



Cytokine Pathways and Investigational Target Therapies in Hidradenitis Suppurativa

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Abstract: Background: Hidradenitis suppurativa (HS) is a chronic inflammatory skin disease affecting areas with a high density of apocrine glands and characterized by subcutaneous nodules that may evolve into fistulas with pus secretion. Methods: The aim of this review is to investigate all current knowledge on cytokine regulation in the pathogenesis of HS. A systematic literature research using the words "cytokine", "interleukin", "pathway", and "hidradenitis suppurativa" was performed in PubMed/Medline and Scopus/Embase databases. A search of the clinicaltrials.gov website for interventional recruiting and completed trials including the term "hidradenitis suppurativa" was also performed up to August 2020. We will discuss the pathogenetic role of various cytokines in HS and potential therapeutic targets for this debilitating disease. Results: The pathophysiology underlying this complex condition has not been clearly defined. An upregulation of various cytokines, such as tumor necrosis factor alpha (TNF- α), interleukin (IL)-1, IL-17, IL-23, and other molecules seems to be related to this inflammatory condition. Various cells, such as lymphocytes T Helper 1 and 17 and keratinocytes seem to be involved in the genesis of this condition. Conclusions: Several future studies and clinical trials are necessary in order to have new knowledge about HS and to properly treat this complex condition.

Keywords: hidradenitis suppurativa; cytokine; interleukin

1. Introduction

Hidradenitis suppurativa (HS) is a chronic inflammatory condition primarily affecting apocrine-gland-rich regions of the body such as the axillary and groin areas [1,2]. HS presents with painful nodules and abscesses that may coalesce and form fistulas where the pus may drain. Lesions often evolve into scars, with a high physical and psychological impact for the patients [3]. Various therapies have been proposed to treat HS. Unfortunately, no therapy has been fully successful in the control of the disease [4–10]. Nowadays, the most effective treatment remains surgery [11]. To better define the possible treatments for this disorder, it is fundamental to assess the cytokine pathways involved in this condition, in order to develop new drugs that may lead to a better control of the disease. In this work, we are going to analyze all the possible cytokine pathways involved in the development of HS and which of these cytokines may be used as a possible target in the development of new drugs [12]. In this review, we aim discuss the pathogenetic role of various cytokines in HS and potential therapeutic targets attempted or currently under investigation for this debilitating disease.

2. Materials and Methods

A systematic literature research was performed in PubMed/Medline, Scopus/Embase, and Google Scholar, in order to find articles suitable to be inserted in this review. Keywords used included "interleukin", "cytokine", "pathway", and "hidradenitis suppurativa". Duplicate articles were discarded before a full reading. Articles that did not bring any new information were excluded after a full reading. Article selection was performed independently by two researchers (L.B. and P.M.). Whenever discrepancies arose, a resolution was achieved by discussion with a third independent author (E.D.D.). The article selection flowchart is better described in Figure 1.



Figure 1. Article selection flowchart.

A search of the website clinicaltrials.gov for interventional recruiting and completed clinical trials with the term "hidradenitis suppurativa" was also performed up to 31 August 2020.

3. Results

A total of 197 non-duplicated citations were identified in the literature review (Figure 1). Eighty of these articles were removed upon review of titles and abstracts against the pre-defined eligibility criteria. Seventeen references were further excluded because they did not add any new information (Figure 1).

A total of 58 clinical trials were retrieved by a search of the clinical trials.gov database with the term "hidradenitis suppurativa" performed on 31 August 2020. Of these trials, 35 were completed and 23 were still recruiting patients. Twenty-seven completed and eight recruiting trials were utilizing immuno-modulatory treatments. Clinical trials' characteristics are listed in Tables 1 and 2.

| NCT Clinical Trial | Intervention | Phase | Study Design | Enrollment |
|--------------------|---------------------|---------|--|------------|
| NCT03512275 [13] | Bermekimab 400 mg | Phase 2 | •Allocation: Non-Randomized •Intervention Model: Single | 42 |
| | | | •Masking: None (Open Label) | |
| NCT03960268 [14] | Brodalumab | Phase 1 | •Allocation: N/A | 10 |
| | | | Intervention Model: Single | |
| | | | Group Assignment | |
| | | | Masking: None (Open Label) | |
| NCT03607487 [15] | INCB054707 | Phase 2 | •Allocation: Randomized | 36 |
| | Placebo | | •Intervention Model: Parallel | |
| | | | •Masking: Triple | |
| NCT03569371 [16] | INCB054707 | Phase 2 | •Allocation: N/A | 10 |
| | | 1111002 | •Intervention Model: Single | 10 |
| | | | Group Assignment | |
| | | | Masking: None (Open Label) | |
| NCT03248531 [17] | Bimekizumab | Phase 2 | •Allocation: Randomized | 90 |
| | Adalimumab | | •Intervention Model: Parallel | |
| | Dlacabo | | Assignment | |
| | riacebo | | •Allocation: N/A | |
| | | | •Intervention Model: Single | |
| NCI01516749 [18] | Anakinra | Phase 2 | Group Assignment | 6 |
| | | | •Masking: None (Open Label) | |
| NCT02421172 [19] | CJM112 | Phase 2 | Allocation: Randomized | 66 |
| | Placebo | | •Intervention Model: Parallel | |
| | | | Assignment | |
| | | | Investigator) | |
| NCT00795574 [20] | Infliximab | Phase 2 | •Allocation: Randomized | 38 |
| | Dlaasha Commanator | | Intervention Model: Crossover | |
| | Placebo Comparator | | Assignment | |
| | - | | Masking: Quadruple | |
| NICT00220822 [21] | Etanercept sc 50 mg | Dhase 2 | Allocation. Non Dandomized | 10 |
| NC100329623 [21] | weeks | rnase z | •Anocation: Non-Kandonnized | 10 |
| | weeks | | •Intervention Model: Single | |
| | | | Group Assignment | |
| | | | Masking: None (Open Label) | |
| NCT03628924 [22] | Guselkumab dose 1 | Phase 2 | •Allocation: Randomized | 184 |
| | Guselkumab dose 2 | | •Intervention Model: Parallel | |
| | | | Assignment Masking: Double (Participant | |
| | Guselkumab dose 3 | | Investigator) | |
| NCT03001622 [23] | IFX-1 | Phase 2 | •Allocation: N/A | 12 |
| | | | Intervention Model: Single | |
| | | | Group Assignment | |
| | A '1 I | | •Masking: None (Open Label) | 20 |
| NC103049267 [24] | Apremilast | Phase 2 | •Allocation: Kandomized | 20 |
| | Placebo Oral Tablet | | Assignment | |
| | | | •Masking: Double (Participant. | |
| | | | Investigator) | |
| | | | •Allocation: N/A | |
| NCT03099980 [25] | Secukinumab | Phase 1 | •Intervention Model: Single | 20 |
| | | | Group Assignment | |
| | | | •masking: mone (Open Label) | |

 Table 1. Completed interventional studies on target therapies.

| Table | 1. C | ont. |
|-------|-------------|------|
|-------|-------------|------|

| NCT00107991 [26] Etanercept Phase 2 •Allocation: N/A 15 Intervention Model: Single Group Assignment •Masking: None (Open Label) •Allocation: N/A 15 NCT02643654 [28] MABp1 Phase 3 •Allocation: N/A 15 NCT02643654 [28] MABp1 Phase 2 •Allocation: N/A 15 NCT02643654 [28] MABp1 Phase 2 •Allocation: Randomized 20 NCT02695212 [29] Apremilast Phase 2 •Allocation: N/A 15 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 NCT013487276 [31] IFX-1 Phase 2 •Allocation: N/A 20 NCT01558375 [32] Anakinra Phase 2 •Allocation: N/A 20 NCT01635764 [33] Adalimumab Phase 3 •Allocation: N/A 179 NCT02808975 [34] Adalimumab Phase 4 •Allocation: N/A 179 NCT01468207 [36] Adalimumab Phase 4 •Allocation: N/A 179 NCT01468207 [36] Adalimumab Phase 3 •Allocation: Randomized 20 NCT01468207 [36] Adalimumab | NCT Clinical Trial | Intervention | Phase | Study Design | Enrollment |
|--|--------------------|---------------------|-------------------|--|------------|
| NCT02904902 [27] Adalimumab Phase 3 • Masking: None (Open Label) NCT02643654 [28] MABp1 Phase 2 Placebo • • Allocation: N/A 15 • Intervention Model: Single Group Assignment • Masking: None (Open Label) Apremilast Phase 2 • Allocation: N/A 20 • Intervention Model: Single Group Assignment • Masking: Quadruple • Allocation: N/A 20 • Intervention Model: Single Group Assignment • Masking: Quadruple • Allocation: N/A 20 • Intervention Model: Single Group Assignment • Masking: Quadruple • Allocation: N/A 20 • Intervention Model: Single Group Assignment • Masking: Quadruple • Allocation: N/A 20 • Intervention Model: Single Group Assignment • Masking: None (Open Label) NCT01704534 [30] Ustekinumab Phase 2 • Allocation: N/A 20 • Intervention Model: Single Group Assignment • Masking: Quadruple • Allocation: NA 20 • Intervention Model: Single Group Assignment • Masking: Quadruple • Allocation: Randomized 179 • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: Randomized 20 • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: NA • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: NA • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: Randomized 20 • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: Randomized 20 • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: Randomized 206 • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: Randomized 206 • Intervention Model: Parallel Assignment • Masking: Quadruple • Allocation: Randomized 307 • Intervention Model: Parallel Assignment • Masking: Double (Participant, Investigator) NCT01468233 [37] Adalimumab Phase 3 • Allocation: Randomized 326 | NCT00107991 [26] | Etanercept | Phase 2 | •Allocation: N/A •Intervention Model: Single | 15 |
| NCT02904902 [27] Adalimumab Phase 3 Allocation: N/A 15 NCT02643654 [28] MABp1 Phase 2 Intervention Model: Single Group Assignment 20 NCT02643654 [28] MABp1 Phase 2 Intervention Model: Parallel Assignment 20 NCT02643654 [28] MABp1 Phase 2 Intervention Model: Parallel Assignment 20 NCT02695212 [29] Apremilast Phase 2 Intervention Model: Single Group Assignment 20 NCT01704534 [30] Ustekinumab Phase 2 Intervention Model: Single Group Assignment 20 NCT01487276 [31] IFX-1 Phase 2 Allocation: N/A 20 Placebo Allocation: N/A 20 NCT01558375 [32] Anakinra Phase 2 Allocation: Randomized 179 NCT01635764 [33] Adalimumab Phase 3 Intervention Model: Parallel Assignment Assignment NCT02808975 [34] Adalimumab Phase 4 Intervention Model: Single Group Assignment 206 NCT00918255 [35] Adalimumab Phase 2 Intervention Model: Parallel Assignment 307 NCT01468207 [36] Adalimumab Phase 3 <td< td=""><td></td><td></td><td></td><td>Group Assignment Masking: None (Open Label)</td><td></td></td<> | | | | Group Assignment Masking: None (Open Label) | |
| NCT02643654 [28] MABp1 Phase 2 •Intervention Model: Single Group Assignment 20 NCT02643654 [28] MABp1 Phase 2 •Allocation: Randomized 20 Placebo •Allocation: Randomized 20 •Intervention Model: Parallel Assignment •Masking: Quadruple 20 •Allocation: N/A •Intervention Model: Single Group Assignment 20 NCT02695212 [29] Apremilast Phase 2 •Intervention Model: Single Group Assignment 20 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 •Intervention Model: Single Group Assignment 20 •Allocation: N/A 20 •NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 •Intervention Model: Single Group Assignment •Masking: None (Open Label) 179 NCT0158875 [32] Anakinra Phase 2 •Allocation: Randomized 20 NCT01635764 [33] Adalimumab Phase 3 •Intervention Model: Parallel Assignment 508 NCT02808975 [34] Adalimumab Phase 4 •Allocation: Randomized 20 NCT01468207 [36] Adalimumab Phase 4 <t< td=""><td>NCT02904902 [27]</td><td>Adalimumab</td><td>Phase 3</td><td>•Allocation: N/A</td><td>15</td></t<> | NCT02904902 [27] | Adalimumab | Phase 3 | •Allocation: N/A | 15 |
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| NCT02643654 [28] MABp1 Phase 2 Placebo Placebo Placeb | | | | Group Assignment | |
| NCT02695054 [25] MPAP1 Placebo •Intervention Model: Parallel Assignment 20 Placebo •Intervention Model: Single Group Assignment 20 NCT02695212 [29] Apremilast Phase 2 •Intervention Model: Single Group Assignment 20 NCT01704534 [30] Ustekinumab Phase 2 •Masking: None (Open Label) 0 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 NCT01704534 [31] IFX-1 Phase 2 •Allocation: Randomized 179 Placebo •Intervention Model: Single Group Assignment 179 •Intervention Model: Parallel Assignment 20 NCT01558375 [32] Anakinra Phase 2 •Allocation: Randomized 20 Water for injection •Intervention Model: Single Group Assignment 508 NCT01635764 [33] Adalimumab Phase 3 •Intervention Model: Parallel Assignment 508 NCT00918255 [35] Adalimumab Phase 4 •Allocation: Randomized 206 Placebo Placebo •Allocation: Randomized 154 | NCT02642654 [28] | MAD. 1 | Dhase 2 | •Masking: None (Open Label) | 20 |
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| NCT02695212 [29] Apremilast Phase 2 Intervention Model: Single Group Assignment 20 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 NCT01704534 [30] Ustekinumab Phase 2 •Allocation: N/A 20 NCT03487276 [31] IFX-1 Phase 2 •Allocation: Randomized 179 Placebo •Allocation: Randomized 179 NCT01558375 [32] Anakinra Phase 2 •Allocation: Randomized 20 NCT01635764 [33] Adalimumab Phase 3 •Allocation: Randomized 20 NCT01635764 [33] Adalimumab Phase 3 •Allocation: N/A 20 NCT01635764 [33] Adalimumab Phase 4 •Allocation: Randomized 206 NCT002808975 [34] Adalimumab Phase 4 •Allocation: Randomized 206 NCT00918255 [35] Adalimumab Phase 2 •Allocation: Randomized 154 NCT01468207 [36] Adalimumab Phase 3 •Allocation: Randomized 307 NCT01468207 [36] Adali | | | | •Allocation: N/A | |
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| NCT01704534 [30] Ustekinumab Phase 2 Allocation: N/A 20 NCT01704534 [30] Ustekinumab Phase 2 Allocation: N/A 20 NCT03487276 [31] IFX-1 Phase 2 Intervention Model: Single Group Assignment 179 NCT03487276 [31] IFX-1 Phase 2 Allocation: Randomized 179 Placebo Intervention Model: Parallel Assignment Assign: Quadruple 20 NCT01558375 [32] Anakinra Phase 2 Intervention Model: Parallel Assignment 20 Water for injection Water for injection Intervention Model: Single Group Assignment 20 NCT01635764 [33] Adalimumab Phase 3 Intervention Model: Single Group Assignment 508 NCT02808975 [34] Adalimumab Phase 4 Allocation: Randomized 206 NCT00918255 [35] Adalimumab Phase 2 Intervention Model: Parallel Assignment 508 NCT01468207 [36] Adalimumab Phase 3 Allocation: Randomized 206 NCT01468207 [36] Adalimumab Phase 3 Intervention Model: Parallel Assignment 307 NCT01468203 [37] Adalimumab Phase 3 | | 1 | | Group Assignment | |
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| Group Assignment•Masking: None (Open Label)NCT03487276 [31]IFX-1Phase 2•Allocation: Randomized179•Placebo•Allocation: Randomized179•Intervention Model: ParallelAssignment•Masking: Quadruple•Allocation: Randomized20NCT01558375 [32]AnakinraPhase 2•Allocation: Randomized20Water for injection•Masking: Quadruple•Allocation: Nandomized20NCT01635764 [33]AdalimumabPhase 3•Intervention Model: Single Group Assignment508NCT02808975 [34]AdalimumabPhase 4•Allocation: Randomized206Placebo•Intervention Model: Parallel Assignment206NCT00918255 [35]AdalimumabPhase 2•Allocation: Randomized206NCT01468207 [36]AdalimumabPhase 3•Allocation: Randomized154NCT01468207 [36]AdalimumabPhase 3•Allocation: Randomized307NCT01468233 [37]AdalimumabPhase 3•Allocation: Randomized326 | NC101704004 [00] | Ostekiitulliad | 11111111111111111 | •Intervention Model: Single | 20 |
| NCT03487276 [31]IFX-1Phase 2•Masking: None (Open Label) •Allocation: Randomized179Placebo•Intervention Model: Parallel Assignment179NCT01558375 [32]AnakinraPhase 2•Allocation: Randomized20Water for injection•Masking: Quadruple20Water for injection•Masking: Quadruple20NCT01635764 [33]AdalimumabPhase 3•Intervention Model: Parallel Assignment508NCT01635764 [33]AdalimumabPhase 3•Intervention Model: Single Group Assignment508NCT02808975 [34]AdalimumabPhase 4•Allocation: N/APlacebo•Allocation: Randomized206NCT00918255 [35]AdalimumabPhase 2•Allocation: RandomizedNCT01468207 [36]AdalimumabPhase 3•Allocation: Randomized154Placebo•Masking: Quadruple·Allocation: Randomized307NCT01468207 [36]AdalimumabPhase 3•Allocation: Randomized307NCT01468233 [37]AdalimumabPhase 3•Allocation: Randomized326 | | | | Group Assignment | |
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| Placebo•Intervention Model: Parallel AssignmentNCT01558375 [32]AnakinraPhase 2•Allocation: Randomized20Water for injection•Masking: Quadruple •Allocation: Randomized20NCT01635764 [33]AdalimumabPhase 3•Intervention Model: Parallel Assignment •Masking: Quadruple •Allocation: N/ANCT01635764 [33]AdalimumabPhase 3•Intervention Model: Single Group Assignment508NCT02808975 [34]AdalimumabPhase 4•Allocation: Randomized206Placebo•Masking: Quadruple •Masking: None (Open Label)206NCT00918255 [35]AdalimumabPhase 2•Allocation: Randomized206NCT01468207 [36]AdalimumabPhase 3•Allocation: Randomized154NCT01468207 [36]AdalimumabPhase 3•Allocation: Randomized307Placebo•Masking: Quadruple Assignment307•Intervention Model: Parallel Assignment307NCT01468233 [37]AdalimumabPhase 3•Allocation: Randomized326 | NCT03487276 [31] | IFX-1 | Phase 2 | Allocation: Randomized | 179 |
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| Intervention Model: Parallel | | placebo | | •Intervention Model: Parallel | |
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| •Masking. Double (raincipant, Investigator) | | | | Investigator) | |
| NCT00827996 [38] Adalimumab Phase 2 •Allocation: N/A 10 | NCT00827996 [38] | Adalimumab | Phase 2 | •Allocation: N/A | 10 |
| Intervention Model: Single | - [] | | | •Intervention Model: Single | |
| Group Assignment | | | | Group Assignment | |
| •Masking: None (Open Label) | | | | •Masking: None (Open Label) | _ |
| NCT04018599 [39] 40 mg MSB11022 Phase 1 •Allocation: Randomized 216 | NCT04018599 [39] | 40 mg MSB11022 | Phase 1 | •Allocation: Randomized | 216 |
| Intervention Model: Parallel | | | | •Intervention Model: Parallel | |
| Accorrector | | | | Assignment | |

IFX, infliximab.

| NCT Clinical Trial | Intervention | Phase | Study Design | Enrollment |
|---------------------|----------------------|----------|---|------------|
| NCT03512275 [13] | CFZ533 | Phase 2 | •Allocation: Randomized | 90 |
| | 1.2006 | | Intervention Model: Parallel | |
| | E1000 | | Assignment | |
| | Placebo | | •Masking: Quadruple | |
| NCT03926169 [40] | Risankizumab | Phase 2 | •Allocation: Randomized | 220 |
| | Placebo | | Intervention Model: Parallel Assignment | |
| | | | Masking: Quadruple | |
| NCT04430855 [41] | Upadacitinib | Phase 2 | •Allocation: Randomized | 60 |
| | Placebo | | Intervention Model: Parallel | |
| | | | Masking: Quadrupla | |
| NCT04242498 [42] | Bimokizumah | Phase 3 | •Allocation: Randomized | 460 |
| NC104242490 [42] | Dimerizumad | 1 Hase 5 | •Intervention Model: Parallel | 400 |
| | Placebo | | Assignment | |
| | | | •Masking: Ouadruple | |
| NCT04179175 [43] | Secukinumab | Phase 3 | •Allocation: Randomized | 745 |
| | | | •Intervention Model: Parallel | |
| | | | Assignment | |
| | | | Masking: Triple | |
| | | | •Allocation: N/A | |
| NCT02712622 [44] | Secukinumab | Phase 2 | Intervention Model: Parallel | 471 |
| INC 1057 15052 [44] | Placebo | 1 Hase 5 | Assignment | 471 |
| | | | Masking: Triple | |
| NCT04092452 [45] | PF-06650833, Placebo | Phase 2 | Allocation: Randomized | 192 |
| | PF-06700841 | | •Intervention Model: Parallel | |
| | DE 0/92//47 | | Assignment | |
| NICT04246272 [46] | PF-06826647 | Dhase 2 | •Masking: Iriple | 16 |
| NC104246372 [46] | Ioracitinib | Phase 2 | •Allocation: IN/A | 46 |
| | | | -Intervention Woder. Single | |
| | | | •Masking: None (Open Label) | |
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Table 2. Recruiting interventional studies on target therapies.

3.1. Immunopathogenesis of Hidradenitis Suppurativa

HS pathogenesis is still largely unknown, and it is probably multifactorial [11]. Nevertheless, it is generally believed that follicular occlusion is the primary event, caused by hyperkeratinization and hyperplasia of the infundibular epithelium associated with defects in keratin production (downregulation of cytokeratin K17 and upregulation of K5 and K6) [12]. Genetics and lifestyle factors including smoking and obesity have been shown to contribute to the development of HS [47]. Follicular occlusion leads to dilatation of the hair follicle followed by rupture and release of contents, including hair-shafts, keratin fibers, microbes, and pathogen-/damage-associated molecular patterns (PAMPs/DAMPs), which leads to an acute and severe inflammatory response [48]. The release of the follicular contents into the dermis activates several inflammatory pathways, particularly (NOD)-like receptor protein 3 (NLRP3) inflammasome and toll-like receptor (TLR) signaling, thereby further aggravating the skin inflammation and the inflammatory loop [49]. Histologically, this event is characterized by cell infiltrates, including neutrophils (neutrophil elastase), T cells (CD3), B cells (CD19, CD20), plasma cells (CD138), natural killer (NK) cells (CD56), mast cells, macrophages (Factor XIIIA, CD68), and dendritic cells (CD11c, CD14). Multinucleate giant cells and body granulomas have also been identified in HS tissues [50]. Finally, as a specific hallmark of advanced HS, chronic inflammation induces sinus tract or tunnel formation. Studies of lesional tissue proposed the involvement of Ki67+ epithelial strands, the elevated proteolytic mechanism of metalloproteinases (MMP1, MMP2, and MMP8), and the increased activity of fibrotic factors such as transforming growth factor (TGF-ß 1-2-3) as fundamental in the sinus/tunnel formation [51–53]. These epithelialized cavities contribute

to create a favorable habitat for biofilm-producing bacteria, which are able to trigger inflammation continuously [54].

3.2. Cytokines' Role

Immune cells and keratinocyte-mediated products are widely accepted as key players in HS pathogenesis, and they appear dysregulated in lesional, perilesional, and normal-appearing tissue, serum and exudate of HS patients [55,56]. However, the exact role of each cytokine is not completely elucidated yet [55]. Cytokine-mediated keratinocyte hyperproliferation have been shown to contribute to the hyperkeratinization and hyperplasia of the infundibular epithelium in HS skin, leading to follicular occlusion with the subsequent formation of cysts [57]. Nonetheless, other trigger factors (e.g., mechanical friction) and/or predisposing factors (e.g., hair follicles of patients with HS seem to be susceptible to rupture due to alterations of the follicular structure) have been reported as contributory factors involved in the inflammatory loop of HS [58]. A representative scheme of the cytokines involved in the pathomechanism of HS is exemplified in Figure 2.



Figure 2. Schematic representation of the inflammatory pathways in hidradenitis suppurativa (HS), as for the pathways identified in this review.

3.2.1. Interleukin (IL)-1 Pathway

It was proposed that the release of follicular content initiates the NLRP3 inflammasome, an innate immune signaling complex and key mediator of IL-1 family cytokine production. Upon activation, NLRP3 recruits the adapter molecule ASC (apoptosis-associated speck-like protein containing a caspase recruitment domain), which binds NLRP3 to pro-caspase-1. Caspase-1 is activated by autoproteolysis and formation of the enzymatically active heterotetramer. Active caspase-1 catalyzes the cleavage of inactive pro-IL-1 β and pro-IL18 into active-form IL-1 β and IL-18, respectively [59]. In several studies, elevated levels of caspase-1 with enhanced mRNA expression of NRLP3 were detected in HS lesions [60,61].

The IL-1 pathway is hyperactive and contributes to cell infiltration and tissue damage in HS. The IL-1 family consists of 11 members, 7 with a pro-inflammatory activity (IL-1 α , IL-1 β , IL-18, IL-33, IL-36 α , IL-36 β , and IL-36 γ), while the remaining 4 have antagonistic (IL-1Ra, IL-36Ra, IL-38) [62] or anti-inflammatory (IL-37) [63] properties [64]. IL-1 α is highly pro-inflammatory and induces a strong downstream of inflammatory cytokines such as tumor necrosis factor (TNF) and IL-18 [65,66].

IL-1 β is produced mainly by monocytes and macrophages and contributes to amplify the inflammatory pathway leading to the induction of chemokines such as CXCL1 and CXCL6 involved in neutrophilic granulocytes' recruitment [67]. Furthermore, IL-1 β enhances the secretion of MMPs, which supports immune cell infiltration and contributes to tissue damage. Overexpression of IL-1 β at mRNA and protein levels has been reported in lesional, perilesional, and normal-appearing HS skin compared to that in healthy control [60], and an active IL-1 β pathway was also found in the serum of patients [68]. As result, IL-1 β signaling induces overexpression of inflammatory markers, mainly IL-8, TNF- α , and IL-17 stimulating chemotaxis of new neutrophils into the damaged skin, pus formation, and triggering of the inflammatory loop [69].

Although the IL-1 pathway is well known to be activated in HS, Ardon et al. showed decreased levels of IL-1 α in HS lesional skin compared with those in uninvolved skin of the same patient. Lower IL-1 α levels in lesional HS skin may be related to the intracellular location and consumption of IL-1 α at sites of inflammation [70]. Nonetheless, several data support that keratinocytes intrinsically produce increased levels of IL-1, thus it is supposed as a positive feedback between IL-1 and IL-17 [67]. In particular, IL-36, a member of IL-1 superfamily, is involved in the inflammasome activation and pro-inflammatory signaling through the activation of nuclear factor-kB (NF-kB) and mitogen-activated protein kinase (MAPK) [71]. In serum and lesional HS skin, several studies have proven increased levels of IL-36α, IL-36β, and IL-36γ and decreased antagonist cytokines (IL-36Ra, IL-37, IL-38) [72]. IL-36 increases dendritic cells activation, neutrophil recruitment, keratinocyte proliferation, and secretion of pro-inflammatory mediators (IL-1 β , TNF- α , IL-6, IL-8) stimulating the production of Th1 cells and Th17 cells and their cytokines (interferon gamma (IFN- γ), IL-17, IL-22, and IL-23) [73]. Regarding IL-36 antagonists, IL-37 and IL-38 levels have been significantly higher in perilesional HS skin than in healthy controls [72]. These cytokines are involved in the negative regulation of the inflammatory response, for example, by suppressing the secretion of the Th17 cells cytokines IL-17 and IL-22 [74,75]. The contributions of IL-37 and IL-38 imbalance in perilesional skin to the inflammatory pathogenesis of HS should be explored. IL-18 is secreted by macrophages and dendritic cells. Unlike IL-1 β , there is a constitutive pool of pro-IL-18 in producer cells, thus the regulation of secretion is determined mainly by inflammasome activation. IL-18 promotes Th1 cell activation and increases the cytotoxic activity of CD8+ T cells and natural killer (NK) cells. In addition, IL-18 induces other inflammatory cytokines, especially IFN- γ . Typically, IL-18 activity is dramatically enhanced by other cytokines, including IL-2, IL-12, IL-15, IL-21, and IL-23. mRNA and protein expression of IL-18 showed high levels in lesional and perilesional HS skin [60,61]. As here depicted, the IL-1 pathway has been found upregulated in a large number of studies that investigated HS immune dysregulation, denoting a strong level of evidence for its role in HS pathogenesis [76].

3.2.2. TNF- α and IFN- γ

TNF- α levels exhibited a positive correlation with HS severity and the therapeutic efficacy of TNF- α inhibitors supports the role of the dysregulated production of these cytokines in the pathophysiology of HS [77,78]. The only Food and drug administration-approved drug available to treat HS is adalimumab, a monoclonal antibody targeting TNF- α . As such, there is a high level of evidence on the upregulation of TNF- α in HS [79]. The TNF- α increases the ratio of Th17 to regulatory T-cells (Treg), resulting in aberrant production of Th17 cells and their cytokines IL-17 and IL-22 [80]. TNF- α in keratinocytes induces expression of CXCL8, CXCL11, CCL20, and CCL2 chemokines, which recruit neutrophils, T cells, and monocytes into the skin [81]. Together, these signals lead to massive immune cell infiltration into damaged tissue. Therefore, HS lesions are characterized by granulocytes,

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T cells, B cells, and monocytes, which differentiate into macrophages and dendritic cells. The dendritic cells mediators, IL-23 and IL-12, support Th17 and Th1 cells to produce their specific cytokines, IL-17 and IFN- γ , respectively.

IFN- γ is secreted by Th1 cells, induces Th1-attracting chemokines such as CXCL10, and activates dermal endothelial cells and macrophages allowing the infiltration of immune cells from the bloodstream [82]. IFN- γ mRNA and protein expression have been shown to be elevated compared to those in healthy controls in skin lesions and wound exudate, and its contribution has been assessed by several high-powered studies [77,83,84].

3.2.3. IL-17/IL-23 Axis

Although it has been proven that Th1 cells plays an important role in the pathogenesis of chronic inflammatory conditions, several studies have found that Th17 exceeds the Th1 pathway in the induction of tissue inflammation [80,85,86]. Th17 cells are abundantly found in the papillary and reticular dermis of HS lesions and may be responsible for excessive neutrophilic inflammation and purulent drainage [86]. Th17 cell development is promoted by IL-23, IL-1 β , and IL-6 produced by innate cells such as dendritic cells [87]. The Th17 axis encompasses several pro-inflammatory mediators such as IL-22, IL-21, IL-6, Granulocyte colony-stimulating factor (GCSF), IL-1 β , TGF β , and TNF- α and antimicrobial peptides (AMPs), in particular β -defensin-2, S100 proteins, and lipocalin-2 [88]. In keratinocytes, IL-17 induces the expression of LL37/cathelicidin, S100A7, S100A8, and S100A9, which are increased in the lesional tissue and serum of HS patients but not in perilesional skin [89,90]. These proteins are involved in keratinocyte proliferation and pro-inflammatory cytokine and chemokine expression. IL-17 overexpression has been detected in lesional, perilesional, and unaffected skin, suggesting that subclinical inflammation is present in HS skin prior to the formation of active lesions. Elevated IL-17 levels have also been registered in the serum of HS patients [91]. While the above-mentioned studies support a central role of Th17 cells in lesion development and driving HS inflammation, conflicting results were reported by one serum study, which showed no significant difference between patients and controls [92].

It has been reported that infiltrate macrophages in the papillary and reticular dermis of HS lesions overexpressed IL-23 [93]. Thus, considering the importance of the aberrant IL-17 expression, the IL-23/IL-17 axis is believed to be crucially involved in the pathogenesis of HS [86]. IL-23 is commonly expressed by macrophages in response to infectious stimuli and as mentioned above leads the differentiation of Th17 cells [94]. IL-23 is a member of the IL-12 family of cytokines, which also includes IL-12 and IL-27, and it is a heterodimer sharing a p40 subunit with IL-12 and having a distinct p19 subunit [95]. Increased mRNA expression of IL-23p19 in HS lesions and overexpression of IL-23p40 in serum has been proven [81,86]. IL-12 has also been observed in HS lesional skin in a limited number of studies. Remarkably, IL-12 and IL-23 cytokines are mainly produced by dendritic cells or macrophages and support the function of Th1 and Th17 cells, respectively forming their specific cytokines (IFN- γ , IL-17) [95].

3.2.4. IL-6

IL-6 is a pleotropic cytokine that plays a key role in a wide variety of immune processes. IL-6 promotes antibody production by activated B cells, induces the expression of acute phase proteins such as C-reactive protein, and affects the function of several other cell types including keratinocytes. IL-6, in combination with TGF- β , IL-1 β , and IL-23, promotes the development of Th17 cells and inhibits TGF- β -induced regulatory T-cell development [96]. At present, there are only a few evidences on the association between IL-6 and HS, but the results are controversial and the evidence is conflicting. Several studies show that IL-6 mRNA expression was increased in lesions of HS patients compared to that in non-lesional areas [56,97]. In contrast, other data revealed that the IL-6 levels in HS skin lesions were lower than those in non-lesional skin [98] and also that monocytes from HS patients exhibited in vitro an impaired ability to secrete IL-6 [55]. Elevated levels of IL-6 in the serum of

HS patients have been described, and it has been suggested that IL-6 not only participates in the maintenance of inflammation but might also promote the formation of granulomas in lesions [99]. Notably, HS coexists with other inflammatory diseases where IL-6 also contributes to the development, including pyoderma gangrenosum and inflammatory bowel diseases, suggesting that they share similar immune–pathogenic pathways [99].

3.2.5. IL-10

Unlike in other immune-mediated skin disorders, it has been observed that in HS there is a high expression not only of pro-inflammatory cytokines but also of the anti-inflammatory mediator IL-10 [84]. IL-10 is secreted by innate and adaptive immune cells, it induces the differentiation of Treg cells and suppresses the development of Th1, Th2, and Th17 cells [100]. IL-10 reduces immune responses by suppressing pro-inflammatory cytokine production by monocytes and macrophages and limiting T cell activation [101]. Several studies have demonstrated that the expression of IL-10 is elevated in HS lesional and perilesional skin [56,89,98]. Thus, the immunosuppressive role of IL-10 seems to be upregulated in HS skin as a compensatory response to the pro-inflammatory process and to the dissemination of skin commensal microbes. Moreover, this increase selectively suppresses not only IL-22 but also IL-17 lesional levels [84]. IL-22 is a member of the IL-10 cytokine family secreted by Th22 and other lymphocytic cells. It is known to promote keratinocyte hyperproliferation and epidermal acanthosis and to enhance AMP expression [102]. IL-22 has antimicrobial and pro-inflammatory functions and contributes to wound healing and the maintenance of epithelial barrier function [84]. The lack of IL-22 is associated with insufficient upregulation of AMPs, even in the presence of high levels of IL-17, which results in microbial spread in HS skin [103]. The decreased expression of IL-22 might be due to reduced infiltration of IL-22-secreting cells and impaired production of IL-22 by these cells [89,104]. IL-22 deficit has also been related to increase of IL-10 production, which might be induced, among others by IL-1 β [84]. It is important to underline that IL-22 production is enhanced by Notch signaling, which is defective in HS patients [89]. On the other hand, some conflicting results have been found on the hyperexpression of the serum level of IL-10 in two studies that show no significant difference between the serum of HS patients and controls raising controversial ideas on the substantial role of this cytokine in the pathogenesis of HS [105,106]. Along with IL-10, other cytokines such as IL-4 and IL-13 have an anti-inflammatory role in HS. These cytokines can inhibit the synthesis of IL-1 β , yet they stimulate the synthesis of IL-1Ra [64]. The specific role of these anti-inflammatory cytokines in the pathogenetic mechanism of HS remains to be further clarified.

3.3. Overview on Therapies and Treatment Possibilities

Adalimumab, a monoclonal antibody directed against tumor necrosis factor- α , already approved for psoriasis and other various rheumatological and gastroenterological diseases [103], is the only biologic agent currently available for the treatment of moderate-to-severe HS resistant to antibiotics. However, several cases reported a certain rate of primary or secondary lack of response in some patients [107]. Numerous other specific anti-interleukins and small-molecules drugs are currently under investigation for the treatment of HS. The main HS-related drugs and their targets are represented in Figure 3.



Figure 3. Schematic representation of the therapeutics targeting the immune products involved in HS pathogenesis.

A total of 21 biologics and other immunomodulatory agents reported in the treatment of HS were identified and categorized according to their mode of action and quality of evidence ranking as previously published [108] (Table 3). Among them, only the newest and the ones whose data are available on PubMed or on clinicaltrials.gov were discussed in this review, and levels of evidence have been added when available. We are providing below an up-to-date review of the most relevant clinical trials targeting the key products related to HS as shown in Tables 1 and 2.

| Cytokines | Drugs | Quality of Evidence |
|---------------|---------------------|---------------------|
| anti-TNF-α | Adalimumab [33–38] | А |
| | Infliximab [20] | В |
| | Etanercept [26] | В |
| anti-IL-1 | Anakinra [18] | В |
| | MEDI8968 | Ongoing Trial |
| | Canakinumab | C |
| | Bermekimab [13] | В |
| anti-IL-12/23 | Ustekinumab [30] | Ongoing Trial |
| anti-IL-23 | Guselkumab [22] | Ongoing Trial |
| | Risankizumab [40] | Ongoing Trial |
| anti-IL-17 | Secukinumab [43,44] | Ongoing Trial |
| | CJM112 [19] | Ongoing Trial |
| | Bimekizumab [42] | Ongoing Trial |
| | Brodalumab [14,16] | Ongoing Trial |
| anti-PDE-4 | Apremilast [24] | B |
| anti-C5a | IFX-1 [31] | Ongoing Trial |
| anti-CD20 | Rituximab | C |
| anti-CD40 | Iscalimab [13] | Ongoing Trial |
| anti-JAK | Upadacitinib [41] | Ongoing Trial |
| | INCB054707 [15,16] | Ongoing Trial |
| | Tofacitinib [46] | Ongoing Trial |

Table 3. Recruiting interventional studies on target therapies.

TNF, tumor necrosis factor; IL, interleukin; JAK, Janus kinase; PDE, phosphodiesterase.

3.3.1. TNF- α Inhibitors

Adalimumab is an IgG1 monoclonal antibody. In HS, it is administrated as an initial dose of 160 mg, followed by a dose of 80 mg for 2 weeks and a maintenance dose of 40 mg weekly. To date, adalimumab is the only FDA-approved biologic agent for the treatment of moderate/severe HS and a strong level of evidence, clinical trials, and case reports corroborate its use [109,110]. Two large double-blind, placebo-controlled, randomized clinical trials (RCTs), PIONEERI and II, demonstrated the safety and efficacy of adalimumab. In these studies, patients received either adalimumab (160 mg at week 0, 80 mg at week 2, and 40 mg weekly starting at week 4) or placebo for 12 weeks [78,110]. The primary endpoint in both studies was the number of patients achieving clinical response according to the Hidradenitis Suppurativa Clinical Response (HiSCR) at week 12 defined as a 50% decrease in total abscess and inflammatory nodule lesions from baseline. This endpoint was achieved by 41.8% and 58.9% of patients in the treatment groups versus 26.0% and 27.6% of patients in the placebo groups, for PIONEER I and II, respectively [110]. Despite providing clinical improvement, HiSCR is only achieved by approximately 50% of patients receiving adalimumab, and nearly 6.5% of patients developed anti-drug antibodies reducing drug efficacy [111].

Infliximab

Infliximab (IFX) is a monoclonal antibody directed against TNF- α that inhibits its downstream effects [112]. A clinical trial on IFX dosed at 5 mg/kg at weeks 0, 2, and 6 followed by maintenance dosing every 8 weeks for 22 weeks, showed a decrease of HSSI >50% (HS-specific severity index) from baseline in 26.7% of patients [113,114]. Although a quite low number of patients met the primary endpoint, IFX undoubtedly presented an advantage over placebo in which the majority of patients (88.9%) showed less than 25% improvement in HSSI where severe disease is defined by an HSSI score \geq 13 [114,115]. Despite the fact that further studies are needed to better define the use of IFX compared with that of adalimumab, IFX remains a valuable tool in the treatment of HS.

• Etanercept

Etanercept is a recombinant human TNF- α receptor that competitively binds TNF- α receptors [116]. Literature regarding the use of etanercept in HS has reported discordant data on its efficacy, and its current use in real life is limited [4,117,118].

3.3.2. IL-1 Inhibitors

• Anakinra

Anakinra is a recombinant IL-1 receptor inhibitor [119]. In HS, it is subcutaneously administrated as a 100 mg daily dose [32,120,121]. A double-blind, randomized, clinical trial showed significantly decreased disease activity in the anakinra group compared to that in the placebo group at week 12 [121]. However, at 24 weeks, the difference in patients achieving HiSCR was not statistically significant (10% vs. 33%). Painful reactions at the injection site were commonly reported in rare cases also linked with drug-induced sarcoidosis [119,121]. Moreover, some reports showed anakinra failure in severe HS patients [122].

• Bermekimab

Bermekimab (MABp1) is an anti-IL-1 α human monoclonal antibody [123]. Forty-two patients were enrolled in a phase 2 clinical trial, reporting that bermekimab was effectively inducing a clinical response after 12 weeks of treatment [123]. A significant reduction in abscesses and inflammatory nodules of 60% (p < 0.004) and 46% (p < 0.001) was seen in anti-TNF-naive and anti-TNF-failure

groups, respectively. IL-1 α could characterize an important clinical target for HS, and bermekimab may represent a new option to treat moderate-to-severe HS.

MEDI8968 and Canakinumab

MEDI8968 [124] and canakinumab [125] are human monoclonal antibodies recently approved for rheumatologic conditions [124,126,127]. MEDI8968 selectively binds the IL-1R1 receptor inhibiting the activation of IL-1 α and IL-1 β , while canakinumab selectively targets IL-1 β . A Phase IIa study was conducted to evaluate MEDI8968 for the treatment of moderate-to-severe HS patients, but it was terminated early due to a lack of efficacy [124]. Canakinumab showed mixed results in several case reports and series [128].

3.3.3. Anti-IL-17 Drugs

Based on the key pathways involved in the pathogenesis of HS, several anti-IL-17 drugs are currently under investigation as possible efficacious therapeutics.

Secukinumab

Secukinumab, an anti-IL-17A IgG1 antibody [103], has currently been studied in two Phase 3 double-blinded, randomized clinical trials (SUNSHINE and SUNRISE) [43,44]. It is given at the dosage of 300 mg subcutaneously per week, then followed by 4-weekly maintenance. Secukinumab was shown to improve HS condition in several case reports [129,130]. Phase 3 trials on secukinumab for treating HS are currently underway (Table 2), but results are still not available.

Bimekizumab

Bimekizumab, an anti-IL-17A and anti-IL-17F IgG1 antibody, is under evaluation with two Phase 3 double-blinded, randomized clinical trials (BE HEARD 1 and 2) [17,42,103], with no results available at the time of writing. Among anti-IL-17 drugs, the ones that simultaneously block more subunits of anti-IL-17, such as Bimekizumab, may be more effective in the treatment of HS, as the various subunits seems equally involved in the development of inflammation, and bimekizumab seems to have a major effectiveness in suppressing inflammation and cytokine production from preclinical studies [103].

Brodalumab

Brodalumab is a recombinant, fully human monoclonal antibody (IgG2), which binds with high affinity to the interleukin (IL)-17 receptor A (IL-17R). Brodalumab is FDA approved for the treatment of moderate-to-severe chronic plaque psoriasis [131].

A recently published study [85] reported promising results using brodalumab for the treatment of moderate-to-severe HS, along with no grade 2/3 adverse events. All patients in the study achieved HiSCR, and 80% achieved IHS4 (Severity Score System) [132] at week 12. HiSCR achievement occurred as early as week 2, likely due to the unique blockade of IL-17A, IL-17C, and IL-17F by brodalumab.

• CJM112

CJM112 is a human monoclonal anti-IL-17A antibody. A Phase II study with moderate-to-severe chronic HS has been completed, but results are not available at the present time [19].

3.3.4. Anti-IL-23 Drugs

Two anti-IL-23 drugs, risankizumab [40,133] and guselkumab [22,134], are currently in phase 2 clinical trials to evaluate their efficacy in the treatment of moderate-to-severe HS. Various case series and isolated reports describe the effectiveness of this category of medication in the treatment of HS, as was reported for ustekinumab [103]. In future years, we expect the results of these trials that may bring a new weapon in the treatment of HS.

3.3.5. Anti-IL-12/23 Drugs

• Ustekinumab

Ustekinumab is a human monoclonal antibody that acts by inhibiting the p40 subunit on IL-12 and IL-23 [127,135]. In a Phase II open-label study involving 17 patients, patients showed moderate-to-marked improvement achieving HiSCR in almost 40% of cases [136]. Several cases series reported positive outcomes using ustekinumab in moderate-to-severe HS patients [136–138].

3.3.6. Janus Kinase (JAK) Inhibitors

• INCB054707

INCB054707 is an orally administered inhibitor of the JAK 1 pathway. There are currently two Phase II trials underway [15,16].

• Tofacitinib and Upadacitinib

Tofacitinib is a potent, selective JAK inhibitor that preferentially inhibits Janus kinase (JAK) 1 and JAK3 [139]. It has been recently shown to be potentially effective in recalcitrant HS by some case series serving as proof of concept for the ongoing clinical trial [46,140]. Upadacitinib is a selective JAK1 inhibitor, with 74- and 58-fold selectivity for JAK1 over JAK2 and JAK3, respectively [141]. A Phase 2, multicenter, randomized, double-blind study is currently recruiting moderate-to-severe patients to evaluate the safety and efficacy of this drug in treating HS [41].

3.3.7. Others

Apremilast

Apremilast is a small-molecule inhibiting phosphodiesterase 4 [142]. It blocks cyclic adenosine monophosphate (cAMP) degradation, which drives the activation of protein kinase A (PKA) and reduces production of TNF, IL-12p40, and IL-17 [143]. Apremilast has been tested in two double-blinded, phase 2 trials in patients with moderate HS. Eight out of 15 patients (53.3%) given apremilast achieved a positive HiSCR at week 16, compared to zero out of five patients in the placebo group (p = 0.055). Patients receiving apremilast also showed a lower rate of nodules and abscesses [144,145].

• IFX-1

IFX-1 is a human C5a-specific monoclonal antibody [31,146]. An open-label clinical trial reported that 75% of patients achieved HiSCR at day and more than 80% at day 134 [31]. No other information is available at the time of writing.

• Iscalimab (CFZ533)

Iscalimab CFZ533 is a fully human, Fc-silenced, non-depleting, IgG1 mAb preventing CD40 pathway signaling and activation of CD40+ cell types [147]. It is proposed for the treatment of several immune diseases [148,149]. A new study is currently ongoing to assess the preliminary efficacy and safety of CFZ533 in patients with moderate-to-severe HS to evaluate the clinical profile for further clinical development. A schematic representation of the main axes involved in the pathogenesis of HS and the most relevant drugs targeting those axes are shown in Figure 3.

3.4. The Role of Microbiome and Biofilms

The role of microbes in the pathogenesis of HS is still discussed. High levels of antimicrobial peptides including β -defensin-2, S100 proteins, lipocalin-2, and LL37/cathelicidin in HS skin [89] and

the efficacy of antibiotics have suggested a strong microbial influence in disease activity [150]. However, it is unclear whether bacteria are initiating- and/or promoting-factors in the progression of HS or if sinus/tunnel formation provides a favorable habitat for biofilm-producing bacteria. Microbes probably trigger a cascade of PAMPs/DAMPs leading to NLRP3 activation and IL-1 β release. In addition, TLR2 plays a central role in innate immunity, by sensing microbial ligands and activating the host defense response through inflammatory mechanisms. Recent findings of the highly increased expression of TLR2 by CD68+ macrophages and CD209+ in HS lesions suggest that microbial colonization might contribute to the tissue inflammation [77,151].

Culture-dependent studies of superficial and deep HS lesions have found the involvement of different microbial species [152]. Gram-positive cocci and rods, including *Staphylococcus aureus*, coagulase-negative Staphylococci (CoNSs), *Streptococcus* spp., and *Corynebacterium* spp. have been isolated from surface swabs and deep tissue samples [153,154]. CoNS species were the most common species found in bacterial cultures of deep HS lesions obtained by carbon dioxide laser treatment [155]. *Staphylococcus lugdunensis* and other CoNS species such as *Staphylococcus epidermidis* were associated with the early stages of HS lesions [156]. Anaerobic bacteria, mainly Gram-negative bacilli *Prevotella* and *Porphyromonas* spp. were also isolated in early and chronic HS lesions [153,154]. In addition, Enterobacteriaceae, particularly *Proteus mirabilis*, were commonly identified in cultures of superficial and deep lesions [153,157].

Thus, the most common species found on bacteriology analysis of superficial and deep lesions of HS patients are CoNS, *Staphylococcus aureus*, and anaerobic bacteria. These bacteria have been shown to be capable of biofilm formation, which might be responsible for resistance to antimicrobial therapy. Studies based on 16S and 18S ribosomal RNA (rRNA) next-generation sequencing have provided new insights into the role of the skin microbiome in the pathophysiology of HS, most likely a biofilm-driven disease [158]. It was demonstrated that there is a significantly different microbiome in HS patients, either lesional or non-lesional, compared to that in healthy controls [158]. In metagenomic studies, Corynebacterium spp., Porphyromonas, and Peptoniphilus ssp. were the predominant species identified from HS lesions, whereas Porphyromonas and Peptoniphilus ssp. were not detected in healthy control. In contrast, healthy skin showed a relatively higher abundance of *Propionibacterium* ssp. [159,160]. Some studies have shown that a mixed anaerobic microbiome was associated with clinical severity, and the abundance of anaerobes Fusobacterium and Parvimonas increases with higher classifications of Hurley staging [161,162]. These studies, while limited, indicate a proliferative bacterial phenotype in HS lesions. Ring et al. have specifically investigated the microbiome in the persistently inflamed sinus tracts of moderate and severe HS patients, finding that these samples were dominated by *Prevotella* ssp., *Porphyromonas* ssp., and other anaerobic species [163].

In conclusion, a pattern of cutaneous dysbiosis appears in HS, which highlights the presence of anaerobic bacteria and *Staphylococcus* spp., but the pathogenic role of the microbiome in HS is still unclear. The presence of microorganisms in HS has led to the condition being considered an infectious disorder. This is reinforced by the current guidelines for therapy focusing on the use of antibiotics [150,164].

4. Discussion

Hidradenitis suppurativa is a condition that involves several, concomitant pathways and, even though its pathogenesis remains unclear, more and more efforts are being made to elucidate the trigger factors of this debilitating disease. Various cytokines seem to be involved in the pathogenesis of HS, and the dysregulation of multiple inflammatory pathways, such as TNF, IL-1, IL-17/23, and anti-inflammatory cytokines, such as IL-10, has been observed.

Although none of the current treatments seem to adequately control this condition, immunomodulatory treatments targeting the Th17 pathway and the JAK/STAT pathways are now explored in HS. Anti-IL-17 drugs such as secukinumab, bimekizumab, or brodalumab and anti-IL-23 drugs such as risankizumab or guselkumab may represent an effective alternative in controlling

this condition. Other biologic drugs directed to neutralize different axes have also been explored. The anti-IL-1 drug, bermekimab, it is currently under investigation, and may also become a treatment opportunity for patients with moderate-to-severe HS. Although the trials for the majority of this cytokine's selective inhibitors are still at early phases (most of the clinical trials are in phase two or three), the pursuit of an effective treatment for the most severe cases of HS seems to have promising alternatives that are at present under investigation.

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