Ku	2011 KG	orea	×	Clear cell	78	0./c	HIF-1α, HIF-2α	IHC	Novus	1:5000 τοτ HIF-1α, 1:150 for HIF-2α	0	w cak or strong staining	AN	9 (0-73)	PF5, C55 1	Jurectly	~
Biswas	2012 UI	×	ы	Clear cell	103 for HIF-1α and 105 for HIF-2α	64 (27–87)	HIF-1α, HIF-2α	IHC	Novus	1:25 for HIF- 1α, NA for HIF-2α	N for HIF-1α, C and N for HIF-2α	Histoscore >2 for HIF-1 α , >0 for HIF- 2α	VA	123.84	CSS 1	Directly	×
Choueiri	2013 Ui	nited States	Ж	Clear cell	65 for HIF-1α and 66 for HIF-2α	NA	HIF-lα, HIF-2α	IHC	Neomarkers for HIF-1α, NA for HIF-2α	1:10000 for HIF-1α, 1:5000 for HIF-2α	Z	Intensity score >1	Pazopanib	ΝA	PFS I	ndirectly	6
Dornbusch	2013 Gé	ermany	R	Clear cell	42 immunoexpression	64 (56–70) Sunitinib	HIF-1 α 39.6	IHC PFS	Novus Directly	1:100	Z	Median					
Garcia-Donas	2013 SF	ain	д.	Clear cell	67	66 (46–82)	HIF-2 α	IHC	Abcam for HIF-1α, Novus for HIF-2α	1:200	NA	Median					
Kroeger	2014 Un	nited States	2	Clear cell	immunoexpression 299	Sunitinib 61	21.2 ΗΙΕ-2α	PFS, OS IHC	Directly NA	7 1:15000	C and N	0% for	N A	27.6	CSS	Directly	2
0												cytoplasmic staining and 32% for nuclear					
Motzer	2014 Ur	nited States	Я	NA	149	NA	HIF-1 α	IHC	Novus	NA	z	50%	Sunitinib	24	PFS, OS 1	Directly	9
Li	2015 Cł	ina	R	Clear cell	52	58 (13-88)	HIF-1 α	IHC	ZSGB-Bio	1:50	C and N	30%	NA	25-84	OS I	ndirectly	9
Minardi	2015 Ita	ıly	К	Clear cell	148	61.03 (38-85)	HIF-1 α	IHC	Santa Cruz	1:100	C and N	25%	NA 2.	17.43 (5-327)	I SO	ndirectly	2
C = cytoplas	m, CSS=	= cancer-spe	cific survi	ival, HR =hazan	d ratio, IHC = immunol	histochemistry, h	V = nucleus	, NA=not a	tvailable, $OS = ov$	erall survival, P=	= prospective, PF	S = progression-:	free survival,]	R = retrospective.			



FIGURE 1. Flowchart of study selection. IHC = immunohistochemistry, HR = hazard ratio.

0.667–3.824, P = 0.293; random-effects model, $I^2 = 86.2\%$, P = 0.000; Figure 3A) and PFS (HR 0.847, 95% CI 0.566–1.266, P = 0.417; fixed-effects model, $I^2 = 45.9\%$, P = 0.158; Figure 3B).

The overall insignificant results and inter-study heterogeneity led us to explore the detailed information in subgroup analyses. We first stratified the studies in each analysis group as nuclear expression and cytoplasmic expression subgroups. Interestingly, a high nuclear expression of HIF-1 α was revealed to be significantly associated with poor OS (HR 2.014, 95% CI 1.206–3.363, P = 0.007; random-effect model, $I^2 = 43.3\%$, P = 0.152; Figure 2A), whereas a high cytoplasmic expression of HIF-2a was significantly associated with poor CSS (HR 2.356, 95% CI 1.629-3.407, P = 0.000; random-effects model, $I^2 = 0.0\%$, P = 0.757; Figure 3A). For the absence of obvious heterogeneity in the subgroup results above, a fixed-effects model was applied for analysis again, and the pooled HRs were 1.760 (95% CI 1.257-2.464, P=0.001) and 2.356 (95% CI 1.629–3.407, P = 0.000) for the HIF-1 α nuclear expression on OS and HIF-2 α cytoplasmic expression on CSS, respectively. Further subgroup analyses were performed in terms of targeted therapy (received or not received) and HR availability (directly or indirectly), and the association of HIF expression and survival outcomes did not reach significance for the stratification (Tables 2 and 3).

Sensitivity analysis indicated that the pooled HR was not significantly influenced if any single study was omitted in subgroup analysis for the effect of HIF-1 α nuclear expression on OS (Table 4). Given that only 2 studies were included in subgroup analysis for the effect of HIF-2 α cytoplasmic expression on CSS and inter-study heterogeneity was not detected ($I^2 = 0.0\%$, P = 0.757), sensitivity analysis was not conducted in this category.

Publication Bias

The publication bias of our meta-analysis was assessed using funnel plots and Begg's and Egger's tests. As shown in Figure 4, no evidence of significant publication bias was found.



FIGURE 2. Forest plots of HR for the association of HIF-1 α expression and survival outcomes of RCC patients and subgroup analysis in terms of different subcellular localization of HIF-1 α expression: (A) effect of HIF-1 α overexpression on OS; (B) effect of HIF-1 α overexpression on PFS. Cl = confidence interval, CSS = cancer-specific survival, HIF = hypoxia-inducible factor, HR = hazard ratio, OS = overall survival, PFS = progression-free survival, HR>1 implied unfavorable prognosis, RCC = renal cell carcinoma.

DISCUSSION

HIF isoforms, which are optimally characterized as markers mediating cellular adaptation to hypoxic stress,^{31,32} have long been associated with alterations to vessel function by transcriptionally activating specific HIF-responsive genes.³³





FIGURE 3. Forest plots of HR for the association of HIF-2 α expression and survival outcomes of RCC patients and subgroup analysis in terms of different subcellular localization of HIF-2 α expression: (A) effect of HIF-2 α overexpression on CSS; (B) effect of HIF-2 α overexpression on PFS. CI = confidence interval, CSS = cancerspecific survival, HIF = hypoxia-inducible factor, HR = hazard ratio, PFS = progression-free survival, HR>1 implied unfavorable prognosis, RCC = renal cell carcinoma.

Under normoxia, HIF is targeted and ubiquitinated by pVHL, leading to subsequent degradation by cellular proteasome. However, given that VHL inactivation was reported in ~80% of sporadic ccRCC,⁶ loss of functional pVHL impairs the destabilization of HIF isoforms even in normoxic conditions. Therefore, inappropriate activation of downstream target genes that would normally only be activated in hypoxic conditions were promoted, contributing to ccRCC development.⁷ Given that the hypoxia-inducible pathway plays a critical role in ccRCC, and clear cell histology represents the most common subtype of RCC, numerous studies were performed to assess the prognostic values of HIF for RCC patients. Nevertheless, inconsistent reports generated inconclusive results. In this regard, we conducted a meta-analysis of all eligible studies to provide a systematic review of HIF expression on the prognosis of RCC patients.

In the present study, we evaluated the prognostic values of HIF-1 α and HIF-2 α independently because of their distinct target genes and potentially nonoverlapping properties. When initially analyzed as a whole, no significant association was identified between HIF-1 α expression and OS (HR 1.637, 95% CI 0.898–2.985, P = 0.108), CSS (HR 1.110, 95% CI 0.595–2.069, P = 0.744), and PFS (HR 1.113, 95% CI 0.675–1.836,

Heterogeneity Heterogeneity Heterogeneity Subgroup Pooled HR Subgroup Pooled HR Subgroup Pooled HR Pooled HR Subgroup Pooled HR Pooled HR Pooled HR Pooled HR Analysis Pooled HR Pooled HR Pooled HR Pooled HR Pooled HR Pooled HR (95%CI) P Value			SC				S	s				ЪĘ	<i>y</i>		
Heterogeneity Heterogeneity Heterogeneity Subgroup Pooled HR Pooled HR Proled HR <th></th> <th>1</th> <th></th> <th></th>													1		
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$				Hetero	geneity				Hetero	geneity				Hetero	geneity
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	Subgroup Analysis Studies	Pooled HR (95%CI)	P Value	$I^2(\%)$	P Value	Studies	Pooled HR (95% CI)	P Value	$I^2(\%)$	P Value	Studies	Pooled HR (95% CI)	<i>P</i> Value	$I^2(\%)$	P Value
$ \begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$	Subcellular location														
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Nucleus 4	2.014(1.206 - 3.363)	0.007	43.3	0.152	4	1.802(0.874 - 3.713)	0.111	86.3	0.000	б	0.996(0.518 - 1.913)	0.990	77.3	0.012
Targeted therapy – – – – – – 0.996(0.518-1.913) 0.990 77.3 Received 1 1.340(0.850-2.110) 0.207 – 0 0 77.3 8 ceived 1 1.551(0.801-3.003) 0.193 77.3 No received 4 1.888(0.776-4.594) 0.161 82 0.001 6 1.110(0.595-2.069) 0.744 85.1 0.000 1 1.551(0.801-3.003) 0.193 – No received 4 1.888(0.776-4.594) 0.161 82 0.001 6 1.110(0.595-2.069) 0.744 85.1 0.000 1 1.551(0.801-3.003) 0.193 – Ro availability 2 1.497(0.887-2.529) 0.131 10.6 0.29 3 0.977(0.781-1.197) 0.753 90.7 0.000 1 1.206(0.51-4.800) 0.296 86.4 0.001 3 1.314(0.239-7.218) 0.753 90.7 0.000 1 1.200(0.700-2.050) 0.506 - 78.9 Indirec	Cytoplasm 1	0.720(0.490 - 1.059)	0.095			2	0.312(0.080 - 1.210)	0.092	71.4	0.061	-	1.551(0.801 - 3.003)	0.193	I	I
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Targeted therapy				I										
Not received 4 1.888(0.776-4.594) 0.161 82 0.001 6 1.110(0.595-2.069) 0.744 85.1 0.000 1 1.551(0.801-3.003) 0.193 — HR availability 2 1.497(0.887-2.529) 0.131 10.6 0.29 3 0.967(0.781-1.197) 0.759 17.6 0.297 3 1.064(0.512-2.213) 0.867 78.9 Indirectly 3 1.726(0.621-4.800) 0.296 86.4 0.001 3 1.314(0.239-7.218) 0.753 90.7 0.000 1 1.200(0.700-2.050) 0.506 —	Received 1	1.340(0.850 - 2.110)	0.207		I	0		I		I	б	0.996(0.518-1.913)	0.990	77.3	0.012
HR availability 2 1.497(0.887–2.529) 0.131 10.6 0.29 3 0.967(0.781–1.197) 0.759 17.6 0.297 3 1.064(0.512–2.213) 0.867 78.9 Directly 3 1.726(0.621–4.800) 0.296 86.4 0.001 3 1.314(0.239–7.218) 0.753 90.7 0.000 1 1.200(0.700–2.050) 0.506 —	Not received 4	1.888(0.776 - 4.594)	0.161	82	0.001	9	1.110(0.595 - 2.069)	0.744	85.1	0.000	1	1.551(0.801 - 3.003)	0.193	Ι	Ι
Directly 2 1.497(0.887-2.529) 0.131 10.6 0.29 3 0.967(0.781-1.197) 0.759 17.6 0.297 3 1.064(0.512-2.213) 0.867 78.9 Indirectly 3 1.726(6.621-4.800) 0.296 86.4 0.001 3 1.314(0.239-7.218) 0.753 90.7 0.000 1 1.200(0.700-2.050) 0.506 -	HR availability														
Indirectly 3 1.726(6.621-4.800) 0.296 864 0.001 3 1.314(0.239-7.218) 0.753 90.7 0.000 1 1.200(0.700-2.050) 0.506 -	Directly 2	1.497(0.887 - 2.529)	0.131	10.6	0.29	3	0.967(0.781-1.197)	0.759	17.6	0.297	ŝ	1.064(0.512 - 2.213)	0.867	78.9	0.009
	Indirectly 3	1.726(0.621 - 4.800)	0.296	86.4	0.001	3	1.314(0.239–7.218)	0.753	90.7	0.000	-	1.200(0.700 - 2.050)	0.506	I	I

		(CSS				1	PFS		
HIF-2α	_			Heter	ogeneity				Heter	ogeneity
Subgroup Analysis	Studies	Pooled HR (95%CI)	P Value	<i>I</i> ² (%)	P Value	Studies	Pooled HR (95%CI)	P Value	$I^{2}(\%)$	P Value
Subcellular location										
Nucleus	2	1.125 (0.313-4.050)	0.857	87.2	0.005	1	1.110 (0.680-1.830)	0.760	_	_
Cytoplasm	2	2.356 (1.629-3.407)	0	0	0.757	1	0.293 (0.039-2.173)	0.213	_	_
Targeted therapy										
Received	0	_	_	_	_	2	0.885 (0.587-1.334)	0.559	61.3	0.108
Not received	4	1.597 (0.667-3.824)	0.293	86.2	0	1	0.293 (0.039-2.173)	0.213	_	_
HR availability										
Directly	4	1.597 (0.667-3.824)	0.293	86.2	0	2	1.029 (0.636-1.663)	0.908	37.1	0.207
Indirectly	0		_	_	_	1	1.537 (0.260-1.130)	0.097	_	_

TABLE 3. Subgroup Analysis of Pooled HR for RCC Patients With HIF-2 α Overexpression

TABLE 4. Pooled HR (95% CI) of Sensitivity Analysis for Effect of HIF-1 α Nuclear Expression on OS

Study Omitted	Pooled HR	Lower Limit of 95% CI	Upper Limit of 95% CI
Minardi (2008)	1.702	1.202	2.410
Motzer (2014)	2.447	1.484	4.034
Li (2015)	1.599	1.126	2.271
Minardi (2015)	1.685	1.125	2.525
Combined	1.760	1.257	2.464

CI = confidence interval, HR = hazard ratio, OS = overall survival.

P = 0.674), although patients with high HIF-1 α level tended to show a relatively unfavorable prognosis. Similar insignificant results were obtained for HIF-2 α between high and low expression in terms of CSS (HR 1.597, 95% CI 0.667–3.824, P = 0.293) and PFS (HR 0.847, 95% CI 0.566–1.266, P = 0.417).

Subgroup analysis was performed to explain the heterogeneity of pooled HRs and reveal detailed information of the HIF function. Considering that recent evidence indicated that different subcellular localizations of HIF expression might exert different effects on RCC^{21,24} and the same mechanism was shown in other proteins, such as p21,³⁴ we performed subgroup analysis by first stratifying studies as the nuclear expression group or cytoplasmic expression group. The high nuclear expression of HIF-1 α was discovered to be significantly associated with poor OS (HR 2.014, 95% CI 1.206-3.363, P = 0.007), whereas patients with high cytoplasmic expression of HIF-1a tended to demonstrate a more favorable prognosis in terms of OS, though not reaching a statistical significance (HR 0.720, 95% CI 0.490–1.059, P = 0.095). The same conflicting trend was also observed in the subgroup analyses of the effects of nuclear and cytoplasmic HIF-1 α expression on CSS, although the difference was not significant. HIF-1 α was considered an unfavorable prognostic marker in various cancers, ^{15,16,35} but the underlying mechanism of the opposing effects of HIF-1a overexpression in different subcellular compartments remains



FIGURE 4. Funnel plots, Begg and Egger' test results for the evaluation of publication bias. (A) HIF-1 α and OS, (B) HIF-1 α and CSS, (C) HIF-1 α and PFS, (D) HIF-2 α and CSS, (E) HIF-2 α and PFS. CSS = cancer-specific survival, HIF = hypoxia-inducible factor, HR = hazard ratio, OS = overall survival, PFS = progression-free survival.

speculative. A possible explanation indicates that HIF-1 α is a nuclear transcription factor that is functional only in the nucleus, and a high cytoplasmic expression of HIF-1 α suggests that this factor has translocated to the cytoplasm and thus unable to activate HIF-responsive genes, leading to a relatively good prognosis.²¹ For HIF-2 α , subgroup analysis revealed that high cytoplasmic expression of HIF-2 α was significantly associated with poor CSS (HR 2.356, 95% CI 1.629–3.407, P=0.000). Accumulating evidence demonstrates that HIF-2 α , rather than HIF-1 α , is the primary oncogenic driver in RCC and can act as a nuclear tumor promoter.^{36,37} However, unlike HIF-1 α , HIF-2 α plays a unique role in the tumor cytoplasm. Uniacke et al³⁸ demonstrated that HIF-2 α can form a complex in the cytoplasm to help initiate protein synthesis in periods of oxygen scarcity and eukaryotic translation initiation factor 4E (eIF4E) inhibition. This mechanism concerning HIF-2 α function in the cytoplasm is very important for tumor development as it evaded hypoxia-induced repression of protein translation, which can explain the association between HIF-2 α with high cytoplasmic expression and unfavorable prognosis in RCC patients.

Given that the HIF expression is associated with the alteration in the VEGF pathway, the HIF expression levels may provide therapeutic implications in the selection of patients who will benefit from new targeted antiangiogenic therapies. Unfortunately, our subgroup analyses for patients who received tyrosine kinase inhibitors (Sunitinib or Pazopanib) did not show any significant benefit for survival outcomes in terms of different levels of HIFs, even though several single studies reached significant statistics with opposing results.^{22,23,30} This discrepancy and insignificant pooled results may be attributed to limited studies and different follow-up time in each study. Thus, future investigations with large cohort and sufficient follow-up are warranted to reach a consensus on patient selection for targeted therapy based on the HIF expression.

The present study comprehensively evaluated the effect of HIFs on the prognosis of RCC patients, but several limitations should be noted. First, although IHC is the standard method to evaluate the HIF expression, the cutoff value for high or low levels of HIFs varied in different studies, which might cause heterogeneity of the overall results. Therefore, a more unanimous cutoff value for the definition of high HIF expression is recommended for future studies. Second, a remarkable heterogeneity was observed in certain categories of analysis. Although the heterogeneity was attributed by subgroup analysis to the subcellular location of HIF expression, this basis cannot cover the entire source. Third, several studies included in the analysis did not directly provide HRs and should be calculated using the methods recommended by Tierney et al,¹⁸ which somehow rendered the extracted data less precise. Fourth, relatively few studies were included in this meta-analysis and it could lead to a premature result. So it may need an update in the future when more eligible studies are published. Finally, as a literature-based analysis, the fact that studies with positive results were more likely to be published can amplify the association between HIF expression and survival outcomes, which can lead to publication bias,³⁹ although this bias was not detected in the current analysis.

In conclusion, this meta-analysis is the first to evaluate the influence of the expression of different HIF isoforms on the prognosis of RCC patients. This study showed that high nuclear expression of HIF-1 α and high cytoplasmic expression of HIF-2 α indicate unfavorable prognosis in patients with RCC, which may potentially serve as risk stratification markers and even therapeutic targets to manage this disease.

ACKNOWLEDGMENTS

We thank Dr. Zhi Mao in Intensive Care Unit of PLA General Hospital for assisting the manuscript revision.

REFERENCES

- Jemal A, Bray F, Center MM, et al. Global cancer statistics. CA Cancer J Clin. 2011;61:69–90.
- Bukowski RM. Natural history and therapy of metastatic renal cell carcinoma: the role of interleukin-2. *Cancer*. 1997;80:1198–1220.
- Zisman A, Pantuck AJ, Wieder J, et al. Risk group assessment and clinical outcome algorithm to predict the natural history of patients with surgically resected renal cell carcinoma. *J Clin Oncol.* 2002;20: 4559–4566.
- Rini BI, Campbell SC, Escudier B. Renal cell carcinoma. *Lancet*. 2009;373:1119–1132.
- Zbar B, Klausner R, Linehan WM. Studying cancer families to identify kidney cancer genes. *Annu Rev Med.* 2003;54:217–233.
- Kaelin WG. Von Hippel-Lindau disease. Annu Rev Pathol. 2007;2:145–173.
- Gossage L, Eisen T. Alterations in VHL as potential biomarkers in renal-cell carcinoma. Nat Rev Clin Oncol. 2010;7:277–288.
- Hu CJ, Wang LY, Chodosh LA, et al. Differential roles of hypoxiainducible factor lalpha (HIF-lalpha) and HIF-2alpha in hypoxic gene regulation. *Mol Cell Biol.* 2003;23:9361–9374.
- Keith B, Johnson RS, Simon MC. HIF1alpha and HIF2alpha: sibling rivalry in hypoxic tumour growth and progression. *Nat Rev Cancer*. 2012;12:9–22.
- Klatte T, Seligson DB, Riggs SB, et al. Hypoxia-inducible factor 1 alpha in clear cell renal cell carcinoma. *Clin Cancer Res.* 2007;13:7388–7393.
- Minardi D, Lucarini G, Santoni M, et al. Survival in patients with clear cell renal cell carcinoma is predicted by HIF-1alpha expression. *Anticancer Res.* 2015;35:433–438.
- Biswas S, Charlesworth PJ, Turner GD, et al. CD31 angiogenesis and combined expression of HIF-1alpha and HIF-2alpha are prognostic in primary clear-cell renal cell carcinoma (CC-RCC), but HIF alpha transcriptional products are not: implications for antiangiogenic trials and HIF alpha biomarker studies in primary CC-RCC. *Carcinogenesis.* 2012;33:1717–1725.
- Shen C, Beroukhim R, Schumacher SE, et al. Genetic and functional studies implicate HIF1alpha as a 14q kidney cancer suppressor gene. *Cancer Discov.* 2011;1:222–235.
- Gordan JD, Simon MC. Hypoxia-inducible factors: central regulators of the tumor phenotype. *Curr Opin Genet Dev.* 2007;17:71–77.
- Chen Z, He X, Xia W, et al. Prognostic value and clinicopathological differences of HIFs in colorectal cancer: evidence from metaanalysis. *PLoS One*. 2013;8:e80337.
- Jin Y, Wang H, Ma X, et al. Clinicopathological characteristics of gynecological cancer associated with hypoxia-inducible factor lalpha expression: a meta-analysis including 6,612 subjects. *PLoS One*. 2015;10:e0127229.
- Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med.* 2009;6:e1000097.
- Tierney JF, Stewart LA, Ghersi D, et al. Practical methods for incorporating summary time-to-event data into meta-analysis. *Trials.* 2007;8:16.
- Stang A. Critical evaluation of the Newcastle–Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur J Epidemiol.* 2010;25:603–605.
- Choueiri TK, Fay AP, Gagnon R, et al. The role of aberrant VHL/ HIF pathway elements in predicting clinical outcome to pazopanib

therapy in patients with metastatic clear-cell renal cell carcinoma. *Clin Cancer Res.* 2013;19:5218–5226.

- Di Cristofano C, Minervini A, Menicagli M, et al. Nuclear expression of hypoxia-inducible factor-lalpha in clear cell renal cell carcinoma is involved in tumor progression. Am J Surg Pathol. 2007;31:1875–1881.
- Dornbusch J, Zacharis A, Meinhardt M, et al. Analyses of potential predictive markers and survival data for a response to sunitinib in patients with metastatic renal cell carcinoma. *PLoS One.* 2013;8:e76386.
- 23. Garcia-Donas J, Leandro-Garcia LJ, Gonzalez Del Alba A, et al. Prospective study assessing hypoxia-related proteins as markers for the outcome of treatment with sunitinib in advanced clear-cell renal cell carcinoma. *Ann Oncol.* 2013;24:2409–2414.
- 24. Kroeger N, Seligson DB, Signoretti S, et al. Poor prognosis and advanced clinicopathological features of clear cell renal cell carcinoma (ccRCC) are associated with cytoplasmic subcellular localisation of Hypoxia inducible factor-2alpha. *Eur J Cancer.* 2014;50:1531–1540.
- Ku JH, Park YH, Myung JK, et al. Expression of hypoxia inducible factor-1alpha and 2 alpha in conventional renal cell carcinoma with or without sarcomatoid differentiation. *Urol Oncol.* 2011;29:731–737.
- Li W, Huang J, Chen J, et al. Expression and significance of F0XP1, HIF-la and VEGF in renal clear cell carcinoma. *J BUON*. 2015;20:188–195.
- Lidgren A, Hedberg Y, Grankvist K, et al. Hypoxia-inducible factor lalpha expression in renal cell carcinoma analyzed by tissue microarray. *Eur Urol.* 2006;50:1272–1277.
- Minardi D, Lucarini G, Filosa A, et al. Prognostic role of tumor necrosis, microvessel density, vascular endothelial growth factor and hypoxia inducible factor-1alpha in patients with clear cell renal carcinoma after radical nephrectomy in a long term follow-up. *Int J Immunopathol Pharmacol.* 2008;21:447–455.
- 29. Minervini A, Di Cristofano C, Gacci M, et al. Prognostic role of histological necrosis for nonmetastatic clear cell renal cell carci-

noma: correlation with pathological features and molecular markers. *J Urol.* 2008;180:1284–1289.

- 30. Motzer RJ, Hutson TE, Hudes GR, et al. Investigation of novel circulating proteins, germ line single-nucleotide polymorphisms, and molecular tumor markers as potential efficacy biomarkers of firstline sunitinib therapy for advanced renal cell carcinoma. *Cancer Chemother Pharmacol.* 2014;74:739–750.
- Wang GL, Jiang BH, Rue EA, et al. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O2 tension. *Proc Natl Acad Sci U S A*. 1995;92:5510–5514.
- Semenza GL. Hypoxia-inducible factors in physiology and medicine. *Cell.* 2012;148:399–408.
- 33. Sutter CH, Laughner E, Semenza GL. Hypoxia-inducible factor lalpha protein expression is controlled by oxygen-regulated ubiquitination that is disrupted by deletions and missense mutations. *Proc Natl Acad Sci U S A*. 2000;97:4748–4753.
- Weiss RH, Borowsky AD, Seligson D, et al. p21 is a prognostic marker for renal cell carcinoma: implications for novel therapeutic approaches. J Urol. 2007;177:63–68discussion 68–69.
- Chen J, Li T, Liu Q, et al. Clinical and prognostic significance of HIF-1alpha, PTEN, CD44v6, and survivin for gastric cancer: a metaanalysis. *PLoS One*. 2014;9:e91842.
- Kaelin WG Jr, Ratcliffe PJ. Oxygen sensing by metazoans: the central role of the HIF hydroxylase pathway. *Mol Cell*. 2008;30:393–402.
- Kondo K, Kim WY, Lechpammer M, et al. Inhibition of HIF2alpha is sufficient to suppress pVHL-defective tumor growth. *PLoS Biol.* 2003;1:E83.
- Uniacke J, Holterman CE, Lachance G, et al. An oxygen-regulated switch in the protein synthesis machinery. *Nature*. 2012;486:126–129.
- Sutton AJ, Song F, Gilbody SM, et al. Modelling publication bias in meta-analysis: a review. Stat Methods Med Res. 2000;9:421–445.