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Relationship of common hemodynamic and respiratory target parameters with brain tissue oxygen tension in the absence of hypoxemia or hypotension after cardiac arrest: A post-hoc analysis of an experimental study using a pig model

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Abstract

Brain tissue oxygen tension (PbtO₂)-guided care, a therapeutic strategy to treat or prevent cerebral hypoxia through modifying determinants of cerebral oxygen delivery, including arterial oxygen tension (PaO₂), end-tidal carbon dioxide (ETCO₂), and mean arterial pressure (MAP), has recently been introduced. Studies have reported that cerebral hypoxia occurs after cardiac arrest in the absence of hypoxemia or hypotension. To obtain preliminary information on the degree to which PbtO₂ is responsive to changes in the common target variables for PbtO2-guided care in conditions without hypoxemia or hypotension, we investigated the relationships between the common target variables for PbtO2-guided care and PbtO₂ using data from an experimental study in which the animals did not experience hypoxemia or hypotension after resuscitation. We retrospectively analyzed 170 sets of MAP, ETCO₂, PaO₂, PbtO₂, and cerebral microcirculation parameters obtained during the 60-min post-resuscitation period in 10 pigs resuscitated from ventricular fibrillation cardiac arrest. PbtO₂ and cerebral microcirculation parameters were measured on parietal cortices exposed through burr holes. Multiple linear mixed effect models were used to test the independent effects of each variable on PbtO2. Despite the absence of arterial hypoxemia or hypotension, seven (70%) animals experienced cerebral hypoxia (defined as PbtO₂ <20 mmHg). Linear mixed effect models revealed that neither MAP nor ETCO₂ were related to PbtO₂. PaO₂ had a significant linear relationship with PbtO₂ after adjusting for significant covariates (P = 0.030), but it could explain only 17.5% of the total PbtO₂ variance (semi-partial $R^2 = 0.175$; 95% confidence interval, 0.086–0.282). In conclusion, MAP and ETCO₂ were not significantly related to PbtO₂ in animals without hypoxemia or hypotension during

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the early post-resuscitation period. PaO_2 had a significant linear association with $PbtO_2$, but its ability to explain $PbtO_2$ variance was small.

Introduction

Despite advances in the treatment of cardiac arrest, most cardiac arrest survivors fail to achieve meaningful neurological recovery due to brain injury [1–3]. Brain injury after cardiac arrest results not only from interruption of cerebral blood flow (CBF) during circulatory arrest but also from various pathophysiological derangements after restoration of spontaneous circulation (ROSC). Several studies have suggested associations between perturbations in cerebral oxygen delivery after ROSC and worse neurological outcome after cardiac arrest [4–10]. In line with these studies [4–10], recent post-cardiac arrest care guidelines have emphasized the importance of maintaining adequate arterial oxygen tension (PaO₂), carbon dioxide tension (PaCO₂), and mean arterial pressure (MAP) to ensure sufficient cerebral oxygenation, which in turn prevents secondary hypoxic-ischemic brain injury [11, 12].

Tissue oxygen tension, the partial pressure of oxygen within the interstitial space of a tissue, has been measured in various clinical and laboratory settings to assess the adequacy of tissue oxygenation. Currently, two types of oxygen sensors are widely used to measure tissue oxygen tension: polarographic electrodes and optical sensors. Polarographic electrodes measure oxygen tension based on the electrical current produced when an anode and cathode submerged in an electrolyte solution come into contact with oxygen [13]. Optical sensors measure the oxygen tension resulting from luminescence quenching in the presence of oxygen [14]. Brain tissue oxygen tension (PbtO₂) monitoring is performed by placing an oxygen sensor within brain tissue through a burr hole. This technique is an established method for direct real-time monitoring of cerebral oxygenation. It has been used as a neuromonitoring modality in the management of patients with severe traumatic brain injury. In patients with severe traumatic brain injury, low PbtO₂ levels have been shown to increase the risk of hypoxic-ischemic brain injury and the likelihood of poor outcomes [15-18]. Recently, PbtO₂-guided care, a therapeutic strategy to treat or prevent cerebral hypoxia through modifying determinants of cerebral oxygen delivery, including PaO₂, end-tidal carbon dioxide (ETCO₂), and MAP, has been introduced in post-cardiac arrest care [19–21]. Although it appears to be a promising strategy, further information regarding PbtO₂ levels after cardiac arrest is needed before PbtO₂-guided care can be trialed or widely implemented in clinical post-cardiac arrest care. In several studies using PbtO₂ monitoring, despite maintaining adequate arterial oxygenation and perfusion pressure, episodes of cerebral hypoxia frequently occurred after resuscitation from cardiac arrest [20-22]. It is evident that correcting hypoxemia and/or hypotension can increase PbtO₂ levels [23-26]. However, no studies have evaluated the impact of increasing PaO₂ and/or MAP on PbtO₂ levels in non-hypoxemic, non-hypotensive cardiac arrest patients during the early post-resuscitation period.

In this study, we sought to determine the extent to which the common target variables for $PbtO_2$ -guided care, including PaO_2 , $ETCO_2$, and MAP, are related to $PbtO_2$ in the early post-resuscitation period using data from an experimental study conducted using a pig model of cardiac arrest in which the animals did not experience hypoxemia or hypotension during the early post-resuscitation period. The results of this analysis would provide preliminary information on the degree to which $PbtO_2$ is responsive to changes in the common target variables for $PbtO_2$ -guided care and, thus, would help identify an optimal management protocol for $PbtO_2$ -

guided care. We hypothesized that in conditions without hypoxemia or hypotension after cardiac arrest, PaO₂, ETCO₂, and MAP would be closely related to the PbtO₂.

Materials and methods

This study was a post-hoc analysis of data derived from a previous study investigating the effects of pralidoxime administration during cardiopulmonary resuscitation (CPR) on hemodynamics and cerebral microcirculation after ROSC in 20 healthy Yorkshire/Landrace cross pigs weighing 24.7 ± 1.7 kg [27]. In this study, data from 10 animals that received only standard advanced cardiovascular life support (control group) were included. All experiments were approved by the Animal Care and Use Committee of Chonnam National University Hospital (CNUH IACUC-18004) and performed in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. The investigators who performed the experiments had completed an Institutional Animal Care and Use Committee training course on animal care and handling. All surgical interventions were performed under sevoflurane anesthesia, and every effort was made to prevent unnecessary suffering of the animals.

Animal preparation

The animals were fasted overnight, but they were given water ad libitum. After an intramuscular injection of ketamine (20 mg/kg) and xylazine (2.2 mg/kg), they were placed in a supine position, and their trachea was intubated with a 7.0-mm internal diameter tracheal tube. The animals were anesthetized using an anesthesia machine with a 70/30 mixture of nitrous oxide/ oxygen and sevoflurane. During surgical intervention, sevoflurane was titrated to maintain adequate anesthetic depth, measured by the absence of reflex withdrawal and no change in heart rate, arterial pressure, or respiratory rate. They were mechanically ventilated in a volume control mode with the following settings: tidal volume, 10 ml/kg; respiratory rate, 14 breaths/ min; positive end-expiratory pressure, 5 cmH₂O; inspiratory/expiratory ratio, 1:2; and fraction of inspired oxygen (FiO₂), 0.3. ETCO₂ was monitored using a sidestream capnography monitor (B40 Patient Monitor; GE Healthcare, Chalfont St. Giles, UK), and the respiratory rate was subsequently adjusted to maintain normocapnia. A 6.0-F introducer sheath was advanced through the right external jugular vein for pacemaker wire insertion and right atrial pressure monitoring. A 7.0-F saline-filled catheter was advanced through the left femoral artery for arterial pressure monitoring and blood sampling. Another 7.0-F saline-filled catheter was inserted into the left internal jugular vein and advanced toward the skull base for jugular venous blood sampling. After subcutaneous infiltration of 2% lidocaine solution, a scalp incision was made to expose the right and left parietal bones. A 10-mm diameter burr hole was drilled bilaterally through the parietal bone (10-mm lateral and 10-mm posterior to the intersection of the coronal and sagittal sutures). The dura underneath the burr hole was opened to visualize the pial vessels. Three adhesive electrodes were applied on limbs to record the electrocardiogram. Normal saline was infused to maintain a mean right atrial pressure of 5-10 mmHg. Throughout the experiment, the rectal temperature was maintained at 38°C using a heating blanket.

Experimental protocol

After baseline measurements were obtained, an electrical current (60 Hz and 30 mA alternating current) was applied through the pacemaker wire placed in the right ventricle to induce ventricular fibrillation (VF). Mechanical ventilation was discontinued immediately after the onset of VF. After 5 min of untreated VF (Fig 1), external mechanical chest compressions (Life-Stat; Michigan Instruments, Grand Rapids, MI, USA) with a compression depth comparable to 20% of the anteroposterior chest diameter and a rate of 100 compressions/min were



Fig 1. Experimental timeline. After 5 min of untreated ventricular fibrillation, cardiopulmonary resuscitation was initiated using a mechanical chest compression device. The lightning marks indicate the onset of a 10-s pause in chest compression for rhythm analysis and a 150-J electric shock, if indicated. After ROSC, the animals were observed for 60 min in a simulated intensive care setting. Arterial pressure, ETCO₂, MFI, %Capillary number, and PbtO₂ were sampled at pre-arrest baseline, 1-min intervals for 15 min after ROSC, and 30, 45, and 60 min after ROSC. Arterial blood gases were measured at pre-arrest baseline and 3, 15, and 60 min after ROSC. MAP and CPP during cardiopulmonary resuscitation were sampled 2 min after the initiation of cardiopulmonary resuscitation since most of the animals (80%) achieved ROSC 2–4 min after cardiopulmonary resuscitation. ETCO₂, end-tidal carbon dioxide; PbtO₂, brain tissue oxygen tension, MFI, microvascular flow index; %Capillary number, percent of counted number of perfused capillaries relative to that at pre-arrest baseline; PaO₂, partial pressure of oxygen in arterial blood; MAP, mean arterial pressure; CPP, coronary perfusion pressure; ROSC, restoration of spontaneous circulation.

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initiated. Simultaneously, ventilation was provided with high-flow oxygen (14 l/min) at a rate of 10 breaths/min using a volume-marked bag devised by Cho et al [28]. During CPR, epinephrine (0.02 mg/kg) was administered intravenously every 3 min, and defibrillation was attempted with a single biphasic 150-J electric shock delivered between the right infraclavicular area and the cardiac apex every 2 min, if indicated. These procedures were continued until ROSC was achieved or until 14 min had elapsed since the initiation of CPR after which CPR was discontinued.

After ROSC, the animals were observed for 60 min in a simulated intensive care setting. General anesthesia with sevoflurane was provided throughout the 60-min period. Immediately after ROSC, mechanical ventilation was resumed with an FiO₂ of 1.0; the other ventilator settings were unchanged. Fifteen minutes after achieving ROSC, the ventilatory rate was adjusted to maintain an ETCO₂ of 45 mmHg. Normal saline (10 ml/kg/h) was intravenously infused to maintain normovolemia during the 60-min period, but no hemodynamic drugs were given during this period. We predetermined the humane endpoints for euthanasia (systolic arterial pressure <60 mmHg, heart rate <40 beats/min, or seizure), but none of the animals reached the endpoints until completion of the experimental protocol. Immediately after completing the experimental protocol, the animals were euthanized with a rapid bolus of 40 mEq potassium chloride under general anesthesia.

Measurements

Arterial pressure and right atrial pressure were continuously monitored (CS/3 CCM; Datex-Ohmeda, Helsinki, Finland) and stored on a personal computer using a data collection software (S/5 Collect software, Datex-Ohmeda, Helsinki, Finland). Coronary perfusion pressure during CPR was calculated from the differences in time-coincident aortic end-diastolic and right atrial pressures. PbtO₂ and cerebral microcirculatory blood flow were assessed in the right and left parietal cortices. The measurement site (right or left) was randomized and counterbalanced. PbtO₂, the study primary outcome, was measured using an optical oxygen sensor (DP-PSt7; PreSens-Precision Sensing GmbH, Regensburg, Germany), which was placed directly on the surface of the parietal cortex. To assess the responsiveness of PbtO₂ to changes in FiO₂ at pre-arrest baseline, FiO₂ was increased from 0.3 to 1.0, maintained at 1.0 until the $PbtO_2$ level reached a plateau, and then decreased to 0.3. Cerebral hypoxia was defined as $PbtO_2 < 20 \text{ mmHg} [29]$, and the duration of exposure to cerebral hypoxia was determined for each animal. Cerebral cortical microcirculation images were recorded using a hand-held digital microscope (G-Scope G5; Genie Tech, Seoul, Korea) placed over the burr hole. The microscope provided ×250 magnification and showed an area of interest of approximately $1800 \times 1000 \ \mu\text{m}^2$. Two investigators analyzed the cerebral microcirculation videos and quantitated the microvascular flow index (MFI) of vessels <20 µm in diameter (representing primarily capillaries) by consensus [30, 31]. They also counted the number of perfused capillaries as previously described by Serné et al [32]. The number of perfused capillaries after ROSC was expressed as a percent of the counted number of perfused capillaries relative to that at the prearrest baseline (%Capillary number) to correct for its substantial inter-animal variation. Arterial pressure, ETCO₂, MFI, %Capillary number, and PbtO₂ were sampled at pre-arrest baseline, 1-min intervals for 15 min after ROSC, and 30, 45, and 60 min after ROSC. Arterial blood gases were measured at pre-arrest baseline and 3, 15, and 60 min after ROSC.

Statistical analysis

Data were analyzed using R version 3.3.3 (R Foundation for Statistical Computing, Vienna, Austria) and T&F program version 3.0 (YooJin BioSoft, Goyang, Republic of Korea). Continuous variables, presented as mean \pm standard deviation or median values with interguartile ranges, were examined for normality using the Shapiro-Wilk test. We used multiple linear mixed effect models to test the independent effects of common target variables for PbtO₂guided care, including PaO₂, ETCO₂, and MAP, on the PbtO₂ during the 60-min post-resuscitation period. In addition, we also tested the independent effects of cerebral microcirculation variables, including MFI and %Capillary number, on the PbtO₂. Since arterial blood gases were measured only at three timepoints (3, 15, and 60 min) after ROSC, PaO₂ data were not available for several timepoints at which the other target variables were sampled. In the original study [27], we found that PaO₂ levels linearly increased over time during the 60-min postresuscitation period. Thus, PaO₂ values at unmeasured timepoints were derived using linear interpolation and extrapolation methods. Mixed models included each predictor variable (PaO₂, ETCO₂, MAP, MFI, and %Capillary number) and time as fixed effect covariates with random effects of intercept and slope for time and each predictor variable. The hemodynamic, blood gas, and cerebral measurement data obtained at pre-arrest baseline and CPR data, including the number of electric shocks, number of epinephrine administrations, duration of CPR, and coronary perfusion pressure and MAP during CPR, were independently examined to evaluate their effectiveness in determining the PbtO₂. Significant variables (P value cutoff = 0.05) were further used for covariate adjustment in the mixed effect models. Semi-partial R-squared values of the fixed effects were computed for predictor variables and time using the r2glmm package of R software. A two-tailed P-value of <0.05 was considered statistically significant.

Results

Pre-arrest baseline measurements were within normal limits, as shown in Table 1. The PbtO₂ was 31.4 ± 6.6 torr at an FiO₂ of 0.3. Increasing the FiO₂ to 1.0 resulted in a marked increase in the PbtO₂ to 63.8 ± 18.1 torr. All animals achieved ROSC after 4 (2–5) min of CPR and survived the 60-min observation period. Thus, a total of 170 sets of MAP, ETCO₂, PaO₂, MFI, % Capillary number, and PbtO₂ data were collected during the 60-min post-resuscitation period

Weight (kg)	25.3 ± 1.3		
Systolic arterial pressure (mmHg)	134 ± 8		
Diastolic arterial pressure (mmHg)	90 ± 11		
Mean arterial pressure (mmHg)	108 ± 9		
Systolic right atrial pressure (mmHg)	13 (10–13)		
Diastolic right atrial pressure (mmHg)	5 (5-6)		
Mean right atrial pressure (mmHg)	9 ± 1		
Heart rate (/min)	101 (88–108)		
End-tidal carbon dioxide (mmHg)	45 (42–45)		
Rectal temperature (°C)	38.0 ± 0.4		
pH	7.502 ± 0.031		
PaCO ₂ (mmHg)	44.1 ± 3.4		
PaO ₂ (mmHg)	126.4 ± 21.4		
HCO ₃ ⁻ (mmol/l)	34.5 ± 2.3		
Lactate (mmol/l)	0.87 ± 0.37		
SaO ₂ (%)	99.0 (99.0-99.3)		
PbtO ₂ at FiO ₂ 0.3 (torr)	31.4 ± 6.6		
PbtO ₂ at FiO ₂ 1.0 (torr)	63.8 ± 18.1		
Microvascular flow index*	3		
Number of perfused capillaries (n)	13 ± 3		
fumber of electric shocks (n) 1 (1-2)			
Number of epinephrine administrations (n) 2 (1-2)			
Duration of CPR (min)	4 (2-5)		
Mean arterial pressure after 2 min of CPR (mmHg)	49.9 ± 10.2		
Coronary perfusion pressure after 2 min of CPR (mmHg)	17.5 ± 4.8		

Table 1. Pre-arrest baseline measurement and cardiopulmonary resuscitation data.

Data are presented as medians with interquartile ranges or mean ± standard deviation.

* The microvascular flow index at pre-arrest baseline was 3 in all animals. PaCO₂, partial pressure of carbon dioxide in arterial blood; PaO₂, partial pressure of oxygen in arterial blood; HCO₃⁻, bicarbonate; SaO₂, oxygen saturation in arterial blood; PbtO₂, brain tissue oxygen tension; FiO₂, fraction of inspired oxygen; CPR, cardiopulmonary resuscitation.

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(17 times in 10 animals) and included in the analyses. Fig 2 shows MAP, ETCO₂, PaO₂, MFI, %Capillary number, and PbtO₂ data during the 60-min post-resuscitation period. Immediately after ROSC, MAP increased rapidly, reaching a plateau within 5 min. It decreased progressively after 15 min, but it was maintained >65 mmHg. ETCO₂ and PbtO₂ progressively decreased during the 60-min period, while PaO₂ progressively increased. Immediately after ROSC, cerebral cortices became hyperemic; hence, there were more perfused capillaries than those at baseline for all animals. After a brief hyperemic period, cerebral cortices turned pale and the %Capillary number and MFI decreased progressively over the next 10 min. S1 Movie shows a representative video of the cerebral microcirculation. Despite the absence of arterial hypoxemia, hypocarbia, or hypotension, seven (70%) animals had cerebral hypoxia for 12.1 (6.6-33.5) min. Table 2 shows the fixed effects of time and each predictor variable on PbtO₂ in linear mixed effect models. In the linear mixed effect models, PaO₂ levels had a significant linear relationship with PbtO₂ after adjusting for significant covariates (semi-partial $R^2 = 0.175$; 95% confidence interval [CI], 0.086-0.282, P = 0.030), but neither MAP nor ETCO₂ were related to PbtO₂. Although the cerebral microcirculation variables were significantly associated with PbtO₂, their ability to explain PbtO₂ variance was limited. When adjusted for significant



Fig 2. Mean arterial pressure (A), arterial oxygen tension (B), end-tidal carbon dioxide (C), brain tissue oxygen tension (D), microvascular flow index (E), and % Capillary number (E) after the restoration of spontaneous circulation. Data are presented as medians with interquartile ranges (A–D) or mean \pm standard deviation (E and F). PaO₂, partial pressure of oxygen in arterial blood; %Capillary number, percent of counted number of perfused capillaries relative to that at the pre-arrest baseline.

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covariates, the MFI and %Capillary number explained 14.3% (semi-partial $R^2 = 0.143$; 95% CI = 0.062–0.247, P = 0.023) and 11.3% (semi-partial $R^2 = 0.113$; 95% CI = 0.040–0.212, P = 0.048) of the total PbtO₂ variance, respectively.

Discussion

Using data from an experimental study in which the animals did not experience hypoxemia or hypotension during the early post-resuscitation period, this study showed that PaO₂, but not

Variable	Coefficient	SE	P-value	Semi-partial R ²	95% CI
Time	-0.165	0.181	0.384	0.038	0.002-0.112
Mean arterial pressure (mmHg)	-0.090	0.155	0.578	0.028	0.001-0.095
Time*	-0.177	0.180	0.349	0.044	0.004-0.121
Mean arterial pressure (mmHg)*	-0.102	0.152	0.522	0.031	0.001-0.101
Time	0.153	0.320	0.643	0.022	0-0.086
PaO ₂	-0.120	0.056	0.053	0.271	0.170-0.380
Time*	0.192	0.324	0.567	0.026	0.001-0.093
PaO ₂ *	-0.135	0.055	0.030	0.175	0.086-0.282
Time	-0.177	0.226	0.452	0.033	0.001-0.103
End-tidal carbon dioxide (mmHg)	0.058	0.298	0.846	0.001	0-0.033
Time*	-0.181	0.227	0.442	0.034	0.002-0.106
End-tidal carbon dioxide (mmHg)*	0.037	0.299	0.901	0	0-0.031
Time	-0.135	0.223	0.559	0.023	0-0.087
Microvascular flow index	5.729	2.161	0.025	0.142	0.060-0.245
Time*	-0.132	0.221	0.564	0.022	0-0.085
Microvascular flow index*	5.777	2.125	0.023	0.143	0.062-0.247
Time	-0.119	0.238	0.628	0.018	0-0.077
%Capillary number	10.623	4.688	0.049	0.127	0.050-0.228
Time*	-0.122	0.235	0.616	0.019	0-0.079
%Capillary number*	10.507	4.632	0.048	0.113	0.040-0.212

Table 2. Fixed effects of time and each predictor variable on brain tissue oxygen tension in mixed effect models.

* Baseline end-tidal carbon dioxide and rectal temperature were used to adjust the mixed effect model. SE, standard error; CI, confidence interval; PaO₂, partial pressure of oxygen in arterial blood; %Capillary number, percent of counted number of perfused capillaries relative to that at the pre-arrest baseline.

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MAP nor $ETCO_2$, had a significant relationship with $PbtO_2$. To the best of our knowledge, this is the first study to examine the relationship between key determinants of cerebral oxygen delivery and $PbtO_2$ in non-hypoxemic, non-hypotensive conditions after cardiac arrest.

Several studies reported the frequent occurrence of cerebral hypoxia after ROSC despite adequate arterial oxygenation and perfusion pressure [20–22]. In a study that included 14 comatose cardiac arrest patients who underwent invasive neuromonitoring [22], 7 patients (50%) had a mean PbtO₂ level \leq 20 torr, which is a widely accepted hypoxic threshold [29]. Consistent with these studies [20–22], 70% of the animals in this study experienced PbtO₂ values < 20 torr in the absence of hypoxemia or hypotension. This finding indicates that a significant percentage of cardiac arrest patients may experience secondary hypoxic insults to the brain if the initial treatments are solely based on PaO₂ and MAP. PbtO₂ monitoring, although highly invasive, has been repeatedly reported as a safe procedure [16, 20, 22, 24, 33, 34]. This technique could help identify patients with ongoing cerebral hypoxia after ROSC who would not otherwise be detected.

Cerebral oxygenation can be monitored noninvasively by near-infrared spectroscopy (NIRS), which estimates regional tissue oxygenation by using light waves of near-infrared wavelength to penetrate the scalp, skull, and brain. Although NIRS is a non-invasive and easy-to-use technique for monitoring cerebral oxygenation, it measures the oxygen saturation of combined arterial, venous, and microcirculatory blood compartments in both cerebral and extracerebral tissue [35–37], which limits its accuracy and reliability for reflecting PbtO₂ levels [38, 39]. However, technological advances continue to improve the accuracy and usefulness of NIRS as a non-invasive tool for estimating the PbtO₂ [40]. Consequently, NIRS technology has the future potential to serve as a reliable and non-invasive surrogate of PbtO₂.

Although data regarding the significance of reduced PbtO₂ levels on clinical outcomes in cardiac arrest patients are unavailable, several studies involving patients with severe traumatic brain injury have suggested associations between reduced PbtO₂ levels and poor outcomes [15–18]. In a study investigating the relationship between low PbtO₂ values and outcomes in 101 comatose head-injured patients [15], the depth and duration of low PbtO₂ values during the first 24 h after injury correlated with unfavorable neurological outcomes and death at 6 mon. Studies have also suggested that reduced PbtO₂ can be a modifiable therapeutic target for neurological recovery [21, 33]. In a pig model of opioid overdose cardiac arrest, Elmer et al. reported that PbtO₂ puided care, compared with standard care, reduced the duration of exposure to PbtO₂ levels below the hypoxic threshold [21]. A phase 2 randomized trial including 119 patients with severe traumatic brain injury conducted by Okonkwo et al. compared a treatment protocol based on intracranial pressure (ICP) plus PbtO₂ monitoring, to a protocol based on PbtO₂ plus ICP monitoring reduced the time of exposure to cerebral hypoxia, with a trend toward lower mortality and favorable neurological recovery.

MAP is the main determinant of cerebral perfusion pressure, which is the driving force of CBF. The lack of a significant relationship between MAP and PbtO₂ in this study is best explained by the presence of cerebral autoregulation. In our data, the overall MAP values were above the suggested autoregulation threshold of 60–75 mmHg [24, 41, 42]. Several studies have suggested that the cerebral autoregulation mechanism, although it is often right shifted, operates in most cardiac arrest patients [20, 43]. In a study that included 51 comatose cardiac arrest patients who underwent MAP and regional cerebral oxygen saturation monitoring during the first 24 h of intensive care unit stay [43], cerebral autoregulation was preserved in about two thirds of the patients, whereas it was right-shifted in one third of the patients. In a study that assessed the relationship between MAP and brain tissue oxygenation in 10 cardiac arrest survivors [20], MAP was positively associated with PbtO₂ when MAP values were below the optimal MAP, but not when MAP values were above the optimal MAP, determined using the pressure reactivity index ($89 \pm 11 \text{ mmHg}$). Our finding suggests that in non-hypotensive cardiac arrest patients, MAP augmentation may not be able to influence PbtO₂ considerably. This finding is consistent with that of a recent randomized clinical trial that compared targeted low-normal (65-75 mmHg) and high-normal MAP (80-100 mmHg) in 123 comatose cardiac arrest patients [44], in which despite a clear separation in MAP between the two MAP target groups, the regional cerebral oxygen saturation (measured by using NIRS monitoring), neurobiomarkers (neuron-specific enolase and S100B), and clinical outcomes did not differ between the two groups.

Under physiological conditions, changes in $PaCO_2$ influence CBF considerably [45]. ETCO₂, a non-invasive surrogate of $PaCO_2$, is widely used to guide ventilator management in critically ill patients and is also related to CBF [46]. A study investigating ETCO₂ response curves for middle cerebral artery blood flow velocity (measured by using the transcranial Doppler technique) in 31 normal subjects showed that ETCO₂ was well correlated with the middle cerebral artery blood flow velocity [46]. Previous studies reported that cerebrovascular reactivity to $PaCO_2$ was preserved in patients resuscitated from cardiac arrest [47, 48]. In a study that investigated cerebrovascular reactivity to changes in $PaCO_2$ for 24 h after intensive care unit admission in 10 comatose out-of-hospital cardiac arrest patients [48], the mean percentage change in middle cerebral artery blood flow velocity per mmHg $PaCO_2$ change (3.6 ± 2.9%) was comparable to that reported in healthy subjects [49]. However, in contrast to these studies [47, 48], our study did not show a significant relationship between ETCO₂ and PbtO₂. The reason for this lack of a significant relationship between ETCO₂ and PbtO₂ is unclear, but it may be due to the blunted cerebrovascular reactivity occurring in the early postischemic period. Several studies have suggested that cerebrovascular reactivity to $PaCO_2$ is markedly blunted in the early post-ischemic period [50, 51]. In a dog model of post-ischemic cerebral hypoperfusion, cerebrovascular reactivity was markedly blunted after recovery from 15 min of global ischemia, while cerebral autoregulation was maintained [50]. In a study that evaluated the reactivity of $PbtO_2$ to changes in $PaCO_2$ over time in patients with severe head injury [52], the reactivity of $PbtO_2$ was initially abolished, but it recovered within the first two days after sustaining an injury.

Among the common target variables for PbtO₂-guided care that we investigated, only PaO₂ had a significant relationship with PbtO₂ in this study. This finding is consistent with that reported in other studies [19, 34]. In rats subjected to asphyxial cardiac arrest, reduced cortical PbtO₂ increased with an increase in FiO₂; however, it did not respond to increasing MAP with epinephrine infusion [19]. Our findings, together with the findings reported by the abovementioned studies [19, 34], suggest that to increase $PbtO_2$ in non-hypoxemic, non-hypotensive cardiac arrest patients, care should primarily focus on increasing PaO₂ rather than increasing ETCO₂ or MAP. However, the following points make the therapeutic efficacy of increasing PaO₂ questionable. Although the PaO₂ level had a significant linear association with the PbtO₂ level, it explained only 17.5% of the total PbtO₂ variance. This finding suggests that the extent to which one can increase PbtO₂ by increasing PaO₂ may not be high enough to be clinically useful. Its limited ability to explain the PbtO₂ variance may be related to impaired cerebral microcirculation, as evidenced by the reduced MFI and number of perfused capillaries during the post-resuscitation period. In several studies, the response of PbtO2 to supplemental oxygen diminished in the presence of reduced CBF [53, 54]. A study investigating the role of regional CBF in determining the response of PbtO₂ to induced hyperoxia in 83 patients with traumatic brain injury showed that the increase in PbtO2 induced by hyperoxia was smaller when regional CBF was < 20 ml/100 g/min than when it was > 20 ml/100 g/min [53]. In the presence of reduced PbtO₂ response to supplemental oxygen, it would be difficult to treat cerebral hypoxia by increasing FiO_2 without exposing the patients to arterial hyperoxemia, which has repeatedly been linked with poor outcomes after cardiac arrest [8, 55, 56].

In this study, the perfusion of cerebral capillaries, which serve as the ultimate exchange vessels for oxygen and vital substrates, explained only a small proportion of PbtO₂ variance. The limited capability of the MFI and %Capillary number to explain PbtO₂ variance, in addition to that of PaO₂, suggests that factors other than cerebral perfusion and blood oxygenation affect the PbtO₂ level. Recent studies have suggested that the diffusion limitation of oxygen delivery resulting from extensive perivascular edema and endothelial swelling is a plausible mechanism for cerebral hypoxia [22, 57]. A study assessing the relationship of the jugular venous oxygen tension-PbtO₂ gradient with cerebral perfusion pressure in 14 comatose cardiac arrest patients showed that increased cerebral perfusion pressure was associated with a decrease in the jugular venous oxygen tension-PbtO₂ gradient in patients without cerebral hypoxia (PbtO₂ > 20 mmHg) [22]. However, patients with cerebral hypoxia (PbtO₂ \leq 20 mmHg) exhibited an increased jugular venous oxygen tension-PbtO₂ gradient unrelated to varying cerebral perfusion pressure, suggesting diffusion limitation of oxygen delivery as a mechanism for cerebral hypoxia in those patients.

In the randomized trial by Okonkwo et al. in which adding PbtO₂-guided care significantly reduced the time of exposure to cerebral hypoxia in patients with severe traumatic brain injury [33], the PbtO₂ treatment protocol consisted of multiple physiological interventions to improve cerebral oxygen delivery, including increasing PaO₂, PaCO₂, and/or MAP, decreasing ICP, and performing red blood cell transfusion. However, our findings together with those of the aforementioned studies suggest that nonspecific interventions such as increasing FiO₂ alone are not likely to sufficiently increase the PbtO₂ level in cardiac arrest patients with cerebral hypoxia secondary to impaired cerebral microcirculation or diffusion limitation

physiology [22, 53, 54]. We believe that therapeutic strategies targeting diffusion limitation physiology and/or impaired cerebral microcirculation may be needed to effectively treat or prevent post-ROSC cerebral hypoxia occurring in the absence of hypoxemia or hypotension. For example, intrathecal sodium nitroprusside administration may be an effective treatment for cerebral microcirculatory impairment after ROSC. We found that it reversed the cerebral microcirculatory impairment after ROSC in pilot experiments. Based on these results, we can postulate that correction of diffusion limitation physiology and/or impaired cerebral microcirculation improves the PbtO₂ level itself, as well as the response of PbtO₂ to supplemental oxygen. We are planning future investigations on the effect of therapeutic strategies that target diffusion limitation physiology and/or impaired cerebral microcircirculation parameters and PbtO₂, These studies could illuminate why the PaO₂ had a limited capacity to explain PbtO₂ variance in the present study, and also help identify effective treatments for treating post-ROSC cerebral hypoxia.

Many further questions regarding the $PbtO_2$ after cardiac arrest should be addressed prior to clinical implementation of PbtO₂-guided care. First, the impacts of varying MAP, PaO₂, and $ETCO_2$ on PbtO₂ levels in other cerebral regions, or on global cerebral oxygenation, must be investigated. Since the cerebral measurements were restricted to a small area of the cerebral cortex in our study, our results cannot be extended to the entire brain. Experimental studies in rats have reported marked regional differences in CBF and cerebral oxygenation after cardiac arrest [19, 58]. Manole et al. investigated cerebral cortical and thalamic PbtO₂ levels in immature rats subjected to 9 min and 12 min asphyxial cardiac arrest [19], and reported that cerebral cortical hypoxia occurred despite the presence of hyperoxia in the thalamus. Second, the impacts of varying MAP, PaO₂, and ETCO₂ on cerebral oxygenation should be studied in different cardiac arrest models, specifically regarding the duration and etiology of cardiac arrest. In the study by Manole et al. [19], the reduction in PbtO₂ was greater, and the PbtO₂ response to supplemental oxygen was less pronounced, after a longer duration of asphyxia cardiac arrest. In addition, several studies have reported significant differences in patterns of CBF reperfusion and cerebral injury between VF and asphyxial cardiac arrest [59, 60]. Third, although a number of studies have suggested the benefits of correcting cerebral hypoxia in traumatic brain injury [15-18, 33], studies assessing whether the correction of cerebral hypoxia is beneficial after cardiac arrest are limited [21, 22]. Therefore, further experiments assessing the relationship between PbtO₂ and cerebral metabolic and neurophysiological markers after cardiac arrest are warranted. Additionally, further studies to determine the effects of correcting cerebral hypoxia on histopathological and functional neurological outcomes after cardiac arrest are crucial.

Our study has several important limitations. First, data were obtained from anesthetized animals free of an underlying disease. Despite similarities in neuroanatomy between pigs and humans, direct extrapolation of our findings to human cardiac arrest patients cannot be assumed. Second, we retrospectively analyzed data derived from a previous study. Our study could not determine causal relationships between the common target variables and PbtO₂. Third, the data were obtained from a study conducted in a well-controlled experimental setting. Variation in factors related to cardiac arrest and resuscitation, including arrest etiology and duration of arrest, in the clinical setting may result in different results. Fourth, other factors influencing PbtO₂, including ICP and oxygen consumption, could confound the relationship between the common target variables and PbtO₂, but they were not included in our study.

Conclusions

In the post-hoc analysis of data from an experimental study in which the animals did not experience hypoxemia or hypotension during the early post-resuscitation period, MAP and ETCO₂

were not significantly related to $PbtO_2$. The PaO_2 level had a significant linear association with the $PbtO_2$ level, but its ability to explain $PbtO_2$ variance was small.

Supporting information

S1 Movie. Representative cerebral microcirculation video. It shows changes in cerebral cortical microcirculation during the first 15 min of the post-resuscitation period. (MP4)

S1 Data. Raw data. (XLSX)

S1 File. ARRIVE guidelines checklist. (DOC)

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References

- Lemiale V, Dumas F, Mongardon N, Giovanetti O, Charpentier J, Chiche JD, et al. Intensive care unit mortality after cardiac arrest: the relative contribution of shock and brain injury in a large cohort. Intensive Care Med. 2013; 39: 1972–1980. https://doi.org/10.1007/s00134-013-3043-4 PMID: 23942856
- Ong ME, Shin SD, De Souza NN, Tanaka H, Nishiuchi T, Song KJ, et al. Outcomes for out-of-hospital cardiac arrests across 7 countries in Asia: The Pan Asian Resuscitation Outcomes Study (PAROS). Resuscitation. 2015; 96: 100–108. https://doi.org/10.1016/j.resuscitation.2015.07.026 PMID: 26234891
- Perkins GD, Ji C, Deakin CD, Quinn T, Nolan JP, Scomparin C, et al. A Randomized Trial of Epinephrine in Out-of-Hospital Cardiac Arrest. N Engl J Med. 2018; 379: 711–721. <u>https://doi.org/10.1056/ NEJMoa1806842</u> PMID: 30021076
- Trzeciak S, Jones AE, Kilgannon JH, Milcarek B, Hunter K, Shapiro NI, et al. Significance of arterial hypotension after resuscitation from cardiac arrest. Crit Care Med. 2009; 37: 2895–2903. <u>https://doi.org/10.1097/ccm.0b013e3181b01d8c PMID: 19866506</u>

- Müllner M, Sterz F, Binder M, Hellwagner K, Meron G, Herkner H, et al. Arterial blood pressure after human cardiac arrest and neurological recovery. Stroke. 1996; 27: 59–62. https://doi.org/10.1161/01. str.27.1.59 PMID: 8553404
- Kilgannon JH, Roberts BW, Jones AE, Mittal N, Cohen E, Mitchell J, et al. Arterial blood pressure and neurologic outcome after resuscitation from cardiac arrest*. Crit Care Med. 2014; 42: 2083–2091. https://doi.org/10.1097/CCM.0000000000406 PMID: 24901606
- Bhate TD, McDonald B, Sekhon MS, Griesdale DE. Association between blood pressure and outcomes in patients after cardiac arrest: A systematic review. Resuscitation. 2015; 97: 1–6. https://doi.org/10. 1016/j.resuscitation.2015.08.023 PMID: 26410569
- Kilgannon JH, Jones AE, Shapiro NI, Angelos MG, Milcarek B, Hunter K, et al. Association between arterial hyperoxia following resuscitation from cardiac arrest and in-hospital mortality. JAMA. 2010; 303: 2165–2171. https://doi.org/10.1001/jama.2010.707 PMID: 20516417
- Roberts BW, Kilgannon JH, Chansky ME, Mittal N, Wooden J, Trzeciak S. Association between postresuscitation partial pressure of arterial carbon dioxide and neurological outcome in patients with post-cardiac arrest syndrome. Circulation. 2013; 127: 2107–2113. https://doi.org/10.1161/CIRCULATIONAHA. 112.000168 PMID: 23613256
- Schneider AG, Eastwood GM, Bellomo R, Bailey M, Lipcsey M, Pilcher D, et al. Arterial carbon dioxide tension and outcome in patients admitted to the intensive care unit after cardiac arrest. Resuscitation. 2013; 84: 927–934. https://doi.org/10.1016/j.resuscitation.2013.02.014 PMID: 23454258
- Callaway CW, Soar J, Aibiki M, Böttiger BW, Brooks SC, Deakin CD, et al. Part 4: Advanced Life Support: 2015 International Consensus on Cardiopulmonary Resuscitation and Emergency Cardiovascular Care Science With Treatment Recommendations. Circulation. 2015; 132: S84–S145. https://doi.org/10.1161/CIR.00000000000273 PMID: 26472860
- Panchal AR, Bartos JA, Cabañas JG, Donnino MW, Drennan IR, Hirsch KG, et al. Part 3: Adult Basic and Advanced Life Support: 2020 American Heart Association Guidelines for Cardiopulmonary Resuscitation and Emergency Cardiovascular Care. Circulation. 2020; 142: S366–S468. <u>https://doi.org/10. 1161/CIR.00000000000916</u> PMID: 33081529
- Clark LC Jr, Kaplan S, Matthews EC, Edwards FK, Helmsworth JA. Monitor and control of blood oxygen tension and pH during total body perfusion. J Thorac Surg. 1958; 36: 488–496. PMID: 13588706
- 14. Peterson JI, Fitzgerald RV, Buckhold DK. Fiber-optic probe for in vivo measurement of oxygen partial pressure. Anal Chem. 1984; 56: 62–67. https://doi.org/10.1021/ac00265a017 PMID: 6696231
- van den Brink WA, van Santbrink H, Steyerberg EW, Avezaat CJ, Suazo JA, Hogesteeger C, et al. Brain oxygen tension in severe head injury. Neurosurgery. 2000; 46: 868–876. <u>https://doi.org/10.1097/00006123-200004000-00018 PMID: 10764260</u>
- Valadka AB, Gopinath SP, Contant CF, Uzura M, Robertson CS. Relationship of brain tissue PO2 to outcome after severe head injury. Crit Care Med. 1998; 26: 1576–1581. <u>https://doi.org/10.1097/</u> 00003246-199809000-00029 PMID: 9751596
- Goodman JC, Valadka AB, Gopinath SP, Uzura M, Robertson CS. Extracellular lactate and glucose alterations in the brain after head injury measured by microdialysis. Crit Care Med. 1999; 27: 1965– 1973. https://doi.org/10.1097/00003246-199909000-00041 PMID: 10507626
- Maloney-Wilensky E, Gracias V, Itkin A, Hoffman K, Bloom S, Yang W, et al. Brain tissue oxygen and outcome after severe traumatic brain injury: a systematic review. Crit Care Med. 2009; 37: 2057–2063. https://doi.org/10.1097/CCM.0b013e3181a009f8 PMID: 19384213
- Manole MD, Kochanek PM, Bayır H, Alexander H, Dezfulian C, Fink EL, et al. Brain tissue oxygen monitoring identifies cortical hypoxia and thalamic hyperoxia after experimental cardiac arrest in rats. Pediatr Res. 2014; 75: 295–301. https://doi.org/10.1038/pr.2013.220 PMID: 24226633
- Sekhon MS, Gooderham P, Menon DK, Brasher PMA, Foster D, Cardim D, et al. The Burden of Brain Hypoxia and Optimal Mean Arterial Pressure in Patients With Hypoxic Ischemic Brain Injury After Cardiac Arrest. Crit Care Med. 2019; 47: 960–969. <u>https://doi.org/10.1097/CCM.00000000003745</u> PMID: 30889022
- Elmer J, Flickinger KL, Anderson MW, Koller AC, Sundermann ML, Dezfulian C, et al. Effect of neuromonitor-guided titrated care on brain tissue hypoxia after opioid overdose cardiac arrest. Resuscitation. 2018; 129: 121–126. https://doi.org/10.1016/j.resuscitation.2018.04.013 PMID: 29679696
- Sekhon MS, Ainslie PN, Menon DK, Thiara SS, Cardim D, Gupta AK, et al. Brain Hypoxia Secondary to Diffusion Limitation in Hypoxic Ischemic Brain Injury Postcardiac Arrest. Crit Care Med. 2020; 48: 378– 384. https://doi.org/10.1097/CCM.00000000004138 PMID: 31789834
- Cavus E, Bein B, Dörges V, Stadlbauer KH, Wenzel V, Steinfath M, et al. Brain tissue oxygen pressure and cerebral metabolism in an animal model of cardiac arrest and cardiopulmonary resuscitation. Resuscitation. 2006; 71: 97–106. https://doi.org/10.1016/j.resuscitation.2006.03.007 PMID: 16942830

- Kiening KL, Unterberg AW, Bardt TF, Schneider GH, Lanksch WR. Monitoring of cerebral oxygenation in patients with severe head injuries: brain tissue PO2 versus jugular vein oxygen saturation. J Neurosurg. 1996; 85: 751–757. https://doi.org/10.3171/jns.1996.85.5.0751 PMID: 8893710
- Leidorf A, Mader MM, Hecker A, Heimann A, Alessandri B, Mayr P, et al. Description of the response of a new multi-parametric brain sensor to physiological and pathophysiological challenges in the cortex of juvenile pigs. Turk Neurosurg. 2014; 24: 913–922. https://doi.org/10.5137/1019-5149.JTN.11808-14.1 PMID: 25448209
- Mader MM, Leidorf A, Hecker A, Heimann A, Mayr PSM, Kempski O, et al. Evaluation of a New Multiparameter Brain Probe for Simultaneous Measurement of Brain Tissue Oxygenation, Cerebral Blood Flow, Intracranial Pressure, and Brain Temperature in a Porcine Model. Neurocrit Care. 2018; 29: 291– 301. https://doi.org/10.1007/s12028-018-0541-9 PMID: 29949006
- Lee HY, Mamadjonov N, Jeung KW, Jung YH, Lee BK, Moon KS, et al. Pralidoxime-Induced Potentiation of the Pressor Effect of Adrenaline and Hastened Successful Resuscitation by Pralidoxime in a Porcine Cardiac Arrest Model. Cardiovasc Drugs Ther. 2020; 34: 619–628. <u>https://doi.org/10.1007/</u> s10557-020-07026-5 PMID: 32562104
- Cho YC, Cho SW, Chung SP, Yu K, Kwon OY, Kim SW. How can a single rescuer adequately deliver tidal volume with a manual resuscitator? An improved device for delivering regular tidal volume. Emerg Med J. 2011; 28: 40–3. https://doi.org/10.1136/emj.2010.099911 PMID: 21131393
- 29. Le Roux P, Menon DK, Citerio G, Vespa P, Bader MK, Brophy GM, et al. Consensus summary statement of the International Multidisciplinary Consensus Conference on Multimodality Monitoring in Neurocritical Care: a statement for healthcare professionals from the Neurocritical Care Society and the European Society of Intensive Care Medicine. Intensive Care Med. 2014; 40: 1189–1209. https://doi. org/10.1007/s00134-014-3369-6 PMID: 25138226
- Spronk PE, Ince C, Gardien MJ, Mathura KR, Oudemans-van Straaten HM, Zandstra DF. Nitroglycerin in septic shock after intravascular volume resuscitation. Lancet. 2002; 360: 1395–1396. https://doi.org/ 10.1016/s0140-6736(02)11393-6 PMID: 12423989
- Boerma EC, Mathura KR, van der Voort PH, Spronk PE, Ince C. Quantifying bedside-derived imaging of microcirculatory abnormalities in septic patients: a prospective validation study. Crit Care. 2005; 9: R601–R606. https://doi.org/10.1186/cc3809 PMID: 16280059
- Serné EH, Gans RO, ter Maaten JC, Tangelder GJ, Donker AJ, Stehouwer CD. Impaired skin capillary recruitment in essential hypertension is caused by both functional and structural capillary rarefaction. Hypertension. 2001; 38: 238–42. https://doi.org/10.1161/01.hyp.38.2.238 PMID: 11509483
- Okonkwo DO, Shutter LA, Moore C, Temkin NR, Puccio AM, Madden CJ, et al. Brain Oxygen Optimization in Severe Traumatic Brain Injury Phase-II: A Phase II Randomized Trial. Crit Care Med. 2017; 45: 1907–1914. https://doi.org/10.1097/CCM.00000000002619 PMID: 29028696
- 34. van Santbrink H, Maas AI, Avezaat CJ. Continuous monitoring of partial pressure of brain tissue oxygen in patients with severe head injury. Neurosurgery. 1996; 38: 21–31. https://doi.org/10.1097/00006123-199601000-00007 PMID: 8747947
- **35.** Ter Minassian A, Poirier N, Pierrot M, Menei P, Granry JC, Ursino M, et al. Correlation between cerebral oxygen saturation measured by near-infrared spectroscopy and jugular oxygen saturation in patients with severe closed head injury. Anesthesiology. 1999; 91: 985–990. https://doi.org/10.1097/00000542-199910000-00018 PMID: 10519501
- Germon TJ, Kane NM, Manara AR, Nelson RJ. Near-infrared spectroscopy in adults: effects of extracranial ischaemia and intracranial hypoxia on estimation of cerebral oxygenation. Br J Anaesth. 1994; 73: 503–506. https://doi.org/10.1093/bja/73.4.503 PMID: 7999492
- Bein B, Cavus E, Stadlbauer KH, Tonner PH, Steinfath M, Scholz J, et al. Monitoring of cerebral oxygenation with near infrared spectroscopy and tissue oxygen partial pressure during cardiopulmonary resuscitation in pigs. Eur J Anaesthesiol. 2006; 23: 501–509. <u>https://doi.org/10.1017/</u> S0265021506000366 PMID: 16507191
- Putzer G, Braun P, Strapazzon G, Toferer M, Mulino M, Glodny B, et al. Monitoring of brain oxygenation during hypothermic CPR—A prospective porcine study. Resuscitation. 2016; 104: 1–5. <u>https://doi.org/ 10.1016/j.resuscitation.2016.03.027</u> PMID: 27095125
- Nelskylä A, Nurmi J, Jousi M, Schramko A, Mervaala E, Ristagno G, et al. The effect of 50% compared to 100% inspired oxygen fraction on brain oxygenation and post cardiac arrest mitochondrial function in experimental cardiac arrest. Resuscitation. 2017; 116: 1–7. https://doi.org/10.1016/j.resuscitation. 2017.04.022 PMID: 28438718
- Wang CC, Kuo JR, Chen YC, Chio CC, Wang JJ, Lin BS. Brain tissue oxygen evaluation by wireless near-infrared spectroscopy. J Surg Res. 2016; 200: 669–675. https://doi.org/10.1016/j.jss.2015.10.005 PMID: 26521677

- Hemphill JC 3rd, Knudson MM, Derugin N, Morabito D, Manley GT. Carbon dioxide reactivity and pressure autoregulation of brain tissue oxygen. Neurosurgery. 2001; 48: 377–383. https://doi.org/10.1097/00006123-200102000-00028 PMID: 11220382
- Jaeger M, Dengl M, Meixensberger J, Schuhmann MU. Effects of cerebrovascular pressure reactivityguided optimization of cerebral perfusion pressure on brain tissue oxygenation after traumatic brain injury. Crit Care Med. 2010; 38: 1343–1347. https://doi.org/10.1097/CCM.0b013e3181d45530 PMID: 20154598
- 43. Ameloot K, Genbrugge C, Meex I, Jans F, Boer W, Vander Laenen M, et al. An observational near-infrared spectroscopy study on cerebral autoregulation in post-cardiac arrest patients: time to drop 'onesize-fits-all' hemodynamic targets? Resuscitation. 2015; 90: 121–126. <u>https://doi.org/10.1016/j.</u> resuscitation.2015.03.001 PMID: 25769511
- 44. Jakkula P, Pettilä V, Skrifvars MB, Hästbacka J, Loisa P, Tiainen M, et al. Targeting low-normal or highnormal mean arterial pressure after cardiac arrest and resuscitation: a randomised pilot trial. Intensive Care Med. 2018; 44: 2091–2101. https://doi.org/10.1007/s00134-018-5446-8 PMID: 30443729
- 45. Reivich M. Arterial PCO2 and cerebral hemodynamics. Am J Physiol. 1964; 206: 25–35. https://doi.org/ 10.1152/ajplegacy.1964.206.1.25 PMID: 14117646
- 46. Markwalder TM, Grolimund P, Seiler RW, Roth F, Aaslid R. Dependency of blood flow velocity in the middle cerebral artery on end-tidal carbon dioxide partial pressure—a transcranial ultrasound Doppler study. J Cereb Blood Flow Metab. 1984; 4: 368–372. https://doi.org/10.1038/jcbfm.1984.54 PMID: 6432808
- Buunk G, van der Hoeven JG, Meinders AE. Cerebrovascular reactivity in comatose patients resuscitated from a cardiac arrest. Stroke. 1997; 28: 1569–1573. https://doi.org/10.1161/01.str.28.8.1569 PMID: 9259750
- Bisschops LL, Hoedemaekers CW, Simons KS, van der Hoeven JG. Preserved metabolic coupling and cerebrovascular reactivity during mild hypothermia after cardiac arrest. Crit Care Med. 2010; 38: 1542– 1547. https://doi.org/10.1097/CCM.0b013e3181e2cc1e PMID: 20453643
- Klingelhöfer J, Sander D. Doppler CO2 test as an indicator of cerebral vasoreactivity and prognosis in severe intracranial hemorrhages. Stroke. 1992; 23: 962–966. https://doi.org/10.1161/01.str.23.7.962
 PMID: 1615545
- Nemoto EM, Snyder JV, Carroll RG, Morita H. Global ischemia in dogs: cerebrovascular CO2 reactivity and autoregulation. Stroke. 1975; 6: 425–431. https://doi.org/10.1161/01.str.6.4.425 PMID: 1154479
- Kågström E, Smith ML, Siesjö BK. Cerebral circulatory responses to hypercapnia and hypoxia in the recovery period following complete and incomplete cerebral ischemia in the rat. Acta Physiol Scand. 1983; 118: 281–291. https://doi.org/10.1111/j.1748-1716.1983.tb07272.x PMID: 6414251
- Dings J, Meixensberger J, Amschler J, Hamelbeck B, Roosen K. Brain tissue pO2 in relation to cerebral perfusion pressure, TCD findings and TCD-CO2-reactivity after severe head injury. Acta Neurochir (Wien). 1996; 138: 425–434. https://doi.org/10.1007/BF01420305 PMID: 8738393
- Hlatky R, Valadka AB, Gopinath SP, Robertson CS. Brain tissue oxygen tension response to induced hyperoxia reduced in hypoperfused brain. J Neurosurg. 2008; 108: 53–58. https://doi.org/10.3171/JNS/ 2008/108/01/0053 PMID: 18173310
- Rossi S, Longhi L, Balestreri M, Spagnoli D, deLeo A, Stocchetti N. Brain oxygen tension during hyperoxia in a swine model of cerebral ischaemia. Acta Neurochir Suppl. 2000; 76: 243–245. <u>https://doi.org/ 10.1007/978-3-7091-6346-7_49 PMID: 11450016</u>
- 55. Janz DR, Hollenbeck RD, Pollock JS, McPherson JA, Rice TW. Hyperoxia is associated with increased mortality in patients treated with mild therapeutic hypothermia after sudden cardiac arrest. Crit Care Med. 2012; 40: 3135–3139. https://doi.org/10.1097/CCM.0b013e3182656976 PMID: 22971589
- 56. Roberts BW, Kilgannon JH, Hunter BR, Puskarich MA, Pierce L, Donnino M, et al. Association Between Early Hyperoxia Exposure After Resuscitation From Cardiac Arrest and Neurological Disability: Prospective Multicenter Protocol-Directed Cohort Study. Circulation. 2018; 137: 2114–2124. https://doi. org/10.1161/CIRCULATIONAHA.117.032054 PMID: 29437118
- Menon DK, Coles JP, Gupta AK, Fryer TD, Smielewski P, Chatfield DA, et al. Diffusion limited oxygen delivery following head injury. Crit Care Med. 2004; 32: 1384–1390. <u>https://doi.org/10.1097/01.ccm.</u> 0000127777.16609.08 PMID: 15187523
- Manole MD, Foley LM, Hitchens TK, Kochanek PM, Hickey RW, Bayir H, et al. Magnetic resonance imaging assessment of regional cerebral blood flow after asphyxial cardiac arrest in immature rats. J Cereb Blood Flow Metab. 2009; 29: 197–205. https://doi.org/10.1038/jcbfm.2008.112 PMID: 18827831
- Vaagenes P, Safar P, Moossy J, Rao G, Diven W, Ravi C, et al. Asphyxiation versus ventricular fibrillation cardiac arrest in dogs. Differences in cerebral resuscitation effects—a preliminary study. Resuscitation. 1997; 35: 41–52. https://doi.org/10.1016/s0300-9572(97)01108-8 PMID: 9259060

60. Drabek T, Foley LM, Janata A, Stezoski J, Hitchens TK, Manole MD, et al. Global and regional differences in cerebral blood flow after asphyxial versus ventricular fibrillation cardiac arrest in rats using ASL-MRI. Resuscitation. 2014; 85: 964–971. https://doi.org/10.1016/j.resuscitation.2014.03.314 PMID: 24727136