



# Transient receptor potential vanilloid subtype 1 depletion mediates mechanical allodynia through cellular signal alterations in small-fiber neuropathy

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## Abstract

Transient receptor potential vanilloid subtype 1 (TRPV1) is a polymodal nociceptor that monitors noxious thermal sensations. Few studies have addressed the role of TRPV1 in mechanical allodynia in small-fiber neuropathy (SFN) caused by sensory nerve damage. Accordingly, this article reviews the putative mechanisms of TRPV1 depletion that mediates mechanical allodynia in SFN. The intraepidermal nerve fibers (IENFs) degeneration and sensory neuronal injury are the primary characteristics of SFN. Intraepidermal nerve fibers are mainly C-polymodal nociceptors and A $\delta$ -fibers, which mediated allodynic pain after neuronal sensitization. TRPV1 depletion by highly potent neurotoxins induces the upregulation of activating transcription factor 3 and IENFs degeneration which mimics SFN. TRPV1 is predominately expressed by the peptidergic than nonpeptidergic nociceptors, and these neurochemical discrepancies provided the basis of the distinct pathways of thermal analgesia and mechanical allodynia. The depletion of peptidergic nociceptors and their IENFs cause thermal analgesia and sensitized nonpeptidergic nociceptors respond to mechanical allodynia. These distinct pathways of noxious stimuli suggested determined by the neurochemical-dependent neurotrophin cognate receptors such as TrkA and Ret receptors. The neurogenic inflammation after TRPV1 depletion also sensitized Ret receptors which results in mechanical allodynia. The activation of spinal TRPV1(+) neurons may contribute to mechanical allodynia. Also, an imbalance in adenosinergic analgesic signaling in sensory neurons such as the downregulation of prostatic acid phosphatase and adenosine A<sub>1</sub> receptors, which colocalized with TRPV1 as a membrane microdomain also correlated with the development of mechanical allodynia. Collectively, TRPV1 depletion-induced mechanical allodynia involves a complicated cascade of cellular signaling alterations.

**Keywords:** P2X3, TrkA receptor, Ret receptor, Prostatic acid phosphatase, Adenosine A<sub>1</sub> receptor, Calcitonin gene-related protein, Activating transcription factor 3

## 1. Introduction

The transient receptor potential vanilloid subtype 1 (TRPV1), also known as capsaicin receptor, is encoded in humans by the *trpv1* gene. The TRP family of receptors (including several subtypes)<sup>61</sup> and their physiological function are well established in the literature.<sup>16,17,61</sup> TRPV1 was first cloned by Caterina et al.<sup>17</sup> and has been a well-known receptor. Transient receptor potential vanilloid subtype 1 is a nonselective ion channel and a polymodal nociceptor<sup>16,153</sup> used to detect and regulate body temperature as well as respond to heat and pain signals.<sup>128</sup> Transient receptor

potential vanilloid subtype 1 acts as a thermoreceptor.<sup>16</sup> Transient receptor potential vanilloid subtype 1 is expressed by small-diameter nociceptors, and TRPV1 depletion has specifically been reported to result in thermal analgesia.<sup>16,61</sup> The molecular mechanisms of TRPV1-mediated thermal analgesia are related to the cytotoxicity induced by increased calcium permeability and the influx of Ca<sup>2+</sup> into TRPV1(+) nociceptors.<sup>39,42</sup> Patients with small-fiber neuropathy (SFN) experience various nociceptive sensations<sup>64,160</sup> such as reduced noxious thermal sensation (thermal analgesia) and mechanical

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hypersensitivity (mechanical allodynia). This suggests that TRPV1(+) nociceptors regulate both thermal and mechanical nociception in SFN. This review article discusses the molecular mechanisms of TRPV1 associated with mechanical allodynia.

### 1.1. Roles of transient receptor potential vanilloid subtype 1 in small-fiber neuropathy models

The skin is innervated by intraepidermal nerve fibers (IENFs) which respond to external stimuli, and skin denervation is a common clinicopathological manifestation of SFN.<sup>10,19,20,26,35,83,147</sup> Neuroanatomically, these IENFs are mainly C-polymodal nociceptors and A $\delta$ -fibers, which mediated allodynic pain confirmed by neuropharmacological blockade,<sup>50,57,59</sup> evoked potentials recordings,<sup>57,93,145</sup> and genetic expression.<sup>57</sup> Particularly, the C-polymodal nociceptors are further categorized as mechano-heat-responsive units and mechano-insensitive units (CMI) according to the different electrical thresholds. Past studies suggested CMI became sensitized by tissue damage (ie, skin denervation),<sup>130</sup> nerve growth factor (NGF),<sup>113,114,141</sup> and capsaicin administration.<sup>140</sup> Therefore, the neuropathology of skin denervation provided a basis for evaluating the alteration of pain perception. The combination of skin biopsy and protein gene product 9.5-immunohistochemistry is a useful clinical diagnostic tool for assessing skin denervation in patients with SFN. It has been reported to provide reliable diagnosis and progress prediction in SFN.<sup>68–71,97</sup> This skin biopsy assessment has also been applied to various animal models of SFN, particularly in cases of degeneration of TRPV1(+) IENFs,<sup>45,51,53,58,126</sup> which may sensitize their neuronal soma in the dorsal root ganglia (DRG).<sup>50</sup> The benefit of these animal models of SFN is that they allow the simultaneous assessment of skin denervation and the degree of neuronal soma injury. For example, the different pathophysiological responses of IENFs and TRPV1 (+) neuronal soma may have paradoxical neurophysiological outcomes.<sup>50,51</sup> However, studies using animal models of diabetes-induced SFN have not addressed the factors affecting the development of neuropathic pain by sensitized and/or irritated large sensory neurons<sup>81,115</sup> and identifying these factors may delineate the role of TRPV1 in the development of mechanical allodynia, which has a high incidence among patients with diabetes-induced SFN.

Studies have used highly selective neurotoxins to TRPV1 such as topical capsaicin<sup>21,36,66,78</sup> and systemic resiniferatoxin (RTX) treatment<sup>14,56,77</sup> to induce skin denervation, confirming the role of thermal transmission in SFN.<sup>16,17,153</sup> Genetic knockout of TRPV1 has resulted in similar outcomes.<sup>78,87,101</sup> In another study, highly selective and highly potent neurotoxins depleted TRPV1, inducing pure SFN that spared large sensory nerves.<sup>51</sup> Furthermore, Pan et al.<sup>118</sup> and our research group<sup>51</sup> demonstrated that TRPV1(+) nociceptors had a paradoxical effect on thermal and mechanical sensitivity after systemic neurotoxicity was induced in the naïve rodent. The genetic depletion of TRPV1 also reversed mechanical allodynia that was induced in a neurogenic inflammation model.<sup>87</sup> Collectively, it is believed the role of TRPV1 in SFN may involve several neuronal and immune responses,<sup>50,52,90,92,118</sup> suggesting that alteration of TRPV1 pathophysiology is a critical factor.

### 1.2. Transient receptor potential vanilloid subtype 1-expressing peptidergic vs nonpeptidergic nociceptors

Small-diameter nociceptors are categorized into 2 types according to their neurotrophic characteristics: peptidergic and nonpeptidergic nociceptors. TRPV1 is predominantly expressed by

peptidergic nociceptors<sup>53,125</sup> and less commonly coexpressed by nonpeptidergic nociceptors.<sup>50</sup> The peripheral processes of these peptidergic and nonpeptidergic nociceptors terminate on skin response to different types of noxious stimuli.<sup>50,52,53</sup> The colocalized ratios of TRPV1 and peptidergic(+) nociceptors are correlated with different neurophysiological outcomes. For example, approximately 20%–50% of calcitonin gene-related peptide (CGRP) (+) nociceptors coexpress TRPV1 in the DRG of rodents,<sup>53,125</sup> and the reinnervation of CGRP (+) IENFs reverses thermal analgesia after TRPV1 depletion,<sup>51,53</sup> which may due to TRPV1 depletion alter the expression of neuronal phenotypes and neurophysiological functions.<sup>151,152</sup> By contrast, denervation of substance P(+) IENFs reportedly results in a loss of thermal response ability because of the low density of IENFs and high colocalization (40%–60%) with TRPV1.<sup>53,125</sup> These peptidergic neuronal soma exhibit the same pathology as their peripheral IENFs.<sup>51</sup> In addition to thermal transmission, the depletion of these peptidergic neurons after TRPV1 depletion improved periodontitis,<sup>13</sup> arthritis,<sup>47</sup> and orthodontic force-derived mechanical irritation<sup>157</sup> by altered inflammatory response. Collectively, the elimination of peptidergic neurons after TRPV1 depletion may alter systemic neurophysiological function which involved inflammatory responses.

Pathological evidence indicates that the ratio of TRPV1 colocalized with different phenotypic nociceptors is a critical factor for different neuropathic behaviors. For example, TRPV1 has limited expression by nonpeptidergic (also called purinergic) P2X3 nociceptors, and specifically, P2X3(+) nociceptors sensitized by highly potent neurotoxins respond to mechanical allodynia<sup>15,62</sup> because of the burst release of adenosine triphosphate (ATP) from denervated skin and injured DRG tissues.<sup>50</sup> The other desensitized nonpeptidergic nociceptors that coexpress with TRPV1 also exhibit similar neuropathic behaviors. For example, the downregulation of the prostatic acid phosphatase (PAP), which is highly coexpressed with P2X3(+) nociceptors (up to 87%) and approximately 50% coexpressed with TRPV1, correlates with the development of mechanical allodynia.<sup>65,162</sup> In addition to TRPV1 (+) neuronal soma and their peripheral IENFs correlated to the noxious stimulation, presynaptic TRPV1 (+) central terminals on the spinal cord suggested modulated the postsynaptic current activities.<sup>55,105</sup> This neurophysiological modulation by TRPV1 is further confirmed by c-fos activation on postsynaptic spinal interneurons after TRPV1 activation at the periphery.<sup>46,119</sup> The expression of TRPV1 (+) central terminals and interneurons showed their neurophysiological plasticity that suggested commonly contribute to painful sensation, which was regulated by the pathways from the ventromedial medulla and thalamus in brainstem and diencephalon, respectively, to spinal TRPV1 (+) interneurons.<sup>22</sup> Particularly, TRPV1 activation expressed by spinal GABAergic interneurons mediated mechanical allodynia by the disinhibition of long-term depression in the spinothalamic tract.<sup>75</sup> Collectively, topographical TRPV1 expression played an important role in plastic neuronal activity for pain transmission.

Notably, the neurophysiological functions of small-diameter nociceptors depend on the regulation of neurotrophin signals; the reinnervation of CGRP (+) peptidergic IENFs normalizes the thermal noxious sensation that underlies the activation of NGF-TrkA signaling.<sup>48,52,53</sup> TrkA is the high-affinity receptor of NGF that regulates the neurophysiological functions of peptidergic nociceptors. NGF-TrkA signaling also regulates the neurophysiological functions of nonpeptidergic PAP (+) nociceptors through high PAP/TrkA colocalization ratios (approximately 70%).<sup>162</sup> However, 2 studies have reported that the activation

of NGF-TrkA signaling has no effect on TRPV1 recovery, despite TRPV1/TrkA colocalization ratios of approximately 30%–50%.<sup>52,79</sup> Furthermore, NGF has been found to normalize neuropathic manifestations through distinct neurotrophin receptor–dependent pathways.<sup>52</sup> For example, NGF has paradoxical effects, such as upregulation of TrkA, the peptidergic nociceptor–dependent neurotrophin receptor, and down-regulation of the nonpeptidergic nociceptor–dependent Ret receptor.<sup>52</sup> Evidence from pharmacological interventions indicates that the activation of Ret receptors mediates both thermal and mechanical noxious sensations and that TrkA receptors have no effect on mechanosensation. Notably, the expression of different neurotrophin cognate receptors such as activating transcription factor 3 (ATF3) is associated with intranuclear signaling, suggesting that these injury-dependent molecules expressed by different neurotrophin cognate receptors are critical factors in the development of neuropathic pain.

The different phenotypes of small-diameter nociceptors have a wide spectrum of profiles colocalized with TRPV1 that exhibit distinct neurophysiological functions, such as the induction of both thermal analgesia and mechanical allodynia after TRPV1 depletion, which is mediated by 2 distinct noxious pathways regulated by specific neurotrophins and their cognate receptors.<sup>52</sup>

### **1.3. Correlation of neuronal soma injury after transient receptor potential vanilloid subtype 1 depletion with neurotrophic receptor expression and pain development**

Transient receptor potential vanilloid subtype 1 depletion and the degeneration of IENFs cause a cascade of responses in the neuronal soma. For example, ATF3 is a member of the ATF/CREB transcription factor superfamily.<sup>40</sup> Activating transcription factor 3 dimerizes with c-Jun, which responds to neuronal irritation and outgrowth on different types of neurons.<sup>91</sup> In addition, ATF3 acts as an effector molecule in small-diameter nociceptors that respond to maladaptive behaviors of neuropathic pain.<sup>12,50,127</sup> Recently, comprehensive studies have demonstrated increased ATF3 upregulation in DRG neurons following various nerve injuries and exposure to noxious stimuli.<sup>12,25,32,41,112,127,156,161</sup> ATF3 upregulation by small-diameter nociceptors is specific and differential, and the phenotypic profiles of ATF3 provide molecular explanations for manifested behaviors. For example, ATF3 has been reported to be preferentially upregulated by small-diameter nociceptors, suggesting the susceptibility and topographical relationship of ATF3 to neuropathic pain.<sup>33,50,52,162</sup> Although ATF3 is preferentially expressed by small-diameter nociceptors after TRPV1 depletion, this ATF3 upregulation is distinct. For example, ATF3 was reported to be predominantly expressed by nonpeptidergic nociceptors such as P2X3 (+) and PAP (+) nociceptors rather than by CGRP (+) peptidergic nociceptors.<sup>50,162</sup> In addition, ATF3 upregulation was linked to skin denervation and phenotypic changes in nonpeptidergic nociceptors. Another study reported opposite results; ATF3 was predominantly expressed by peptidergic CGRP (+) nociceptors rather than isolectin B<sub>4</sub> (+) nonpeptidergic neurons.<sup>108</sup>

Transient receptor potential vanilloid subtype 1 depletion by neurotoxicity is a comprehensive neuropathological effect of the skin denervation response to transcription factors in neuronal soma, suggesting that ATF3 activity as a pain indicator directly reflects nerve injury and that expression by nonpeptidergic nociceptors is necessary for pain development. However, ATF3 has also been documented to act as a simplified marker for injury rather than a pain marker.<sup>134</sup> Furthermore, extracellular signaling

is coordinated with intracellular ATF3 responses. The enhancement of extracellular purinergic signaling occurs in parallel with TRPV1 depletion and the burst release of ATP from injured DRG tissues.<sup>50</sup> Notably, extracellular ATP also acts as a gliotransmitter to mediate glial activation in pain development<sup>60,80</sup> through communication with microglia and astrocytes and neuronal interactions, which are also factors affecting pain development.<sup>60</sup> Thus, the burst release of ATP from injured tissues is also associated with the neuroinflammation that mediates neuropathic manifestations.

Intracellular signal cascades of ATF3 involve activation of the c-Jun/c-Jun N-terminal kinase (JNK) signal pathway during axonal transportation activity, which may mediate neuronal stress signals.<sup>91</sup> In addition to ATF3, activating transcription factor 2 (ATF2) acts as a responding molecule in pathological manifestations.<sup>158</sup> It is activated by JNK in diabetic neuropathy.<sup>99</sup> Although few studies have reported an association between ATF2 expression and TRPV1 expression,<sup>30</sup> ATF2 is a potential upstream molecule that regulates TRPV1-mediated inflammatory pain.

### **1.4. Transient receptor potential vanilloid subtype 1 depletion results in neuroinflammatory pain**

Skin denervation and neuronal soma injury are typical neuropathological characteristics of SFN. They result in neurogenic inflammation, also referred to as neuroinflammation, which is associated with neuropathic behavior. Tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) is a major pleiotropic cytokine that mediates neuroinflammation through activation of the TNF receptor 1 (TNFR1) ligand receptor.<sup>143,163</sup> TNF $\alpha$  and TRPV1 are considered 2 critical mediators of inflammatory pain, suggesting that functional TRPV1 is required for the development of inflammatory pain mediated by TNF $\alpha$ .<sup>120</sup> Blocking TNF $\alpha$ /TNFR1 signaling is a new therapeutic strategy for inflammatory diseases and is particularly effective in alleviating injury-induced neuropathic pain.<sup>23,28,31,110</sup> Notably, one study reported that Ret receptor–mediated nociceptive behavior was reversed by TNFR1 loss,<sup>159</sup> suggesting that the interaction of TNF $\alpha$  with nociceptors rather than with TRPV1 is a more critical factor for the development of neuropathic behavior.<sup>92</sup> However, colocalized studies of TNFR1 (+)/TRPV1 (+) neurons have demonstrated that TNFR1 may play a silent role in neuropathic behavior after TRPV1 depletion by the highly potent neurotoxin RTX. Instead, TNF $\alpha$  sensitizes Ret (+) neurons, which mediate mechanical allodynia after TRPV1 depletion by neurotoxins; for example, RTX induces a burst of TNF $\alpha$  that further sensitizes and upregulates Ret (+) neurons in addition to TRPV1.<sup>92</sup> Moreover, TNF $\alpha$ -deficient mice have been found to exhibit fewer nonpeptidergic Ret (+) neurons, suggesting that Ret/TNF $\alpha$  signaling is required in neurotoxin-induced neuropathy, which is mediated by a TRPV1-independent pathway. One study also demonstrated that TNF receptor 2 (TNFR2), another receptor of TNF $\alpha$ , is a major responding effector that mediates TRPV1-dependent inflammatory pain.<sup>24</sup> The collective evidence indicates that TNF $\alpha$  initiates upstream signaling and that, in concert with its cognate receptors, has a broad spectrum of roles in the development of neuropathic pain.

Interleukin-6 (IL-6) and interleukin-1 $\beta$  (IL-1 $\beta$ ) belong to another category of cytokines involved in neuroinflammation. Reduction of IL-6 and IL-1 $\beta$  may relieve neuropathic pain through attenuated TRPV1 expression.<sup>5,86</sup> In several neuropathic models, the parallel expression of TRPV1 and cytokines was correlated with pain development.<sup>29,86,95,109,139,144</sup> These findings suggest

that proinflammatory cytokine-mediated pain sensation requires TRPV1 activation.

TNF $\alpha$  neutralization is currently used for clinical treatment of autoimmune diseases,<sup>85</sup> and TNF $\alpha$ -inhibitory drugs are also used to inhibit glial cell activation in neuropathic pain treatment.<sup>100</sup> Neuroinflammation induces neuropathic behavior through TNF $\alpha$ -mediated activation of glial cells.<sup>7,135</sup> TNF $\alpha$  sensitizes Ret (+) neurons, which mediate neuropathic behavior through an alternative pathway. Generally, glial cell line-derived neurotrophic factor (GDNF) is used as an analgesic agent that acts as a ligand to the Ret receptor, regulating its function and morphology.<sup>43,98,133</sup> An NGF inducer, 4-methylcatechol (4-MC),<sup>63,76,121</sup> has been demonstrated to suppress neuropathic behavior<sup>52</sup> and enhance neuroregeneration.<sup>51,54,63</sup> However, NGF also induced neurogenic inflammation followed by mechanical allodynia, which mediated by unsilencing nicotinic acetylcholine receptor subunit  $\alpha$ -3 (CHRNA3) (+) peptidergic nociceptors<sup>124</sup> and activated previous silent nociceptors.<sup>44</sup> Noteworthy, the elimination of CHRNA3 (+) nociceptor after TRPV1 depletion<sup>72</sup> sparing NGF-induced neurogenic inflammation.

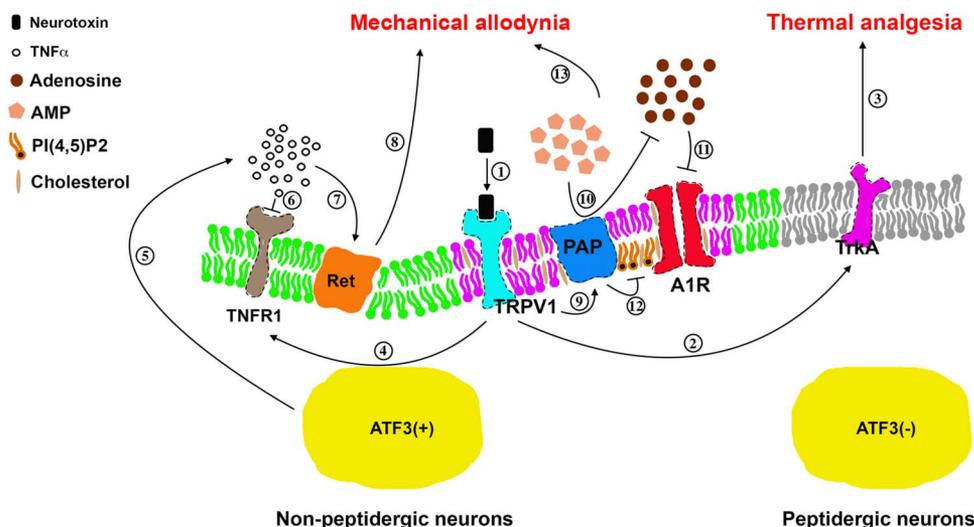
Studies have also shown that 4-MC induces GDNF synthesis<sup>54</sup> and normalizes the upregulation of Ret (+) neurons and neuropathic behavior.<sup>52</sup> The normalization of TNF $\alpha$  and Ret receptors could be targeted in the development of a pharmacological intervention that could provide a new therapeutic direction in the treatment of SFN beyond glial cell activation. The signaling interactions between TNF $\alpha$  and Ret require further investigation.

### 1.5. Role of microdomains in transient receptor potential vanilloid subtype 1 signaling transduction

The cell membrane microdomain is a microstructure whose structural integrity is essential in functional physiology. The structural alteration of microdomains is correlated with neuronal

pathogenesis, such as that of peripheral pathology,<sup>34,84,148</sup> antagonizes hyperalgesia,<sup>27</sup> and inhibits endocannabinoid-mediated analgesic systems.<sup>129</sup> Microdomains have a complex composition of lipid derivatives<sup>132</sup> and have received increasing attention for their relation to nociceptive modulation.<sup>18,37,82,88,132,136</sup> Microdomains are also involved in the modulation of nociceptive development.<sup>96,132,150</sup> Growing evidence suggests that TRPV1 is a microdomain component that modulates nociceptive transmission, particularly through interaction with nociceptive molecules.<sup>11,96,131,149</sup> Microdomain signaling requires the hydrolysis of phosphatidylinositol 4,5-bisphosphate [PI(4,5)P2],<sup>149</sup> a process also involved in PAP-mediated antinociception through the prevention of TRPV1-sensitized noxious sensations.<sup>65,149</sup> Molecular intervention between TRPV1 and PAP involves a PI (4,5) P2 signal. TRPV1 depletion is associated with PAP downregulation, which results in higher PI (4,5) P2 availability, which acts as an agonist of TRPV1 to modulate TRPV1 activity.<sup>4,123</sup> PI (4,5) P2 metabolism is also involved in inhibiting the mechanosensitive receptors modulated by TRPV1 activation.<sup>8</sup> Furthermore, PAP downregulation is associated with an imbalance in analgesic adenosinergic signaling.<sup>65,162</sup> For example, adenosine receptors mediate cellular analgesia through adenosine ligand-activated adenosine receptors.<sup>9,138,164</sup> In particular, adenosine A<sub>1</sub> receptor (A1R) activation plays a key role in SFN.<sup>65,67,90</sup> Downregulation of PAP (+) neurons reduces their ectonucleotidase activity, which in turn reduces the hydrolysis of AMP to adenosine, resulting in the inhibition of cellular analgesia.<sup>65</sup>

One study found that microdomain disruption by cholesterol depletion with methyl- $\beta$ -cyclodextrin (M $\beta$ C) preserves PAP-mediated antinociception through PI (4,5) P2 hydrolysis,<sup>90</sup> indicating that TRPV1 and PAP are located in cholesterol-rich microdomains. These findings suggest that intracellular signal alterations also contribute to pain modulation. However, another study demonstrated that TRPV1 depletion had paradoxical



**Figure 1.** Mechanism of thermal analgesia and mechanical allodynia induced by transient receptor potential vanilloid subtype 1 (TRPV1) depletion in small-fiber neuropathy. TRPV1 receptors were depleted by highly selective neurotoxins such as resiniferatoxin (1) and induced thermal analgesia and mechanical allodynia through 2 distinct pathways. The depletion of TrkA receptors, which were expressed by peptidergic calcitonin gene-related peptides and substance P (+) neurons (2), resulted in thermal analgesia (3). By contrast, nonpeptidergic TRPV1 (+) neurons upregulated activating transcription factor 3 (ATF3) expression, which reflected underlying injury of neuronal soma (4), resulting in a burst of TNF $\alpha$  (5). TNF $\alpha$  had no effect on TNFR1 because it was depleted as a result of its high degree of colocalization with TRPV1 (6) but sensitized Ret receptors (7), leading to the development of mechanical allodynia (8). TRPV1 colocalized with prostatic acid phosphatase and A1R within the same cell membrane microdomain. TRPV1 depletion caused an imbalance in analgesic adenosinergic signaling that induced the downregulation of prostatic acid phosphatase (9), which resulted in reduced hydrolysis of adenosine by AMP (10). This in turn reduced the capacity of A1R-mediated cellular analgesia (11). Structural disruption of microdomains by cholesterol depletion is associated with reduced PI (4,5) P2 hydrolysis (12), which leads to mechanical allodynia (13).

effects on nociceptive transmission; for example, TRPV1 depletion from microdomains induced neuropathic pain.<sup>65</sup> Microdomains labelled by flotillin 1 (FLOT1) and flotillin 2 (FLOT2) as well as microdomains within sensory neurons have an abundance of FLOT1 and FLOT2. A1R also coexpresses with FLOT1 and FLOT2, and these FLOT1 (+) and FLOT2 (+) neurons are depleted after TRPV1 depletion associated with the development of mechanical allodynia.<sup>90</sup> These findings suggest that microdomains containing cellular analgesic molecules on sensory neurons act as functional units for pain transduction.

In addition, microdomains contain nociceptive receptors such as P2X<sub>3</sub>,<sup>37,106,154</sup> and P2X<sub>3</sub> may act as the downstream molecule of PI (4,5) P2.<sup>102,103</sup> Moreover, PI (4,5) P2 modulation of P2X<sub>3</sub> may occur through autocrine signaling because of the high colocalization of PAP and P2X<sub>3</sub> in DRG neurons.<sup>162</sup> These collective findings indicate that TRPV1 depletion initiates a cascade of signaling alterations caused by the neuronal injury response to degeneration of peripheral IENFs after neurotoxin-induced SFN.

### 1.6. Clinical implications for pain management of lipid derivatives of microdomains

Most SFN studies have focused on investigating the responses and contributions of sensitized small-diameter nociceptors after those nociceptors suffer injury and irritation.<sup>49,50,111,117,142</sup> Microdomains act as functional units of nociceptive transmission and could contribute to the development of new first-line pharmacotherapeutic treatments. Microdomains are composed of cholesterol, sphingomyelin, and gangliosides,<sup>122,132</sup> and studies have demonstrated that the TRPV1 activity is affected by altered ganglioside synthesis<sup>132,137,150</sup> and sphingomyelin inhibition.<sup>132,150</sup> Sphingomyelin signaling modulates nociception through the activation of p75 neurotrophin receptors,<sup>73,74</sup> and NGF regulates TrkA receptor activation through alteration of these lipid derivatives.<sup>89</sup> In addition, some G protein-coupled receptors are intracellular components of microdomains and are involved in nociceptive development.<sup>104,107</sup> The elimination of lipid metabolic constituents is a potential target for the pain management of lipid metabolism disorders related to microdomain-attributed peripheral neuropathy.<sup>34,89,116,146</sup> Lipid derivatives in microdomains were demonstrated to be sensitive to drug-induced disruption of microdomains.<sup>38</sup> Therefore, targeting the cytoplasm membranes surrounding microdomains could constitute a new therapeutic direction. Multiple doses of drugs may be required for the depletion of lipid derivatives because of the dynamic replenishment of cholesterol from intracellular stores.<sup>94</sup>

## 2. Conclusions

Transient receptor potential vanilloid subtype 1 acts as a polymodal nociceptor that responds to different noxious stimuli. Commercial capsaicin dermal patch (Qutenza) with 8% (wt/wt) capsaicin has high efficacy for treatment SFN<sup>3,155</sup> such as diabetic peripheral neuropathy.<sup>1</sup> Local analgesia by a high concentration of capsaicin because of axonal degeneration by cytoskeleton disassembling<sup>17</sup> and mitochondrial fission,<sup>21</sup> which showed “defunctionalization” of nociceptor peripheral fibers including the transient loss of membrane potential, inability to axonal transport of neurotrophic factors, and reduction of IENFs.<sup>2</sup> Noteworthy, administration routes determined the survival of TRPV1 (+) neuronal soma and nociceptive-related neuropeptides expression at different nervous tissues.<sup>6</sup> It believed TRPV1 agonist administration through different

routes had distinct effects on neurophysiological responses. This article reviews the role of TRPV1 in SFN. In particular, it discusses the depletion of TRPV1 by highly potent neurotoxins such as RTX, which induces a cascade of extracellular and intracellular signaling alterations caused by peripheral skin denervation and injury to central sensory neurons. **Figure 1** summarizes the putative mechanisms of neuropathic manifestations induced by TRPV1 depletion.

## Disclosures

The authors have no conflicts of interest to declare.

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## References

- [1] Abrams RMC, Pedowitz EJ, Simpson DM. A critical review of the capsaicin 8% patch for the treatment of neuropathic pain associated with diabetic peripheral neuropathy of the feet in adults. *Expert Rev Neurother* 2021. doi: 10.1080/14737175.2021.1874920 [Epub ahead of print].
- [2] Anand P, Bley K. Topical capsaicin for pain management: therapeutic potential and mechanisms of action of the new high-concentration capsaicin 8% patch. *Br J Anaesth* 2011;107:490–502.
- [3] Backonja M, Wallace MS, Blonsky ER, Cutler BJ, Malan P Jr, Rauck R, Tobias J, Group N-CS. NGX-4010, a high-concentration capsaicin patch, for the treatment of postherpetic neuralgia: a randomised, double-blind study. *Lancet Neurol* 2008;7:1106–12.
- [4] Bevan S, Quallo T, Andersson DA. TRPV1. *Handb Exp Pharmacol* 2014; 222:207–45.
- [5] Bishnoi M, Bosgraaf CA, Abooj M, Zhong L, Premkumar LS. Streptozotocin-induced early thermal hyperalgesia is independent of glycemic state of rats: role of transient receptor potential vanilloid 1 (TRPV1) and inflammatory mediators. *Mol Pain* 2011;7:52.
- [6] Bishnoi M, Bosgraaf CA, Premkumar LS. Preservation of acute pain and efferent functions following intrathecal resiniferatoxin-induced analgesia in rats. *J Pain* 2011;12:991–1003.
- [7] Bleich D, Chen S, Zipser B, Sun D, Funk CD, Nadler JL. Resistance to type 1 diabetes induction in 12-lipoxygenase knockout mice. *J Clin Invest* 1999;103:1431–6.
- [8] Borbiri I, Badheka D, Rohacs T. Activation of TRPV1 channels inhibits mechanosensitive Piezo channel activity by depleting membrane phosphoinositides. *Sci Signal* 2015;8:ra15.
- [9] Borea PA, Gessi S, Merighi S, Vincenzi F, Varani K. Pharmacology of adenosine receptors: the state of the art. *Physiol Rev* 2018;98: 1591–625.
- [10] Boruchow SA, Gibbons CH. Utility of skin biopsy in management of small fiber neuropathy. *Muscle Nerve* 2013;48:877–82.
- [11] Botschuijver S, van Diest SA, van Thiel IAM, Saia RS, Strik AS, Yu Z, Maria-Ferreira D, Welting O, Keszthelyi D, Jennings G, Heinsbroek SEM, Elferink RPO, Schuren FHJ, de Jonge WJ, van den Wijngaard RM. Miltefosine treatment reduces visceral hypersensitivity in a rat model for irritable bowel syndrome via multiple mechanisms. *Sci Rep* 2019;9: 12530.
- [12] Braz JM, Basbaum AI. Differential ATF3 expression in dorsal root ganglion neurons reveals the profile of primary afferents engaged by diverse noxious chemical stimuli. *PAIN* 2010;150:290–301.

- [13] Breivik T, Gundersen Y, Gjermo P, Fristad I, Opstad PK. Systemic chemical desensitization of peptidergic sensory neurons with resiniferatoxin inhibits experimental periodontitis. *Open Dent J* 2011;5: 1–6.
- [14] Brown DC, Iadarola MJ, Perkowski SZ, Erin H, Shofer F, Laszlo KJ, Olah Z, Mannes AJ. Physiologic and antinociceptive effects of intrathecal resiniferatoxin in a canine bone cancer model. *Anesthesiology* 2005; 103:1052–9.
- [15] Burnstock G. Purinergic receptors and pain. *Curr Pharm Des* 2009;15: 1717–35.
- [16] Caterina MJ, Leffler A, Malmberg AB, Martin WJ, Trifonov J, Petersen-Zeitl KR, Koltzenburg M, Basbaum AI, Julius D. Impaired nociception and pain sensation in mice lacking the capsaicin receptor. *Science* 2000;288:306–13.
- [17] Caterina MJ, Schumacher MA, Tominaga M, Rosen TA, Levine JD, Julius D. The capsaicin receptor: a heat-activated ion channel in the pain pathway. *Nature* 1997;389:816–24.
- [18] Certal M, Vinhas A, Barros-Barbosa A, Ferreirinha F, Costa MA, Correia-de-Sa P. ADP-induced Ca<sup>2+</sup> signaling and proliferation of rat ventricular myofibroblasts depend on phospholipase C-linked TRP channels activation within lipid rafts. *J Cell Physiol* 2017;232:1511–26.
- [19] Chao CC, Hsueh HW, Kan HW, Liao CH, Jiang HH, Chiang H, Lin WM, Yeh TY, Lin YH, Cheng YY, Hsieh ST. Skin nerve pathology: biomarkers of premanifest and manifest amyloid neuropathy. *Ann Neurol* 2019;85: 560–73.
- [20] Chao CC, Tseng MT, Hsieh ST. Pathophysiology of neuropathic pain in type 2 diabetes: skin denervation and contact heat evoked potentials. *J Peripher Nerv Syst* 2011;16:S21.
- [21] Chiang H, Ohno N, Hsieh YL, Mahad DJ, Kikuchi S, Komuro H, Hsieh ST, Trapp BD. Mitochondrial fission augments capsaicin-induced axonal degeneration. *Acta Neuropathol* 2015;129:81–96.
- [22] Choi SI, Lim JY, Yoo S, Kim H, Hwang SW. Emerging role of spinal cord TRPV1 in pain exacerbation. *Neural Plast* 2016;2016:5954890.
- [23] Coelho SC, Bastos-Pereira AL, Fraga D, Chichorro JG, Zampronio AR. Etanercept reduces thermal and mechanical orofacial hyperalgesia following inflammation and neuropathic injury. *Eur J Pain* 2014;18: 957–67.
- [24] Constantine CE, Mair N, Sailer CA, Andratsch M, Xu ZZ, Blumer MJ, Scherbakov N, Davis JB, Bluethmann H, Ji RR, Kress M. Endogenous tumor necrosis factor alpha (TNF $\alpha$ ) requires TNF receptor type 2 to generate heat hyperalgesia in a mouse cancer model. *J Neurosci* 2008; 28:5072–81.
- [25] Dahlin LB, Stenberg L, Luthman H, Thomsen NO. Nerve compression induces activating transcription factor 3 in neurons and Schwann cells in diabetic rats. *Neuroreport* 2008;19:987–90.
- [26] Devigili G, Tugnoli V, Penza P, Camozzi F, Lombardi R, Melli G, Broglio L, Granieri E, Lauria G. The diagnostic criteria for small fibre neuropathy: from symptoms to neuropathology. *Brain* 2008;131:1912–25.
- [27] Dina OA, Hucho T, Yeh J, Malik-Hall M, Reichling DB, Levine JD. Primary afferent second messenger cascades interact with specific integrin subunits in producing inflammatory hyperalgesia. *PAIN* 2005; 115:191–203.
- [28] Dogrul A, Gul H, Yesilyurt O, Ulas UH, Yildiz O. Systemic and spinal administration of etanercept, a tumor necrosis factor alpha inhibitor, blocks tactile allodynia in diabetic mice. *Acta Diabetol* 2011;48: 135–42.
- [29] Fang D, Kong LY, Cai J, Li S, Liu XD, Han JS, Xing GG. Interleukin-6-mediated functional upregulation of TRPV1 receptors in dorsal root ganglion neurons through the activation of JAK/PI3K signaling pathway: roles in the development of bone cancer pain in a rat model. *PAIN* 2015; 156:1124–44.
- [30] Fang JQ, Du JY, Liang Y, Fang JF. Intervention of electroacupuncture on spinal p38 MAPK/ATF-2/VR-1 pathway in treating inflammatory pain induced by CFA in rats. *Mol Pain* 2013;9:13.
- [31] Fischer R, Kontermann R, Maier O. Targeting sTNF/TNFR1 signaling as a new therapeutic strategy. *Antibodies* 2015;4:48.
- [32] Flatters SJ, Bennett GJ. Studies of peripheral sensory nerves in paclitaxel-induced painful peripheral neuropathy: evidence for mitochondrial dysfunction. *PAIN* 2006;122:245–57.
- [33] Fukuoka T, Yamanaka H, Kobayashi K, Okubo M, Miyoshi K, Dai Y, Noguuchi K. Re-evaluation of the phenotypic changes in L4 dorsal root ganglion neurons after L5 spinal nerve ligation. *PAIN* 2012;153: 68–79.
- [34] Gambert S, Gabrielle PH, Masson E, Leger-Charnay E, Ferrero A, Vannier A, Gendrait C, Lachot M, Creuzot-Garcher C, Bron A, Gregoire S, Leclere L, Martine L, Lucchi G, Truntzer C, Pecqueur D, Bretilon L. Cholesterol metabolism and glaucoma: modulation of Muller cell membrane organization by 24S-hydroxycholesterol. *Chem Phys Lipids* 2017;207:179–91.
- [35] Giannoccaro MP, Donadio V, Incensi A, Pizzi F, Cason E, Di Stasi V, Martinelli P, Scaglione C, Capellari S, Treglia G, Liguori R. Skin biopsy and I-123 MIBG scintigraphy findings in idiopathic Parkinson's disease and parkinsonism: a comparative study. *Mov Disord* 2015;30:986–9.
- [36] Gibbons CH, Wang N, Freeman R. Capsaicin induces degeneration of cutaneous autonomic nerve fibers. *Ann Neurol* 2010;68:888–98.
- [37] Gnanasekaran A, Sundukova M, van den Maagdenberg AM, Fabbretti E, Nistri A. Lipid rafts control P2X3 receptor distribution and function in trigeminal sensory neurons of a transgenic migraine mouse model. *Mol Pain* 2011;7:77.
- [38] Gomide AB, Thome CH, dos Santos GA, Ferreira GA, Faca VM, Rego EM, Greene LJ, Stabeli RG, Ciancaglini P, Itri R. Disrupting membrane raft domains by alkylphospholipids. *Biochim Biophys Acta* 2013;1828: 1384–9.
- [39] Goswami C, Dreger M, Jahnel R, Bogen O, Gillen C, Hucho F. Identification and characterization of a Ca<sup>2+</sup>-sensitive interaction of the vanilloid receptor TRPV1 with tubulin. *J Neurochem* 2004;91: 1092–103.
- [40] Hai T, Hartman MG. The molecular biology and nomenclature of the activating transcription factor/cAMP responsive element binding family of transcription factors: activating transcription factor proteins and homeostasis. *Gene* 2001;273:1–11.
- [41] Hai T, Wolford CC, Chang YS. ATF3, a hub of the cellular adaptive-response network, in the pathogenesis of diseases: is modulation of inflammation a unifying component? *Gene Expr* 2010;15:1–11.
- [42] Han P, McDonald HA, Bianchi BR, Kouhen RE, Vos MH, Jarvis MF, Faltynek CR, Moreland RB. Capsaicin causes protein synthesis inhibition and microtubule disassembly through TRPV1 activities both on the plasma membrane and intracellular membranes. *Biochem Pharmacol* 2007;73:1635–45.
- [43] Hedstrom KL, Murtie JC, Albers K, Calcutt NA, Corfas G. Treating small fiber neuropathy by topical application of a small molecule modulator of ligand-induced GFR $\alpha$ /RET receptor signaling. *Proc Natl Acad Sci U S A* 2014;111:2325–30.
- [44] Hirth M, Rukwied R, Gromann A, Turnquist B, Weinkauff B, Francke K, Albrecht P, Rice F, Hagglof B, Ringkamp M, Engelhardt M, Schultz C, Schmelz M, Obreja O. Nerve growth factor induces sensitization of nociceptors without evidence for increased intraepidermal nerve fiber density. *PAIN* 2013;154:2500–11.
- [45] Hofmann L, Hose D, Griesshammer A, Blum R, Doring F, Dib-Hajj S, Waxman S, Sommer C, Wischmeyer E, Uceyler N. Characterization of small fiber pathology in a mouse model of Fabry disease. *Elife* 2018;7: e39300. doi: 10.7554/eLife.39300.
- [46] Hohmann AG, Neely MH, Pina J, Nackley AG. Neonatal chronic hind paw inflammation alters sensitization to intradermal capsaicin in adult rats: a behavioral and immunocytochemical study. *J Pain* 2005;6: 798–808.
- [47] Horvath A, Borbely E, Bolcskei K, Szentos N, Kiss T, Belak M, Rauch T, Glant T, Zakary R, Juhasz T, Karanyicz E, Boldizsar F, Helyes Z, Botz B. Regulatory role of capsaicin-sensitive peptidergic sensory nerves in the proteoglycan-induced autoimmune arthritis model of the mouse. *J Neuroinflammation* 2018;15:335.
- [48] Hsiao TH, Fu YS, Ho WY, Chen TH, Hsieh YL. Promotion of thermal analgesia and neuropeptidergic skin reinnervation by 4-methylcatechol in resiniferatoxin-induced neuropathy. *Kaohsiung J Med Sci* 2013;29: 405–11.
- [49] Hsieh ST. Pathology and functional diagnosis of small-fiber painful neuropathy. *Acta Neurol Taiwan* 2010;19:82–9.
- [50] Hsieh YL, Chiang H, Lue JH, Hsieh ST. P2X3-mediated peripheral sensitization of neuropathic pain in resiniferatoxin-induced neuropathy. *Exp Neurol* 2012;235:316–25.
- [51] Hsieh YL, Chiang H, Tseng TJ, Hsieh ST. Enhancement of cutaneous nerve regeneration by 4-methylcatechol in resiniferatoxin-induced neuropathy. *J Neuropathol Exp Neurol* 2008;67:93–104.
- [52] Hsieh YL, Kan HW, Chiang H, Lee YC, Hsieh ST. Distinct TrkA and Ret mediated negative and positive neuropathic behaviors in a mouse model of resiniferatoxin-induced small fiber neuropathy. *Exp Neurol* 2018;300:87–99.
- [53] Hsieh YL, Lin CL, Chiang H, Fu YS, Lue JH, Hsieh ST. Role of peptidergic nerve terminals in the skin: reversal of thermal sensation by calcitonin gene-related peptide in TRPV1-depleted neuropathy. *PLoS One* 2012;7:e50805.
- [54] Hsieh YL, Lin WM, Lue JH, Chang MF, Hsieh ST. Effects of 4-methylcatechol on skin reinnervation: promotion of cutaneous nerve regeneration after crush injury. *J Neuropathol Exp Neurol* 2009;68: 1269–81.
- [55] Huang Y, Chen SR, Chen H, Pan HL. Endogenous transient receptor potential ankyrin 1 and vanilloid 1 activity potentiates glutamatergic input

- to spinal lamina I neurons in inflammatory pain. *J Neurochem* 2019;149:381–98.
- [56] Iadarola MJ, Mannes AJ. The vanilloid agonist resiniferatoxin for interventional-based pain control. *Curr Top Med Chem* 2011;11:2171–9.
- [57] Jankowski MP, Lawson JJ, Mcllwraith SL, Rau KK, Anderson CE, Albers KM, Koerber HR. Sensitization of cutaneous nociceptors after nerve transection and regeneration: possible role of target-derived neurotrophic factor signaling. *J Neurosci* 2009;29:1636–47.
- [58] Jankowski MP, Rau KK, Soneji DJ, Anderson CE, Koerber HR. Enhanced artemin/GFR $\alpha$ 3 levels regulate mechanically insensitive, heat-sensitive C-fiber recruitment after axotomy and regeneration. *J Neurosci* 2010;30:16272–83.
- [59] Jarvis MF. Contributions of P2X3 homomeric and heteromeric channels to acute and chronic pain. *Expert Opin Ther Targets* 2003;7:513–22.
- [60] Jeremic A, Jeftinija K, Stevanovic J, Glavaski A, Jeftinija S. ATP stimulates calcium-dependent glutamate release from cultured astrocytes. *J Neurochem* 2001;77:664–75.
- [61] Julius D. TRP channels and pain. *Annu Rev Cell Dev Biol* 2013;29:355–84.
- [62] Kaan TK, Yip PK, Patel S, Davies M, Marchand F, Cockayne DA, Nunn PA, Dickenson AH, Ford AP, Zhong Y, Malcangio M, McMahon SB. Systemic blockade of P2X3 and P2X2/3 receptors attenuates bone cancer pain behaviour in rats. *Brain* 2010;133:2549–64.
- [63] Kaechi K, Ikegami R, Nakamura N, Nakajima M, Furukawa Y, Furukawa S. 4-Methylcatechol, an inducer of nerve growth factor synthesis, enhances peripheral nerve regeneration across nerve gaps. *J Pharmacol Exp Ther* 1995;272:1300–4.
- [64] Kalliomaki M, Kieseritzky JV, Schmidt R, Hagglof B, Karlsten R, Sjogren N, Albrecht P, Gee L, Rice F, Wiig M, Schmelz M, Gordh T. Structural and functional differences between neuropathy with and without pain? *Exp Neurol* 2011;231:199–206.
- [65] Kan HW, Chang CH, Lin CL, Lee YC, Hsieh ST, Hsieh YL. Downregulation of adenosine and adenosine A1 receptor contributes to neuropathic pain in resiniferatoxin neuropathy. *PAIN* 2018;159:1580–91.
- [66] Kang S, Wu C, Banik RK, Brennan TJ. Effect of capsaicin treatment on nociceptors in rat glabrous skin one day after plantar incision. *PAIN* 2010;148:128–40.
- [67] Katz NK, Ryals JM, Wright DE. Central or peripheral delivery of an adenosine A1 receptor agonist improves mechanical allodynia in a mouse model of painful diabetic neuropathy. *Neuroscience* 2015;285:312–23.
- [68] Kennedy WR. Opportunities afforded by the study of unmyelinated nerves in skin and other organs. *Muscle Nerve* 2004;29:756–67.
- [69] Kennedy WR, Said G. Sensory nerves in skin: answers about painful feet?. *Neurology* 1999;53:1614–15.
- [70] Kennedy WR, Wendelschafer-Crabb G. The innervation of human epidermis. *J Neurol Sci* 1993;115:184–90.
- [71] Kennedy WR, Wendelschafer-Crabb G. Utility of the skin biopsy method in studies of diabetic neuropathy. *Electroencephalogr Clin Neurophysiol Suppl* 1999;50:553–9.
- [72] Khan IM, Wennerholm M, Singletary E, Polston K, Zhang L, Deerinck T, Yaksh TL, Taylor P. Ablation of primary afferent terminals reduces nicotinic receptor expression and the nociceptive responses to nicotinic agonists in the spinal cord. *J Neurocytol* 2004;33:543–56.
- [73] Khodorova A, Nicol GD, Strichartz G. The p75NTR signaling cascade mediates mechanical hyperalgesia induced by nerve growth factor injected into the rat hind paw. *Neuroscience* 2013;254:312–23.
- [74] Khodorova A, Nicol GD, Strichartz G. The TrkA receptor mediates experimental thermal hyperalgesia produced by nerve growth factor: modulation by the p75 neurotrophin receptor. *Neuroscience* 2017;340:384–97.
- [75] Kim YH, Back SK, Davies AJ, Jeong H, Jo HJ, Chung G, Na HS, Bae YC, Kim SJ, Kim JS, Jung SJ, Oh SB. TRPV1 in GABAergic interneurons mediates neuropathic hyperalgesia and disinhibition of the nociceptive circuitry in the spinal cord. *Neuron* 2012;74:640–7.
- [76] Kimura N, Nishizaki K, Orita Y, Masuda Y. 4-methylcatechol, a potent inducer of nerve growth factor synthesis, protects spiral ganglion neurons from aminoglycoside ototoxicity—preliminary report. *Acta Otolaryngol* 1999;540:12–15.
- [77] Kissin I, Szallasi A. Therapeutic targeting of TRPV1 by resiniferatoxin, from preclinical studies to clinical trials. *Curr Top Med Chem* 2011;11:2159–70.
- [78] Knotkova H, Pappagallo M, Szallasi A. Capsaicin (TRPV1 Agonist) therapy for pain relief: farewell or revival? *Clin J Pain* 2008;24:142–54.
- [79] Kobayashi K, Fukuoka T, Obata K, Yamanaka H, Dai Y, Tokunaga A, Noguchi K. Distinct expression of TRPM8, TRPA1, and TRPV1 mRNAs in rat primary afferent neurons with adelta/c-fibers and colocalization with trk receptors. *J Comp Neurol* 2005;493:596–606.
- [80] Koizumi S, Fujishita K, Inoue K. Regulation of cell-to-cell communication mediated by astrocytic ATP in the CNS. *Purinergic Signal* 2005;1:211–17.
- [81] Kroin JS, Buvanendran A, Williams DK, Wagenaar B, Moric M, Tuman KJ, Kerns JM. Local anesthetic sciatic nerve block and nerve fiber damage in diabetic rats. *Reg Anesth Pain Med* 2010;35:343–50.
- [82] Kumari S, Kumar A, Sardar P, Yadav M, Majhi RK, Kumar A, Goswami C. Influence of membrane cholesterol in the molecular evolution and functional regulation of TRPV4. *Biochem Biophys Res Commun* 2015;456:312–19.
- [83] Lauria G, Devigili G. Skin biopsy as a diagnostic tool in peripheral neuropathy. *Nat Clin Pract Neurol* 2007;3:546–57.
- [84] Lee S, Amici S, Tavori H, Zeng WM, Freeland S, Fazio S, Notterpek L. PMP22 is critical for actin-mediated cellular functions and for establishing lipid rafts. *J Neurosci* 2014;34:16140–52.
- [85] Li P, Zheng Y, Chen X. Drugs for autoimmune inflammatory diseases: from small molecule compounds to anti-TNF biologics. *Front Pharmacol* 2017;8:460.
- [86] Li R, Zhao C, Yao M, Song Y, Wu Y, Wen A. Analgesic effect of coumarins from *Radix angelicae pubescentis* is mediated by inflammatory factors and TRPV1 in a spared nerve injury model of neuropathic pain. *J Ethnopharmacol* 2017;195:81–8.
- [87] Liao HY, Hsieh CL, Huang CP, Lin YW. Electroacupuncture attenuates CFA-induced inflammatory pain by suppressing Nav1.8 through S100B, TRPV1, opioid, and adenosine pathways in mice. *Sci Rep* 2017;7:42531.
- [88] Licon Y, Leandro D, Romero-Mendez C, Rodriguez-Menchaca AA, Sanchez-Armass S, Meza U. Inhibition of CaV2.3 channels by NK1 receptors is sensitive to membrane cholesterol but insensitive to caveolin-1. *Pflugers Arch* 2015;467:1699–709.
- [89] Limpert AS, Karlo JC, Landreth GE. Nerve growth factor stimulates the concentration of TrkA within lipid rafts and extracellular signal-regulated kinase activation through c-Cbl-associated protein. *Mol Cell Biol* 2007;27:5686–98.
- [90] Lin CL, Chang CH, Chang YS, Lu SC, Hsieh YL. Treatment with methyl-beta-cyclodextrin prevents mechanical allodynia in resiniferatoxin neuropathy in a mouse model. *Biol Open* 2018;8:bio039511.
- [91] Lindwall C, Kanje M. The Janus role of c-Jun: cell death versus survival and regeneration of neonatal sympathetic and sensory neurons. *Exp Neurol* 2005;196:184–94.
- [92] Lu SC, Chang YS, Kan HW, Hsieh YL. Tumor necrosis factor- $\alpha$  mediated pain hypersensitivity through Ret receptor in resiniferatoxin neuropathy. *Kaohsiung J Med Sci* 2018;34:494–502.
- [93] Madsen CS, Finnerup NB, Baumgartner U. Assessment of small fibers using evoked potentials. *Scand J Pain* 2014;5:111–18.
- [94] Mahammad S, Parmyrd I. Cholesterol homeostasis in T cells. Methyl-beta-cyclodextrin treatment results in equal loss of cholesterol from Triton X-100 soluble and insoluble fractions. *Biochim Biophys Acta* 2008;1778:1251–8.
- [95] Malek N, Pajak A, Kolosowska N, Kucharczyk M, Starowicz K. The importance of TRPV1-sensitisation factors for the development of neuropathic pain. *Mol Cell Neurosci* 2015;65:1–10.
- [96] Marchenkova A, Vilotti S, Ntamati N, van den Maagdenberg AM, Nistri A. Inefficient constitutive inhibition of P2X3 receptors by brain natriuretic peptide system contributes to sensitization of trigeminal sensory neurons in a genetic mouse model of familial hemiplegic migraine. *Mol Pain* 2016;12:1744806916646110. doi: 10.1177/1744806916646110.
- [97] McCarthy BG, Hsieh ST, Stocks A, Hauer P, Macko C, Cornblath DR, Griffin JW, McArthur JC. Cutaneous innervation in sensory neuropathies: evaluation by skin biopsy. *Neurology* 1995;45:1848–55.
- [98] Merighi A. Targeting the glial-derived neurotrophic factor and related molecules for controlling normal and pathologic pain. *Expert Opin Ther Targets* 2015;20:193–208.
- [99] Middlemas A, Delcroix JD, Sayers NM, Tomlinson DR, Fernyhough P. Enhanced activation of axonally transported stress-activated protein kinases in peripheral nerve in diabetic neuropathy is prevented by neurotrophin-3. *Brain* 2003;126:1671–82.
- [100] Mika J, Zychowska M, Popiolek-Barczyk K, Rojewska E, Przewlocka B. Importance of glial activation in neuropathic pain. *Eur J Pharmacol* 2013;716:106–19.
- [101] Mishra SK, Hoon MA. Ablation of TrpV1 neurons reveals their selective role in thermal pain sensation. *Mol Cell Neurosci* 2010;43:157–63.
- [102] Mo G, Bernier LP, Zhao Q, Chabot-Dore AJ, Ase AR, Logothetis D, Cao CQ, Seguela P. Subtype-specific regulation of P2X3 and P2X2/3 receptors by phosphoinositides in peripheral nociceptors. *Mol Pain* 2009;5:47.

- [103] Mo G, Peleshok JC, Cao CQ, Ribeiro-da-Silva A, Seguela P. Control of P2X3 channel function by metabotropic P2Y2 utp receptors in primary sensory neurons. *Mol Pharmacol* 2013;83:640–7.
- [104] Monastyrskaya K, Hostettler A, Buergi S, Draeger A. The NK1 receptor localizes to the plasma membrane microdomains, and its activation is dependent on lipid raft integrity. *J Biol Chem* 2005;280:7135–46.
- [105] Mrozkova P, Spicarova D, Palecek J. Hypersensitivity induced by activation of spinal cord PAR2 receptors is partially mediated by TRPV1 receptors. *PLoS One* 2016;11:e0163991.
- [106] Murrell-Lagnaud RD. Regulation of P2X purinergic receptor signaling by cholesterol. *Curr Top Membr* 2017;80:211–32.
- [107] Nakagawa T, Takahashi C, Matsuzaki H, Takeyama S, Sato S, Sato A, Kuroda Y, Higashi H. N-glycan-dependent cell-surface expression of the P2Y2 receptor and N-glycan-independent distribution to lipid rafts. *Biochem Biophys Res Commun* 2017;485:427–31.
- [108] Nascimento D, Pozza DH, Castro-Lopes JM, Neto FL. Neuronal injury marker ATF-3 is induced in primary afferent neurons of monoarthritic rats. *Neurosignals* 2011;19:210–21.
- [109] Nees TA, Rosshirt N, Reiner T, Schiltewolf M, Moradi B. Inflammation and osteoarthritis-related pain. *Schmerz* 2019;33:4–12.
- [110] Oaklander AL. Immunotherapy prospects for painful small-fiber sensory neuropathies and ganglionopathies. *Neurotherapeutics* 2016;13:108–17.
- [111] Oaklander AL, Fields HL. Is reflex sympathetic dystrophy/complex regional pain syndrome type I a small-fiber neuropathy? *Ann Neurol* 2009;65:629–38.
- [112] Obata K, Yamanaka H, Dai Y, Mizushima T, Fukuoka T, Tokunaga A, Noguchi K. Differential activation of MAPK in injured and uninjured DRG neurons following chronic constriction injury of the sciatic nerve in rats. *Eur J Neurosci* 2004;20:2881–95.
- [113] Obreja O, Ringkamp M, Turnquist B, Hirth M, Forsch E, Rukwied R, Petersen M, Schmelz M. Nerve growth factor selectively decreases activity-dependent conduction slowing in mechano-insensitive C-nociceptors. *PAIN* 2011;152:2138–46.
- [114] Obreja O, Rukwied R, Nagler L, Schmidt M, Schmelz M, Namer B. Nerve growth factor locally sensitizes nociceptors in human skin. *PAIN* 2018;159:416–26.
- [115] Obrosova IG, Stavniichuk R, Drel VR, Shevalye H, Vareniuk I, Nadler JL, Schmidt RE. Different roles of 12/15-lipoxygenase in diabetic large and small fiber peripheral and autonomic neuropathies. *Am J Pathol* 2010;177:1436–47.
- [116] Ogawa Y, Rasband MN. Proteomic analysis of optic nerve lipid rafts reveals new paranodal proteins. *J Neurosci Res* 2009;87:3502–10.
- [117] Orstavik K, Namer B, Schmidt R, Schmelz M, Hilliges M, Weidner C, Carr RW, Handwerker H, Jorum E, Torebjork HE. Abnormal function of C-fibers in patients with diabetic neuropathy. *J Neurosci* 2006;26:11287–94.
- [118] Pan HL, Khan GM, Alloway KD, Chen SR. Resiniferatoxin induces paradoxical changes in thermal and mechanical sensitivities in rats: mechanism of action. *J Neurosci* 2003;23:2911–19.
- [119] Pang Z, Sakamoto T, Tiwari V, Kim YS, Yang F, Dong X, Guler AD, Guan Y, Caterina MJ. Selective keratinocyte stimulation is sufficient to evoke nociception in mice. *PAIN* 2015;156:656–65.
- [120] Park CK, Lu N, Xu ZZ, Liu T, Serhan CN, Ji RR. Resolving TRPV1- and TNF-alpha-mediated spinal cord synaptic plasticity and inflammatory pain with neuroprotectin D1. *J Neurosci* 2011;31:15072–85.
- [121] Perez-Perez M, Garcia-Suarez O, Esteban I, Germana A, Farinas I, Naves FJ, Vega JA. p75NTR in the spleen: age-dependent changes, effect of NGF and 4-methylcatechol treatment, and structural changes in p75NTR-deficient mice. *Anat Rec A Discov Mol Cell Evol Biol* 2003;270:117–28.
- [122] Pike LJ. Lipid rafts: bringing order to chaos. *J Lipid Res* 2003;44:655–67.
- [123] Poblete H, Oyarzun I, Olivero P, Comer J, Zuniga M, Sepulveda RV, Baez-Nieto D, Gonzalez Leon C, Gonzalez-Nilo F, Latorre R. Molecular determinants of phosphatidylinositol 4,5-bisphosphate (PI(4,5)P2) binding to transient receptor potential V1 (TRPV1) channels. *J Biol Chem* 2015;290:2086–98.
- [124] Prato V, Taberner FJ, Hockley JRF, Callejo G, Arcourt A, Tazir B, Hammer L, Schad P, Heppenstall PA, Smith ES, Lechner SG. Functional and molecular characterization of mechanoinsensitive silent nociceptors. *Cell Rep* 2017;21:3102–15.
- [125] Price TJ, Flores CM. Critical evaluation of the colocalization between calcitonin gene-related peptide, substance P, transient receptor potential vanilloid subfamily type 1 immunoreactivities, and isolectin B4 binding in primary afferent neurons of the rat and mouse. *J Pain* 2007;8:263–72.
- [126] Puglia C, Santonocito D, Bonaccorso A, Musumeci T, Ruozi B, Pignatello R, Carbone C, Parenti C, Chiechio S. Lipid nanoparticle inclusion prevents capsaicin-induced TRPV1 defunctionalization. *Pharmaceutics* 2020;12:339.
- [127] Rau KK, Hill CE, Harrison BJ, Venkat G, Koenig HM, Cook SB, Rabchevsky AG, Taylor BK, Hai T, Petruska JC. Cutaneous tissue damage induces long-lasting nociceptive sensitization and regulation of cellular stress- and nerve injury-associated genes in sensory neurons. *Exp Neurol* 2016;283:413–27.
- [128] Rosenbaum T, Simon SA. TRPV1 receptors and signal transduction. In: Liedtke WB, Heller S, editors. *TRP ion channel function in sensory transduction and cellular signaling cascades*. Boca Raton, FL: CRC Press/Taylor & Francis, 2007.
- [129] Rossi S, Sacchetti L, Napolitano F, De Chiara V, Motta C, Studer V, Mussella A, Barbieri F, Bari M, Bernardi G, Maccarrone M, Usiello A, Centonze D. Interleukin-1beta causes anxiety by interacting with the endocannabinoid system. *J Neurosci* 2012;32:13896–905.
- [130] Ruehle BS, Handwerker HO, Lenner JK, Ringler R, Forster C. Brain activation during input from mechanoinsensitive versus polymodal C-nociceptors. *J Neurosci* 2006;26:5492–9.
- [131] Saghy E, Payrits M, Biro-Suto T, Skoda-Foldes R, Szanti-Pinter E, Erostyak J, Makkai G, Setalo G Jr, Kollar L, Koszegi T, Csepregi R, Szolcsanyi J, Helyes Z, Szoke E. Carboxamido steroids inhibit the opening properties of transient receptor potential ion channels by lipid raft modulation. *J Lipid Res* 2018;59:1851–63.
- [132] Saghy E, Szoke E, Payrits M, Helyes Z, Borzsei R, Erostyak J, Janosi TZ, Setalo G Jr, Szolcsanyi J. Evidence for the role of lipid rafts and sphingomyelin in Ca-gating of Transient Receptor Potential channels in trigeminal sensory neurons and peripheral nerve terminals. *Pharmacol Res* 2015;100:101–16.
- [133] Sakai A, Asada M, Seno N, Suzuki H. Involvement of neural cell adhesion molecule signaling in glial cell line-derived neurotrophic factor-induced analgesia in a rat model of neuropathic pain. *PAIN* 2008;137:378–88.
- [134] Salinas-Abarca AB, Velazquez-Lagunas I, Franco-Enzastiga U, Torres-Lopez JE, Rocha-Gonzalez HI, Granados-Soto V. ATF2, but not ATF3, participates in the maintenance of nerve injury-induced tactile allodynia and thermal hyperalgesia. *Mol Pain* 2018;14:1744806918787427.
- [135] Santello M, Bezzi P, Volterra A. TNFalpha controls glutamatergic gliotransmission in the hippocampal dentate gyrus. *Neuron* 2011;69:988–1001.
- [136] Santha P, Dobos I, Kis G, Jancso G. Role of gangliosides in peripheral pain mechanisms. *Int J Mol Sci* 2020;21:1005.
- [137] Santha P, Oszlacs O, Dux M, Dobos I, Jancso G. Inhibition of glucosylceramide synthase reversibly decreases the capsaicin-induced activation and TRPV1 expression of cultured dorsal root ganglion neurons. *PAIN* 2010;150:103–12.
- [138] Sawynok J. Adenosine receptor targets for pain. *Neuroscience* 2016;338:1–18.
- [139] Schaible HG, von Banchet GS, Boettger MK, Brauer R, Gajda M, Richter F, Hensellek S, Brenn D, Natura G. The role of proinflammatory cytokines in the generation and maintenance of joint pain. *Ann N Y Acad Sci* 2010;1193:60–9.
- [140] Schmelz M, Schmid R, Handwerker HO, Torebjork HE. Encoding of burning pain from capsaicin-treated human skin in two categories of unmyelinated nerve fibres. *Brain* 2000;123(pt 3):560–71.
- [141] Schnakenberg M, Thomas C, Schmelz M, Rukwied R. Nerve growth factor sensitizes nociceptors to C-fibre selective supra-threshold electrical stimuli in human skin. *Eur J Pain* 2021;25:385–97.
- [142] Serra J, Collado A, Sola R, Antonelli F, Torres X, Salgueiro M, Quiles C, Bostock H. Hyperexcitable C nociceptors in fibromyalgia. *Ann Neurol* 2014;75:196–208.
- [143] Shen CH, Tsai RY, Shih MS, Lin SL, Tai YH, Chien CC, Wong CS. Etanercept restores the antinociceptive effect of morphine and suppresses spinal neuroinflammation in morphine-tolerant rats. *Anesth Analg* 2011;112:454–9.
- [144] Shi J, Jiang K, Li Z. MiR-145 ameliorates neuropathic pain via inhibiting inflammatory responses and mTOR signaling pathway by targeting Akt3 in a rat model. *Neurosci Res* 2018;134:10–17.
- [145] Smith AK, O'Hara CL, Stucky CL. Mechanical sensitization of cutaneous sensory fibers in the spared nerve injury mouse model. *Mol Pain* 2013;9:61.
- [146] Smith B, Galbiati F, Castelvetri LC, Givogri MI, Lopez-Rosas A, Bongarzone ER. Peripheral neuropathy in the Twitcher mouse involves the activation of axonal caspase 3. *ASN Neuro* 2011;3:e00066.
- [147] Sommer C, Lauria G. Skin biopsy in the management of peripheral neuropathy. *Lancet Neurol* 2007;6:632–42.

- [148] Sonnino S, Aurell M, Grassi S, Mauri L, Prioni S, Prinetti A. Lipid rafts in neurodegeneration and neuroprotection. *Mol Neurobiol* 2014;50:130–48.
- [149] Sowa NA, Street SE, Vihko P, Zylka MJ. Prostatic acid phosphatase reduces thermal sensitivity and chronic pain sensitization by depleting phosphatidylinositol 4,5-bisphosphate. *J Neurosci* 2010;30:10282–93.
- [150] Szoke E, Borzsei R, Toth DM, Lengi O, Helyes Z, Sandor Z, Szolcsanyi J. Effect of lipid raft disruption on TRPV1 receptor activation of trigeminal sensory neurons and transfected cell line. *Eur J Pharmacol* 2010;628:67–74.
- [151] Tender GC, Li YY, Cui JG. Brain-derived neurotrophic factor redistribution in the dorsal root ganglia correlates with neuropathic pain inhibition after resiniferatoxin treatment. *Spine J* 2010;10:715–20.
- [152] Tender GC, Li YY, Cui JG. The role of nerve growth factor in neuropathic pain inhibition produced by resiniferatoxin treatment in the dorsal root ganglia. *Neurosurgery* 2013;73:158–65; discussion 165–156.
- [153] Tominaga M, Caterina MJ, Malmberg AB, Rosen TA, Gilbert H, Skinner K, Raumann BE, Basbaum AI, Julius D. The cloned capsaicin receptor integrates multiple pain-producing stimuli. *Neuron* 1998;21:531–43.
- [154] Vacca F, Amadio S, Sancesario G, Bernardi G, Volonte C. P2X3 receptor localizes into lipid rafts in neuronal cells. *J Neurosci Res* 2004;76:653–61.
- [155] Wallace M, Pappagallo M. Qutenza(R): a capsaicin 8% patch for the management of postherpetic neuralgia. *Expert Rev Neurother* 2011;11:15–27.
- [156] Wang H, Jiang M, Cui H, Chen M, Buttyan R, Hayward SW, Hai T, Wang Z, Yan C. The stress response mediator ATF3 represses androgen signaling by binding the androgen receptor. *Mol Cell Biol* 2012;32:3190–202.
- [157] Wang S, Kim M, Ali Z, Ong K, Pae EK, Chung MK. TRPV1 and TRPV1-expressing nociceptors mediate orofacial pain behaviors in a mouse model of orthodontic tooth movement. *Front Physiol* 2019;10:1207.
- [158] Watson G, Ronai ZA, Lau E. ATF2, a paradigm of the multifaceted regulation of transcription factors in biology and disease. *Pharmacol Res* 2017;119:347–57.
- [159] Wheeler MA, Heffner DL, Kim S, Espy SM, Spano AJ, Cleland CL, Deppmann CD. TNF-alpha/TNFR1 signaling is required for the development and function of primary nociceptors. *Neuron* 2014;82:587–602.
- [160] Wildgaard K, Ringsted TK, Hansen HJ, Petersen RH, Werner MU, Kehlet H. Quantitative sensory testing of persistent pain after video-assisted thoracic surgery lobectomy. *Br J Anaesth* 2012;108:126–33.
- [161] Wright DE, Ryals JM, McCarron KE, Christianson JA. Diabetes-induced expression of activating transcription factor 3 in mouse primary sensory neurons. *J Peripher Nerv Syst* 2004;9:242–54.
- [162] Wu CH, Ho WY, Lee YC, Lin CL, Hsieh YL. EXPRESS: NGF-trkA signaling modulates the analgesic effects of prostatic acid phosphatase in resiniferatoxin-induced neuropathy. *Mol Pain* 2016;12:1744806916656846. doi: 10.1177/1744806916656846.
- [163] Zhang L, Berta T, Xu ZZ, Liu T, Park JY, Ji RR. TNF-alpha contributes to spinal cord synaptic plasticity and inflammatory pain: distinct role of TNF receptor subtypes 1 and 2. *PAIN* 2011;152:419–27.
- [164] Zylka MJ. Pain-relieving prospects for adenosine receptors and ectonucleotidases. *Trends Mol Med* 2011;17:188–96.